

Figure S1. Nur77^{GFP} expression among unconventional T cells after TCR engagement. MLN single cell suspensions from *Lm*-immune Nur77^{GFP} and non-transgenic (NTg) mice were left unstimulated (medium) or stimulated with plate-bound α CD3 ϵ and soluble α CD28 mAbs or with plate-bound GL4 mAb for 6 hours. (a to c) Nur77^{GFP} expression was analyzed among conventional T cells. (a) Representative histograms of Nur77^{GFP} expression in the indicated populations. (b) Nur77^{GFP} MFI among unstimulated CD44^{hi} CD4⁺ and activated CD44^{hi} CD4⁺ T cells producing either IL-17A or IFN γ after stimulation. (c) Nur77^{GFP} expression among unstimulated and activated IFN γ ⁺ CD44^{hi} CD8 α ⁺ T cells. (d and e) Nur77^{GFP} expression was analyzed among CD44^{hi} CD27^{neg} CD19⁻ CD4⁻ CD8 α ⁻ lymphocytes. (d) Representative histograms of Nur77^{GFP} expression in unstimulated and IL-17A⁺ cells upon stimulation with 1 or 10 μ g/ml of coated GL4 mAb. (e) Scatter plots show Nur77^{GFP} MFI among the groups indicated in (d). (f and g) TCR δ expression among CD19⁻ CD4⁻ CD8 α ⁻ cells left unstimulated (medium) or stimulated with the indicated concentration of GL4 mAb. (f) Representative histograms of TCR δ expression are shown. (g) Scatter plots show the frequency of TCR δ ⁺ cells among CD19⁻ CD4⁻ CD8 α ⁻ cells. All data are from 1 experiment with 5 Nur77^{GFP} and 2 NTg mice. Statistical analyses were only performed for Nur77^{GFP} mice. *, $p \leq 0.05$; **, $p \leq 0.01$; ***, $p \leq 0.0001$

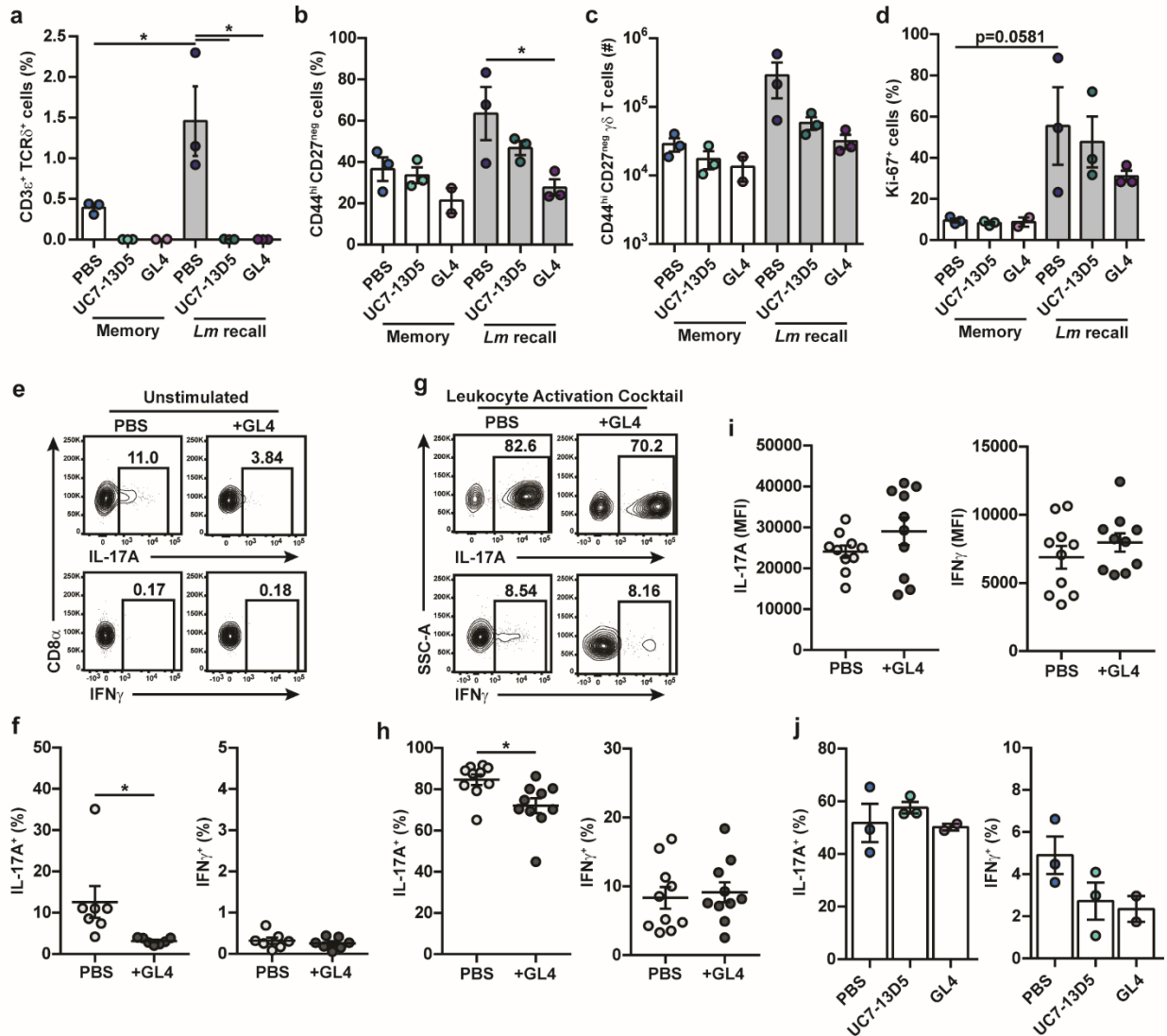


Figure S2. Antibody-mediated $\gamma\delta$ TCR internalization diminishes CD44^{hi} CD27^{neg} $\gamma\delta$ T cell proliferation while minimally impacting cytokine production during recall response to *Lm*. *Lm*-immune *Tcrd*-H2B-eGFP (Balb/c background) mice were i.p. injected with either PBS, UC7-13D5 or GL4 mAb on days -3, -1 and +1 relative to recall infection. On day 0, mice were challenged with *Lm* ("*Lm* recall") or given PBS ("*Lm* Memory"). $\gamma\delta$ T cells (identified as TCR β ^{neg} CD8 α ^{neg} GFP⁺ single live lymphocytes) were analyzed 4 to 5 days later in the MLN. (a) The graph shows the mean \pm SEM (n=2-3 mice/group) of the frequency of CD3 ϵ^+ TCR δ^+ cells among single live lymphocytes. (b-c) The frequency (b) and number (c) of CD44^{hi} CD27^{neg} $\gamma\delta$ T cells are shown as the mean \pm SEM. (d) CD44^{hi} CD27^{neg} $\gamma\delta$ T cells were analyzed for Ki-67 expression. The graph shows the mean \pm SEM of Ki-67⁺ cell frequency. (e-j) Single cell suspensions from MLN were left unstimulated in the presence of BD GolgiPlug (e and f) or stimulated with BD Leukocyte activation cocktail (g-j) for 4 hours. CD44^{hi} CD27^{neg} $\gamma\delta$ T cell cytokine production was analyzed by flow cytometry. Representative flow plots of IL-17A and IFN γ production are shown (e and g). Cumulative data are shown as the mean \pm SEM (n=3-4 mice/group) of cytokine⁺ cell frequency (f and h) and MFI (i) and depict 2-3 independent experiments. (j) Graphs show the mean \pm SEM of cytokine⁺ cell frequency after 4-hour stimulation with BD Leukocyte activation cocktail. *, $p \leq 0.05$

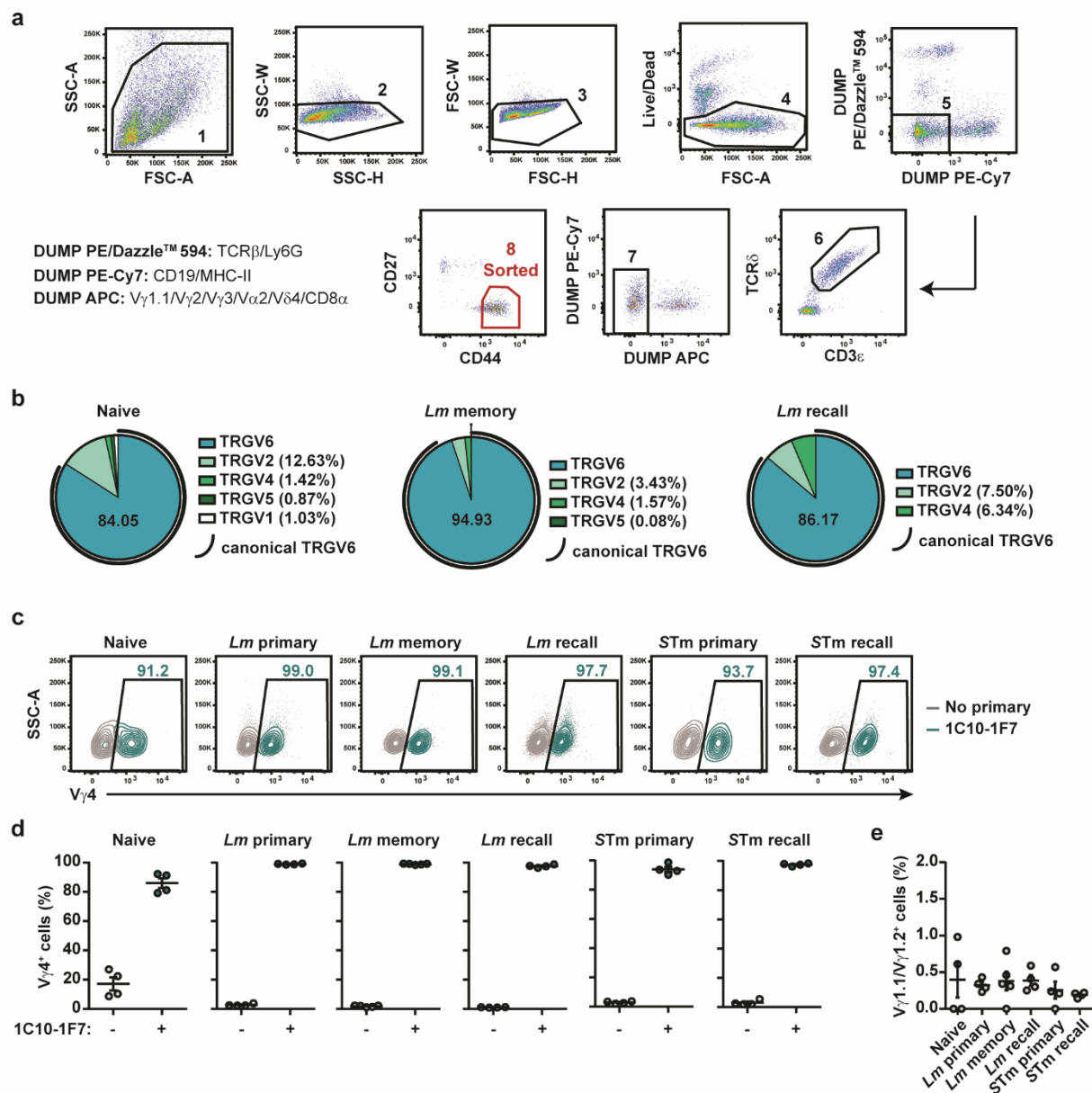


Figure S3. The vast majority of sorted CD44^{hi} CD27^{neg} $\gamma\delta$ T cells express the V γ 4 TCR. (a) Schematic of the gating strategy used to sort purify adaptive $\gamma\delta$ T cells. The numbers within the plots correspond to the gating hierarchy. (b) *Lm*-elicited $\gamma\delta$ T cells in the MLN of infected Balb/c mice were sort-purified >30 days after primary infection (Memory) or 5 days after secondary infection (Recall). Naïve adaptive $\gamma\delta$ T cells were sorted from the MLN, spleen and pLN (n=4-34 mice/group, 1 biological sample). CDR3 γ transcripts were sequenced. Pie charts represent the percent of total reads mapping to the TCR γ usage in each sample. (c-e) MLN cells (*Lm* and STm samples) and mixed MLN, pLN and splenocytes (Naïve) were stained with the V γ 4-specific antibody 1C10-1F7 or left unstained. Adaptive $\gamma\delta$ T cells were identified using the strategy shown in (a) and analyzed for 1C10-1F7 or V γ 1.1/V γ 1.2 staining. Representative flow plot overlays of 1C10-1F7 and no primary control staining are shown in (c). Scatter plots in (d) and (e) show the mean \pm SEM (n=4-6 mice/group) of the percentage of V γ 4⁺ and V γ 1.1/V γ 1.2⁺ cells, respectively.

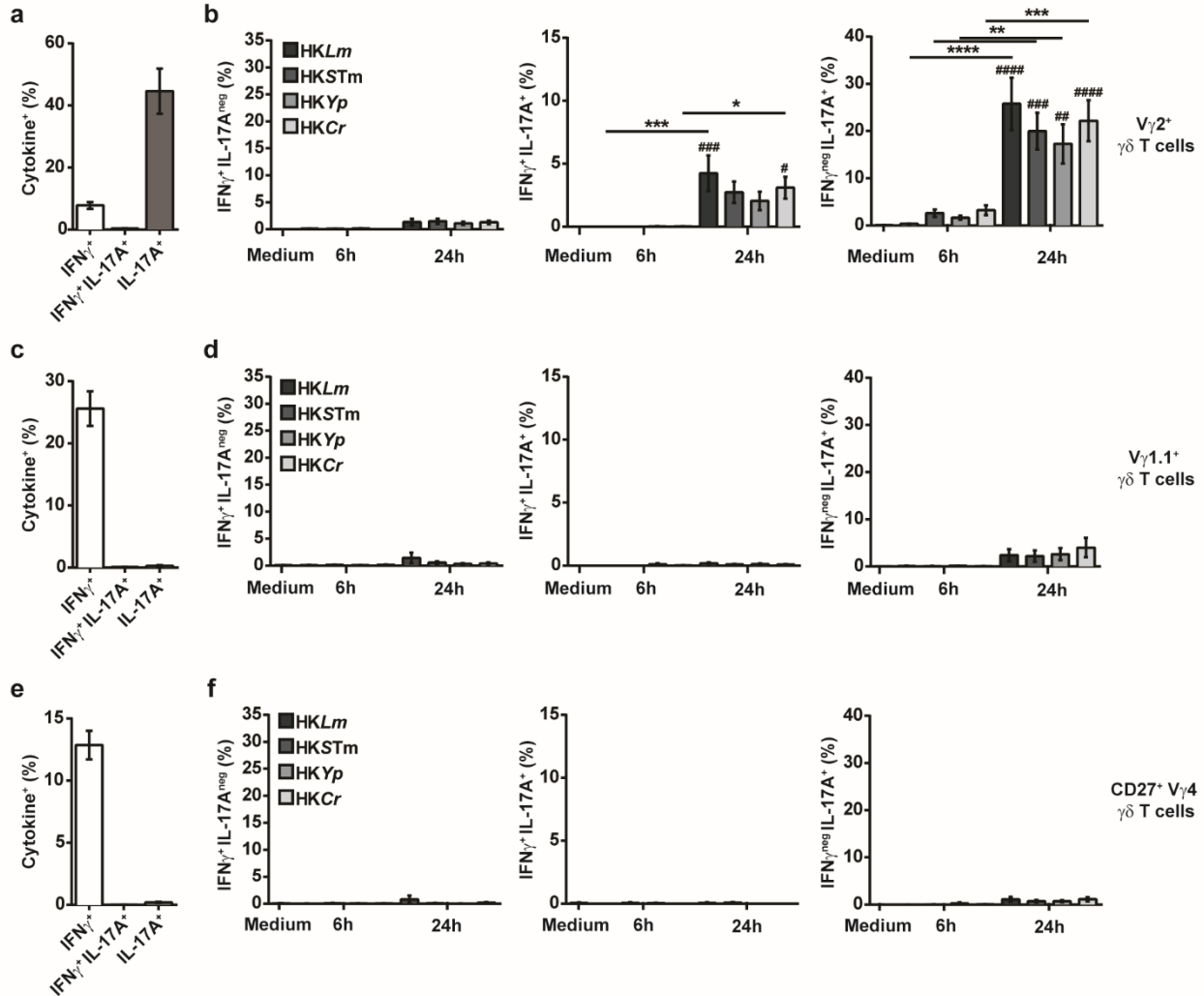


Figure S4. Cytokine production among $\gamma\delta$ T cell subsets in response to heat-killed bacteria. Single cell suspensions of MLN from *Lm*-immune mice were stimulated for 6 hours with Leukocyte Activation Cocktail (PMA, ionomycin, and brefeldin A; a, c and e) or 6 and 24 hours with the indicated heat-killed (HK) bacteria at MOI 10 (b, d and f). (a and b) $V\gamma 2^+$ T cells were analyzed for IL-17A and IFN γ production. (c and d) $V\gamma 1.1^+$ T cells were analyzed for IL-17A and IFN γ production. (e and f) CD27 $^+$ $V\gamma 4$ T cells were analyzed for IL-17A and IFN γ production. Bar graphs show the mean \pm SEM ($n=4$ biological replicates/condition/experiment) of the percentage of cytokine-producing cells and depict the cumulative data of 2 independent experiments. *, $p \leq 0.05$; **, $p \leq 0.01$; ***, $p \leq 0.001$; ****, $p \leq 0.0001$

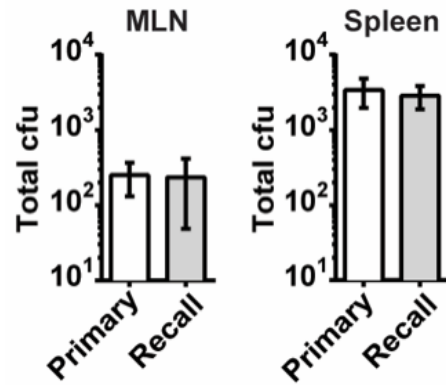


Figure S5. Intraperitoneal *Cr* infection leads to early colonization of the MLN and spleen. Naïve or *Lm*-immune mice were i.p. infected with *Cr*. Two days later, MLN and spleen colonization was evaluated. Bar graphs show the mean \pm SEM (n=3-4 mice/group) of the total cfu per organ.

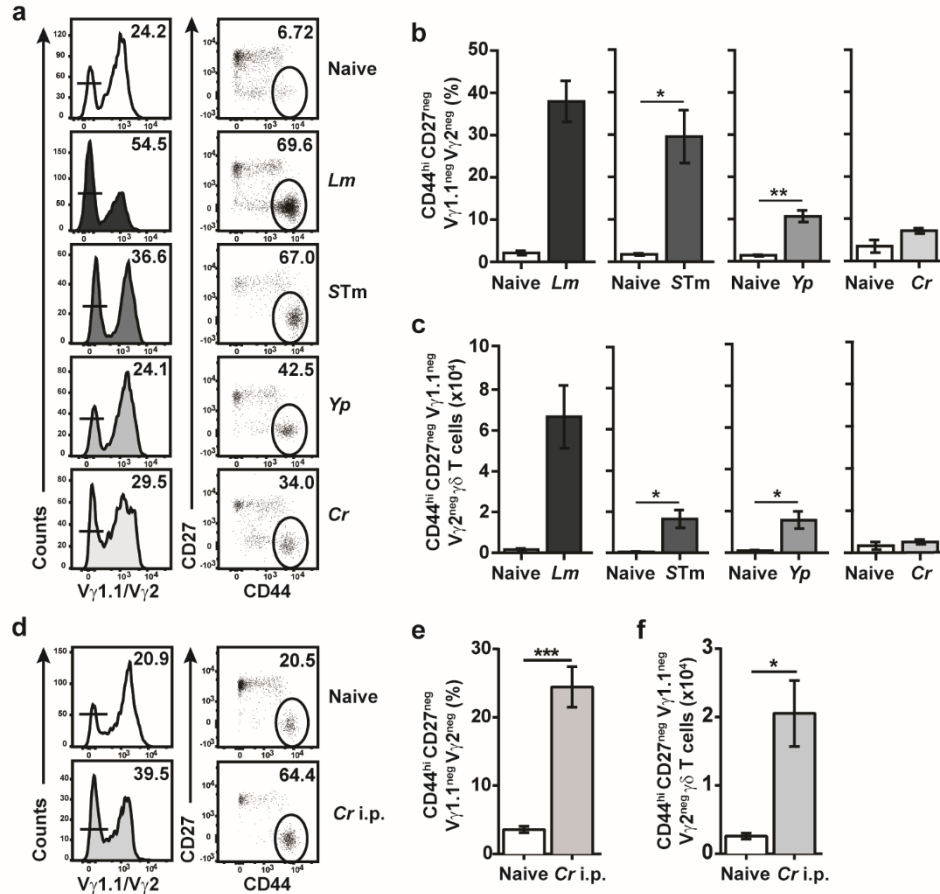


Figure S6. Diverse bacterial infection can elicit adaptive $\gamma\delta$ T cell responses. (a-c) Mice were left uninfected or infected with *Lm*, *STm*, *Yp* and *Cr*. $\gamma\delta$ T cells were analyzed by flow cytometry at 4 dpi (*STm*) or 9 dpi (*Lm*, *Yp*, and *Cr*). Representative flow plots are shown in (a). The naive mouse depicted was a control for the primary response to *Lm*. Data represent the mean \pm SEM ($n=3-6$ mice/group, except for the naive group in the *Lm* panels which comprised 2 mice) of the percentage (b) and absolute number (c) of CD44^{hi} CD27^{neg} V γ 4 T cells and are representative of 2 independent experiments. (d-f) Naïve mice were left uninfected or i.p. infected with *Cr* and $\gamma\delta$ T cells analyzed in the MLN 9 days later. Representative flow plots are shown in (d). Data depict the mean \pm SEM ($n=3-4$ mice/group) of the percentage (e) and absolute numbers (f) of CD44^{hi} CD27^{neg} V γ 4 T cells from the pool of 2 independent experiments. *, $p \leq 0.05$; **, $p \leq 0.01$; ***, $p \leq 0.001$

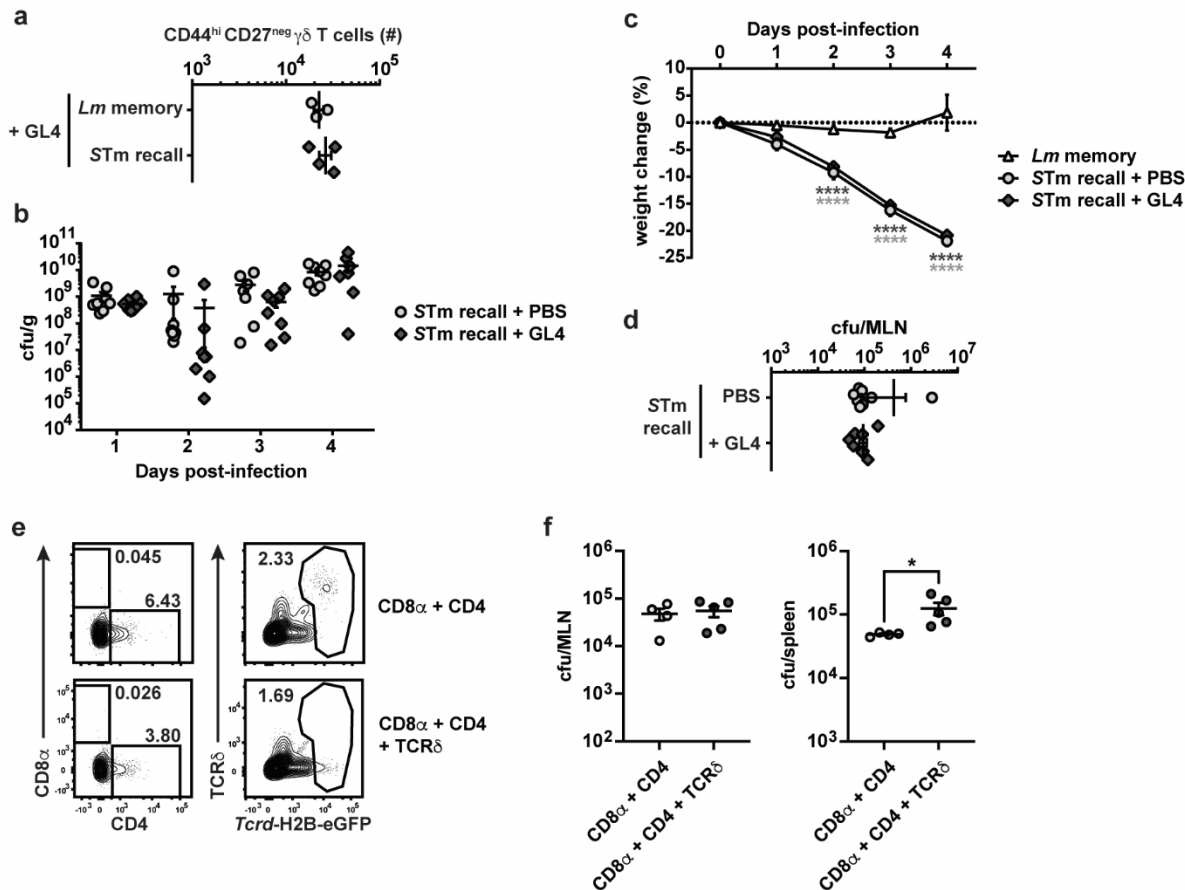


Figure S7. GL4 treatment does not affect the course of STm infection in *Lm*-immune mice. *Lm*-immune *Tcrd*-H2B-eGFP mice received 3 i.p. injections of PBS or GL4 mAb on days -3, -1 and +1 relative to STm infection. (a) $\gamma\delta$ T cell numbers were evaluated from the MLN of GL4 treated mice at 4 dpi. Data show the mean \pm SEM of 3-4 mice/group. (b-c) Mice were followed daily for (b) STm fecal shedding and (c) weight loss. (d) At 4 dpi, STm colonization was evaluated in the MLN. Data are shown as mean \pm SEM and depict the cumulative data of 2 independent experiments with 3-4 mice/group. (e and f) *Lm*-immune *Tcrd*-H2B-eGFP mice received 3 i.p. injections of the indicated antibody cocktails on days -3, -1 and +1 relative to STm challenge infection. (e) Representative flow plots of CD4 and CD8 α expression among live single lymphocytes (left panel) and TCR δ and *Tcrd*-H2B-eGFP expression among CD4⁻ CD8 α ⁺ lymphocytes (right panel) are shown. Analysis was performed on circulating cells. (f) STm colonization of the indicated tissues. Graphs show the mean \pm SEM of 1 experiment with 4-5 mice/group. *, $p \leq 0.05$; ****, $p \leq 0.0001$

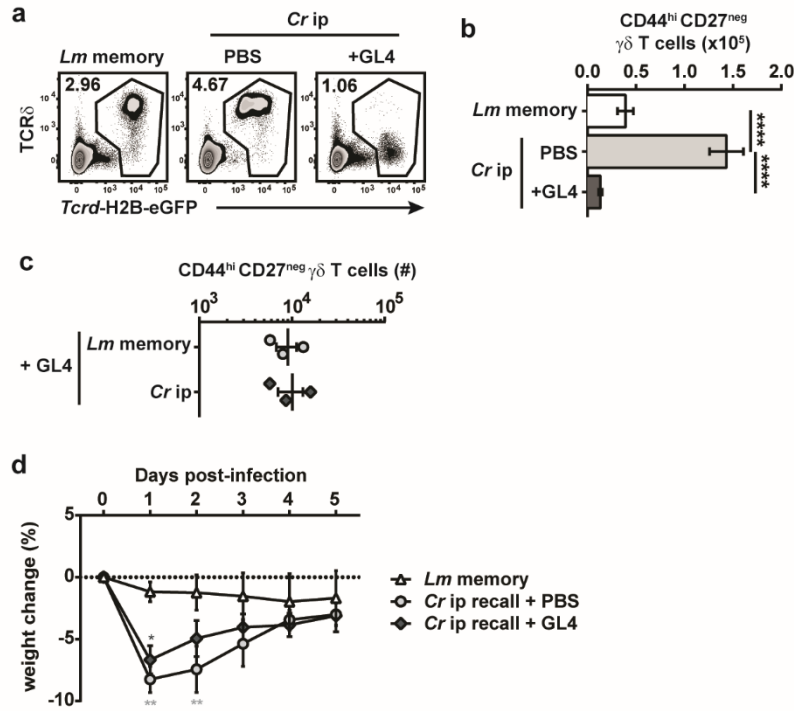


Figure S8. *Lm*-elicited $\gamma\delta$ T cell response to *Cr* i.p. is mediated by the $\gamma\delta$ TCR. *Lm*-immune *Tcrd*-H2B-eGFP mice received 3 i.p. injections of PBS (*Lm* memory and PBS groups) or GL4 mAb on days -3, -1 and +1 relative to i.p. *Cr* challenge (+GL4). Memory mice received 100 μ l PBS i.p. on day 0. Recall groups were challenged i.p. with *Cr* on day 0. (a) CD44^{hi} CD27^{neg} $\gamma\delta$ T cells were analyzed after 5 days. $\gamma\delta$ T cells were identified as CD4^{neg} CD8 α ^{neg} GFP⁺ single live lymphocytes. Representative flow plots are shown. (b) Data show the mean \pm SEM (n=3-4 mice/group/experiment) of CD44^{hi} CD27^{neg} $\gamma\delta$ T cell absolute numbers at 5 dpi and depict the cumulative data of 2 independent experiments. (c) Comparison of CD44^{hi} CD27^{neg} $\gamma\delta$ T cell absolute numbers in the MLN of GL4-treated *Lm*-immune and *Cr* challenged mice at 5 dpi (n=3 mice/group). (d) Mice were followed daily for weight loss (n=4 mice/group). *, $p \leq 0.05$; **, $p \leq 0.01$; ****, $p \leq 0.0001$

Table S1. Frequency of the non-canonical clones selected by STm primary infection in Figure 7f at each analyzed time point. Sequences are sorted by abundance at the STm primary time point. Sequences that increase in both STm primary and recall conditions are shown in bold.

Sequence	% of other non-canonical reads					
	Naïve	<i>Lm</i> primary	STm primary	<i>Lm</i> memory	<i>Lm</i> recall	STm recall
GSDRRDTTDKLV	4.506	8.232	7.171	8.644	9.667	4.070
GSDTEGSSWDTRQMF	0	0.440	0.070	0	0.004	0
GSDIGGIRVTDKLV	0.023	0.048	0.047	0.059	4.167	0.043
GSGRRDTTDKLV	0.013	0.025	0.033	0.010	0.027	0.007
GSDRGDATDKLV	0	0.031	0.031	0	0.004	0
GSGRRDSDKLV	0.004	0.016	0.029	0.026	0.028	0
GSDRGDTTDKLV	0.013	0.025	0.023	0.013	0.012	0
GSDVGGTDKLV	0	0.008	0.016	0.009	0.010	0.009
GSDIGIRAADKLV	0	0.004	0.006	0	0.020	0.017
GSDIGGITNKLV	0	0.003	0.004	0	0.003	0
GSDIGGSPRDTRQMF	0	0.003	0.004	0	0.003	0.004
GSDRRDTGSDWTRQMF	0	0.003	0.004	0.005	0.005	0
GSDVGGTTDKLV	0	0.008	0.004	0	0.008	0.011
GSGRDTSSWDTRQMF	0	0.007	0.004	0	0.005	0
GSDIGGTIDKLV	0	0.008	0.004	0	0.013	0.013
GSDRRDTADKLV	0	0.014	0.004	0	0.004	0.010

Table S2. Frequency of the non-canonical clones selected by STm recall infection in Figure 7f at each analyzed time point. Sequences are sorted by abundance at the STm recall time point. Sequences that increase in both STm primary and recall conditions are shown in bold.

Sequence	% of other non-canonical reads					
	Naïve	<i>Lm</i> primary	STm primary	<i>Lm</i> memory	<i>Lm</i> recall	STm recall
GSDIGGTDKLV	9.147	10.231	7.258	6.257	8.102	19.865
GSDIGGSFWDTRQMF	0.406	0.420	0.353	0.388	0.400	0.409
GSDIGGSYWDTRQMF	0.143	0.161	0.111	0.160	0.166	0.172
GSDFGGSSWDTRQMF	0.061	0.0786	0.054	0.045	0.055	0.072
GSDIGGIATDKLV	0.004	3.850	0.004	0	1.044	0.030
GSDRGDTDKLV	0.019	0.046	0	0.018	0.036	0.026
GSDIGIRAADKLV	0	0.004	0.006	0	0.020	0.017
GSDIGGTIDKLV	0	0.008	0.004	0	0.013	0.013
GSGIGGSSWDTRQMF	0.004	0.010	0	0.005	0.016	0.013
GSDRRDTADKLV	0	0.014	0.004	0	0.004	0.011
GSDVGGTTDKLV	0	0.008	0.004	0	0.008	0.011
GSGTGGSSWDTRQMF	0	0.014	0	0	0.004	0.004
GSDIGGIGKLV	0	0.013	0	0	0.010	0.004
GSDIGGSPRDTRQMF	0	0.003	0.004	0	0.003	0.004

Table S3. Bacterial strains and infection doses.

BACTERIA	STRAIN	INFECTION DOSE (CFU)	REFERENCES
<i>InlA^M Listeria monocytogenes</i>	EGDe	Primary: 2×10^9 Secondary: 2×10^{10}	(1)
<i>InlA^M Listeria monocytogenes</i>	10403s	GF mice - Primary: 1×10^8 gavage Nur77 ^{GFP} and NTg – Primary: 2×10^9 – Secondary: 2×10^{11}	(2)
<i>Salmonella enterica</i> serovar Typhimurium	IR715	Primary and secondary: 1×10^8	(3)
<i>Yersinia pseudotuberculosis</i> serogroup O1	32777	Primary: 5×10^7 Secondary: 1×10^8	(4)
<i>Citrobacter rodentium</i>	DBS100	Primary and secondary: 1×10^9 oral 1×10^7 i.p.	(5)

Table S4. List of antibodies used in this study.

Marker	Conjugate	Clone	Company
anti-mouse IgG	FITC	Poly4060	BioLegend
CD3 ϵ	PE/Cy7	145-2C11	BioLegend
CD3 ϵ	BV421	145-2C11	BioLegend
CD3 ϵ	PE/Dazzle™ 594	145-2C11	BioLegend
CD3 ϵ	BV711	145-2C11	BioLegend
CD4	Purified	GK1.5	Bio X Cell
CD4	Biotin	GK1.5	BioLegend
CD4	PE/Cy7	RM4-4	BioLegend
CD8 α	Purified	2.43	Bio X Cell
CD8 α	Biotin	53-6.7	BioLegend
CD8 α	APC	53-6.7	BioLegend
CD8 α	PE	53-6.7	BioLegend
CD8 α	BV785	53-6.7	BioLegend
CD16/CD32	Purified	2.4G2	Bio X Cell
CD19	PE/Cy7	6D5	BioLegend
CD27	PerCP/Cy5.5	LG.3A10	BioLegend
CD44	APC-eFluor 780	IM7	eBioscience
B220	Biotin	RA3-6B2	BioLegend
I-A/I-E	PE/Cy7	M5/114.15.2	BioLegend
IFN γ	PE/Cy7	XMG1.2	BioLegend
IL-17A	APC	TC11-18H10.1	BioLegend
Ki-67	PE/Dazzle™ 594	16A8	BioLegend
Ly6G	PE/Dazzle™ 594	1A8	BioLegend
Phospho-Zap 70/Syk	PE	n3kobu5	Invitrogen
TCR β	PE/Dazzle™ 594	H57-597	BioLegend
TCR β	BV711	H57-597	BioLegend
TCR δ	BV421	GL3	BioLegend
TCR δ	PE	GL3	BioLegend
TCR δ	APC	GL3	BioLegend
TCR δ	Purified	GL4	Bio X Cell (Custom)
TCR δ	Purified	UC7-13D5	Bio X Cell
V α 2	APC	B20.1	BioLegend
V δ 4	APC	REA372	Miltenyi Biotec
V γ 1.1	FITC	2.11	BioLegend
V γ 1.1	APC	2.11	BioLegend
V γ 1.1/V γ 1.2	PE	4B2.9	BioLegend
V γ 2	FITC	UC3-10A6	BioLegend
V γ 2	PE	UC3-10A6	BioLegend
V γ 2	APC	UC3-10A6	BioLegend
V γ 3	APC	536	BioLegend
V γ 4	Purified	1C10-1F7	Provided by Dr. Hatano and Dr. Yoshikai

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