

Supplementary Material

Structural and molecular insight into piperazine and piperidine derivatives as histamine H₃ and sigma-1 receptor antagonists with promising antinociceptive properties

Katarzyna Szczepańska^{+,a,b}, Sabina Podlewska^{+,a,b,*}, Maria Dichiarà^c, Davide Gentile^c, Vincenzo Patamia^c, Niklas Rosier^d, Denise Mönnich^d, M^a Carmen Ruiz Cantero^e, Tadeusz Karcz^a, Dorota Łażewska^a, Steffen Pockes^d, Agata Siwek^f, Enrique J. Cobos^e, Agostino Marrazzo^c, Holger Stark^g, Antonio Rescifina^c, Andrzej J. Bojarski^b, Emanuele Amata^{c,*}, Katarzyna Kieć-Kononowicz^{a,*}

^a Department of Technology and Biotechnology of Drugs, Faculty of Pharmacy, Jagiellonian University Medical College, Medyczna 9, Kraków 30-688, Poland

^b Maj Institute of Pharmacology, Polish Academy of Sciences, Smełna 12, Kraków 31-343, Poland

^c Department of Drug and Health Sciences, University of Catania, V.le A. Doria, 95125 Catania, Italy

^d Institute of Pharmacy, Faculty of Chemistry and Pharmacy, University of Regensburg, Universitätsstraße 31, D-93053 Regensburg, Germany

^e Department of Pharmacology and Neurosciences Institute (Biomedical Research Center), University of Granada, and Biosanitary Research Institute *ibs.GRANADA*, Avenida de la Investigación 11, 18016 Granada, Spain

^f Department of Pharmacobiology, Faculty of Pharmacy, Jagiellonian University Medical College, Medyczna 9, Kraków 30-688, Poland

^g Institute of Pharmaceutical and Medicinal Chemistry, Heinrich Heine University Düsseldorf, Universitätsstr. 1, 40225 Duesseldorf, Germany

* Corresponding authors. E-mail addresses: smusz@if-pan.krakow.pl (Sabina Podlewska), [mfkonono@cyf-kr.edu.pl](mailto:mfkono@cyf-kr.edu.pl) (Katarzyna Kieć-Kononowicz), eamata@unict.it (Emanuele Amata).

[*] These authors contributed equally to this work.

Contents

1. Purity confirmation data.....	S2
2. Chemistry.....	S3
3. Calibration of parameters for the Smina scoring function.....	S4
4. Molecular modeling – 3D docking poses of compounds 4, 5, and 11 in the binding site of σ_1 R, σ_2 R, and H ₃ R S5	
5. Outcome of correlational studies between frequency of interaction of ligands with particular amino acid residues of σ R.....	S6
6. References.....	S8
7. ¹ H- and ¹³ C-NMR spectra of compounds 11 and 12. HPLC traces of all the final compounds.....	S9

1. Purity confirmation data

Mass spectra (LC/MS) were performed on Waters TQ Detector mass spectrometer. High-resolution mass spectra (HRMS) were performed on UltrafleXtreme maldi-tof/tof mass spectrometer. HPLC analyses were performed on Waters Alliance 2695 Separations Module with Waters 2998 Photodiode Array Detector (Chromolith® SpeedROD RP-18 endcapped 50-4.6 HPLC column was used)¹⁻⁴. Elemental analyses (C, H, N) were performed on an Elemental AnalyserVarioEl III (Hanau, Germany) and agreed with theoretical values within $\pm 0.4\%$ ^{1,2}.

Cpd	LC/MS		HRMS		RP-HPLC		References	
	Purity [%]	Retention time [min]	ESI <i>m/z</i> [M+H] ⁺	MALDI <i>m/z</i> [M+H] ⁺	Purity [%]	Retention time [min]		<i>k</i>
1	100	3.23	326	326.217	99.21	0.923	1.64	3
2	100	4.02	355	-	99.30	1.082	2.09	1
3	100	1.87	341	-	100	0.723	1.07	2
4	100	3.04	366	366.203	100	0.756	1.16	4
5	100	3.28	365	365.209	100	0.788	1.25	4
6	100	3.89	374	374.224	99.64	1.111	2.17	3
7	100	3.67	399	399.234	99.40	0.971	1.77	4
8	97.83	3.18	402	402.212	95.42	0.971	1.77	3
9	97.28	4.07	436	436.183	95.56	1.040	1.97	4
10	95.61	3.61	420	420.195	95.76	0.928	1.65	4
11	100	5.24	296	296.209	100	1.280	2.66	-
12	100	4.47	324	324.201	100	1.140	2.26	-
13	100	4.18	369	-	100	1.168	2.34	1
14	98.46	4.66	383	-	98.06	1.252	2.58	1
15	100	2.26	355	-	-	-	-	2
16	93.73	2.90	382	382.253	92.19	0.942	1.69	3
17	100	3.43	396	396.266	100	1.029	1.94	3
18	100	5.26	396	396.303	100	1.334	2.81	3
19	94.79	3.69	410	410.284	94.47	1.124	2.21	3
20	100	4.03	424	424.296	100	1.202	2.43	3

Cpd 2: Anal. Calcd for C₂₂H₃₁N₃O x C₂H₂O₄: C, 64.99, N, 9.47, H, 7.50%. Found: C, 64.96, N, 9.52, H, 7.48.

Cpd 13: Anal. Calcd for C₂₃H₃₃N₃O x C₂H₂O₄: C, 65.62, N, 9.18, H, 7.71%. Found: C, 65.60, N, 9.19, H, 7.72.

Cpd 14: Anal. Calcd for C₂₄H₃₅N₃O x C₂H₂O₄: C, 66.22, N, 8.91, H, 7.91%. Found: C, 66.19, N, 8.89, H, 7.91.

Cpd 15: Anal. Calcd for C₂₁H₂₇N₃O₂ x C₂H₂O₄: C, 62.29, N, 9.47, H, 6.59%. Found: C, 62.32, N, 9.49, H, 6.63.

2. Chemistry

Compounds **11** and **12** were obtained by *N*-alkylation of piperidine with proper bromides by the method described by Kuder *et al.*⁵ Both compounds have been previously described in the literature^{6,7}.

(Biphenyl-4-yloxy)propyl)piperidine hydrogen oxalate (11)

4-(3-Bromopropoxy)-1,1'-biphenyl (CAS113795-28-1) (1.46 g; 5 mmol), piperidine (0.21 g; 2.5 mmol) was refluxed for 24 h in the mixture of ethanol-water (5:1) with the powdered potassium carbonate (0.52 g; 3.8 mmol) and catalytic amount of potassium iodide. After purification oily product was converted into hydrogen oxalate. White solid, yield (28%), m.p. 213-215 °C, C₂₂H₂₇NO₅ (MW=385.46). ¹H NMR (500 MHz, DMSO-*d*₆) δ ppm: 7.54 - 7.59 (m, 4H), 7.36 - 7.42 (m, 2H), 7.27 (tt, *J*=7.30, 1.15 Hz, 1H), 6.96 - 7.01 (m, 2H), 4.05 (t, *J*=6.01 Hz, 2H), 3.02 - 3.16 (m, 4H), 2.46 - 2.47 (m, 2H), 2.04 - 2.14 (m, 2H), 1.68 (br. s., 4H), 1.49 (br. s., 2H). ¹³C NMR (DMSO-*d*₆): 164.85, 158.43, 140.27, 133.34, 129.42, 128.32, 126.71, 115.48, 65.62, 54.09, 52.82, 24.22, 23.36, 22.10. LC-MS: purity 100% t_R= 5.24, (ESI) *m/z* [M+H]⁺ 296.20.

Phenyl (4-(3-(piperidin-1-yl)propoxy)phenyl)phenyl)methanone hydrogen oxalate (12)

4-(3-Bromopropoxy)phenyl(phenyl)methanone (CAS108357-63-7) (0.998 g; 3.85 mmol), piperidine (0.328g; 3.85 mmol) was refluxed for 24 h in the mixture of ethanol-water (5:1) with the powdered potassium carbonate (0.69 g; 5 mmol) and catalytic amount of potassium iodide. After purification oily product was converted into hydrogen oxalate. White solid, yield (45%), m.p. 129-132 °C, C₂₃H₂₇NO₆ (MW=413.47). ¹H NMR (300 MHz, DMSO-*d*₆) δ ppm: 7.69 - 7.73 (m, 2H), 7.60 - 7.66 (m, 3H), 7.49 - 7.54 (m, 2H), 7.03 - 7.08 (m, 2H), 4.11 (t, *J*=6.16 Hz, 2H), 2.95 - 3.19 (m, 6H), 2.08 - 2.17 (m, 2H), 1.65 - 1.74 (m, 4H), 1.49 (br. s., 2H). ¹³C NMR (DMSO-*d*₆): 194.97, 165.21, 162.52, 138.25, 132.73, 129.79, 129.01, 114.90, 66.01, 53.85, 52.67, 23.94, 23.18, 22.06. LC-MS: purity 100% t_R= 4.47, (ESI) *m/z* [M+H]⁺ 324.18.

None of the final structures has the Pan-assay interference compounds properties⁸.

3. Calibration of parameters for the Smina scoring function

Table S1. Well-known σ_1 and σ_2 receptor ligands used to test the new calibrated Smina scoring function, and their experimental and calculated binding constant values.

Compound	Exp. σ_1 K_i (nM)	Calcd K_i σ_1 (nM)	Exp. σ_2 K_i (nM)	Calcd K_i σ_2 (nM)
Ifenprodil	1.02	2.93	18.8	26.62
PRE-084	2.2	3.68	13091	14691.11
Haloperidol	2.6	3.24	77	89.57
Pentazocine	4.3	6.30	1465	1285.65
BD1063	13.7	16.70	204	231.24
S1RA	17	24.00	9300	9795.90
DTG	124	102.65	18	22.23
F-ISO	330	357.87	6.95	8.35
RHM-4	2150	2071.76	0.26	1.61
Pitolisant	0.5	1.09	6.5	8.14

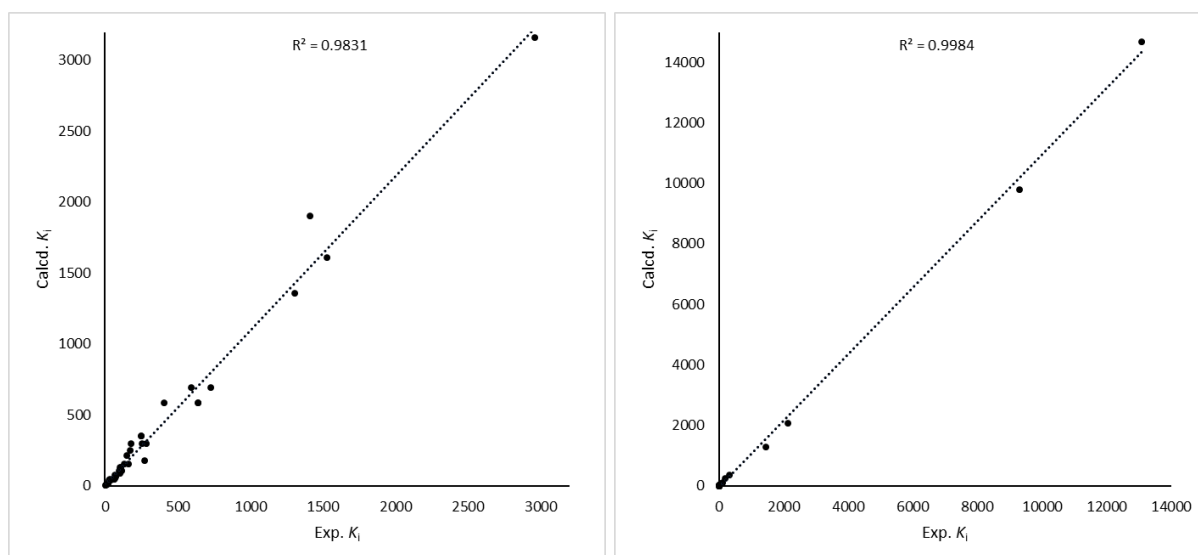


Figure S1. 2D plot of the linear regression fit between experimental and calculated σ_1 and σ_2 binding constants values for (left) compounds 1–14, and 16–20 (the training set), reported in Table 1, and (right) those of the chosen well-known ligands (the test set), reported in Table S1. All values have been calculated by the calibrated Smina scoring function employing the optimized parameters reported in Table 2.

4. Molecular modeling – 3D docking poses of compounds 4, 5, and 11 in the binding site of σ_1R , σ_2R , and H₃R

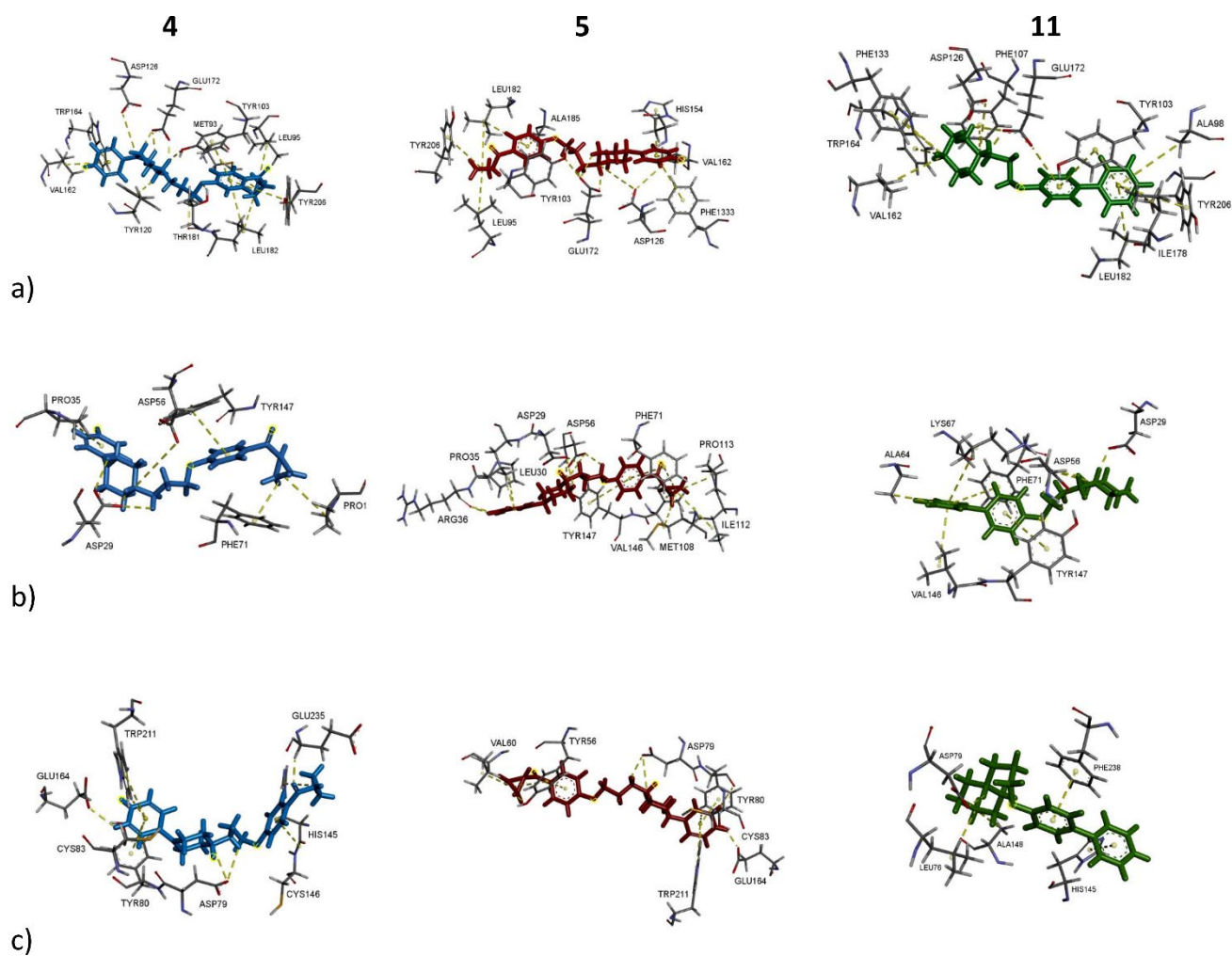


Figure S2. 3D docking poses of compounds 4, 5, and 11 in the binding site of a) σ_1R , b) σ_2R , c) H₃R.

5. Outcome of correlational studies between frequency of interaction of ligands with particular amino acid residues of σ Rs

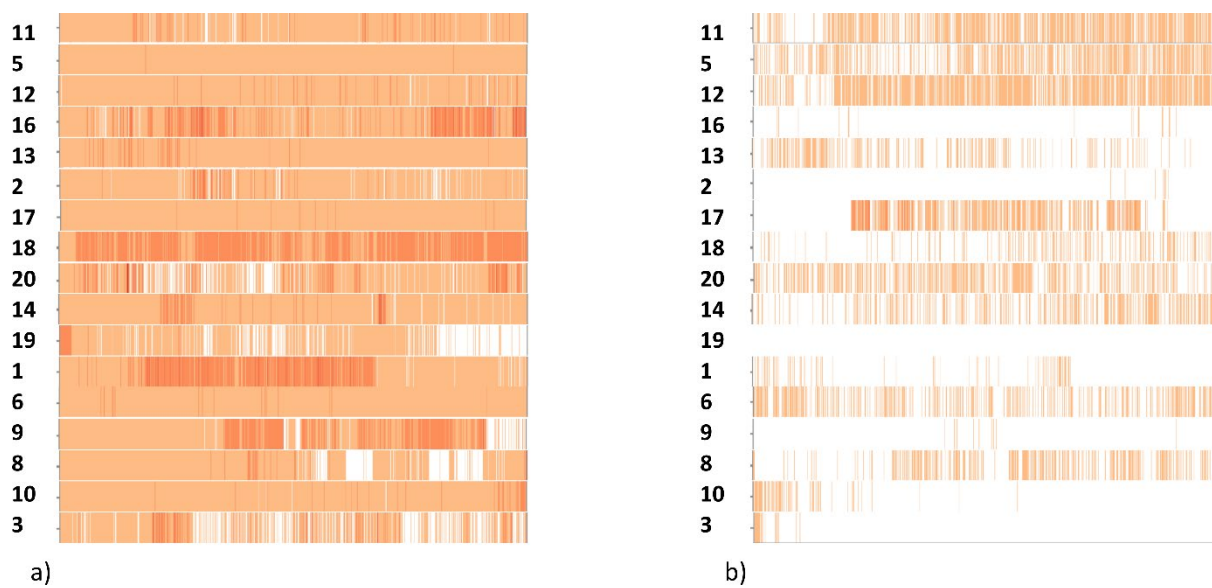


Figure S3. Ligand-protein interaction diagrams obtained during MD simulations for the highest correlated residues of σ_1 R: a) E172, b) Y206.

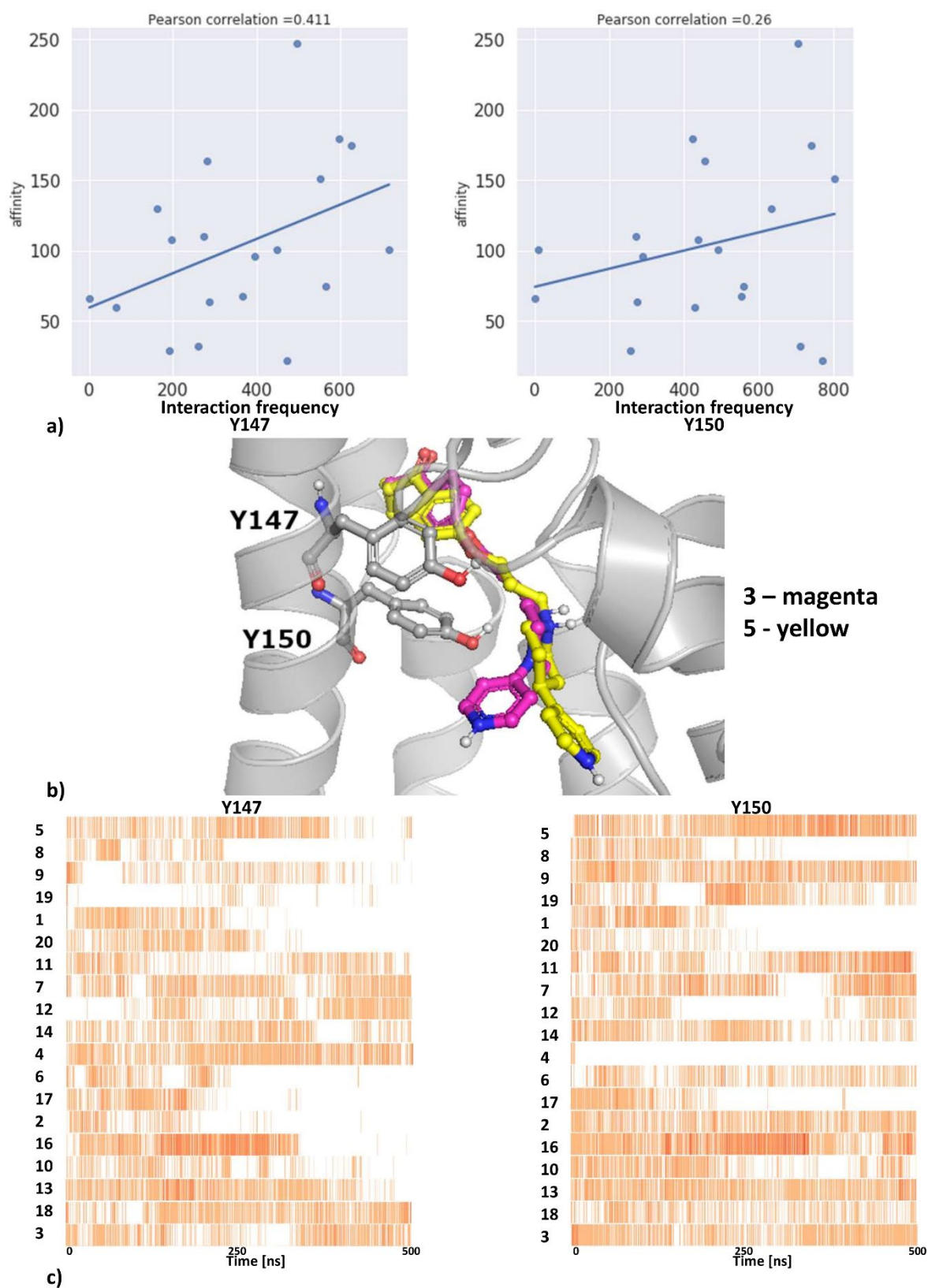


Figure S4. Outcome of correlational studies between frequency of interaction of ligands with particular amino acid residues of σ_2R , a) Pearson correlation coefficients for the highest correlated residues, b) visualization of the highest correlated residues with examples of docked compounds, c) ligand-protein interaction diagrams obtained during md simulations for the highest correlated residues.

6. References

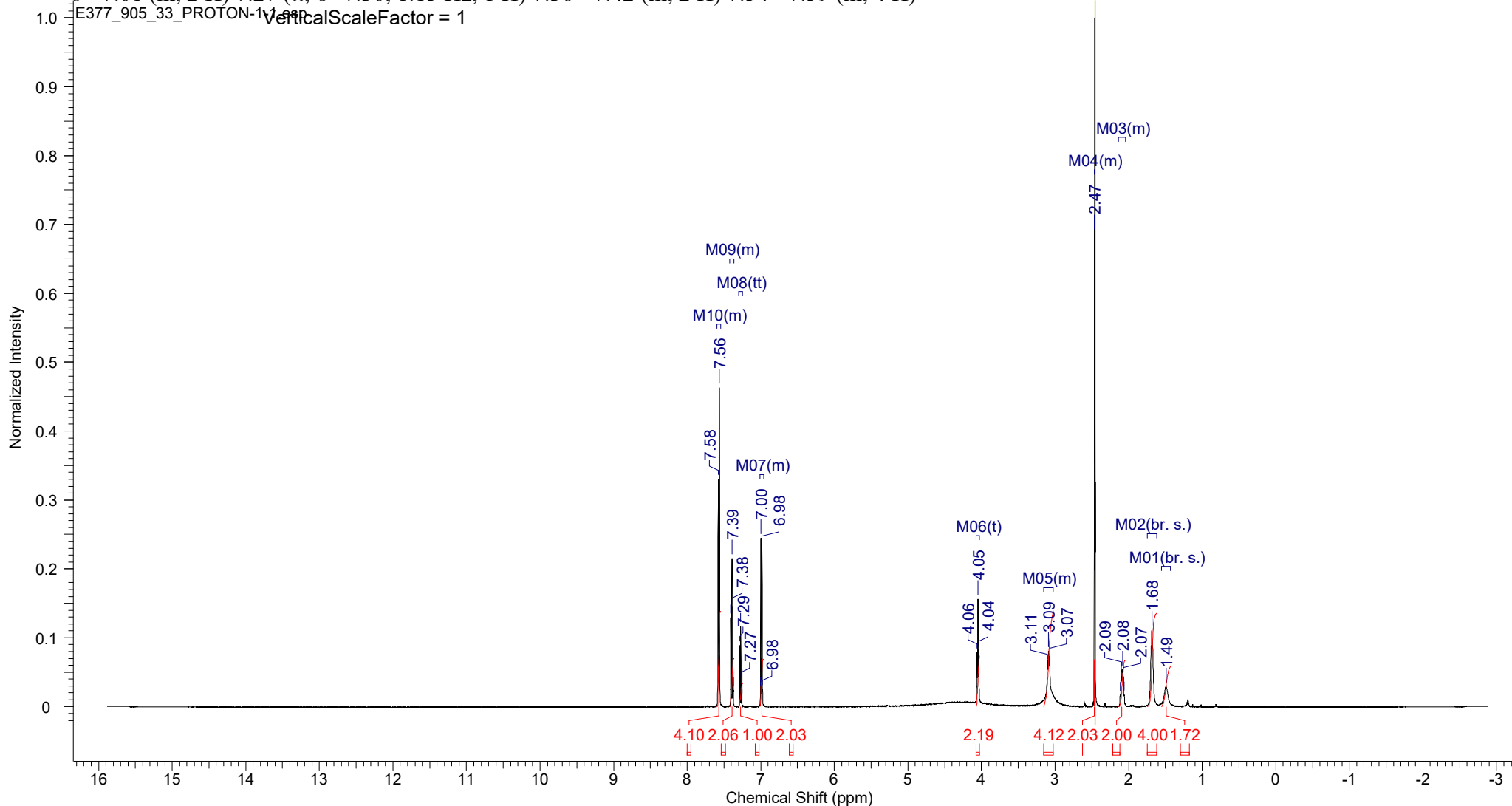
1. Szczepańska K, Karcz T, Mogilski S, Siwek A, Kuder KJ, Latacz G, Kubacka M, Hagenow S, Lubelska A, Olejarz A, Kotańska M, Sadek B, Stark H, Kieć-Kononowicz K. Synthesis and biological activity of novel *tert*-butyl and *tert*-pentylphenoxyalkyl piperazine derivatives as histamine H₃R ligands. *Eur J Med Chem.* 2018; 152: 223-234.
2. Szczepańska K, Karcz T, Kotańska M, Siwek A, Kuder KJ, Latacz G, Mogilski S, Hagenow S, Lubelska A, Sobolewski M, Stark H, Kieć-Kononowicz K. Optimization and preclinical evaluation of novel histamine H₃ receptor ligands: Acetyl and propionyl phenoxyalkyl piperazine derivatives. *Bioorganic Med Chem.* 2018; 26(23-24): 6056-6066.
3. Szczepańska K, Karcz T, Siwek A, Kuder KJ, Latacz G, Bednarski M, Szafarz M, Hagenow S, Lubelska A, Olejarz-Maciej A, Sobolewski M, Mika K, Kotańska M, Stark H, Kieć-Kononowicz K. Structural modifications and *in vitro* pharmacological evaluation of 4-pyridyl-piperazine derivatives as an active and selective histamine H₃ receptor ligands. *Bioorg Chem.* 2019; 103071.
4. Szczepańska K, Pockes S, Podlewska S, Höring C, Mika K, Latacz G, Bednarski M, Siwek A, Karcz T, Nagl M, Bresinsky M, Mönnich D, Seibel U, Kuder KJ, Kotańska M, Stark H, Elz S, Kieć-Kononowicz K. Structural modifications in the distal, regulatory region of histamine H₃ receptor antagonists leading to the identification of a potent anti-obesity agent. *Eur J Med Chem.* 2020; 113041. doi:10.1016/j.ejmech.2020.113041.
5. Kuder K, Łażewska D, Latacz G, Schwed JS, Karcz T, Stark H, Karolak-Wojciechowska J, Kieć-Kononowicz K. Chlorophenoxy aminoalkyl derivatives as histamine H₃R ligands and antiseizure agents. *Bioorganic Med Chem.* 2016, 24(2): 53-72.
6. Bertrand I, Capet M, Lecomte JM, Levoine N, Ligneau X, Poupardin-Olivier O, Robert P, Schwartz JC, Labeeuw O. BIOPROJET. Phenoxypropylpiperidines and -pyrrolidines and their use as histamine H₃ -receptor ligands. WO2006117609 (2006)
7. Levoine N, Labeeuw O, Calmels T, Poupardin-Olivier O, Berrebi-Bertrand I, Lecomte JM, Schwartz JC, Capet M. Novel and highly potent histamine H₃ receptor ligands. Part 1: Withdrawing of hERG activity. *Bioorganic Med Chem Lett.* 2011, 21(18): 5378-5383.
8. Baell JB, Holloway GA. New substructure filters for removal of pan assay interference compounds (PAINS) from screening libraries and for their exclusion in bioassays. *J Med Chem.* 2010, 53(7): 2719-2740.

7. ^1H - and ^{13}C -NMR spectra of compounds 11 and 12. HPLC traces of all the final compounds

Cpd11

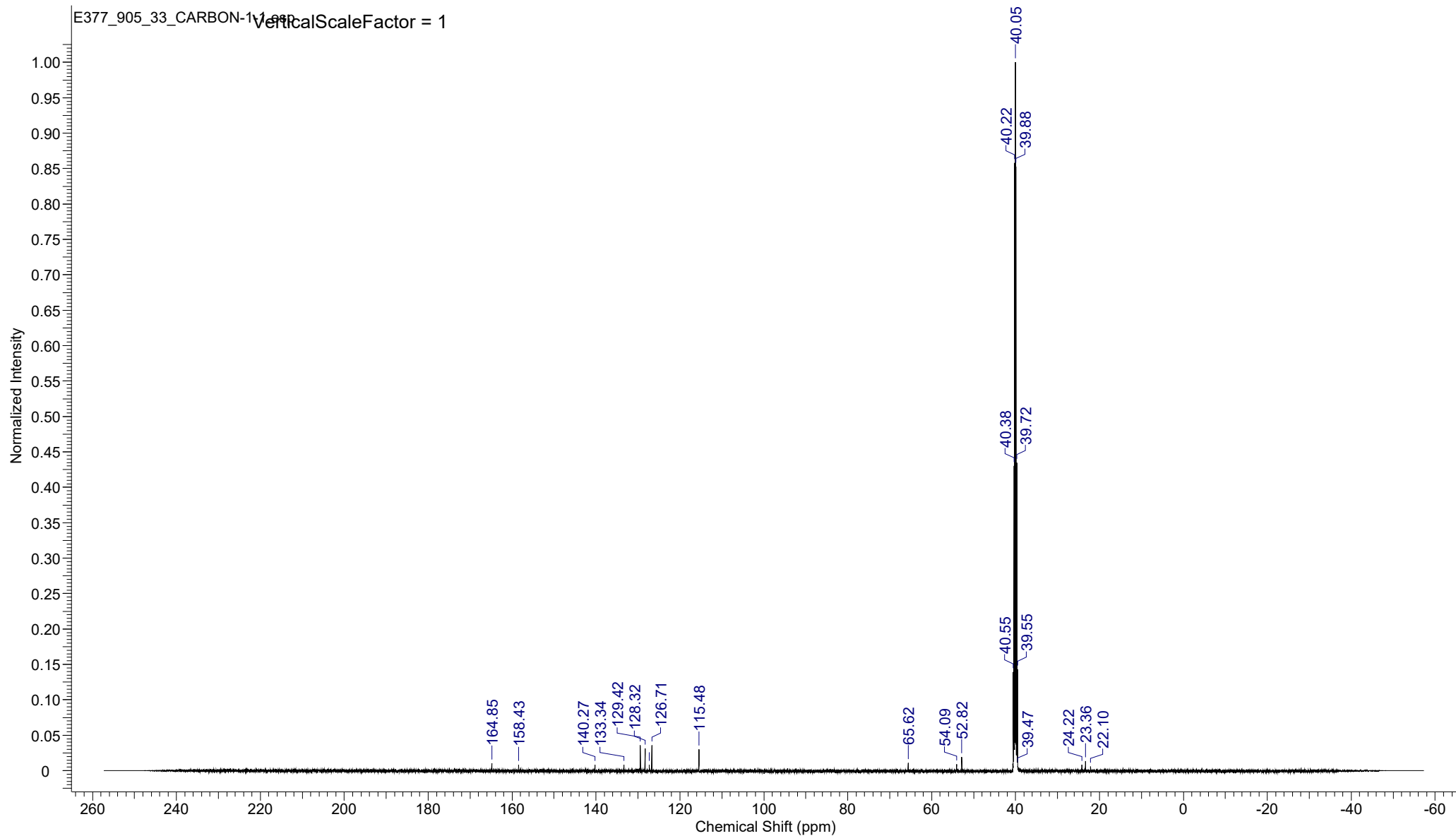
Acquisition Time (sec)	3.4918	Date	30 Oct 2020 10:57:56	Date Stamp	30 Oct 2020 10:57:08		
File Name	C:\Users\turbo\Desktop\E377_905_33_PROTON-1-1.jdf		Frequency (MHz)	500.16	Nucleus	1H	
Origin	ECA	Original Points Count	32768	Owner	delta	Number of Transients	8
Solvent	DMSO-d6	Spectrum Offset (Hz)	3251.0396	Sweep Width (Hz)	9384.38	Pulse Sequence	proton.jxp
				Points Count	32768	Temperature (degree C)	22.500

¹H NMR (500 MHz, DMSO-d₆) δ ppm 1.49 (br. s., 2 H) 1.68 (br. s., 4 H) 2.04 - 2.14 (m, 2 H) 2.46 - 2.47 (m, 2 H) 3.02 - 3.16 (m, 4 H) 4.05 (t, *J*=6.01 Hz, 2 H) 6.96 - 7.01 (m, 2 H) 7.27 (tt, *J*=7.30, 1.15 Hz, 1 H) 7.36 - 7.42 (m, 2 H) 7.54 - 7.59 (m, 4 H)



Cpd11

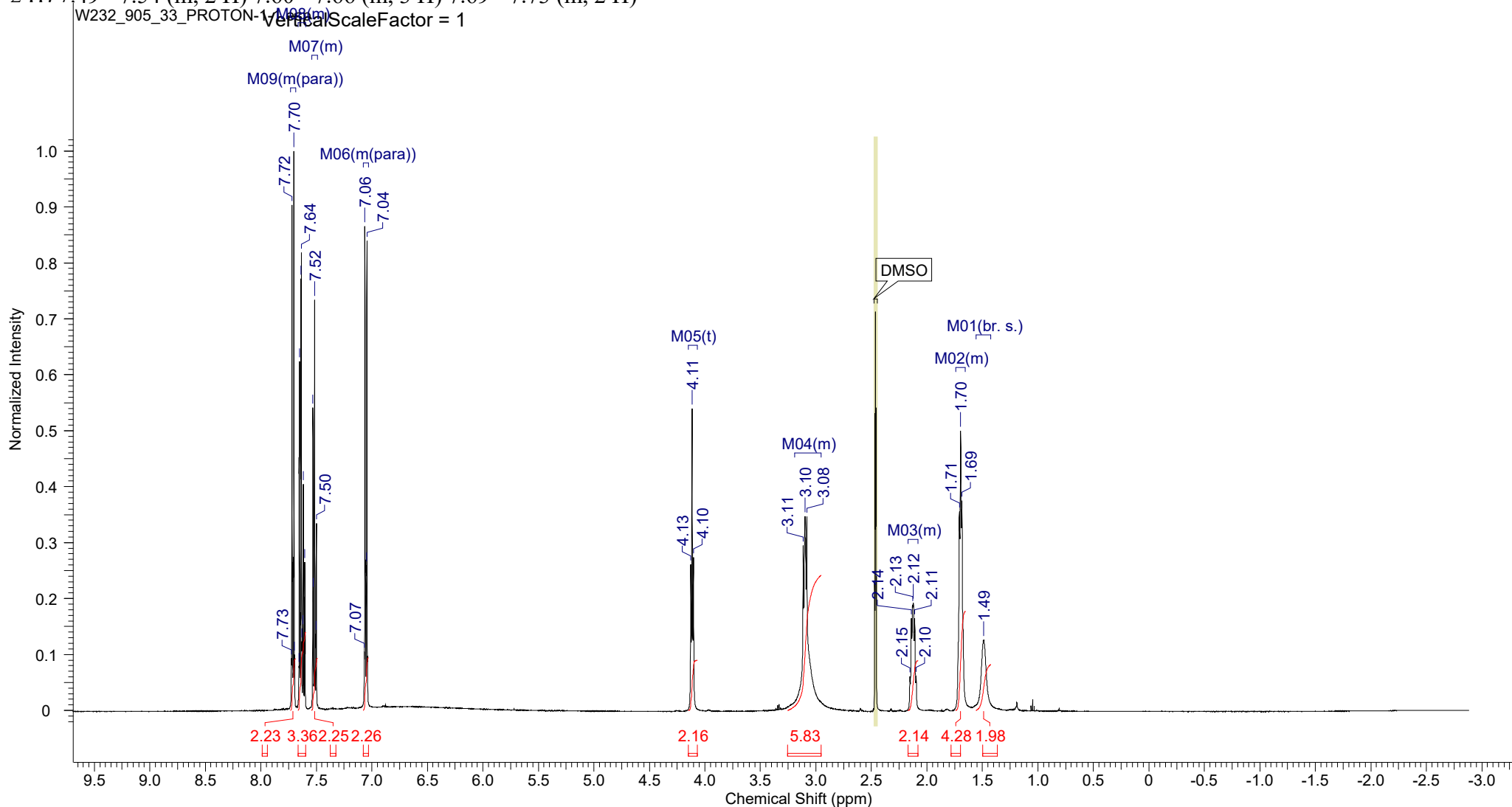
Acquisition Time (sec)	1.6568	Date	30 Oct 2020 11:50:08	Date Stamp	30 Oct 2020 10:58:27				
File Name	C:\Users\turbo\Desktop\E377_905_33 CARBON-1-1.jdf	Frequency (MHz)	125.77	Nucleus	13C	Number of Transients	1024		
Origin	ECA	Original Points Count	65536	Owner	delta	Points Count	65536	Pulse Sequence	carbon.jxp
Solvent	DMSO-d6	Spectrum Offset (Hz)	12576.5293	Sweep Width (Hz)	39556.96	Temperature (degree C)	22.500		



Cpd12

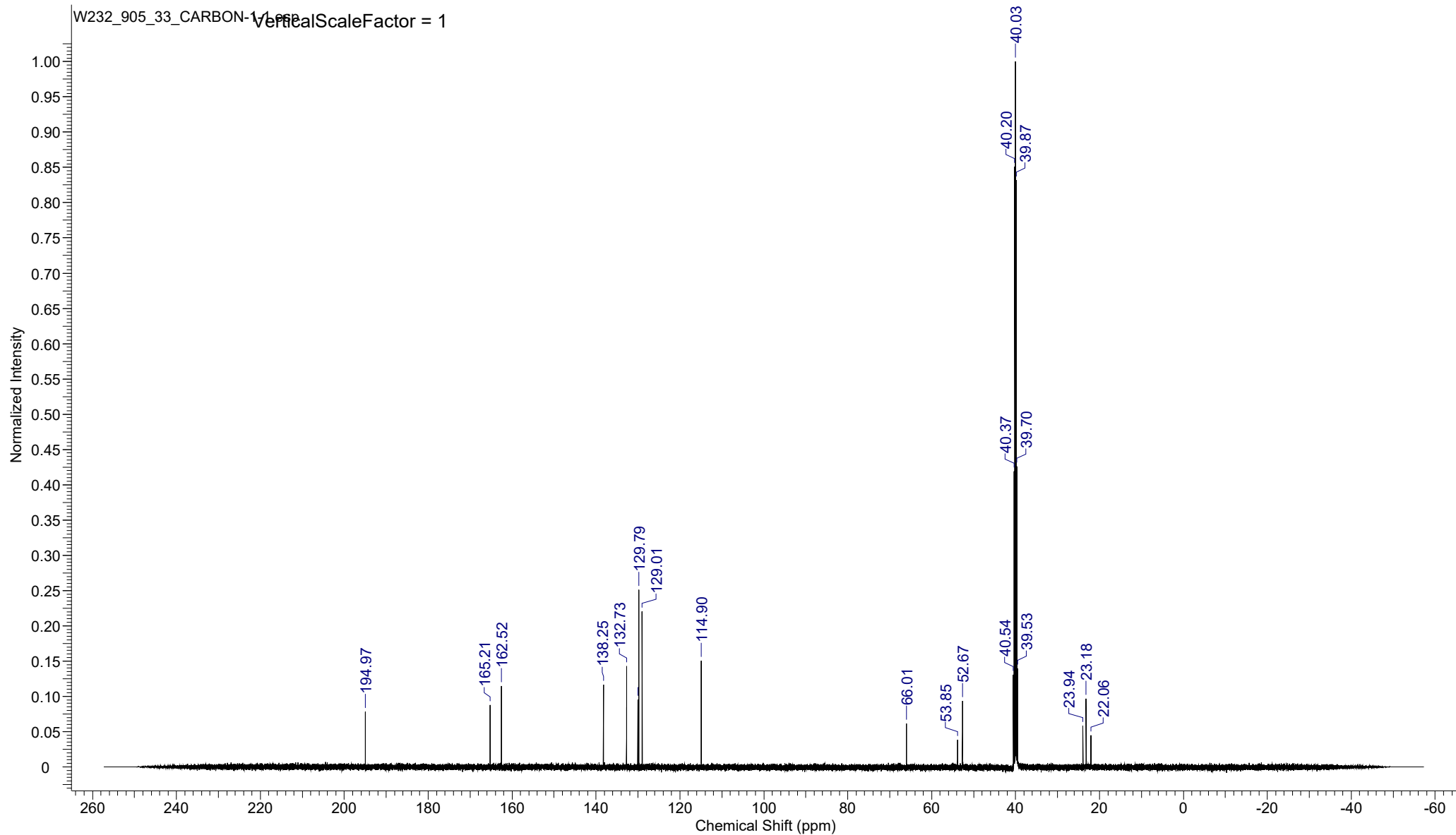
Acquisition Time (sec)	3.4918	Date	24 Jun 2021 08:39:15	Date Stamp	24 Jun 2021 08:38:27
File Name	C:\Users\turbo\Desktop\W232_905_33_PROTON-1-1.jdf			Frequency (MHz)	500.16
Number of Transients	8	Origin	ECA	Original Points Count	32768
Pulse Sequence	proton.jxp	Solvent	DMSO-d6	Spectrum Offset (Hz)	3251.0396
				Sweep Width (Hz)	9384.38
				Temperature (degree C)	21.000

^1H NMR (500 MHz, DMSO- d_6) δ ppm 1.49 (br. s., 2 H) 1.65 - 1.74 (m, 4 H) 2.08 - 2.17 (m, 2 H) 2.95 - 3.19 (m, 6 H) 4.11 (t, $J=6.16$ Hz, 2 H) 7.03 - 7.08 (m, 2 H) 7.49 - 7.54 (m, 2 H) 7.60 - 7.66 (m, 3 H) 7.69 - 7.73 (m, 2 H)

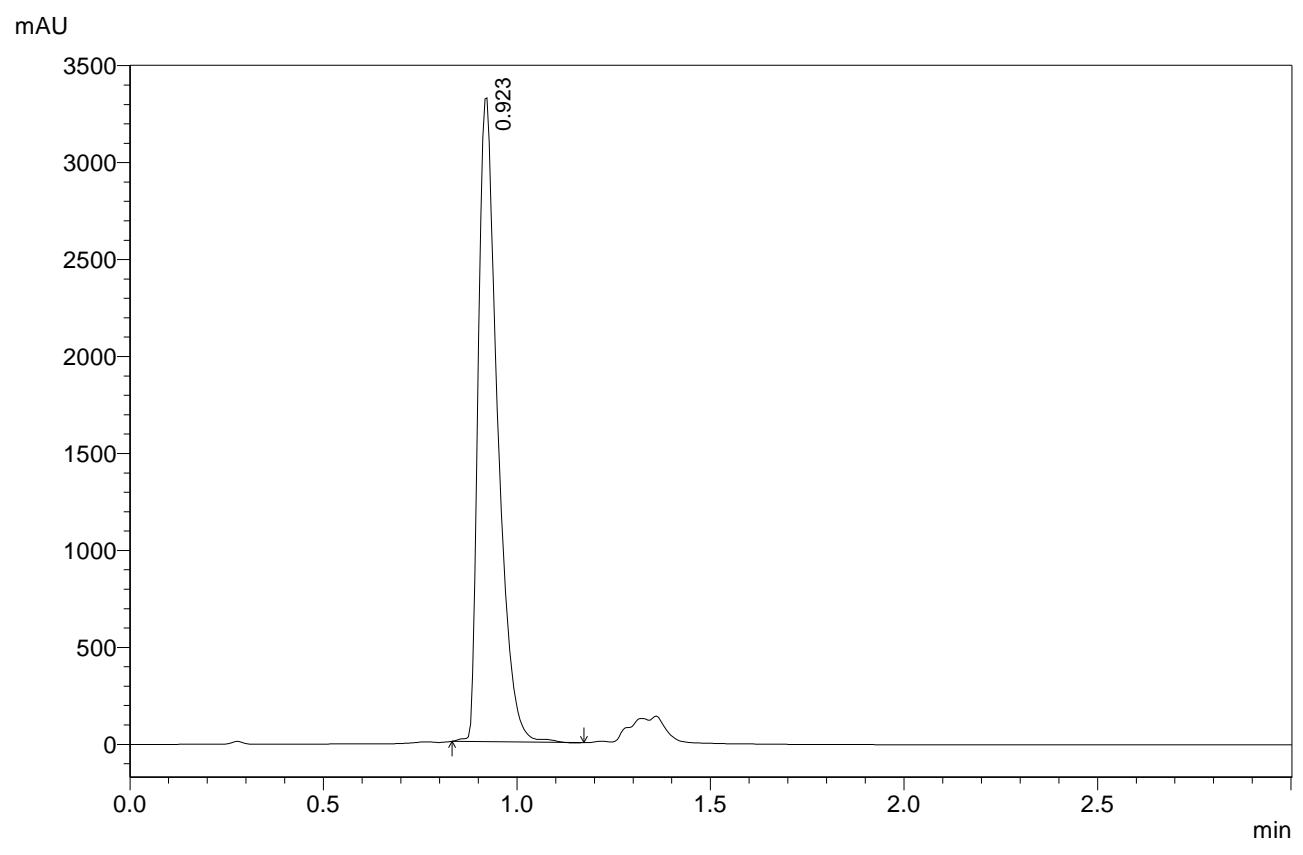


Cpd12

Acquisition Time (sec)	1.6568	Date	24 Jun 2021 09:31:35	Date Stamp	24 Jun 2021 08:39:46		
File Name	C:\Users\turbo\Desktop\W232_905_33 CARBON-1-1.jdf	Frequency (MHz)	125.77	Nucleus	13C	Number of Transients	1024
Origin	ECA	Original Points Count	65536	Owner	delta	Points Count	65536
Solvent	DMSO-d6	Spectrum Offset (Hz)	12576.5293	Sweep Width (Hz)	39556.96	Temperature (degree C)	21.000



Cpd1



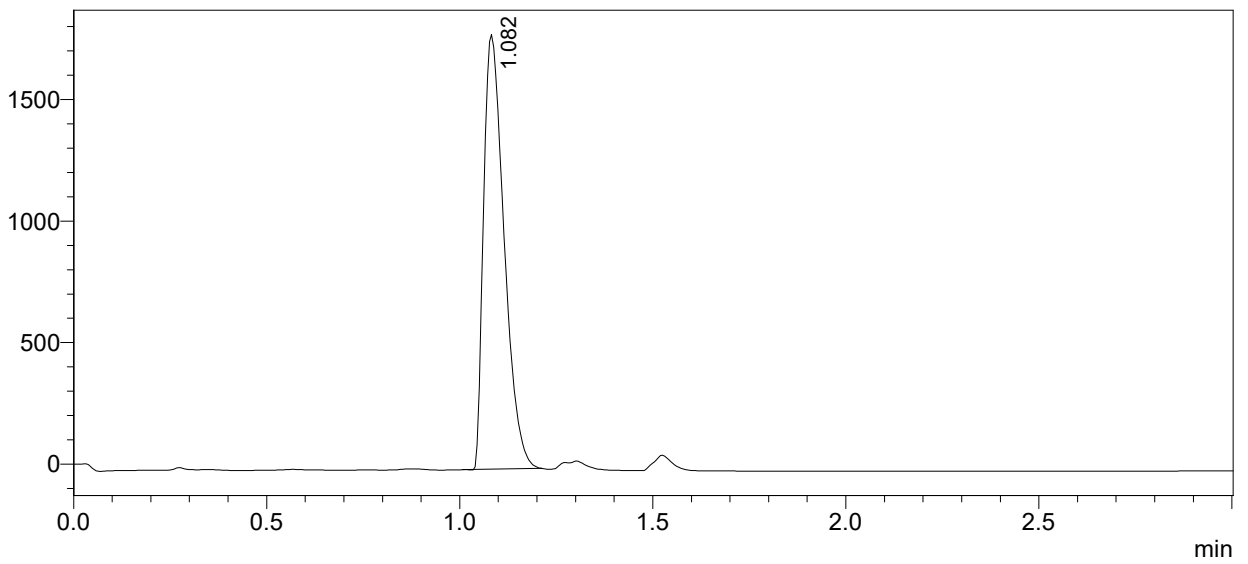


SHIMADZU
LabSolutions

Cpd2

<Chromatogram>

mAU



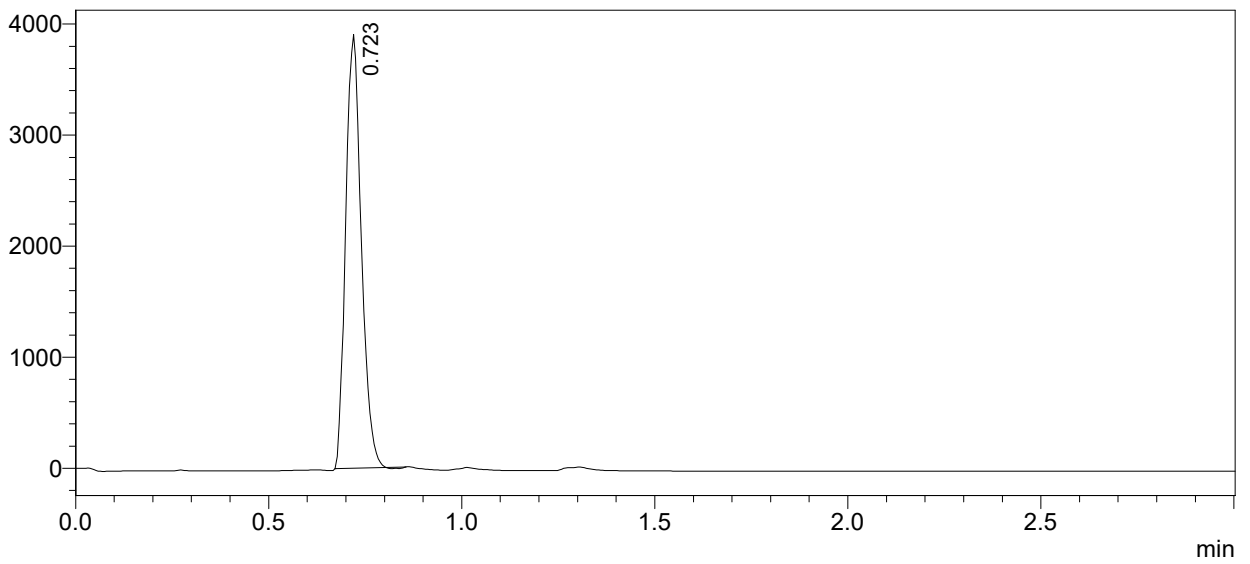


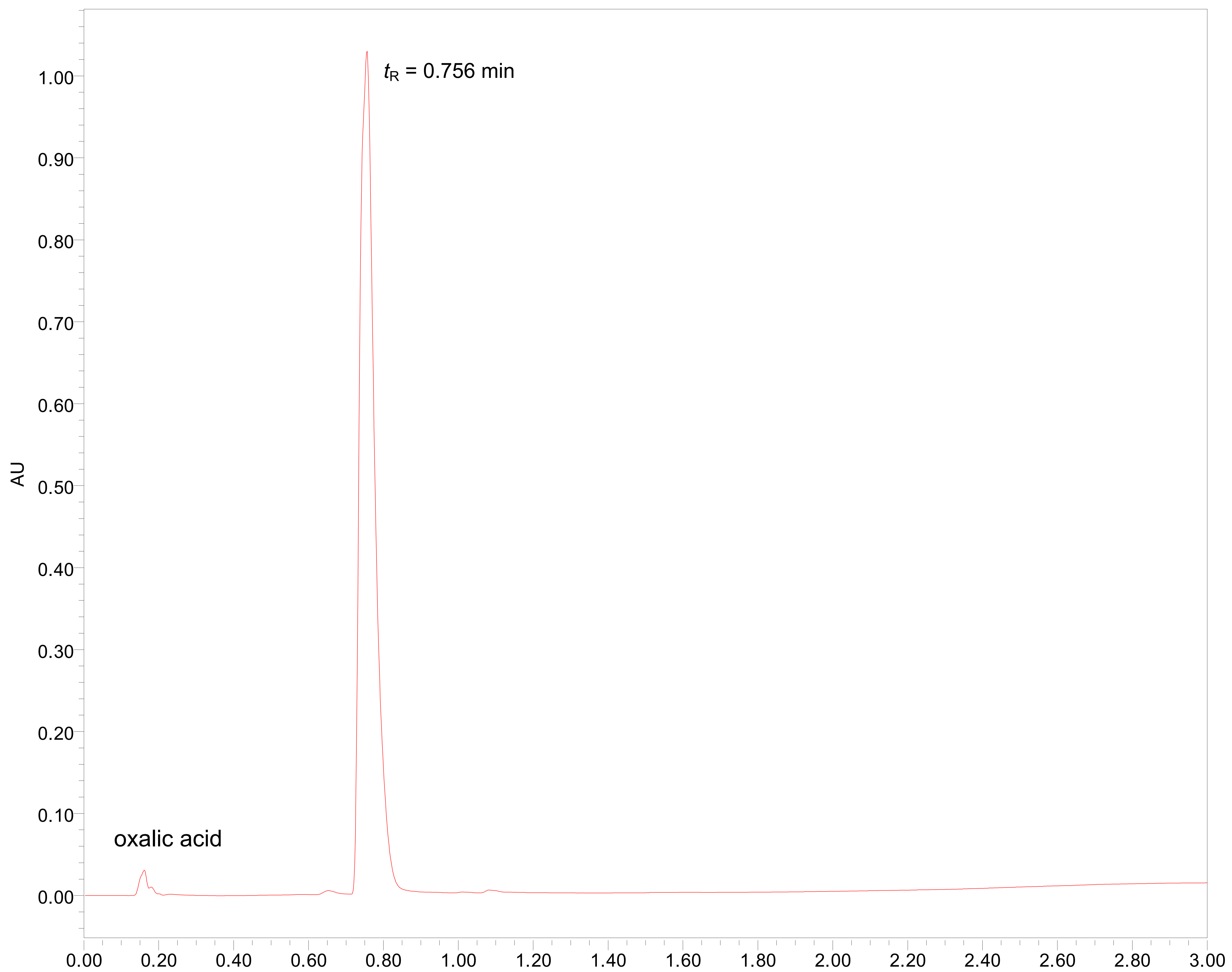
SHIMADZU
LabSolutions

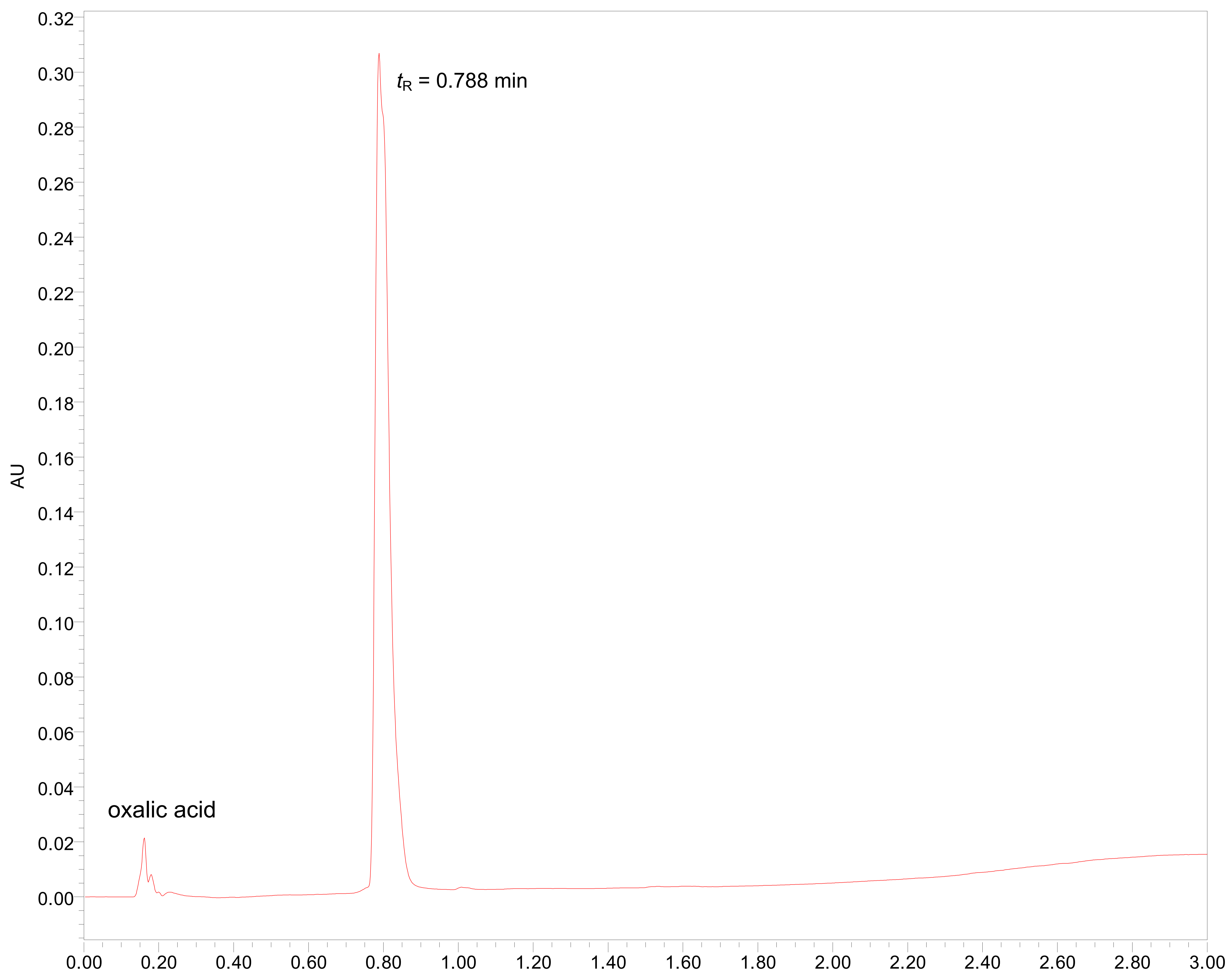
Cpd3

<Chromatogram>

mAU







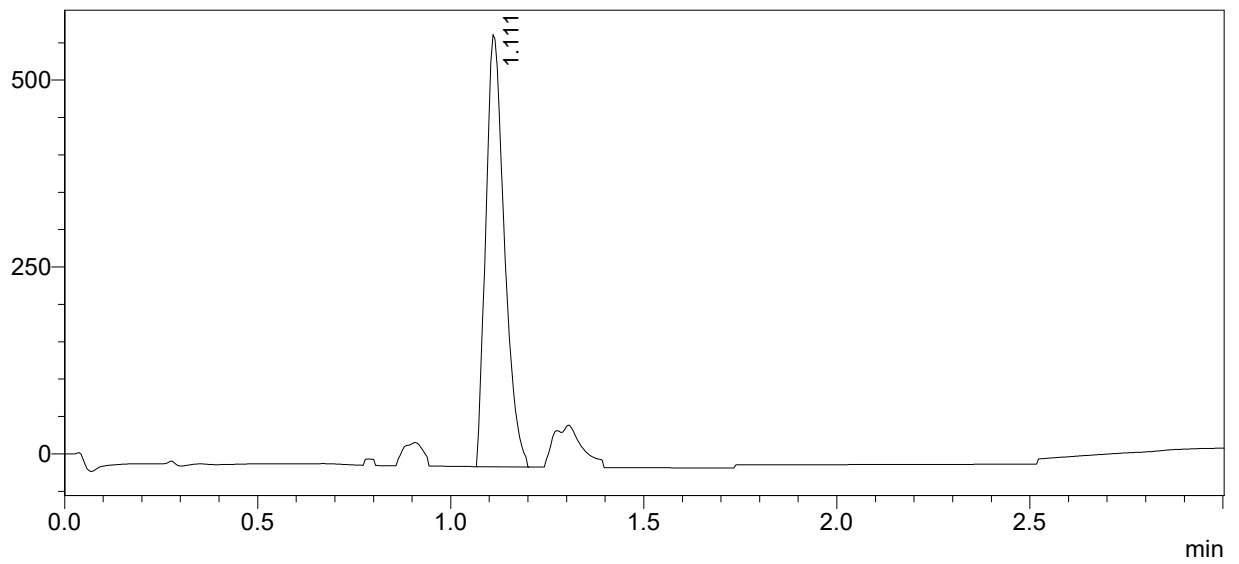


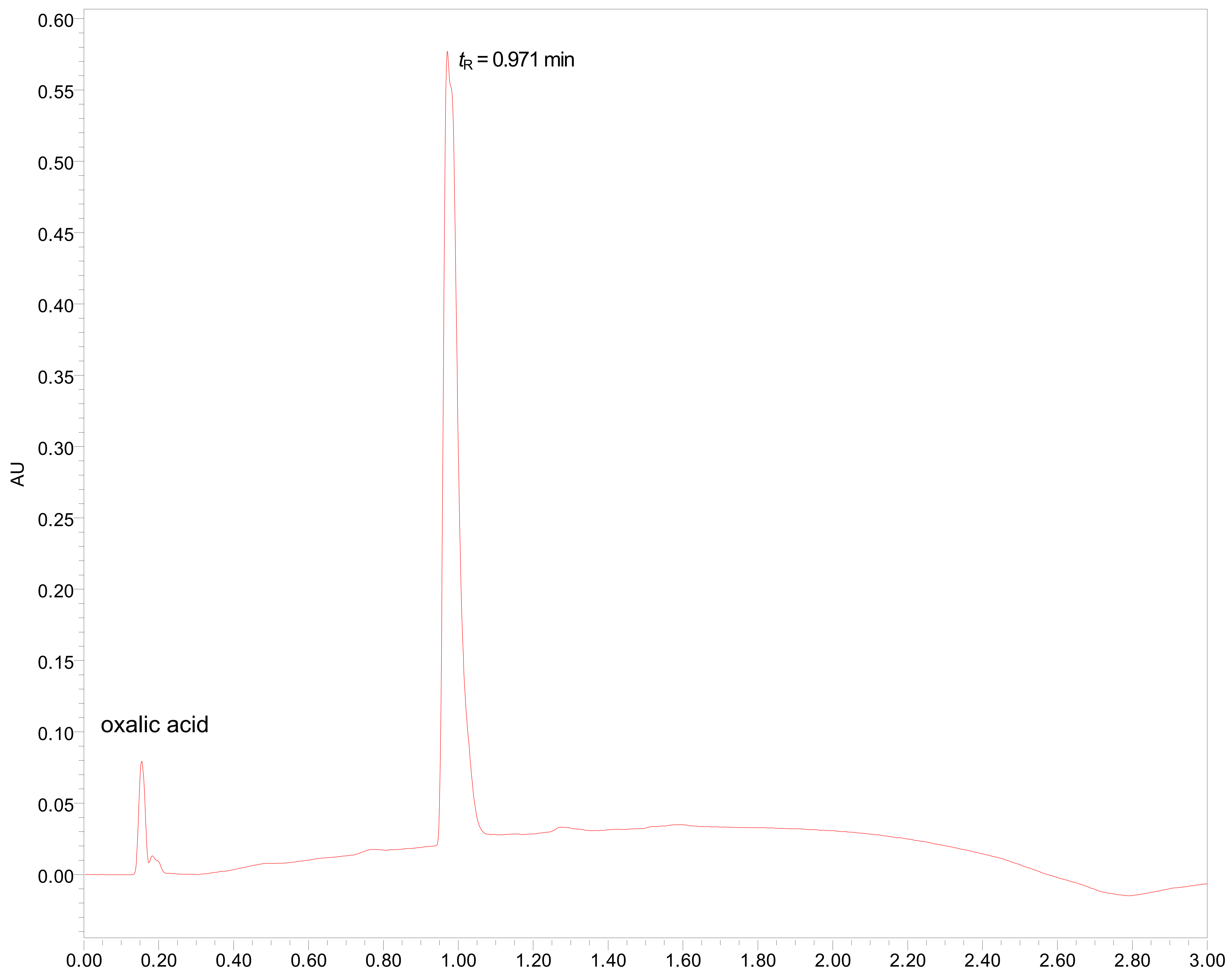
SHIMADZU
LabSolutions

Cpd6

<Chromatogram>

mAU





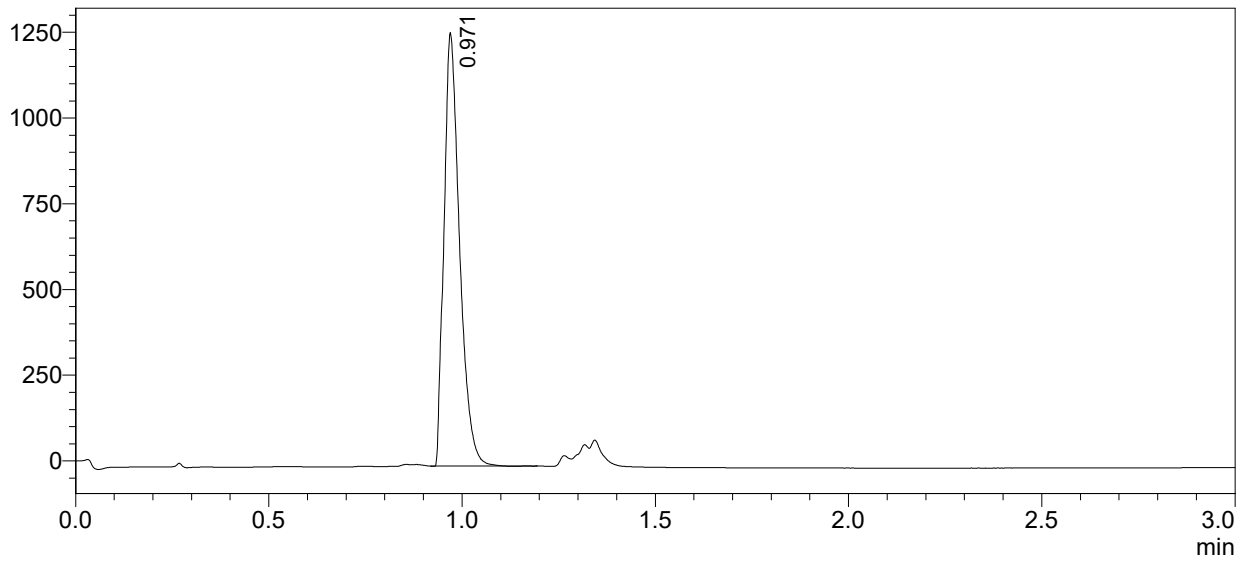


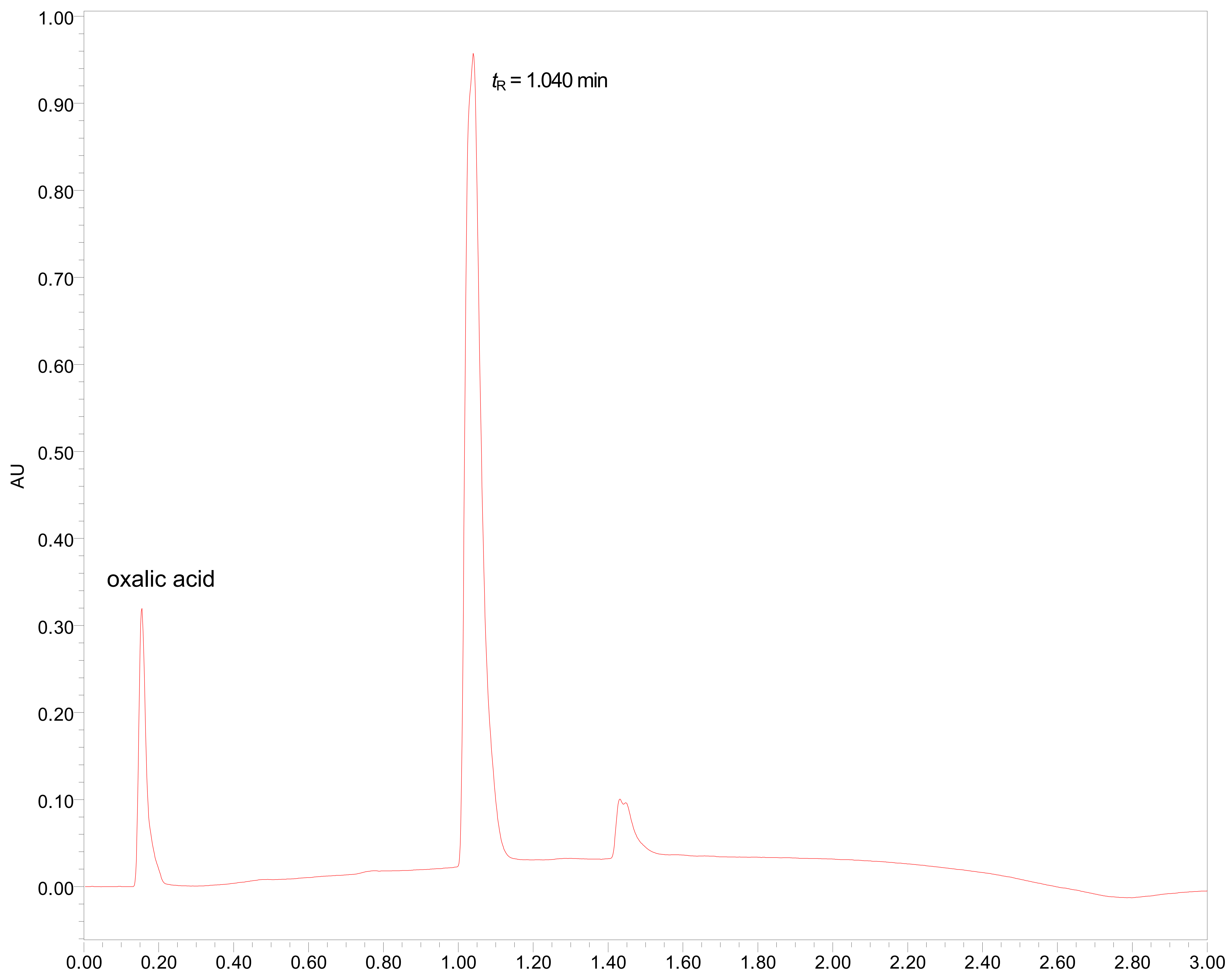
SHIMADZU
LabSolutions

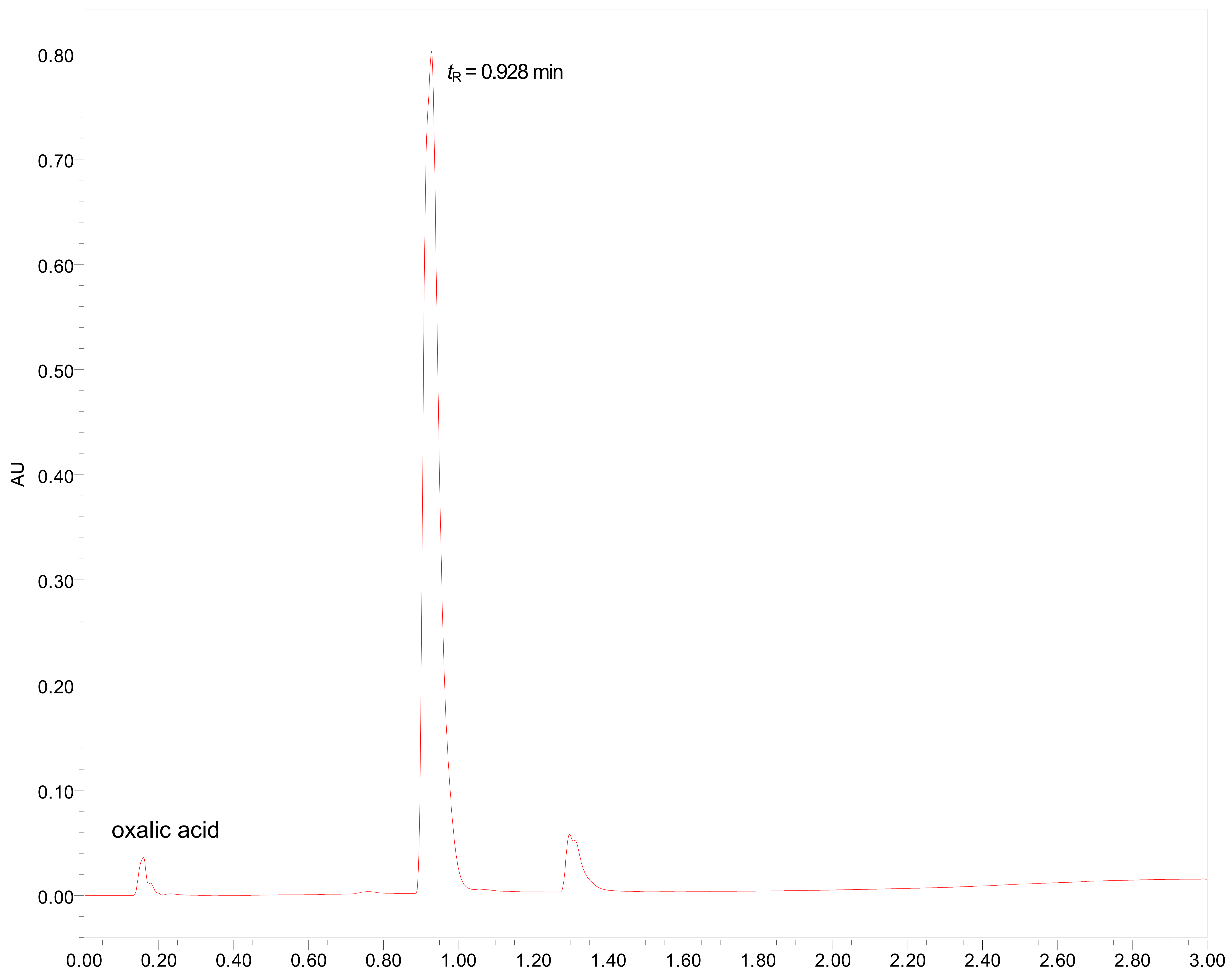
Cpd8

<Chromatogram>

mAU







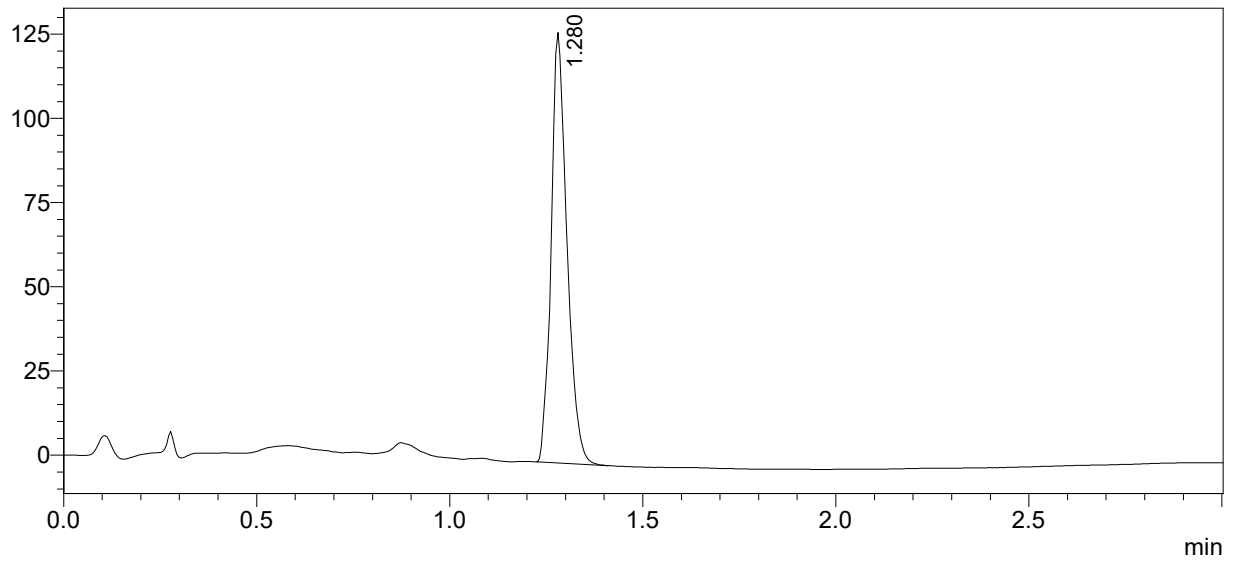


SHIMADZU
LabSolutions

Cpd11

<Chromatogram>

mAU



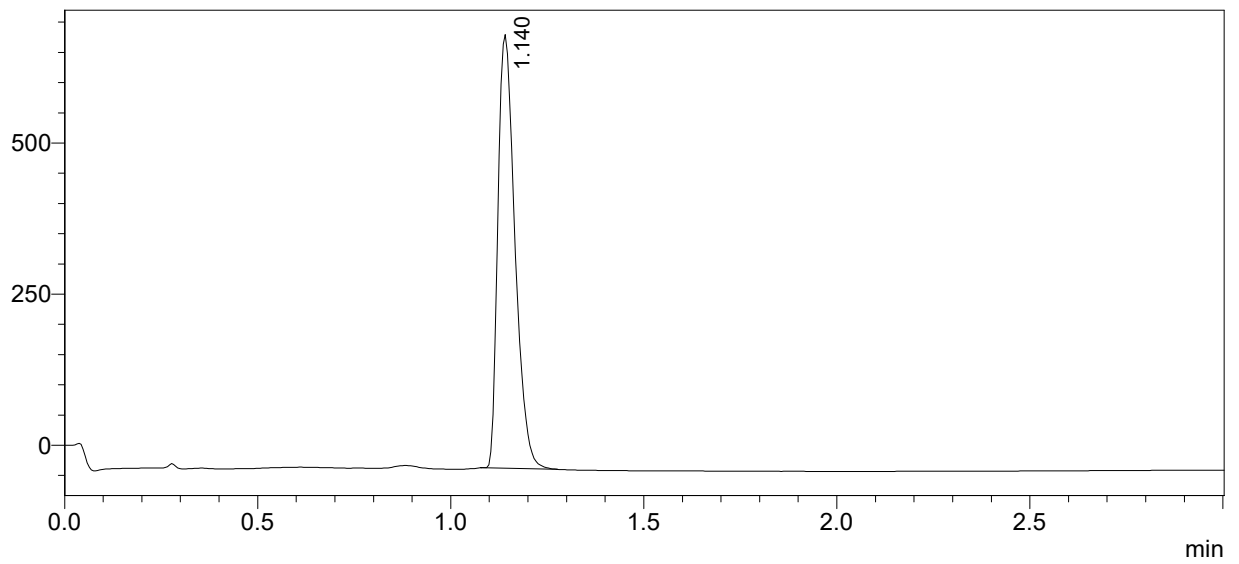


SHIMADZU
LabSolutions

Cpd12

<Chromatogram>

mAU



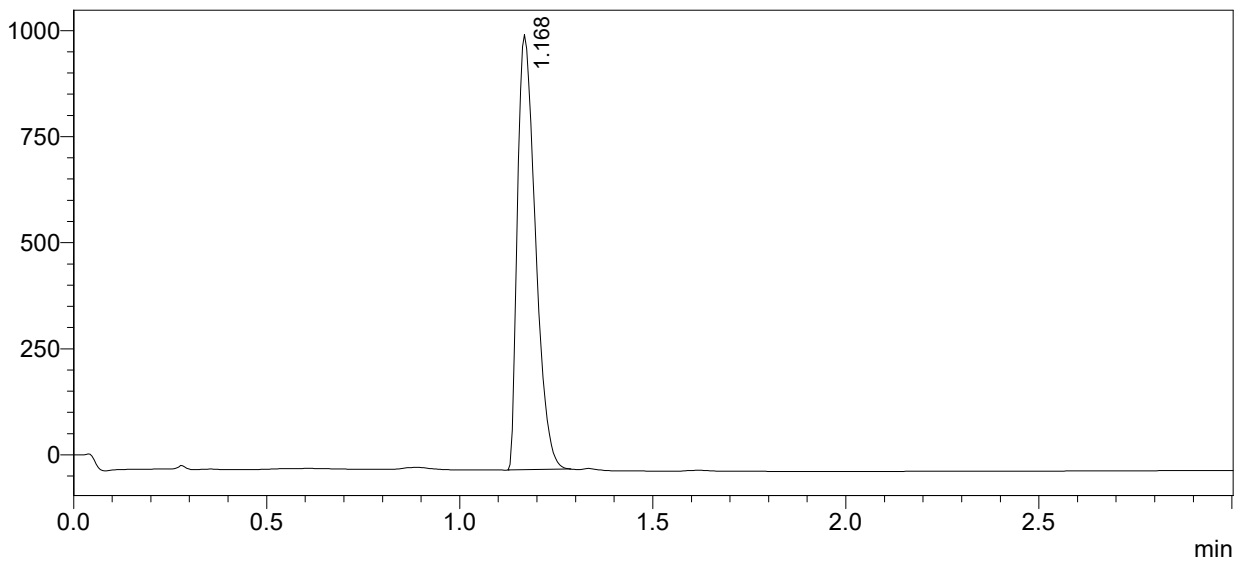


SHIMADZU
LabSolutions

Cpd13

<Chromatogram>

mAU



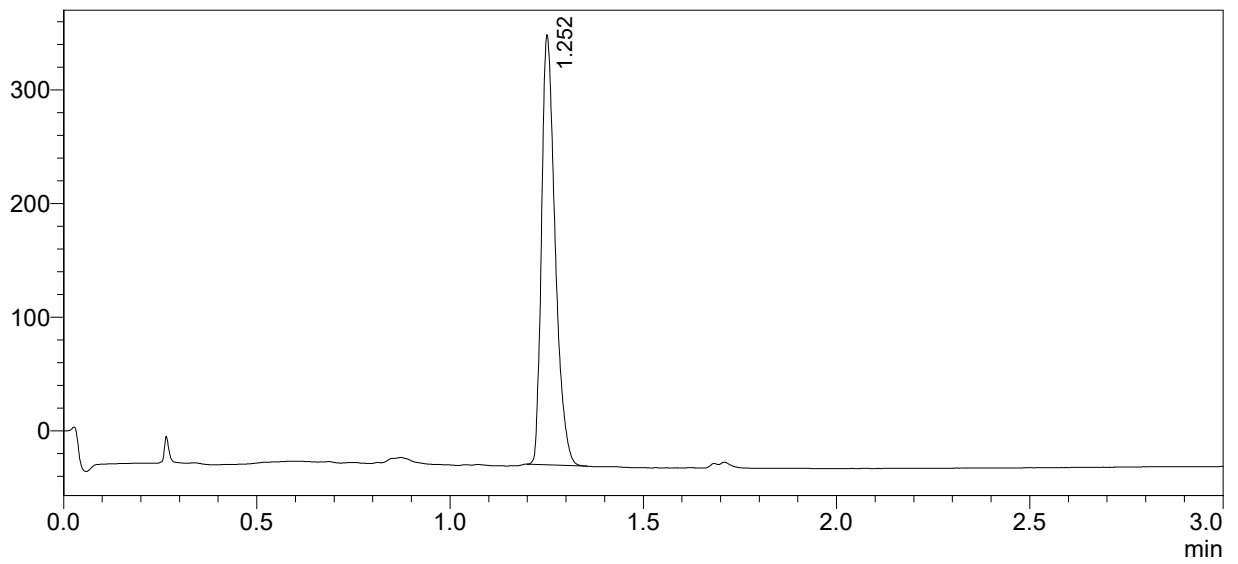


SHIMADZU
LabSolutions

Cpd14

<Chromatogram>

mAU



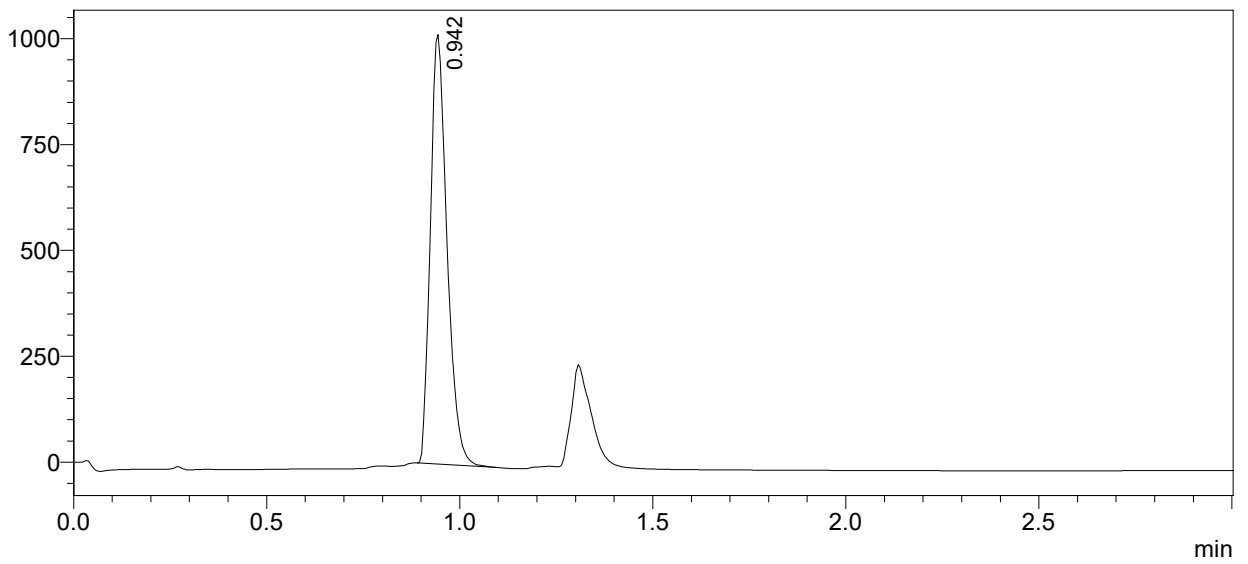


SHIMADZU
LabSolutions

Cpd16

<Chromatogram>

mAU



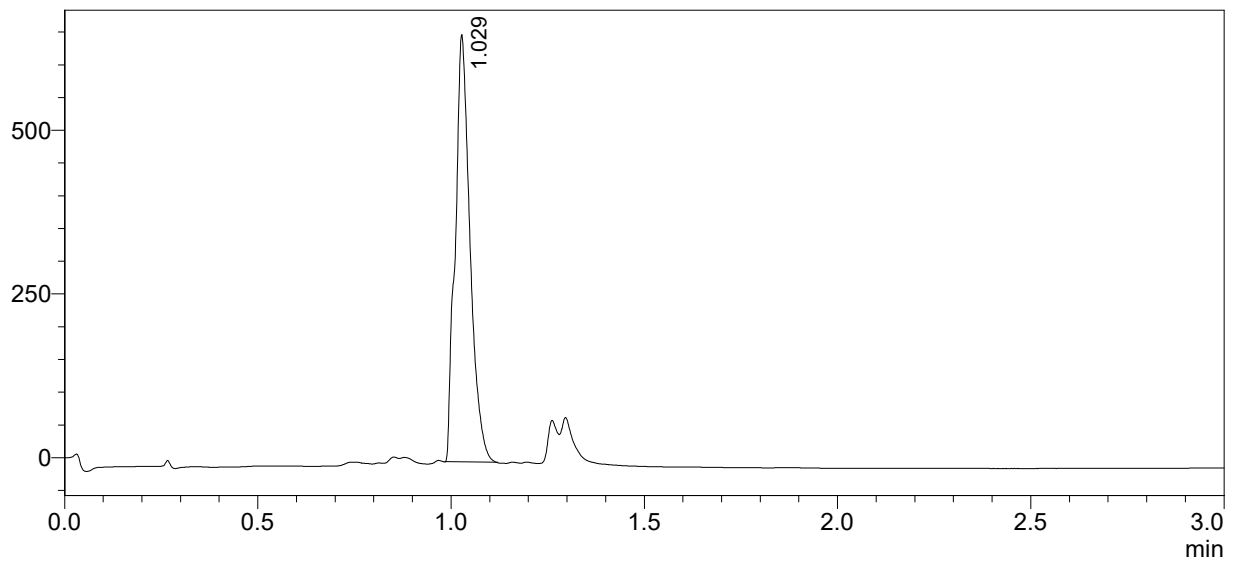


SHIMADZU
LabSolutions

Cpd17

<Chromatogram>

mAU



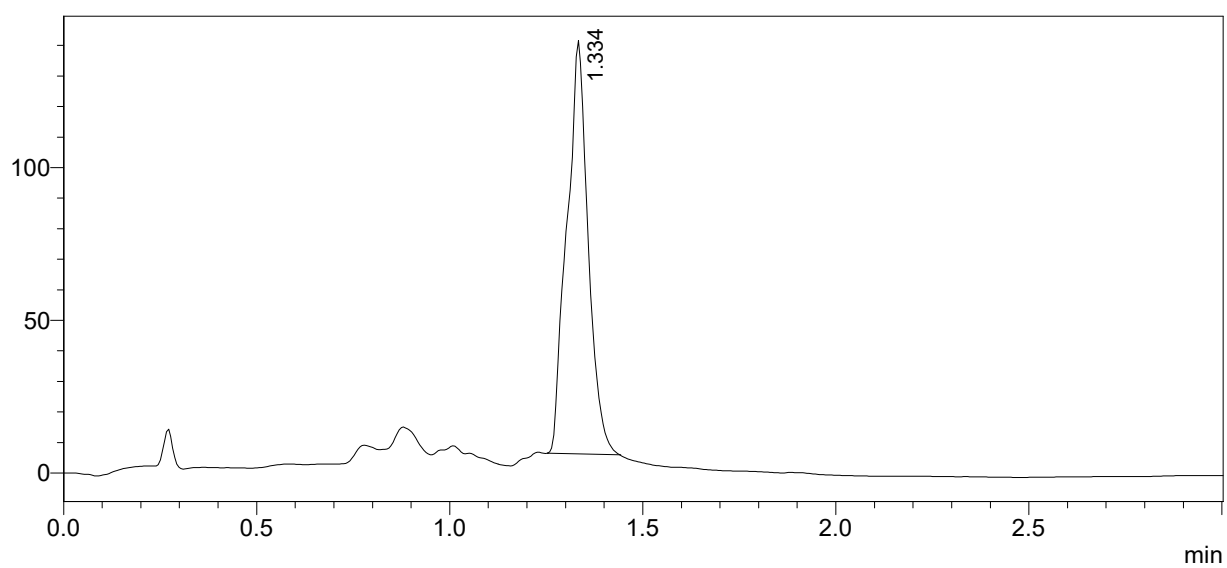


SHIMADZU
LabSolutions

Cpd18

<Chromatogram>

mAU



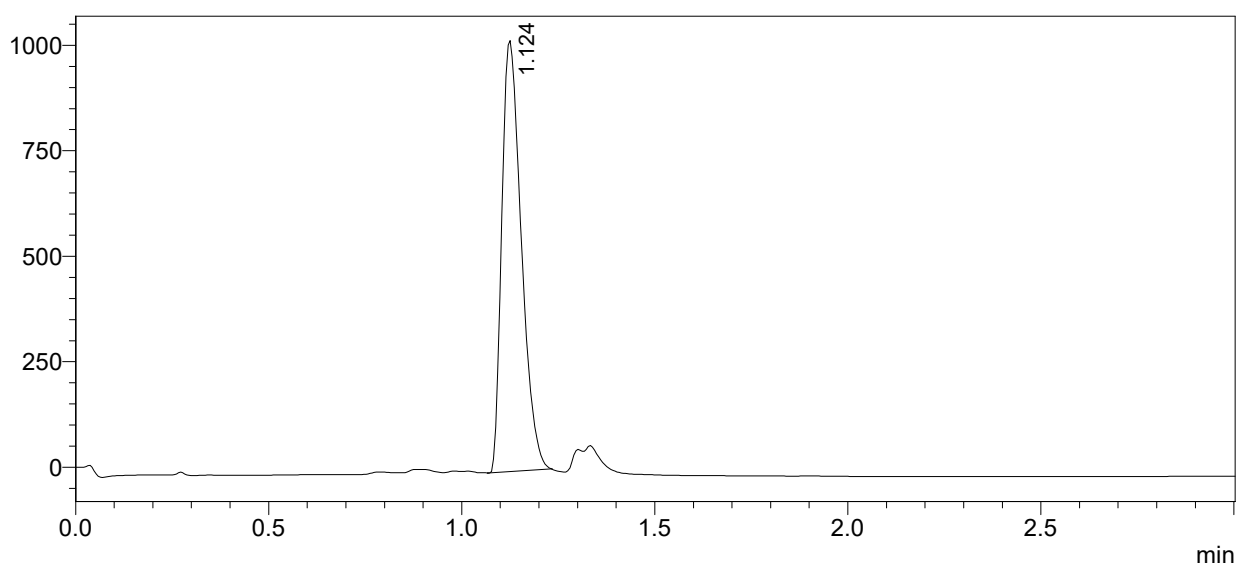


SHIMADZU
LabSolutions

Cpd19

<Chromatogram>

mAU





SHIMADZU
LabSolutions

Cpd20

<Chromatogram>

mAU

