Yeast-Derived Nanoparticles Remodel the Immunosuppressive Microenvironment in Tumor and Tumor-draining Lymph Nodes to Suppress Tumor Growth

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Antibodies	Manufacturer	Catalog Number	Dilutions	
FITC anti-mouse CD3	Biolegend	100204	1:200	
APC anti-mouse CD4	Biolegend	100412	1:200	
PE anti-mouse CD8a	Biolegend	100708	1:200	
FITC anti-mouse CD11c	Biolegend	117306	1:200	
APC anti-mouse Ly-6G/Ly-6C (Gr-1)	Biolegend	108412	1:200	
FITC anti-mouse F4/80	Biolegend	123108	1:200	
PE anti-mouse CD206 (MMR)	Biolegend	141706	1:200	
FITC anti-mouse CD45	Biolegend	103108	1:200	
PerCP anti-mouse/human CD11b	Biolegend	101230	1:200	
PE anti-mouse FOXP3	Biolegend	126403	1:200	
APC anti-mouse CD80	Biolegend	104714	1:200	
PE anti-mouse CD86	Biolegend	105008	1:200	
PE anti-mouse CD274 (B7-H1, PD-L1)	Biolegend	124308	1:200	
APC anti-mouse CD279 (PD- 1)	Biolegend	135209	1:200	
PE anti-mouse I-A/I-E	Biolegend 107608		1:200	
APC anti-mouse CD40	Biolegend	124612	1:200	
APC anti-mouse CD69	Biolegend	104514	1:200	
PE anti-mouse CD19	Biolegend	15508	1:200	
Anti-CD4	BioXcell	BE0004-1; Clone: 53-6.7	Diluted to 100 g/mL	
Anti-CD8a	BioXcell	BE0003-1;	Diluted to 100 g/mL	
		Clone: GK1.5		
Anti-PD-L1	BioXcell	BE0101;	Diluted to 100 g/mL	
		Clone: 10F. 9G2;		
Anti n D65	Abalanal	Lot: 751219D1	1.1000	
Anu-p-Pos		AP0475	1:1000	
Anti-GAPDH	Servicebio	GB11002	1:1000	
Anti-TLR-2	Abcam	ab209216	1:1000	
Anti-p-Syk	Cell Signaling Technology	#2710	1:1000	
Anti-MyD88	Abcam	ab219413	1:1000	
Anti-Dectin-1	Absin	abs124190	1:1000	
Goat anti-Rabbit IgG (H+L)- HRP	Ray Antibody	RM3002	1:5000	

Supplementary Table 1. Materials used in study.

Yeast			
Yeast from	Sigma-aldrich	ysc2-500g	
Saccharomyces Cerevisiae			
Inhibitor			
Laminarin	Sigma-aldrich	9008-22-4	
C29	Cayman	27029	



Supplementary Fig. 1. The composition of YCW NPs. (A) The composition of YCW nanoparticles.(B) Monosaccharide standard high performance liquid chromatography (HPLC) curve. (C) Monosaccharide curve of YCW NPs.

Protein Group	Protein D	A ccession	-101gP	PTM	Avg. M ass	Description
23	15	P60010 ACT_YEAST	156.24		41690	Actin 0 S=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=ACT1 PE=1 SV=1
20	24	P07251 ATPA_YEAST	139.36	0 xidation (HW)	58608	ATP synthase subunitabha m itochondria10 S=Sacchanom yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=ATP1 PE=1 SV=5
12	42	P02992 EFTU_YEA ST	116.47	A cetylation (N-term); O xidation (HW)	47972	Ebngation factor Tu m inchondria10 S=Saccharom yces cenevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=TUF1 PE=1 SV=1
30	50	P05030 PM A1_YEAST	110.87	Carban ilon ethylation; 0 xilation (HW)	99619	Plasm a m em brane ATPase 1 0 S=Saccharon yces cerevixiae (strain ATCC 204508 / S288c) 0 X=559292 G X=PM A1 PE=1 SV=2
35	105	P00925 EN 0 2_YEA ST	101.23		46914	Eno lase 2 O S=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) O X=559292 G N=EN O 2 PE=1 SV=2
35	113	P00924 EN 0 1_YEA ST	101.23		46816	Eno lase 1 0 S=Saccharom yoes cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=EN 0 1 PE=1 SV=3
31	66	POO331 ADH 2_YEAST	96.07	Carban ilon ethylation; Acetylation (N-tem)	36732	A boholdehydrogenase 2 0 S=Saccharon yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=ADH2 PE=1 SV=3
31	36	POO330 ADH 1_YEAST	96.07	Carban idom ethylation;Acetylation (N-term);O xidation (HW)	36849	A koholdehydrogenase 1 0 S=Saccharon yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=ADH1PE=1 SV=5
37	252	POO830 A TPB_YEA ST	95.58		54794	ATP synthase subunit beta m inchondrialOS=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) OX=559292 GN=ATP2 PE=1 SV=2
38	160	P39954 SAHH_YEAST	89.76	Carban idom ethylation	49126	A denosyhom ocysteinase OS=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) OX=559292 GN=SAH1 PE=1 SV=1
32	168	P02994 EF1A_YEA ST	88.56		50033	Ebngation factor I-abha 0 S=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=TEF2 PE=1 SV=1
40	99	P02309 H 4_YEA ST	87.2		11368	H istone H 4 0 S=Saccharon yces cerevisiae (stuain ATCC 204508 / S288c) 0 X=559292 G N=HHF2 PE=1 SV=2
34	138	P00549 KPYK1_YEAST	84.34	Carban idon ethylation	54545	Pynuvate kinase 1 0 S=Saccharom yces cerevisie (stmin ATCC 204508 / S288c) 0 X=559292 GN=CDC19 PE=1 SV=2
36	98	P00560 PGK_YEA ST	83.01		44738	Phosphog prenate kinase 0 S=Saccharon yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=PGK1 PE=1 SV=2
41	1031	Q 12692 H 2AZ_YEAST	71.6		14283	H istone H2A Z 0 S=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 G N=HTZ1 PE=1 SV=3
56	335	P05750 RS3_YEAST	58.02		26503	40S ribosom alprotein S3 0 S=Saccharom yces cenevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN =RPS3 PE=1 SV=5
48	204	P39708 DHE5_YEAST	52.46	0 xidation (HW)	49627	NADP-specific glutam ate dehydrogenase 2 0 S=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=GDH3 PE=3 SV=1
48	293	P07262 DHE4_YEAST	52.46		49570	NADP-specific glutam ate dehydrogenase 1 0 S=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=GDH1 PE=1 SV=2
53	2043	P25515 VATL1_YEAST	51.44	Carban idon ethylation	16351	V-type proton A TPase subunit c 0 S=Saccharom yces cerevisiae (strain A TCC 204508 / S288c) 0 X=559292 GN=VM A 3 PE=1 SV=1
60	6467	Q 08692 0 SW 1_YEA ST	50.67		32758	0 uter spore wallprotein 1 0 S=Saccharon yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN = 0 SW 1 PE=4 SV=1
44	278	P22202 HSP74_YEAST	48.58	A cetylation (N-term)	69651	Heat shock protein SSA4 0 S=Saccharon yces cerevisie (stain ATCC 204508 / S288c) 0 X=559292 G N=SSA4 PE=1 SV=3
44	127	P09435 HSP73_YEAST	48.58	A cetylation (N-term)	70547	Heat shock protein SSA3 0 S=Saccharon yces cerevisie (stain ATCC 204508 / S288c) 0 X=559292 G N=SSA3 PE=1 SV=3
47	74	P00360 G 3P1_YEA ST	44.68	Carban idon ethylation	35750	G hycerablehyde-3-phosphate dehydrogenase 1 0 S=Sacchanon yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 G X=TDH1 PE=1 SV=3
55	226	P80210 PURA_YEAST	43.86		48279	A denyb succhate synthetase O S=Saccharon yces cerevisiae (strain ATCC 204508 / S288c) O X=559292 G N=ADE12 PE=1 SV=3
42	180	Q 06108 RG C1_YEA ST	42.56		120395	Regulator of the glycerolchannell OS=Saccharom yces cerevisiae (stmin ATCC 204508 / S288c) OX=559292 GN=RGC1PE=1SV=1
49	283	P15179 SYDM_YEAST	41.88		75461	A spartateHRN A ligase n itochondrialOS=Saccharon yces cerevisiae (strain ATCC 204508 / S288c) 0X=559292 GN=M SD 1 PE=1 SV=1
45	2778	P53312 SUCB_YEA ST	41.27		46901	Succinate—CoA ligase [ADP-fom ing] subunit beta m itochondrialO S=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 GN=LSC2 PE=1 SV=1
46	86	P19882 HSP60_YEAST	41.23		60752	Heatshock protein 60 m inchondrial0S=Saccharom yces cerevisie (stuain ATCC 204508 / S288c) 0X=559292 GN=HSP60 PE=1 SV=1
51	1996	P38156 M AL31_YEAST	40.55		68263	M altose penn ease M AL31 0 S=Saccharon yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 G N=M AL31 PE=1 SV=1
54	985	P02294 H 2B2_YEA ST	39.98		14237	H istone H 2B.2 0 S=Saccharom yces cerevisiae (strain ATCC 204508 / S288c) 0 X=559292 G N=H TB2 PE=1 SV=2

Supplementary Table 2. Specific information on the proteins contained in YCW NPs.

Number		Retention Time (min)	Peak Area (mAU*min)	Peak Hight (mAU)	Relative peak area (%)	Relative peak height (%)	Sample Amount (ug/g)
1	Mannose	24.547	76.931	147.967	8.17	11.20	30.5912
2	Ribose	32.843	80.046	119.705	8.50	9.06	30.7059
3	Rhamnose	33.957	68.725	101.261	7.30	7.67	37.8932
4	Glucosamine	36.543	81.359	105.265	8.64	7.97	36.1403
5	Glucose	45.287	58.693	108.620	6.23	8.22	29.5305
6	Galacturonic Acid	48.975	64.962	112.869	6.90	8.54	32.1512
7	Galactosamine	50.873	71.417	117.822	7.58	8.92	29.5746
8	Glucuronic Acid	53.685	75.380	117.666	8.00	8.91	40.5044
9	Galactose	59.113	82.683	89.621	8.78	6.78	32.4719
10	Xylose	61.437	94.660	107.369	10.05	8.13	33.4831
11	Arabinose	63.068	103.582	112.464	11.00	8.51	35.7434
12	Fucose	70.398	83.388	80.295	8.85	6.08	31.5859
Total			941.827	1320.923	100.00	100.00	

Number		Retention Time (min)	Peak Area (mAU*min)	Peak Hight (mAU)	Relative peak area (%)	Relative peak height (%)	Sample Amount (ug/g)
n.a.	Mannose	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Ribose	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Rhamnose	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Glucosamine	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
1	Glucose	45.340	2.006	2.157	100.0	100.00	221.1620
n.a.	Galacturonic Acid	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Galactosamine	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Glucuronic Acid	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Galactose	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Xylose	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Arabinose	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
n.a.	Fucose	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
Total			2.006	2.157	100.00	100.00	

Supplementary Table 3. Specific information of mass spectrometry data of monosaccharide. Standards (up) and YCW NPs (down).



Supplementary Fig. 2 (related to Figure 1). Characterization of YCW NPs with different sizes. (A) TEM of mixture nanoparticles (Scale bar = 500 nm). We found that most of the broken particles were spherical or quasi-spherical, whereas NPs with irregular shape were also observed. (B) TEM of large size of YCW NPs (Scale bar = 500 nm). (C) TEM of middle size of YCW NPs (Scale bar = 100 nm). (D) TEM of small size of YCW NPs (Scale bar = 50 nm). (E) Zeta potential of YCW NPs with different sizes (n = 3). (F-G) Diameter and zeta potential of three different size of YCW NPs within two weeks to monitor the stability of YCW NPs (n = 3). (F) RT. (G) 4°C. Data are means \pm SD.



Supplementary Fig. 3. In vitro cytotoxicity assessment evaluated by MTT assay. (A) Dendritic cells. (B) Macrophages. (C) B16 (the concentration of YCW NPs was 150 μ g/mL) (n=6). Data are means \pm SD.



Supplementary Fig. 4. Activation of BMDCs and RAW264.7 after incubation with YCW NPs with different sizes. (A) Representative flow cytometric analysis of Cy5.5 expression and (B) corresponding quantitative analysis of Cy5.5 expression on BMDCs after incubation Cy5.5-labelled YCW NPs with different sizes for 24 h (n = 3). (C) Confocal imaging of Cy5.5-labelled YCW particles with different sizes after incubation with BMDCs for 24 h (blue: DAPI; red: Cy5.5; Scale bar = 10 μ m, n = 3). (D) Representative flow cytometric analysis of Cy5.5 expression on RAW264.7 after incubation Cy5.5-labelled YCW particles (including MPs, Large NPs, Middle NPs, Small NPs) for 24 h. (E) The corresponding quantification of Cy5.5 (MFI) after RAW264.7 incubation with Cy5.5-labelled particles for 24 h (n = 3). (F) Confocal imaging of Cy5.5; Scale bar = 20 μ m, n = 3). (G) Representative flow cytometric analysis and (H) corresponding quantification of Cy5.5 (MFI) after incubation of DC2.4 with Cy5.5-labelled YCW particles for 24 h (n = 3). (I)

Representative flow cytometric analysis and (**J**) corresponding quantification of Cy5.5 (MFI) after incubation of RAW264.7 with Cy5.5-labelled YCW particles for 24 h (n = 3). Statistical significance was obtained by one-way ANOVA using the Tukey post-test. **** P < 0.0001; *** P < 0.005; ** P < 0.01; * P < 0.05. Data are means ± SD. MFI: mean fluorescence intensity; arb. units: arbitrary units.



Supplementary Fig. 5 (related to Figure 2). Activation of BMDCs induced by YCW particles. (A-B) Activation of BMDCs after BMDCs incubation with YCW particles, including MPs, large NPs, middle NPs, small NPs and LPS (positive control) for 24 h. (A) Representative dot plots of costimulatory molecules CD80 and CD86 expression on BMDCs and (B) corresponding quantification of BMDCs maturation (n = 3). (C-F) Concentration of pro-inflammatory cytokines secreted by BMDCs after incubation with YCW particles as indicated. (C) TNF- α ; (D) IL-12p70; (E) IL-1 β ; (F) IL-6 (n = 3). (G) The corresponding quantification of CD40 expression on BMDCs after YCW NPs incubation with BMDCs for 24 h (n = 3). (H) The corresponding quantification of MHC II expression on BMDCs after YCW NPs incubation with BMDCs for 24 h (n = 3). (J) The corresponding quantification of PD-L1 expression on BMDCs after YCW NPs incubation with BMDCs for 24 h (n = 3). (K) The corresponding quantification of CTLA-4 expression on BMDCs for 24 h (n = 3). (K) The corresponding quantification of CTLA-4 expression on

BMDCs after YCW NPs incubation with BMDCs for 24 h (n = 3). (L) The corresponding quantification of LAG-3 expression on BMDCs after YCW NPs incubation with BMDCs for 24 h (n = 3). (M) The corresponding quantification of TIM-3 expression on BMDCs after YCW NPs incubation with BMDCs for 24 h (n = 3). Statistical significance was obtained by one-way ANOVA using the Tukey post-test. **** P < 0.0001; *** P < 0.005; ** P < 0.01; * P < 0.05. Data are means ± SD. MFI: mean fluorescence intensity; arb. units: arbitrary units.



Supplementary Fig. 6 (related to Figure 2). Flow cytometry gating strategy for analysis of BMDCs maturation.



Supplementary Fig. 7 (related to Figure 2). Complete initial data of western blotting analysis of Dectin-1 / Syk pathway and TLR2 / MyD88 pathway. (A)Western blotting analysis of Dectin-1 / Syk pathway and TLR2 / MyD88 pathway from proteins of BMDCs after incubation with three YCW NPs and LPS for 24 h, including TLR2, p-Syk, p-P65, MyD88, Dectin-1. (B) After utilizing Dectin-1 competitor laminarin for 2 h, western blotting analysis of Dectin-1 / Syk pathway from proteins of BMDCs after incubation with Small NPs for 24 h, including p-Syk, p-P65, Dectin-1. (C) After utilizing TLR2 inhibitor C29 for 2 h, western blotting analysis of TLR2 / MyD88 pathway from proteins of BMDCs after incubation with Small NPs for 24 h, including p-P65, TLR2, MyD88 (n = 3, each experiment was repeated three times independently with similar results, and we chose one results to show).



Supplementary Fig. 8. The influence of YCW NPs on T cells in vitro. (**A**) Representative flow cytometric analysis and (**B**) corresponding quantitative analysis of Cy5.5 intensity on T cells after incubation of T cells with Cy5.5-labelled YCW NPs (including large NPs, middle NPs, small NPs) for 24 h (n = 3). (**C-I**) Corresponding quantitative analysis of CD3 T cells (**C**); CD4⁺ cells in CD3 cells (**D**); PD-1 expression on CD4⁺ T cells (**E**); CD69 expression on CD4⁺ T cells (**F**); CD8 expression on CD3 cells (**G**); PD-1 expression on CD8⁺ T cells (**H**); CD69 expression on CD8⁺ T cells (**I**) (n = 3). Statistical significance was obtained by one-way ANOVA using the Tukey post-test. Data are means \pm SD. MFI: mean fluorescence intensity; arb. units: arbitrary units.



Supplementary Fig. 9. Dose-response experiment for antitumor response. (A) Average and (B-F) individual tumor growth curves in different dose group, including 0 mg/kg, 0.09 mg/kg, 0.18 mg/kg, 0.375 mg/kg and 0.75 mg/kg (n = 5). Data are means \pm SD.



Supplementary Fig. 10 (related to Figure 3). YCW NPs inhibited tumor growth by remodeling immunosuppressive tumor microenvironment. (A) Proportion of CD8⁺ T cells in CD3⁺ T cells. (B) Absolute numbers of the CD8⁺ of the tumor in UnTx group and small NPs group. (C) Representative flow cytometric analysis for CD45⁺ in tumors and (D) corresponding quantitative analysis of untreated group and Small NPs group. (E) Absolute numbers of the CD45⁺ of the tumor. (F) Corresponding quantitative analysis for FOXP3⁺ CD4⁺ in tumors of untreated group and small NPs group. (G) Absolute numbers of the CD4⁺ FOXP3⁺ of the tumor. (H) Corresponding quantitative analysis of untreated group and Small NPs group. (I) Absolute numbers of the CD11c⁺ of the tumor (n = 4). Statistical significance was obtained by Student's t tests (two-tailed). *** *P* < 0.005; ** *P* < 0.01; * *P* < 0.05. Data are means ± SD.



Supplementary Fig. 11 (related to Figure 3). Flow cytometry gating strategy for analysis of different cells in tumor. (A) T cells; (B) MDSCs; (C) Tregs; (D) TAMs; (E) DC maturation.



Supplementary Fig. 12. Distribution of YCW micro-particles (MPs). (A) Fluorescence imaging of TDLNs ex vivo after injection of Cy5.5 and Cy5.5-labelled MPs for 48 h. (B) Signal quantification of Cy5.5-labelled MPs in TDLNs (n = 3). (C) Confocal imaging of TDLNs to monitor the signal of Cy5.5 (blue: DAPI; red: Cy5.5; Scale bar = 80 μ m, n = 3). Statistical significance was obtained by one-way ANOVA using the Tukey post-test. Data are means ± SD.



Supplementary Fig. 13 (related to Figure 4). Distribution of YCW NPs in vivo. (A) Fluorescence imaging of tumor, TDLN and non-TDLN ex vivo after injection of Cy5.5-labelled NPs for 48 h. (B) Proportions of NPs in tumor, TDLN and non-TDLN after injection of Cy5.5-labelled NPs for 48 h (n = 3). (C-F) Representative flow cytometric analysis of immune cells with TDLNs after injection of Cy5.5-labelled YCW NPs for 48 h. (C) DCs; (D) Macrophages; (E) B cells; (F) T cells. (G) Proportions of particles associated cells of tumor after intratumorally injection for 48 h (n = 3). (H) Proportions of particles associated cells of non-tumor draining lymph nodes after intratumorally injection for 48 h (n = 3). Statistical significance was obtained by one-way ANOVA using the Tukey post-test. **** P < 0.0001; *** P < 0.005; ** P < 0.01; * P < 0.05. Data are means ± SD.



Supplementary Fig. 14 (related to Figure 5). Flow cytometry gating strategy for analysis of T cells in blood.



Supplementary Fig. 15 (related to Figure 6). H&E staining and flow cytometry analysis of TME after therapeutic. (A) H&E staining of major organs of each group after combination therapy (Scale bar = $200 \mu m$, n = 3). (B) Representative flow cytometry analysis of MDSCs and (C) representative flow cytometry analysis of TAMs within the tumors.



Supplementary Fig. 16 (related to Figure 7). Combination therapeutic with small size YCW NPs and anti-PD-L1 for B16. (A-B) Quantification of bioluminescence signal of primary tumors (A) and distant tumors (B) observed on day 8, 11, 14, 17 (UnTx: n = 4; aPD-L1: n = 3; Small NPs: n = 4; Combine: n = 4). (C) Weight curves of mice in different groups during treatment (n = 4). (D) Imaging of mice in four groups on day 14 and day 17. Statistical significance was obtained by Student's t tests (two-tailed). *** *P* < 0.005; * *P* < 0.05. Data are means ± SD.



Supplementary Fig. 17. Treatment efficiency of YCW small NPs and YCW MPs (GPs). (A) Average and (B) individual tumor growth curves in different treatment group, including UnTx, YCW MPs (GPs), YCW small NPs (n = 4). (C) Weight of mice in three groups during treatment (n = 4). Statistical significance was obtained by one-way ANOVA using the Tukey post-test. **** P < 0.0001. Data are means ± SD.