

Supplemental information

**Inhibition of IKK β /NF- κ B signaling facilitates
tendinopathy healing by rejuvenating
inflamm-aging induced tendon-derived
stem/progenitor cell senescence**

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Supporting Information

Inhibition of IKK β /NF- κ B signaling facilitates tendinopathy healing by rejuvenating
inflamm-aging induced tendon-derived stem/progenitor cell senescence

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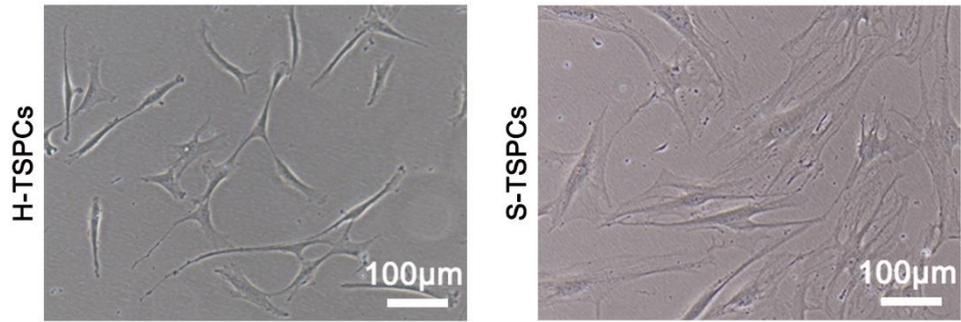


Fig. S1. Cell morphology of S-TSPCs and H-TSPCs. H-TSPCs, TSPCs isolated from degenerative supraspinatus tendons. S-TSPCs, TSPCs isolated from healthy hamstring tendons.

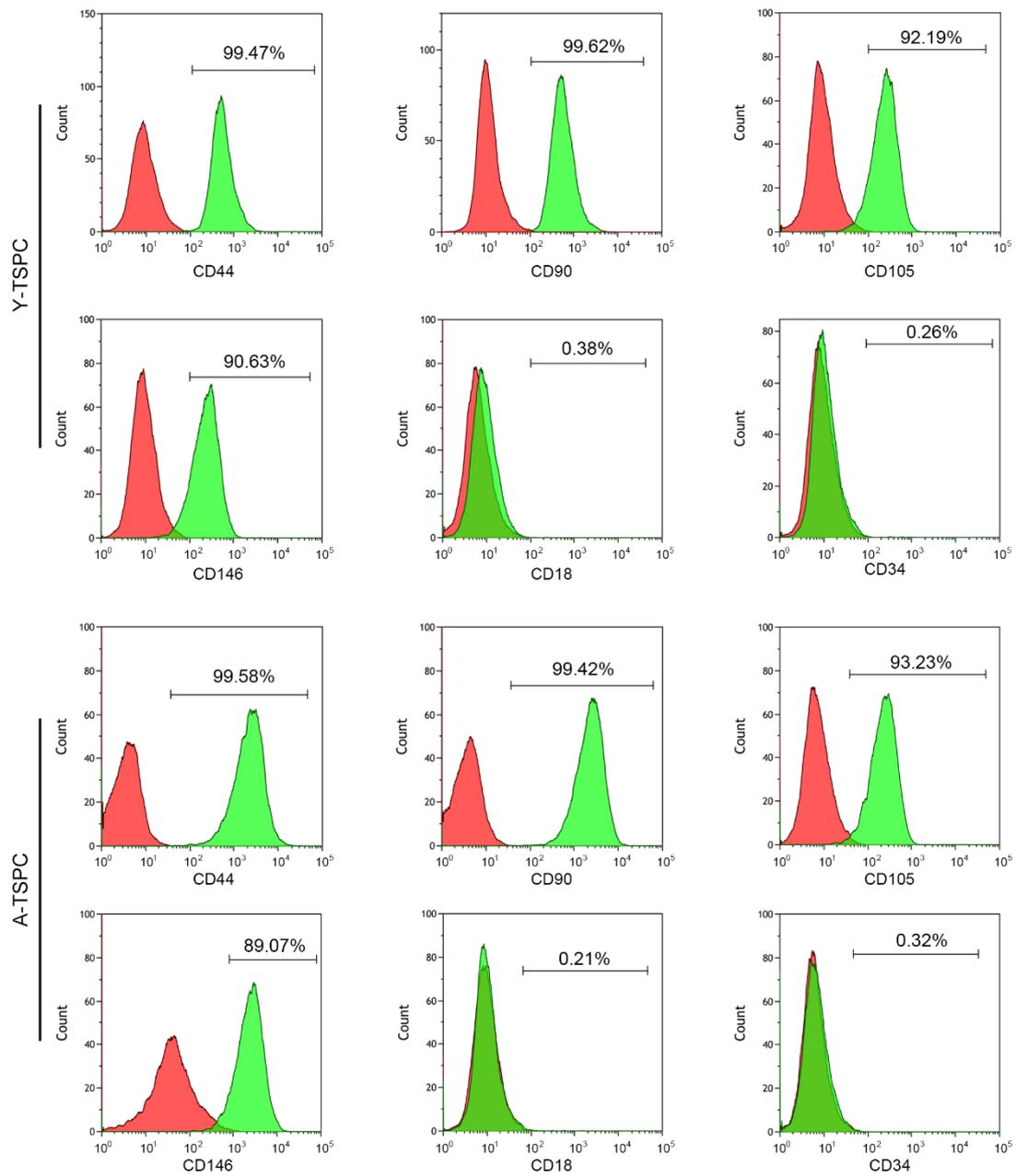


Fig. S2. Flow cytometry analysis of TSPCs surface markers. The red curves represent isotype controls, and the green curves represent measured surface markers (CD44, CD90, CD105, CD146, CD18, and CD34). H-TSPCs, TSPCs isolated from healthy hamstring tendons. S-TSPCs, TSPCs isolated from degenerative supraspinatus tendons.

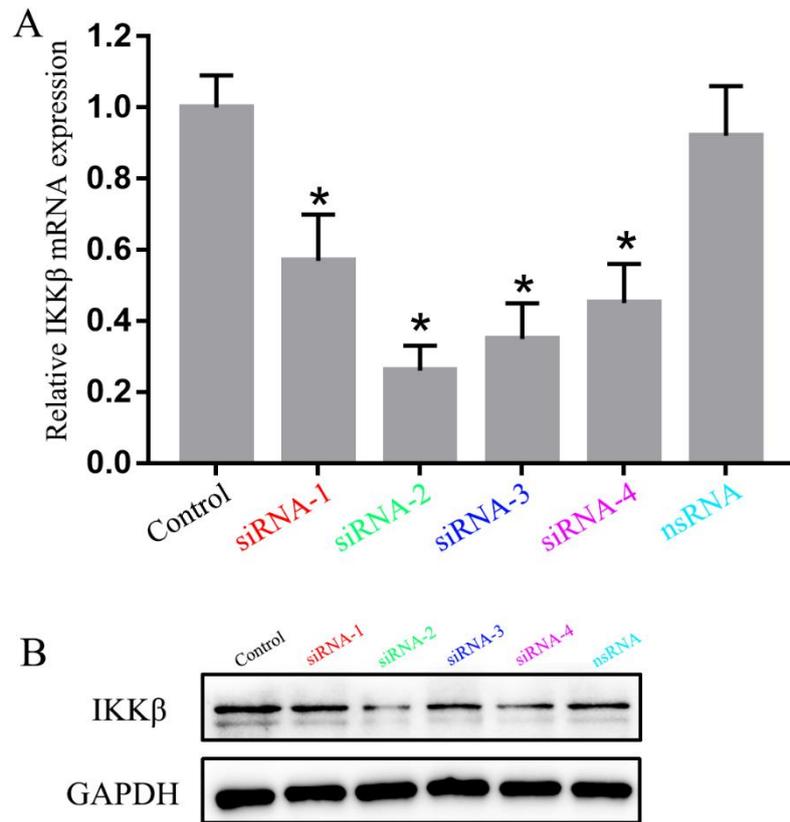


Fig. S3. IKK β gene silencing in TSPCs with different siRNA sequences. (A) Expression of IKK β mRNA in TSPCs. (B) Levels of IKK β protein in TSPCs.

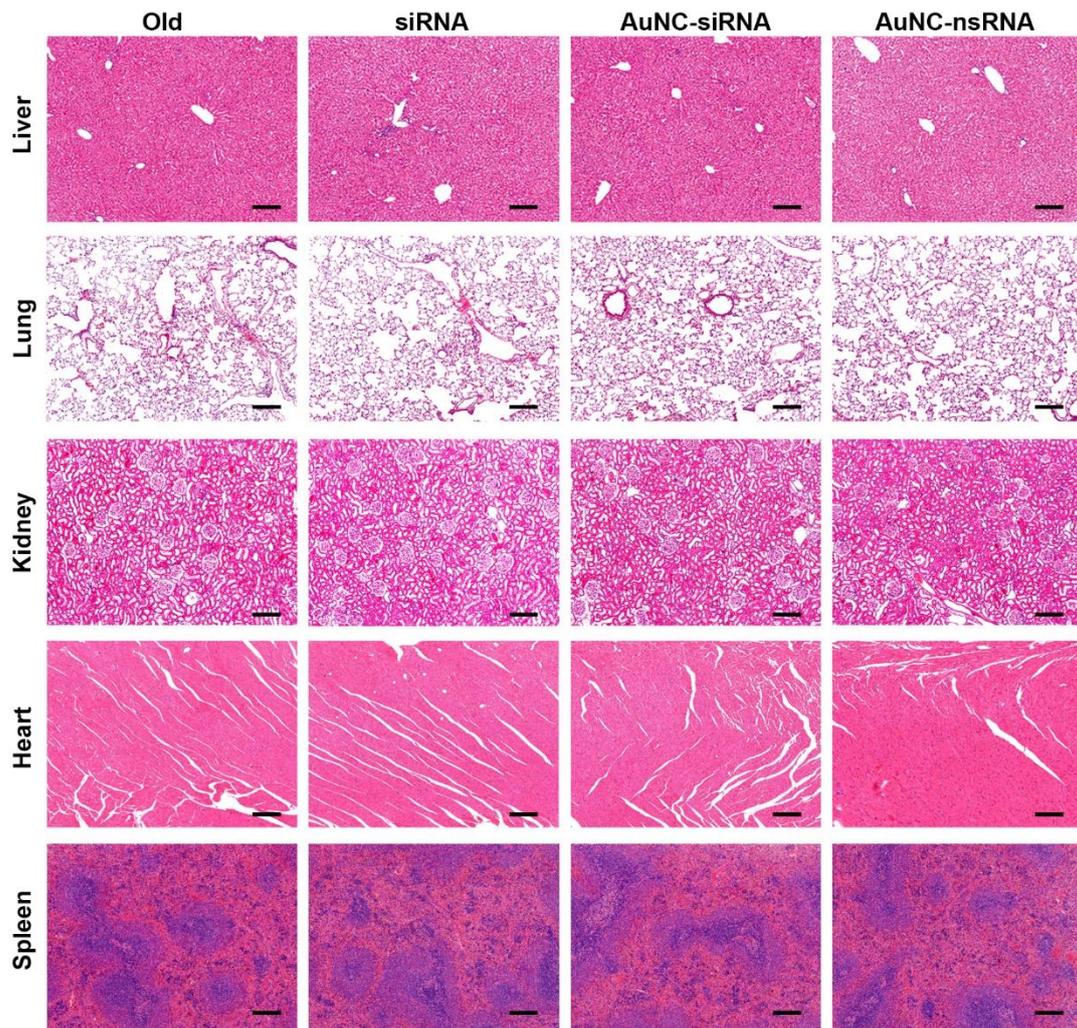


Fig. S4. (A) In vivo toxicity of AuNCs. Representative photomicrographs of H&E staining on the major organs including liver, lung, kidney, heart and spleen. Scale bars represent 200 μm.

Table S1. The exclusion criteria for patients to isolate torn supraspinatus tendons

Previous significant shoulder trauma
Systemic steroid use or tobacco smoking
Previous or current shoulder problems other than rotator cuff tendinopathy
Previous shoulder surgery
Local glucocorticoid injections in the last 6 weeks
Significant systemic medical conditions, e.g., diabetes or a rheumatological condition

Table S2. List of primers used for human TSPCs RT-PCR analysis.

Target	Specie	Forward primer (5' → 3')	Reverse primer (5' → 3')
IL6	Human	AGACAGCCACTCACCTCTTCAG	TTCTGCCAGTGCCTCTTTGCTG
IL1A	Human	TGTATGTGACTGCCCAAGATGAAG	AGAGGAGGTTGGTCTCACTACC
CXCL1	Human	AGCTTGCCTCAATCCTGCATCC	TCCTTCAGGAACAGCCACCAGT
CXCL2	Human	GGCAGAAAGCTTGCTCAACCC	CTCCTTCAGGAACAGCCACCAA
CDKN2A (p16)	Human	CTCGTGCTGATGCTACTGAGGA	GGTCGGCGCAGTTGGGCTCC
SCX	Human	AGACCGCACCAACAGCGTGAAC	CAGGTGCGAGATGTAGCTGGAG
BGN	Human	TTGAACCTGGAGCCTTCGATGG	TTGGAGTAGCGAAGCAGGTCCT
TNMD	Human	GGACTGGTGTTTGGTATCCTGG	CTCCATTGCTGTAGAAAGTGTGC
DCN	Human	GCTCTCCTACATCCGCATTGCT	GTCCTTTCAGGCTAGCTGCATC
COL1	Human	GATTCCTGGACCTAAAGGTGC	AGCCTCTCCATCTTTGCCAGCA
COL3	Human	TGGTCTGCAAGGAATGCCTGGA	TCTTCCCTGGGACACCATCAG
IKK β	Human	ACTTGGCGCCCAATGACCT	CTCTGTTCCTTGCTGCA
GAPDH	Human	CGACCACTTTGTCAAGCTCA	AGGGGTCTACATGGCAACTG

Table S3. Sequences of different IKK β siRNA.

Sequence	Forward primer (5' → 3')	Reverse primer (5' → 3')
siRNA-1	GGAAGUACCUGAACCAGUUTT	AACUGGUUCAGGUACUUCCTT
siRNA-2	GGACAUUGUUGUUAGCGAATT	UUCGCUAACAACAAUGUCCTT
siRNA-3	CCAUGUCUCAGCAGCUCAATT	UUGAGCUGCUGAGACAUGGTT
siRNA-4	GCUGCUGCUUCAGGCAAUUTT	AAUUGCCUGAAGCAGCAGCTT
nsRNA	UUCUUCGAACGUGUCACGUTT	ACGUGACACGUUCGGAGAATT

Table S4. List of primers used for rat TSPCs RT-PCR analysis.

Target	Specie	Forward primer (5' → 3')	Reverse primer (5' → 3')
Sex	Rat	CAGCGGCACACGGCGAAC	CGTTGCCCAGGTGCGAGATG
Tnmd	Rat	GCGACAATGTGACTATGTAC	GTCTTCTCCACCTTCACTTG
Col1	Rat	AGAGGCATAAAGGGTCATCGTG	AGACCGTTGAGTCCATCTTGC
Gapdh	Rat	CTGGAGAAACCTGCCAAGTATG	GGTGAAGAATGGGAGTTGCT

Table S5. List of antibodies for flow cytometry analysis of TSPCs surface markers.

Antibodies	Source	Cat#
PE-CF594 Mouse Anti-Human CD105	BD Biosciences	562380
PE-CF594 Mouse IgG1, κ Isotype Control RUO	BD Biosciences	562292
Hu CD90 APC 5E10 25ug	BD Biosciences	561971
APC Mouse IgG1, κ Isotype Control	BD Biosciences	550854
BV421 Mouse Anti-Human CD146	BD Biosciences	566226
BV421 Mouse IgG1, κ Isotype Control RUO	BD Biosciences	562438
BV786 Mouse Anti-Human CD44	BD Biosciences	564942
BV786 Mouse IgG2b, κ Isotype Control RUO	BD Biosciences	564090
FITC Mouse Anti-Human CD18	BD Biosciences	557156
FITC Mouse IgG1, κ Isotype Control	BD Biosciences	556649
APC Mouse Anti-Human CD34	BD Biosciences	560940
APC Mouse IgG1, κ Isotype Control	BD Biosciences	550854