

### Altered succinylation links abnormal metabolism to APP and tau in Alzheimer's Disease



**Open Access** This file is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. In the cases where the authors are anonymous, such as is the case for the reports of anonymous peer reviewers, author attribution should be to 'Anonymous Referee' followed by a clear attribution to the source work. The images or other third party material in this file are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

## REVIEWER COMMENTS

Reviewer #1 (Remarks to the Author):

Yang et al. describes a link between succinylation and amyloid and tau pathology in Alzheimer's disease (AD). The authors start by identifying differentially-changed succinyl-containing peptides by isobaric tandem mass tagging in two separate cohorts containing control and AD brain tissues (n= 5 cases each). They briefly describe characteristics of identified, succinylated proteins by in silico analyses. By stressing HEK293 cells with rotenone, the authors observe the translocation of KGDHC proteins to the cytosol and the loss of succinylation in mitochondrial proteins, highlighting a possible mechanism of metabolic change in AD. Authors also identify A $\beta$  and Tau succinylation exclusively in AD brain. APP and Tau transgenic mouse models were also used to assess lysine succinylation on amyloid and tau pathology. The authors show increased rates of fibrillization in purified, succinylated A $\beta$  and Tau peptides. Importantly for Tau, they show that the K311Succ site identified in AD brains shows a compromised ability to bind tubulin, similar to the effect of hyperphosphorylation and acetylation previously observed. Overall this is an interesting report that highlights the role of a new PTM in modifying Tau and amyloid structure. However, despite the many intriguing results, there remain many limitations of the data presented. This would include an improved vetting of the proteomic data, starting with sharing of peptide data from Proteome Discoverer to aid in the review of this manuscript. As succinylation in the realm of AD has been understudied, the levels of succinylation, especially on tau, should be compared to acetylation and ubiquitination. There were no comparisons of the spectra of A $\beta$  or MAPT peptides identified in brain to synthetic standards. Also, the imaging has many labeling errors and does not strictly agree with the interpretations of the authors. An independent validation of the A $\beta$  and Tau succinylation by some other means (WB, immunogold microscopy, etc.) is also warranted.

Major Comments:

- In what manner was the subcellular localization determined for Fig.2a/Supplementary Table 2? Gene ontology?
- Was the sample arrangement per each TMT batch ordered 5 CTL in a row then 5 AD in a row?
- Given the low number of succinylated peptides corroborated by both cohorts, please share a supplementary table that include the peptide data following database search (including Xcorr, PEP, etc.)
  - o What are the succinylation localization scores for each succinylated peptide?
  - o What was the succinylation localization score threshold that you set your Proteome Discoverer to?
  - o What methods were used to filter out falsely-localized succinylated peptides?
- ♣ Please provide the succinylation mass shift (+100.0160) used in the database searches as well as masses for the other PTMs.
  - o What was the average number of MS2 ions identified in succinylated peptides compared with non-succinylated peptides? The % enrichment
  - o What were the number of succinylated peptides identified in the total proteome dataset without enrichment?
  - o What was the average number of missed cleavages for each succinylated peptide?
- Please provide spectra for the A $\beta$  HDSGYEVHHQKLVFFAEDVGSNK succinylated peptide from brain and compare this to the synthetic standard (preferably heavy labeled). The authors do a nice job characterizing the in vitro succinylation of amyloid and Tau by MRM. Can the authors show side by side the MS/MS or MRM profiles of these standards compared to the peptides identified in the discovery proteomics screen?
- It is surprising that the VQIVYK Tau hexapeptide was identified with the C-terminal lysine (residue 311) succinylated (Fig. 6b). It has been previously reported that trypsin/LysC cleavage is blocked by succinylation ([https://link.springer.com/protocol/10.1007/978-1-60327-259-9\\_53](https://link.springer.com/protocol/10.1007/978-1-60327-259-9_53)). The authors need to confirm the MS/MS spectrum in AD with standards or use a targeted MS approach to map the more likely modified VQIVYK(succ)PVDSLK peptide in AD brain that they identified in their in vitro assays.
- In most published proteomic studies mitochondria proteins are decreased in AD within the total proteome. How do the authors reconcile this with the succinylome? Are differences in protein abundance rather than site specific changes due to the PTMs driving the decrease in AD. Can the

authors provide or estimate any changes in stoichiometry measurements?

- It is not clear why the pan-lysine succinyl antibody labeling so drastically attenuated in WT 10 month old mice? And why there such an increase in succinylation in Tg19959 mice? Authors need to resolve.
- Are the genotype and time point labels switched in Figure 5d? The images do not agree with the quantitation Also, why is there so little co-localization with A $\beta$  plaque and the succinyl-lysine antibodies? Authors need to explain.
- In Figure 6c the authors label the images with a mouse model (Tg19959) that does not match the mouse model described in the text (TgP301S).
- Again, there is little co-localization with aggregated Tau antibody IF labeling and pan-lysine succinylation. One would expect extensive co-localization if succinylation labels the core of PHFs, where this PTM maps.
- The authors state on Page 13, Lines 309-310 to Page 14, Lines 311-312: “a weak signal for succinylated tau occurred in 10-month-old TgP301S” ... “indicating a desuccinylation process may exist in the final states of tau deposition”. Also, with no evidence, the authors offer a phosphorylation-succinylation switch as a possible reason.
- Authors state that Both APP and Tau were highly succinylated at critical sites in nine out of ten AD brain samples, but no succinylation of APP or tau was detectable in any control brains. Just because these sites were detected only in AD does not necessarily mean they are highly succinylated. What about lysine acetylation or ubiquitination? Are these sites on tau modified by these PTMs? Did the authors search their data for these PTMs at the same sites? Lysines are highly modified residues and one would expect succinylation to be a relatively minor pool of modified Tau in the AD brain. The authors should measure the pools of these PTMs (succinylation, ubiquitination, acetylation) to assess the relative frequency of succinylation vis a vis acetylation and ubiquitination.
- The weak signals on Tau contradict the human data of increased succinylated Tau. How do the authors reconcile these findings? Could other PTMs on these sites (ubiquitin, acetylation) also occur or do mice have increased SIRT5 (or other SIRT/desuccinylase enzymes) abundances? The global human proteomes analyzed showed no changes in SIRT5 as the authors note in the manuscript in Extended Data Fig. 2c. Could this differ in the mice?
- The authors should visit larger proteomic datasets for more accurate steady-state protein level quantification in AD cohorts that can fully describe ADAM/SIRT family levels in AD as compared with controls.
- Can the authors isolate AD brain PHF tau aggregates then perform immunogold labeling with the pan lysine succinylation antibody? Also for Abeta plaques? Then with a non-AD tauopathy control (ex: Corticobasal degeneration with prominent K311 Acetylation?) Describing that your in vitro tau fibrils look like AD brain fibrils doesn't stand up. This would be an independent validation of your mass spectrometry experiments
- The authors should include PHF6-K311Acetyl as a positive control to compare the succinylated peptide for the Tau Self-aggregation assay.

Minor Comments:

- Fig. 2A: change “succinlyated” to “succinylated”
- Fig. 2B: change “succinlated” to “succinylated”
- Change APP770 to APP695 (as well as accompanying notations), as APP695 is the major neuronal species (Fig. 5)
- Specify the mass shift specific to lysine succinylation in the methods
- There are some very minor fold changes between the control and AD brain sample proteins in Extended Data Figure 2. Is this due to MS/MS spectral interference and compression. Can the authors discuss.
- Page 11, Line 252-253: The Tg19959 line contains three mutations:
  - o Swedish: K670N/M671L
  - o Indiana: V717F
- Page 9, Lines 205-206. What was the correlation between change in succinylation and change in protein abundance? Please illustrate/state
- Has acetylation been observed at A $\beta$  K16?
- The authors should provide High-resolution images for Extended Data Figure 3a-b?

- Please provide a representative image for cleaved A $\beta$  6-29 fragment precursor ion peaks in Extended Data Figure 3. The plot in Extended Figure 3f-g is confusing
- What is “Percentage Change (%)” quantified in Figure 5f?
- Page 13, Line 300: effects of what?
- Page 13, Line 302: “IF staining to compare the presence or absence of succinylation with that of tau” – What does this mean
- Page 13, Line 304: “but in 4-month-old TgP301S mice”.
  - o What occurred in 4-month-old TgP301S mice?
- Figure 6d: “d” is covered up by panel c
- Please label Fig.6f-h with the peptide at hand (f=PHF6, g=S-PHF6, h=9:1 mix)
- Page 17, Line 394: “bot amyloidosis”?
- Page 8 Line 179: “Since no specific motifs for lysine succinylation in human cells have been reported” – They have been reported in HeLa cells in Weinert et al., Cell Reports (2013) <http://dx.doi.org/10.1016/j.celrep.2013.07.024>. Since they have reported previously, the authors should compare Motifs in AD brain to theirs, which upon first glance, don’t exactly match.
- Page 16, Line 375: fix spelling errors

Reviewer #2 (Remarks to the Author):

The manuscript “Succinylation Links Metabolic Reductions to Amyloid and Tau Pathology” compares succinylation in AD versus controls and surprisingly identifies AB and tau as targets that succinylated exclusively in AD. As a potential rationale for why these proteins may be succinylated in AD, the paper shows that mitochondrial dysfunction in cells leads to escape of proteins from the mitochondria that may be functioning in the succinylation of pathological AB and tau. In addition, the authors perform a series of biochemical experiments which suggest a way that succinylation may facilitate pathological AB and tau.

In the end, I think they can say that this paper demonstrates that a new protein modification is found on APP and tau that may correlate with AD status. In addition, some nice biochemical experiments raise a potential way in which this modification can potentially influence the aggregation of these proteins. There is no evidence that it is actually functionally doing so in vivo, nor is there any evidence that it correlates with progression in humans, and the correlation in mice is the weakest part of the paper. It remains possible that the modification is simply a consequence of mitochondrial dysfunction in AD patients and the biochemistry not actually relevant to what is functionally occurring in vivo—perhaps because the succinylation moiety is rather large. Nevertheless, even in this case, succinylation could serve as a marker, so it is still potentially relevant. Also, there is sufficient data to warrant following up the work. Thus, overall the findings are interesting. However, the level of over-interpretation and over-blown claims are reckless and unwarranted, so the text needs major revisions. There are also some experimental concerns.

Major comments:

The introduction is really short for an unknown topic and the Nature Communication format. A lot more needs to be added for the reader to understand succinylation, mitochondrial dysfunction, and AD pathology.

In Figure 3C, demonstrating that the change in succinylation is not just due to changes in protein levels is a critical point. The correlation that is shown, though weak, is a bit troubling. There is a brand new paper (Johnson et al, Nature Medicine 2020) focusing on proteomics in neurodegenerative diseases. I think it would be important to compare the changes in succinylated proteins to the changes in proteins presented in an independent paper, such as this one to make this point more convincing.

Figure 4A, without a control for protein loading, the overall change in levels of succinylation are

meaningless. This is somewhat mitigated by the B-actin control for the individual proteins in B. However, is the B-actin from 4B also being used to normalize 4C? The B-actin should be shown on the same blot in C. Also, there probably should be controls showing that the fractions have been sorted intact. In Figure 4D, it looks like there is less colocalization in the Rotenone treatment. Are the images reversed? In either case, the resolution of the images is too low to comment on the localization. Also, why was only 100nm Rotenone shown? What about 5uM? Overall, I probably buy the interpretation, but the data could be cleaned up.

In Figure 5C and 5D, it would be nice to have an unaffected staining control to show that the change succinylation is specific. More importantly, in WT there is a dramatic decrease in succinylation between 4 months and 10 months. Why is this? Could this be due to mitochondrial changes in normal aging. This should probably be commented on in the discussion. In the Tg mouse, there a lot more succinylation. However, there is also a decrease in succinylation from 4 to 10 months that is similar to WT, despite the fact that AB is definitely increasing in the Tg mice over this time period. Thus the two do not seem to be particularly well correlated. Also, is the level of succinylation increase in the Tg mice prior to appearance of AB? This should be checked because they already observe a dramatic increase in succinylation at 4 months when AB is first forming. Thus, it is possible that succinylation is changing in the Tg model well before this. Overall, though it is clear that succinylation is responding to the Tg, it is not at all clear that it correlates with the build up of AB. The exact same thing is true for tau in Figure 6. The fact that the same phenomenon is true in both AB and Tau models is perhaps even more disconcerting, because it suggests that the phenomenon is not specific. That is to say, the overall increase in succinylation seems to occur regardless of the pathological insult, which does not cause mitochondrial dysfunction exactly the same way in the two models, and the timing of that pathological insult, which is not exactly the same in the two mouse models. This should at least be discussed. In particular, it would be nice to know how the changes in succinylation relate to the changes in mitochondrial dysfunction in the two Tg models in their hands. Overall, the data supports a change in succinylation that occurs in the two pathological models, but does not support the conclusion that the change in succinylation truly correlates with pathology. At a minimum, this should be clearly noted in the results and discussion.

There is a slight concern that the peptide used for the tubulin polymerization assay is succinylated throughout, when they only detected succinylation at K311 in AD. Nevertheless, the loss of Tau polymerization function is impressive. This should be more clearly stated and used to qualify the interpretation. This is particularly true because the succinylation moiety is quite large. The authors should definitely discuss how such a large modification could affect proteins and the biochemical assays that they perform on them. The decrease in Tau-tubulin interactions is more convincing, particularly because they also performed this assay with Tau only succinylated at K311.

Overall, it is possible that succinylation is simply a consequence of mitochondrial dysfunction, and not necessarily functional in AD. To mitigate this, I think it might be good to provide some additional negative controls if possible. ie do AB and tau accumulate any other post-translational modifications that might just be due to disruption of mitochondria or the abnormal appearance of the pathological versions of these proteins in the cytosol. Or is it specific to succinylation? Is there any way to truly rule out that the observations are simply due to mitochondrial dysfunction and not necessarily functional?

There is a lot of wild speculation:

Example: The last sentence of the abstract is wildly overexaggerated- While there is a possibility that succinylation could contribute to pathologies in AD, the data presented in the manuscript certainly are not by themselves enough to even raise the possibility that succinylation must be addressed therapeutically for meaningful clinical benefit

Example: The last sentence of the introduction is overstated and unnecessary

Example: line 311- this reflected a potential existence of succinylation-phosphorylation switch as in the case of acetylation- the paper provides absolutely no evidence for this. This could be speculated about in the discussion, but is completely inappropriate for the results section.

Example: line 283- taken together the accumulated data strongly suggest that succinylation of K678

might lead to an early-onset enhanced generation, oligomerization and plaque biogenesis, consistent with the effects of known genetic disease mutations at this site. While the data suggest a potential was for succinylation to affect AB cleavage, there is no functional evidence that it does so.

Example line 369: Notably, these results demonstrate for the first time that succinylation is the key link between the signature metabolic reductions and amyloid plaques and neurofibrillary tangles in AD.-

Again, although this is possible, there is absolutely no evidence for this in the manuscript

Example line 371: The current results reveal that varied in protein succinylation, as a molecular signal, correlates with altered cerebral metabolic function in AD as the disease progresses.- While there may be a small amount of evidence of this in the mice data (if you compare to previous analysis of the mice strains employed and you ignore the fact that the mouse data don't truly correlate), they do not show this on their own and they certainly did not examine succinylation across the progression of the disease in human cases, so this statement is not warranted.

Minor comments:

I really don't understand the math in figure 1, and there is no description of what the 29 proteins in B are? Are they differential between AD vs control, as in D? And what percentages are up vs down? An effort should be made to make the numbers more clear.

The antibody used in extended data figure 3B needs to be clearly shown in the figure.

In extended figure 3F and G, why is the full length protein so different when the production of the cleavage products remains the same? Perhaps I don't quite understand the assay, but this should be clarified.

The label of the graph in 5F needs to be clarified as percentage change from 0hrs.

Line 309 refers to succinylated tau. However, it should refer to succinylation in the Tau Tg mice. There is a big difference.

The Y-axis in Figure 4B says evel instead of level

The Tg19959 and P301S mouse models need to be defined for the reader.

Line 194 should say from rather than form

PDHA1 is mentioned with no context

Line 257: paralleled should be parallel

The logic in line 266-267 needs to be better spelled out for the reader with regard to the competition of the two enzymes. Likewise in line 271, ADAM10 needs to be introduced as a secretase for the reader.

The S and C labels in 5C need to be defined in the main figure.

Line 302: abeta should be absence?

Line 371: varied should be variation

Line 375: involvon should be involved in?

Line 394: bot should be both?

Line 385: The decline in succinylation of mitochondrial proteins suggests that activation of descuccinylases- The alternative, that there could be a failure to maintain succinylation levels, should

be mentioned.

The manuscript should also be edited for grammar.

David Katz  
Emory University

Reviewer #3 (Remarks to the Author):

The authors have investigated the potential role of succinylation and Amyloid and Tau pathology using brain tissue from AD cases and controls. They analysed brain tissue cell lysate proteomes using 10 plex TMT. They also analyzed the same 10 controls and 10 AD patients' brain samples Succinylome using Cell Signaling Tech IP-MS kit and ran LCMS of the PTM enriched tryptic peptides. Comments:

- 1- In addition to bioinformatics analysis of succinylome IP-MS data it would be useful to analyze and show the biological significance of whole tissue lysate 10- plex TMT data as well. It would be useful to cover the global proteome analysis done which may be relevant to disease pathogenesis in addition to the succinylome targeted concept.
- 2- It is important to pinpoint sites of protein succinylation. Succinylome localization shown in Figure1a is not clear.
- 3- The authors considered impaired mitochondrial function resulted in succinylome localization to be pushed out of mitochondria to cytosol by leakage (Figure 4, Line 244-247), however, whole tissue lysate mass spec succinylome data suggested an overall decrease in AD. These findings need to be reconciled
- 4- The K687 site in the middle of APP is the interaction site of  $\alpha$ alpha-secretase and the cleavage was inhibited when the K was succinylated in vitro (Figure 5e). However, it cannot be assumed that succinylated A $\beta$  has more aggregation property since the comparison shown in Figure 5f did not show a significant difference. The AD patient succinylome mass spec identified succinylated K687 peptide (S Table 5). It is not clear whether the succinylome IP conditions favored solubilizing Abeta plaque and protofibril oligomers.
- 5- APOE4 mutation is a risk factor of AD. Does the proteomics data reveal mutation in the 20 patient brains analyzed?

## RESPONSE TO THE REVIEWERS' COMMENTS

The reviewers' comments are in RED.

Our answers are inserted in blue immediately after each question. The sections from the manuscript showing the change with line numbers were inserted after our rationale.

The line numbers refer to the respective document (manuscript, methods etc of final documents). A copy showing all changes is included.

### REVIEWER COMMENTS

#### Reviewer #1 (Remarks to the Author)

Yang et al. describes a link between succinylation and amyloid and tau pathology in Alzheimer's disease (AD). The authors start by identifying differentially changed succinyl-containing peptides by isobaric tandem mass tagging in two separate cohorts containing control and AD brain tissues (n= 5 cases each). They briefly describe characteristics of identified, succinylated proteins by in silico analyses. By stressing HEK293 cells with rotenone, the authors observe the translocation of KGDHC proteins to the cytosol and the loss of succinylation in mitochondrial proteins, highlighting a possible mechanism of metabolic change in AD. Authors also identify A $\beta$  and Tau succinylation exclusively in AD brain. APP and Tau transgenic mouse models were also used to assess lysine succinylation on amyloid and tau pathology. The authors show increased rates of fibrillization in purified, succinylated A $\beta$  and Tau peptides. Importantly for Tau, they show that the K311Succ site identified in AD brains shows a compromised ability to bind tubulin, similar to the effect of hyperphosphorylation and acetylation previously observed. Overall, this is an interesting report that highlights the role of a new PTM in modifying Tau and amyloid structure. However, despite the many intriguing results, there remain many limitations of the data presented.

This would include an improved vetting of the proteomic data, starting with sharing of peptide data from Proteome Discoverer to aid in the review of this manuscript.

As detailed in response to specific comments below (lines 157-167, 721-727) in this response), we improved the vetting of the proteomic data including making the data from Proteome Discoverer software available (see Supplementary Table 7).

As succinylation in the realm of AD has been understudied, the levels of



38 succinylation, especially on tau, should be compared to acetylation and ubiquitination.

39

40 The interaction of succinylation with other modifications is critical. The new  
41 paragraph in the introduction places our findings in the context of the field. In the  
42 current manuscript we focused on succinylation, because each of these post-  
43 translational modifications require specific immuno-enrichment.

44 Manuscript lines 166-175

45

166 Post-translational modifications (PTMs) of proteins provide an efficient and rapid biological regulatory  
167 mechanism that links metabolism to protein and cell functions. PTMs contribute to the functional  
168 diversity of proteomes without the formation of new proteins or a change in their abundance by covalent  
169 addition of functional groups that can alter protein charge, structure, and their interactions. Protein PTMs  
170 play a central role in the pathology of neurological diseases. The function of tau can be altered via its  
171 phosphorylation<sup>10</sup>, acetylation<sup>11</sup>, methylation<sup>12</sup> and O-GlcNAcylation<sup>13</sup>. Protein succinylation of lysine  
172 residues is a relatively novel PTM and changes the charge from positive to negative. The interactions of  
173 lysine succinylation and acetylation play an important role in metabolic pathways<sup>14</sup>. However,  
174 succinylation is poorly studied in the nervous system; our previous work demonstrated that lysine  
175 succinylation functionally modifies enzymes of energy metabolism<sup>15</sup>.

46

47

48 There were no comparisons of the spectra of A $\beta$  or MAPT peptides identified in brain  
49 to synthetic standards.

50

51 These mass spec are in this response and Lines 225-245 in this response.

52

53 Also, the imaging has many labeling errors and does not strictly agree with the  
54 interpretations of the authors.

55

56 Our apologies, poor coordination of co-authors led to many errors in the imaging  
57 section. All of these have been corrected

58 Lines 368 - Lines 376

59

368 To characterize tau succinylation in a transgenic mouse model of tangle formation, we used  
369 immunofluorescence staining to compare the presence of lysine succinylation within tau oligomers (T-22)  
370<sup>46</sup> and phospho-tau (AT8) in the hippocampus from wild type and TgP301S mice. No phosphorylated tau  
371 and few tau oligomers were present in the brain of wild type mice (Figure 6c, d and Extended Data  
372 Figure 4a, b). The immunofluorescence signal of tau oligomers and phospho-tau was significantly  
373 augmented in the hippocampal region of TgP301S mice<sup>47</sup>. A parallel increase in lysine succinylation and  
374 oligomeric tau T-22 (green) and phospho-tau (green) was observed in tau mice compared to wild type  
375 animals (Figure 6c, d and Extended Data Figure 4a, b). Thus, tau succinylation associates with tau  
376 aggregates in a transgenic mouse model of tauopathy.

60

61

62

63 Lines 304-312

304 AD-associated succinylation of APP occurred at a critical site (K612) in nine of ten brains from AD  
305 patients but not in brains from age-matched subjects with no dementia (Figure 5a, b), and the following  
306 experiments demonstrated it to be pathologically important. We observed an increase in the levels of  
307 lysine succinylation and severity of amyloid burden in a transgenic mouse model of AD (Tg19959 mice),  
308 which carries the human APP with two mutations (Figure 5c and Extended Data Figure 3a). Double  
309 immunofluorescence staining with antibodies to pan-lysine-succinylation and to A $\beta$  oligomers (NU-4)<sup>36</sup>  
310 or to A $\beta$  plaque ( $\beta$ -Amyloid, D3D2N) revealed an early increase in lysine succinylation that appeared to  
311 parallel oligomer formation and subsequent plaque deposition in the hippocampus. These findings suggest  
312 that the APP succinylation might be involved in A $\beta$  oligomerization and plaque formation *in vivo*.

64  
65  
66 An independent validation of the A $\beta$  and Tau succinylation by some other means  
67 (WB, immunogold microscopy, etc.) is also warranted.  
68

69 The paper contains extensive evidence by mass spectrometry to support the  
70 unequivocal direct interaction of succinylation with A $\beta$  and tau in human brains by  
71 mass spectrometry. While immunogold may strengthen the conclusions in mouse  
72 brains, we think that this should be part a complete time course study of the mouse  
73 pathology and mass spectrometry to assess the precise labelling. Thus, to do it  
74 properly is beyond the scope of this manuscript.  
75

#### 76 Major Comments:

77  
78 We would like to thank Reviewer 1 for the detailed and pertinent questions raised  
79 about the mass spectrometry datasets of both succinylome and global proteome  
80 studies. The authors agree that important information was missing in the  
81 supplementary documents and that it should be presented to provide a better  
82 understanding of the specific succinylation sites and peptides, their identity  
83 confidence, their biological importance and parallel comparison with the global  
84 proteome results. Please find below a point by point response to Reviewer 1's  
85 comments on our manuscript.  
86

87 • In what manner was the subcellular localization determined for  
88 Fig.2a/Supplementary Table 2? Gene ontology?  
89

90 Methods section line numbers 310-313  
91

310 Subcellular localization of the identified candidates was determined using Cytoscape (version  
311 3.6.1)<sup>16</sup> and stringAPP (version 1.4.0)<sup>17</sup> software. All the parameters were set to the default values, but  
312 only these highest compartment scores equal 5 as the high confidence localization were kept. The result  
313 was visualized in FunRich (version 3.1.3).

92  
93 • Was the sample arrangement per each TMT batch ordered 5 CTL in a row then 5  
94 AD in a row?

95 No, for global proteome per each TMT batch with 5 AD and 5 CTL, we randomized  
96 the order of labeling each of the 10 samples by TMT10plex, we added this point in  
97 the revised method section.

98 Methods line numbers 35-37

35 over 1 hour at r.t. The peptides from the 10 samples (5 controls and 5 AD cases) were mixed each tag  
36 respectively with 126-tag, 127N-tag, 127C-tag, 128N-tag, 128C-tag, 129N-tag, 129C-tag, 130N-tag,  
99 37 130C-tag and 131-tag. The order of labeling each of the 10 samples by TMT10plex was randomized.

100

101 • Given the low number of succinylated peptides corroborated by both cohorts,  
102 please share a supplementary table that include the peptide data following database  
103 search (including Xcorr, PEP, etc.)

104

105 The requested supplementary table that includes all the peptide data following  
106 database search (including Xcorr, PEP, etc.) information was added as Supplementary  
107 Table 7.

108

109 What are the succinylation localization scores for each succinylated peptide?

110

111 The threshold of succinylation localization score was set at PSM grouper node of  
112 consensus workflow in Proteome Discoverer (PD) 2.2 database searching software,  
113 and succinylation localization scores for each succinylated peptide must be greater  
114 than 75 and it lies in between 75-100. ptmRS node was not used in processing  
115 workflow for determination of localization scores for other PTMs because ptmRS  
116 node was more designated for phosphorylation in PD 2.2. We added this point in the  
117 revised method section.

118

119 Methods section lines 122-134

120

122 output to Microsoft Excel software for further data analysis. The threshold of succinylation localization  
123 score was set at PSM grouper node of consensus workflow in Proteome Discoverer (PD) 2.2 database  
124 searching software, and succinylation localization scores for each succinylated peptide must be greater  
125 than 75 and it lies in between 75-100. In combination with threshold score ( $\geq 75$ ) for succinylation  
126 localization, falsely-localized succinylated peptides were further filtered out under peptide validator node  
127 in consensus workflow where q values and PEPs are validated for available PSMs and assigned the PSMs  
128 confidences based on the user defined target FDRs in percolator node (Target FDR-0.01). In addition,  
129 data analysis for all identified succinylated peptides indicated that the average number of missed  
130 cleavages for each succinylated peptides = 1 (Supplementary Table 7). This is not surprising, as lysine  
131 succinylation would prohibit trypsin from cut the modification site creating one missed cleavage. As a  
132 result, we found that over 90% of succinylated peptides being identified were equivalent to 0 miss-  
133 cleavage and 10% contained 1 miss-cleavage site in our data, which is consistent with what we observed  
134 in our regular global proteomics.

121

122

123

124 • What was the succinylation localization score threshold that you set your Proteome  
125 Discoverer to?

126

127 As stated in the above response, threshold score for site probability was set to 75

128 at PSM grouper node of consensus workflow in PD 2.2, which implies  
129 modifications (variable) with lower site probability than the specified threshold  
130 will not be shown in the final list of succinylated peptides. We added this point in  
131 the revised method section.

132 Methods section line numbers 122-135

133

122 **output** to Microsoft Excel software for further data analysis. The threshold of succinylation localization  
123 score was set at PSM grouper node of consensus workflow in Proteome Discoverer (PD) 2.2 database  
124 searching software, and succinylation localization scores for each succinylated peptide must be greater  
125 than 75 and it lies in between 75-100. In combination with threshold score ( $\geq 75$ ) for succinylation  
126 localization, falsely-localized succinylated peptides were further filtered out under peptide validator node  
127 in consensus workflow where q values and PEPs are validated for available PSMs and assigned the PSMs  
128 confidences based on the user defined target FDRs in percolator node (Target FDR-0.01). In addition,  
129 data analysis for all identified succinylated peptides indicated that the average number of missed  
130 cleavages for each succinylated peptides = 1 (Supplementary Table 7). This is not surprising, as lysine  
131 succinylation would prohibit trypsin from cut the modification site creating one missed cleavage. As a  
132 result, we found that over 90% of succinylated peptides being identified were equivalent to 0 miss-  
133 cleavage and 10% contained 1 miss-cleavage site in our data, which is consistent with what we observed  
134 in our regular global proteomics.

134

135

136 • What methods were used to filter out falsely-localized succinylated peptides?

137

138 In combination with threshold score ( $\geq 75$ ) for succinylation localization, falsely-  
139 localized succinylated peptides were further filtered out under peptide validator node  
140 in consensus workflow where q values and PEPs are validated for available PSMs and  
141 assigned the PSMs confidences based on the user defined target FDRs in percolator  
142 node (Target FDR – 0.01). We added this point in the revised method section at

143

144 Methods lines 122-127

145

122 **output** to Microsoft Excel software for further data analysis. The threshold of succinylation localization  
123 score was set at PSM grouper node of consensus workflow in Proteome Discoverer (PD) 2.2 database  
124 searching software, and succinylation localization scores for each succinylated peptide must be greater  
125 than 75 and it lies in between 75-100. In combination with threshold score ( $\geq 75$ ) for succinylation  
126 localization, falsely-localized succinylated peptides were further filtered out under peptide validator node  
127 in consensus workflow where q values and PEPs are validated for available PSMs and assigned the PSMs  
128 confidences based on the user defined target FDRs in percolator node (Target FDR-0.01). In addition,

146

147 • Please provide the succinylation mass shift (+100.0160) used in the database  
148 searches as well as masses for the other PTMs.

149 We used the following modifications with specific mass shift as variable

150 modifications:-1. Dynamic Modification: Oxidation / +15.995 Da (M)

151 -2. Dynamic Modification: Acetyl / +42.011 Da (K) and N-terminal of proteins.

152 -3. Dynamic Modification: Succinyl / +100.016 Da (K)

153 -4. Dynamic Modification: Deamidated / +0.984 Da (N, Q)

154 We have added all the information in the revised method section

155

156 Methods **section lines 116-120**

157

116 For label-free quantitative data analysis of succinylated peptides, fragment ion tolerance 0.5 Da was used  
117 for the ion trap analyzer and an additional succinylation on Lys residue with mass shift (+100.0160 Da)  
118 was specified as variable modifications. In addition, methionine oxidation (+15.995 Da), acetylation  
119 (+42.011) on N-terminal proteins and deamidation (+0.984 Da) on asparagines/glutamine were also set up  
120 as variable modifications. For each relative ratio of succinylated peptides/sites, no normalization was

158

159

160 • **What was the average number of MS2 ions identified in succinylated peptides**  
161 **compared with non-succinylated peptides? The % enrichment**

162

163 **For 1st cohort of succinylome study, the average number of MS2 ions identified**  
164 **in succinylated peptides compared with non succinylated peptides =**  
165 **51779/150384. The % enrichment = 34.4 %.**

166 **For 2nd cohort of succinylome study, the average number of MS2 ions identified**  
167 **in succinylated peptides compared with non succinylated peptides =**  
168 **49901/149319. The % enrichment = 33.4 %.**

169

170 **Manuscript lines 243-246** were inserted.

171

243 identifying the succinylated peptides in large cohorts. After enrichment, we found that  
244 the average enrichment of succinylated peptides was found to be 33.9% in two cohorts while 0.2% of  
245 succinylated peptide was identified in global proteome without enrichment. Of 1,908 succinylated  
246 peptides identified in two independent analyses, 932 succinylated peptides were quantifiable (**Figure 1a**).

172

173

174 • **What were the number of succinylated peptides identified in the total proteome**  
175 **dataset without enrichment?**

176

177 For global proteome analysis in 1st cohort, the number of succinylated peptide  
178 identified = 126 out of total 94,263 peptides (0.13%). For global proteome analysis in  
179 2nd cohort, the number of succinylated peptide identified = 201 out of 71,367  
180 (0.28%).

181 The notable difference in ratio of succinylated peptides over total peptides without  
182 enrichment between the two cohorts of global proteome datasets is not surprising, as  
183 we know that the succinylation has relatively low occupancy level. Therefore, there will  
184 be an anticipated variation between two cohorts' datasets for detection of those low  
185 abundance succinylated peptides under global and complex quantitative proteomics analysis.  
186 This assessment also indicates that the enrichment is important for reliably identifying  
187 the succinylated peptides in large cohorts.

188 **Manuscript line numbers 234-243**

234 **Succinylome and proteome changes in AD brains**

235 Completion of the human brain succinylome and global proteome analyses allowed direct comparison  
236 between brains from controls and AD patients. Without enrichment of succinylated peptide in global  
237 proteome data, the number of succinylated peptides identified is 0.13% total peptides for cohort 1 and  
238 0.28% for cohort 2. The notable difference in ratio of succinylated peptides over total peptides between  
239 the two cohorts of global proteome datasets is not surprising, as we know that the succinylation has  
240 relatively low occupancy level. Therefore, there will be an anticipated variation between two cohorts'  
241 datasets for detection of those low abundance succinylated peptides under global and complex  
242 quantitative proteomics analysis. This assessment also indicates that the enrichment is important for  
243 identifying the succinylated peptides in large cohorts. After enrichment, we found that  
244 the average enrichment of succinylated peptides was found to be 33.9% in two cohorts while 0.2% of  
245 succinylated peptide was identified in global proteome without enrichment. Of 1,908 succinylated

189  
190  
191

- What was the average number of missed cleavages for each succinylated peptide?

193

194 The average number of missed cleavages for each succinylated peptides = 1 (the  
195 requested Supplementary Table 7 for succinylome data has the missed cleavage  
196 information for each succinylated peptide). This is expected as lysine succinylation  
197 will prohibit trypsin from cutting the modification site, creating one missed cleavage.  
198 While small percentage (~7%) with 0 miss cleavage reflects the succinylated lysines  
199 are located at either protein C-terminus or with the Pro residue in its carboxyl side.  
200 Therefore, the miss cleavage ratios for the succinylated peptides we identified are  
201 equivalent to 90% with 0 miss cleavage and 10% with 1 miss cleavage, similar to  
202 what we observed in our regular global proteomics.

203

For 1<sup>st</sup> cohort:

204

0 missed cleavage = 175 succ peptides (7.5%)

205

1 missed cleavage = 1935 succ peptides (83.2%)

206

2 missed cleavage = 215 succ peptides (9.2%)

207

For 2<sup>nd</sup> cohort:

208

0 missed cleavage = 163 succ peptides (7.3%)

209

1 missed cleavage = 1849 succ peptides (82.7%)

210

2 missed cleavage = 224 succ peptides (10.0%)

211

212

See supplementary data 2 and a brief summary in [Methods lines 129-134](#)

213

129 data analysis for all identified succinylated peptides indicated that the average number of missed  
130 cleavages for each succinylated peptides = 1 (Supplementary Table 7). This is not surprising, as lysine  
131 succinylation would prohibit trypsin from cut the modification site creating one missed cleavage. As a  
132 result, we found that over 90% of succinylated peptides being identified were equivalent to 0 miss-  
133 cleavage and 10% contained 1 miss-cleavage site in our data, which is consistent with what we observed  
134 in our regular global proteomics.

214

215

Please provide spectra for the A $\beta$  HDSGYEVHHQKLVFFAEDVGSNK succinylated  
216 peptide from brain and compare this to the synthetic standard (preferably heavy  
217 labeled). The authors do a nice job characterizing the in vitro succinylation of amyloid

218 and Tau by MRM. Can the authors show side by side the MS/MS or MRM profiles of  
219 these standards compared to the peptides identified in the discovery proteomics  
220 screen?

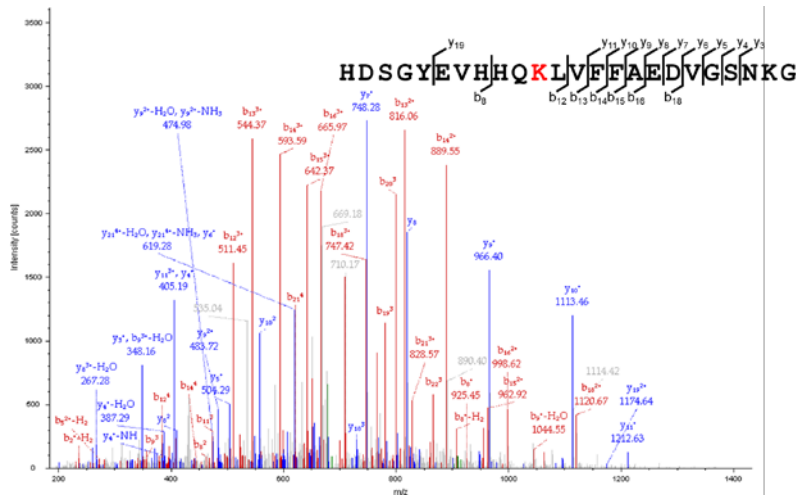
221

222 We have added the figures. Please see Figures and tables lines 76-83 in the Figure 5a  
223 and lines 304-306.

224

225 Fig.5a The MS/MS spectrum of the succinylated peptide from brain  
226 (Succinylation lysine residue is highlighted in red text)

227



228

229

230

231 Figures and tables legends lines 76-83

232

76 **Figure 5.** Succinylation occurs uniquely on APP from AD patients, in early stages of plaque  
77 formation in mouse models and disrupts APP processing.  
78 **a.** Location and identity of succinylation K612 near the A $\beta$  region. Residues are numbered  
79 according to APP695 sequence. Purple amino acids refer to  $\alpha$ - or  $\beta$ - or  $\gamma$ - cleavage sites. The red  
80 underlined lysine refers to succinylated K612. Purple arrow represents the two central strands of  
81 the  $\beta$ -sheet (Leu613-Asp619 and Ala626-Val632). Green highlights the peptide identified in the  
82 MS. The MS spectra of the succinylated peptide from brain (Succinylation lysine residue is  
83 highlighted in red text).

233

234

235 Manuscript lines 304-306

304 AD-associated succinylation of APP occurred at a critical site (K612) in nine of ten brains from AD  
305 patients but not in brains from age-matched subjects with no dementia (**Figure 5a, b**), and the following  
306 experiments demonstrated it to be pathologically important. We observed an increase in the levels of

236

237

238 **Extended Data Figure 3d.** The MS/MS spectrum of the synthetic standard peptide

239 (Succinylation lysine residue is highlighted in red text)

240 Extended data line 22-23

22 d. The MS/MS spectra of the synthetic succinylated A $\beta$ <sub>6-29</sub> peptide used in assay (Succinylation lysine residue is  
23 highlighted in red text).

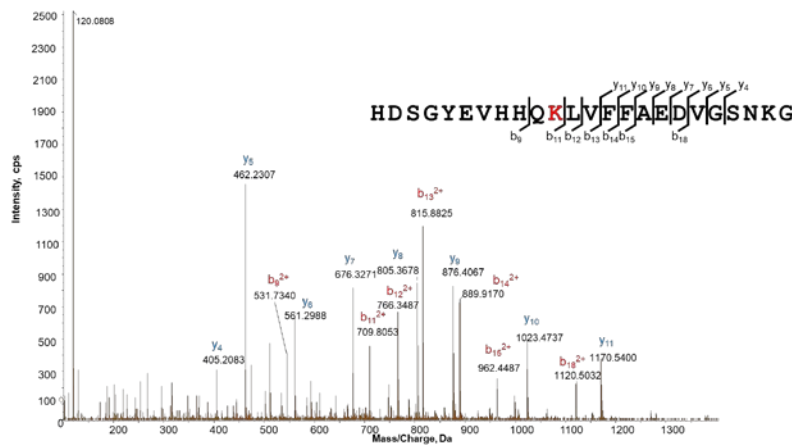
241

242

243

244

245



246

247 Extended Data Figure 3d

248

249

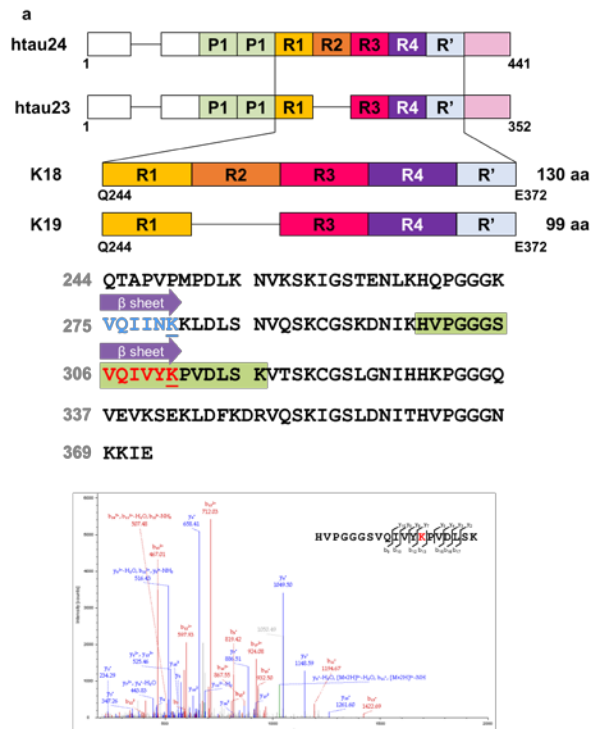
250

251 • It is surprising that the VQIVYK Tau hexapeptide was identified with the C-  
252 terminal lysine (residue 311) succinylated (Fig. 6b). It has been previously reported  
253 that trypsin/LysC cleavage is blocked by succinylation  
254 ([https://link.springer.com/protocol/10.1007/978-1-60327-259-9\\_53](https://link.springer.com/protocol/10.1007/978-1-60327-259-9_53)). The authors need  
255 to confirm the MS/MS spectrum in AD with standards or use a targeted MS approach  
256 to map the more likely modified VQIVYK(succ)PVDSLK peptide in AD brain that  
257 they identified in their in vitro assays.

258

259 The highlighted sequence VQIVYK (named PHF6) highlighted in red in Figure 6a  
260 (not Figure 6b) was intended to indicate beta-sheet structure only. The peptide  
261 “HVPGGGSVQIVYKPVDSLK” highlighted in green was the one identified by MS  
262 in brains. And the MS/MS spectra were added for this peptide in Figure 6a (Line 116-  
263 125 in the Figure).





116 **Figure 6.** The unique succinylation of K311 on tau in brains from patients with AD promotes AD  
 117 like features in tau pathology.

118 **a.** Domain structure of tau and the location of succinylation K311. The diagram shows the domain  
 119 structure of htau23 and 24, which contain three and four repeats, respectively. The constructs K18  
 120 and K19 comprise four repeats and three repeats, respectively. Residues are numbered according  
 121 to tau441 sequence. Purple arrow represents the two central strands of the  $\beta$ -sheet (PHF6\*:  
 122 Val275-Lys280, highlighted in blue, the blue underlined lysine refers to acetylated K280; PHF6:  
 123 Val306-Lys311, highlighted in red, the red underlined lysine refers to succinylated K311). Green  
 124 highlights the peptide identified by MS. The MS spectra of the succinylated peptide from brain  
 125 (Succinylation lysine residue is highlighted in red text).

265  
 266  
 267  
 268 • In most published proteomic studies mitochondria proteins are decreased in AD  
 269 within the total proteome. How do the authors reconcile this with the succinylome?  
 270 changes in mitochondrial protein levels and succinylation are occurring, but the  
 271 decline in protein abundance cannot account for the changes in succinylation  
 272 peptides/sites. There are some succinylated proteins where succinylation  
 273 peptide/site levels were increased or decreased in AD much more than the abundance  
 274 of the corresponding proteins determined in global proteomic data. We have added the  
 275 following to reconcile these findings.

276 **Manuscript lines 258-264**

258 patients while 73 protein levels were increased (**Extended Data Figure 2a**). In a recent large-scale  
 259 proteomic scan, the protein abundance of PDHA, PDHB, and DLD were all decreased in AD, which is  
 260 consistent with our finding, representing a decreased abundance of proteins in impaired mitochondrial

8

261 states<sup>17</sup>. Thus, changes in protein levels and succinylation may be important in AD. Relatively small fold  
 262 changes found between control and AD brain samples, were probably due to a well-known ratio  
 263 compression caused by the co-isolation of isobaric-labeled background ions in MS2-based TMT  
 264 quantitative proteomics.

277

278

279

280 **Are differences in protein abundance rather than site specific changes due to the**  
 281 **PTMs driving the decrease in AD. Can the authors provide or estimate any changes in**  
 282 **stoichiometry measurements?**

283

284 The comparison of the AD-related changes from our proteomics and succinylomics  
 285 indicates the changes in the succinylome are likely independent of protein changes  
 286 (Fig. 3c). The heatmap shows the magnitude of variation in the succinylome/proteome  
 287 as color in two dimensions. Each cell's color indicates the value of the fold change  
 288 ( $\text{Log}_2(\text{Fold Change})$ ). The variation in the succinylome is much larger ( $|\text{Log}_2(\text{Fold Change})| > 0.3$ )  
 289 than the abundance changes of the same protein that happens in the  
 290 proteome ( $|\text{Log}_2(\text{Fold Change})| < 0.2$ ).

291 A total of 213 out of 229 succinylated mitochondria proteins was identified in the  
 292 proteome. Only 37 succinylated mitochondria proteins were significantly changed  
 293 ( $p < 0.05$ ), in which 27 proteins (73%) were decreased. 959 quantifiable succinylated  
 294 peptides were found in 208 succinylated mitochondria proteins. Only 21 succinylated  
 295 peptides from mitochondria proteins were significantly changed ( $p < 0.05$ ), in which 21  
 296 succinylated peptides (71%) were decreased. Only 4 mitochondria proteins  
 297 significantly change at protein level accompanied by a significant alteration of the  
 298 succinylated peptide level (5 succinylated peptides), which are listed below.

GI Number	UniProtKB	Entry name	Succinylome		Proteome	
			Log <sub>2</sub> FC	p-value	Log <sub>2</sub> FC	P-value
129379	P10809	HSPD1	0.52	0.04294	-0.08	0.0255
21542295	Q9NVH6	TMLHE	0.39	0.01607	0.16	0.0054
			0.49	0.04928		
20455474	P24539	ATP5F1	-0.32	0.04433	-0.14	0.0182
			-0.47	0.04788		

<b>3.08E+08</b>	<b>P00505</b>	<b>GOT2</b>	<b>-0.63</b>	<b>0.00561</b>	<b>-0.11</b>	<b>0.0126</b>
-----------------	---------------	-------------	--------------	----------------	--------------	---------------

299

300 Manuscript line numbers 197-200

197 peptides being identified were equivalent to 0 miss-cleavage and 10% contained 1 miss-cleavage site in  
 198 our data, which is consistent with what we observed in our regular global proteomics. The parallel global  
 199 proteomic analysis detected 4,678 proteins (Figure 1d). Nearly all of the succinylated proteins identified  
 200 during the study were found in the global proteome of the same samples (Figure 1e).  
 201

301

302

303 Manuscript line numbers 253-264

253 controls (Figure 1d and Extended Data Figure 2a, b). A comparison of the succinylome with the  
 254 proteome demonstrated little AD-related changes in protein levels of those succinylated proteins, and  
 255 therefore the succinylation variations are most likely independent from the changes of the corresponding  
 256 protein abundance (Figure 3c). The proteomic analysis showed that 81 proteins changed significantly  
 257 (two-tailed Student's *t*-test,  $p < 0.05$  and  $|\log_2FC| > 0.25$ ). Eight proteins decreased in brains from AD  
 258 patients while 73 protein levels were increased (Extended Data Figure 2a). In a recent large-scale  
 259 proteomic scan, the protein abundance of PDHA, PDHB, and DLD were all decreased in AD, which is  
 260 consistent with our finding, representing a decreased abundance of proteins in impaired mitochondrial

8

261 states<sup>17</sup>. Thus, changes in protein levels and succinylation may be important in AD. Relatively small fold  
 262 changes found between control and AD brain samples, were probably due to a well-known ratio  
 263 compression caused by the co-isolation of isobaric-labeled background ions in MS2-based TMT  
 264 quantitative proteomics.  
 265

304

305

306 It is not clear why the pan-lysine succinyl antibody labeling so drastically attenuated  
 307 in WT 10 month old mice?

308

309

310

311

312

313

314

315

316

317

318

319

320

321

322

Manuscript line numbers 308-312, Lines 372-376

Our chemistry test tube experiments are clearly consistent with our hypothesis that succinylation can promote plaques and tangles. The goal of the mouse studies is to show an association of succinylation in the brain to APP or tau. The data clearly show that in mice that are four months old. We do not know the precise relation of succinylated tau or APP to the final pathology (i.e., tangles and plaques). One can imagine scenarios where they promote formation but not be high in final product.

The four-month data clearly shows an association of succinylation to tau and amyloid. Interpretation of the ten-month data, which includes the maturation process, adds confusion not clarity. Thus, we have chosen to omit the ten-month data. These results enhance our enthusiasm for the current study, because they reveal new exciting areas to be developed. We changed the manuscript to reflect the changes and the interpretation.

308 which carries the human APP with two mutations (Figure 5c and Extended Data Figure 3a). Double  
309 immunofluorescence staining with antibodies to pan-lysine-succinylation and to A $\beta$  oligomers (NU-4)<sup>36</sup>  
310 or to A $\beta$  plaque ( $\beta$ -Amyloid, D3D2N) revealed an early increase in lysine succinylation that appeared to  
311 parallel oligomer formation and subsequent plaque deposition in the hippocampus. These findings suggest  
312 that the APP succinylation might be involved in A $\beta$  oligomerization and plaque formation *in vivo*.

323  
324  
325  
326  
327  
328  
329  
330  
331  
332  
333  
334  
335  
336  
337  
338  
339  
340  
341  
342  
343  
344  
345  
346  
347  
348  
349  
350  
351  
352  
353  
354  
355  
356  
357  
358  
372 **Figure 4a, b).** The immunofluorescence signal of tau oligomers and phospho-tau was significantly  
373 augmented in the hippocampal region of TgP301S mice<sup>47</sup>. A parallel increase in lysine succinylation and  
374 oligomeric tau T-22 (green) and phospho-tau (green) was observed in tau mice compared to wild type  
375 animals (Figure 6c, d and Extended Data Figure 4a, b). Thus, tau succinylation associates with tau  
376 aggregates in a transgenic mouse model of tauopathy.  
377

Nevertheless, we have chosen to add a speculative answer to the reviewer's concern, but this has not been added to the text because we omitted the 10 month values.

Succinylation is a post-translational modification and several factors can regulate the balance between succinylation and desuccinylation. Most of these are unknown in brain. Our findings show that KGDHC is a major succinyl transferase in neurons. Brain KGDHC is not altered with age out to 30 months (Freeman, Nielsen et al. 1987) suggesting that the age-related change is not a reduction in active succinylation. Whether aging may alter KGDHC migration to the cytosol has never been studied. The desuccinylases in brain remain unknown. A prominent paper by two of our co-authors have shown that sirtuin 5 (SIRT5) plays a central role in modulating heart metabolism and function (Sadhukhan, Liu et al. 2016). SIRT5 is localized in the mitochondria and shows a weak deacetylase activity but a potent desuccinylase activity on lysine residues both *in vitro* and *in vivo* (Park, J 2013 Mol. Cell; Du et al., 2011; Peng et al., 2011). The catalytic reaction involves the removal of a succinyl group from the lysine side chain of protein substrates, a process that consumes NAD<sup>+</sup> as a co-substrate and generates nicotinamide (NAM) and 2'-O-succinyl-ADP-ribose (Rardin MJ 2013 Cell Metabol). SIRT5 KO mouse embryonic fibroblasts display an increase in lysine succinylation but not acetylation (Du et al. 2011 Science). We have used SIRT5 to desuccinylate enzymes such as the pyruvate dehydrogenase complex. We have also looked at succinylation in SIRT5 KO mice, which show a significant increase in succinylation levels in the liver while trivial changes were found in the brain. Liver succinylation, but not that in brain responds to fasting. Furthermore, the data from the AD samples suggest that different desuccinylases are likely important in the cytosol and mitochondria. We believe that our current results justify further studies on the regulation of succinylation in the brain during aging and in neurodegenerative diseases.

And why there such an increase in succinylation in Tg19959 mice? Authors need to resolve.

Please note that this also occurs in P301S mice. We think that the widespread increase

359 in succinylation shows that the transgenes are causing widespread changes in multiple  
360 proteins including tau and APP. We know the pathological implications for tau and  
361 APP but it is likely changes in other proteins are likely important. This is yet another  
362 important area of research opened by the current results. We have added the following  
363 sentences to the paper.

364  
365 To the results section Manuscript lines 306-312

306 experiments demonstrated it to be pathologically important. We observed an increase in the levels of  
307 lysine succinylation and severity of amyloid burden in a transgenic mouse model of AD (Tg19959 mice),  
308 which carries the human APP with two mutations (Figure 5c and Extended Data Figure 3a). Double  
309 immunofluorescence staining with antibodies to pan-lysine-succinylation and to A $\beta$  oligomers (NU-4)<sup>36</sup>  
310 or to A $\beta$  plaque ( $\beta$ -Amyloid, D3D2N) revealed an early increase in lysine succinylation that appeared to  
311 parallel oligomer formation and subsequent plaque deposition in the hippocampus. These findings suggest  
312 that the APP succinylation might be involved in A $\beta$  oligomerization and plaque formation *in vivo*.

366  
367  
368 The P301S mice reveals a widespread increase in background succinylation.

369 Lines 372-376 in the manuscript.

370  
371  
372 Figure 4a, b). The immunofluorescence signal of tau oligomers and phospho-tau was significantly  
373 augmented in the hippocampal region of TgP301S mice<sup>47</sup>. A parallel increase in lysine succinylation and  
374 oligomeric tau T-22 (green) and phospho-tau (green) was observed in tau mice compared to wild type  
375 animals (Figure 6c, d and Extended Data Figure 4a, b). Thus, tau succinylation associates with tau  
376 aggregates in a transgenic mouse model of tauopathy.

372  
373  
374 To the discussion section

375 The results reveal that both transgenic mice strains reveal widespread increases in  
376 succinylation, which suggests that many proteins in addition to tau and APP are  
377 altered. Determining whether this is an artifact of the transgene or a down- stream  
378 consequence of the abnormal tau and APP remains to be determined.

379 The manuscript lines 439-447 now read

380  
381  
439 We show that transgenic mouse strains of either tauopathy or amyloidosis phenotype including plaques  
440 exhibit widespread increases in lysine succinylation, which is not exclusive to tau and APP, but parallels  
441 the formation of pathological species. While proteins in addition to tau or APP maybe altered, APP and  
442 tau are only succinylated in brains from AD patients, which suggests that increased tau and APP  
443 succinylation may play a role in the development of AD pathology. Thus, the modification of metabolism  
444 in disease may lead to critical succinyl-mediated modifications of extra-mitochondrial proteins including  
445 APP and tau leading to aggregation and deposition. Preventing APP and tau succinylation and/or  
446 increasing mitochondrial succinylation may provide novel therapeutic targets for the prevention and/or  
447 treatment of AD and tauopathies.

382  
383 • Are the genotype and time point labels switched in Figure 5d? The images do not  
384 agree with the quantitation

385

386 We apologize for the mistake. They are all correct in the revised version. This has  
387 been amended in the revised version. See Figures 5c and 5d.

388

389 Also, why is there so little co-localization with A $\beta$  plaque and the succinyl-lysine  
390 antibodies? Authors need to explain.

391

392 We have now provided improved images at low and high magnification to show the  
393 co-localization (white) better. We have also omitted the data from the 10 month-old  
394 mice. We do not postulate that APP and tau are the only cytosolic proteins  
395 succinylated. We used a pan succinylation antibody. As shown by the succinylomics  
396 data, hundreds of proteins are succinylated. Succinylation is not evenly distributed  
397 among all proteins as some contain a higher number of succinylated sites than others  
398 despite containing a similar amount of total lysines, suggesting site-specificity of  
399 succinylation. Nevertheless, we do see co-localization of succinylation with tau and  
400 APP oligomers.

401

402 We added the following sentences to the manuscript lines 442-450.

442 We show that transgenic mouse strains of either tauopathy or amyloidosis phenotype including plaques  
443 exhibit widespread increases in lysine succinylation, which is not exclusive to tau and APP, but parallels  
444 the formation of pathological species. While proteins in addition to tau or APP maybe altered, APP and  
445 tau are only succinylated in brains from AD patients, which suggests that increased tau and APP  
446 succinylation may play a role in the development of AD pathology. Thus, the modification of metabolism |  
447 in disease may lead to critical succinyl-mediated modifications of extra-mitochondrial proteins including  
448 APP and tau leading to aggregation and deposition. Preventing APP and tau succinylation and/or  
449 increasing mitochondrial succinylation may provide novel therapeutic targets for the prevention and/or  
450 treatment.<sup>24</sup>

403

427 Since a pan anti-succinylation antibody was used, many proteins are labelled and the immunostaining for  
428 succinylation shows broad distribution. Thus, the staining would not be expected to specific to A $\beta$   
429 plaques and NFTs. The results support that there is an association and co-localization of succinylation  
430 with plaques and NFTs, which is particularly prominent at early stages. Further, succinylation is not  
431 evenly distributed among all proteins, as some contain a higher number of succinylated sites than others  
432 despite containing a similar amount of total lysines. Thus, we would not expect succinylation to be  
433 limited to plaques and tangles. APP and tau were only succinylated in brains from AD patients. Thus, the

404

405

406 • In Figure 6c the authors label the images with a mouse model (Tg19959) that does  
407 not match the mouse model described in the text (TgP301S).

408

409 We apologize for the mistake. We have now labeled the figures correctly.

410 Lines 128-134 in Figures and Tables.

128 c. High resolution images acquired using confocal laser microscopy display the co-localization of  
129 succinylation and tau oligomers in the hippocampus of TgP301S and WT mice. A T22 antibody  
130 (green) was used to stain tau oligomers while a pan-succinyl-lysine antibody (magenta) labeled

---

131 the levels 131 of succinylation. Results were expressed as the mean with SEM representative of  
132 the average of ~900-1000 pyramidal neurons comprised in 3-4 different brain sections per  
133 animal (n = 4 per each group). \*\*\*\*:  $p < 0.0001$ , two-way ANOVA followed by Tukey's multiple  
134 comparisons test.

411

412

413 • Again, there is little co-localization with aggregated Tau antibody IF labeling and  
414 pan-lysine succinylation. One would expect extensive co-localization if succinylation  
415 labels the core of PHFs, where this PTM maps.

416

417 The white shows clear co-localization. Since we used a pan succinylation antibody,  
418 many other succinylated proteins are present. Further, succinylation is not evenly  
419 distributed among all proteins as some contain a higher number of succinylated sites  
420 than others despite containing a similar amount of total lysines, Thus, we would not  
421 expect succinylation to be limited to tangles.

422

423 Further, we do not know the relation of PTM to tangle maturation. Tau  
424 acetylation has been studied for decades and detailed mechanisms are still unknown,  
425 but acetylation has many consequences on brain function besides just tangle  
426 formation (Tracy, Claiborn et al. 2019) Tau acetylation-induced pathogenesis may  
427 involve regulation of toxic forms of the protein, such as hyperphosphorylated protein,  
428 in which the consequences are site-specific. Tau acetylation can modulate tau toxicity  
429 by altering the formation of cleaved-caspase tau fragments. Acetylation at some sites  
430 is sufficient to drive synaptic and cognitive deficits without producing tau fragments.  
431 Tau acetylation may also affect the formation of tau oligomers and aggregates.  
432 Acetylation of tau lysines blocks those residues from being targeted for  
433 ubiquitination, slowing the rate of protein turnover and leading to accumulation.  
434 Whether acetylated tau propagates from cell-to-cell in the brain is unknown, and  
435 understanding if this property underlies its toxicity is an area of active research  
436 (Tracy, Claiborn et al. 2019). Since our paper is the first paper on succinylation of tau,  
437 it is not surprising that we do not know how all the pieces fit together. This means this  
438 paper is opening a whole new area of research.

438

439 We have added the following to the text on **Manuscript lines 442-450**

442 We show that transgenic mouse strains of either tauopathy or amyloidosis phenotype including plaques  
443 exhibit widespread increases in lysine succinylation, which is not exclusive to tau and APP, but parallels  
444 the formation of pathological species. While proteins in addition to tau or APP maybe altered, APP and  
445 tau are only succinylated in brains from AD patients, which suggests that increased tau and APP  
446 succinylation may play a role in the development of AD pathology. Thus, the modification of metabolism  
447 in disease may lead to critical succinyl-mediated modifications of extra-mitochondrial proteins including  
448 APP and tau leading to aggregation and deposition. Preventing APP and tau succinylation and/or  
449 increasing mitochondrial succinylation may provide novel therapeutic targets for the prevention and/or  
450 treatment.<sup>24</sup>

440

441

442 This the first study of tau succinylation, and the precise relation to tangle formation is  
443 unknown. Tau acetylation has been well-documented for over a decade, and its  
444 precise role is still unknown. **Manuscript lines 461-470**

445

461 Overall, our data represent the first report of the human brain succinylome and its implications for  
462 mitochondrial function as well as for the molecular pathogenesis of amyloidosis and tauopathy, two of the  
463 cardinal features of AD. We provide a rich resource for functional analyses of lysine succinylation, and  
464 facilitate the dissection of metabolic networks in AD. The current studies also lay the foundation for  
465 future investigation into the crosstalk between different PTMs, including acetylation, phosphorylation,  
466 ubiquitination and succinylation associated with AD and tau pathology. The discovery that succinylation  
467 links mitochondrial dysfunction to amyloidosis and tauopathy may provide new molecular diagnostics as  
468 well as potential targets for therapies. Since both succinylated A $\beta$  and tau are closely associated with  
469 disease state, future investigations may reveal additional succinylated proteins that are associated with  
470 AD or other neurodegenerative diseases.

446

447

448 • The authors state on Page 13, Lines 309-310 to Page 14, Lines 311-312: “a weak  
449 signal for succinylated tau occurred in 10-month-old TgP301S” ... “indicating a  
450 desuccinylation process may exist in the final states of tau deposition”. Also, with no  
451 evidence, the authors offer a phosphorylation-succinylation switch as a possible  
452 reason.

453 The decline in succinyl lysine signal in 10-month-old mice has been discussed  
454 elsewhere in this response above **(lines 309-352 and 327-352)**. The data has been  
455 withdrawn from the manuscript.

456

457 As discussed in more detail **above (Lines 309-352 of this response)** in the response),  
458 acetylation facilitates phosphorylation, one of the defining features of tangles. No  
459 such studies exist for succinylation, and we are hoping our findings will encourage  
460 these studies.

461

462 • Authors state that Both APP and Tau were highly succinylated at critical sites in  
463 nine out of ten AD brain samples, but no succinylation of APP or tau was detectable in  
464 any control brains. Just because these sites were detected only in AD does not  
465 necessarily mean they are highly succinylated.

466



467 **We agree. We have deleted the words highly succinylated from the manuscript.**

468

469 **What about lysine acetylation or ubiquitination? Are these sites on tau modified by**  
470 **these PTMs? Did the authors search their data for these PTMs at the same sites?**

471 **Lysines are highly modified residues and one would expect succinylation to be a**  
472 **relatively minor pool of modified Tau in the AD brain. The authors should measure**  
473 **the pools of these PTMs (succinylation, ubiquitination, acetylation) to assess the**  
474 **relative frequency of succinylation vis a vis acetylation and ubiquitination.**

475

476 **Studies of PTM** require immuno-enrichment of for each modification. It is  
477 difficult to identify the lysine acetylation or ubiquitination in the global proteome  
478 analysis without enrichment as those modifications generally have relatively low  
479 abundance. Hence, to study the succinylation effect in AD patients, it is required and  
480 necessary for enrichment of succinylated peptides prior to nano LC-MS/MS analysis.

481 Consequently, we did not search these modifications on tau in their data sets. We  
482 also did not search our datasets for other PTMs because we used the specific  
483 enrichment strategy specifically for succinylated peptides. Hence, it would not be  
484 possible to detect other modifications than succinylation even if they are present in  
485 the original samples.

486 We have searched the uniprot database, the lysine311 has been identified with  
487 ubiquitination, dimethylation, and acetylation in transgenic mice (Morris, Knudsen et  
488 al. 2015), not human brains. So far, these modifications (ubiquitination,  
489 dimethylation, and acetylation) of this site (K311) have not been reported on relevant  
490 to the mechanism of Tau protein pathology.

491 In additional study from our group we enriched with an anti-acetylation antibody  
492 to identify lysine acetylation modification and its change with AD. That is a whole  
493 new paper is being reviewed now.

494 We added the following to the discussion **(lines 452-459 in the manuscript)**

452 Lysines are highly modified residues. Understanding the relationship of succinylation to the other PTM is  
453 critical to a complete understanding of its role in AD pathology. A direct comparison is practically  
454 difficult because each PTM requires a different enrichment strategy. Lysine311 has been associated with  
455 ubiquitination, dimethylation, and acetylation in transgenic mice<sup>59</sup>, but not human brains. Our  
456 experiments raise the question of whether precise changes in mitochondria are required to alter  
457 modification of specific proteins. Succinylation appears directly linked to KGDHC and mitochondria.  
458 Whether other post-translational modifications are also linked to mitochondrial dysfunction remains to be  
459 determined.

495

496

497 • **The weak signals on Tau contradict the human data of increased succinylated Tau.**  
498 **How do the authors reconcile these findings? Could other PTMs on these sites**  
499 **(ubiquitin, acetylation) also occur or do mice have increased SIRT5 (or other**  
500 **SIRT/desuccinylase enzymes) abundances? The global human proteomes analyzed**  
501 **showed no changes in SIRT5 as the authors note in the manuscript in Extended Data**  
502 **Fig. 2c. Could this differ in the mice?**

503

504 **The succinyl lysine signal in plaques and tangles in mice has been discussed above**

505 (Lines 295-338 of this response). As discussed previously, the succinylases and  
 506 desuccinylases in brain are not known. Tau is well-known to be phosphorylated and  
 507 acetylated. We have an active program looking at these interactions. Studies of  
 508 phosphorylation have shown the degree of phosphorylation is not necessarily related  
 509 to the functional implications. Protein levels of SIRT5 do not necessarily reflect  
 510 activity, which can be regulated by substrates and post-translational modification.  
 511 Whether or not SIRT5 is the primary desuccinylase in brain is unknown.  
 512 The following is in the manuscript lines 181-187 and manuscript lines 436-440

181 which the succinyl donor is presumably succinyl-CoA, both in yeast<sup>21</sup> and cultured neurons<sup>15,23,24</sup>. Studies  
 182 of organisms deficient in NAD<sup>+</sup>-dependent desuccinylase sirtuin 5 (SIRT5)<sup>25</sup> provide evidence of the  
 183 regulatory importance of succinylation in metabolic processes<sup>26-30</sup>. However, the role of succinylation in  
 184 metabolic pathways of the human nervous system or in neurodegenerative diseases is unknown. Our  
 185 report represents the first investigation of the human brain succinylome and its changes in AD. The  
 186 results suggest that succinylation may link AD-related metabolic deficits to structural, functional, and  
 187 pathological alterations involving APP and tau.  
 188

513

436 from mitochondria to other cellular compartments<sup>58</sup>. The decline in succinylation of mitochondrial  
 437 proteins, appears due to a failure in maintaining succinylation levels, and may suggest that activation of  
 438 desuccinylases (e.g., Sirtuins) or general increases in NAD<sup>+</sup> should be reconsidered. The large increase  
 439 in succinylation in 4-month-old Tg19959 mice agrees with our hypothesis, in which abnormal  
 440 mitochondrial function in AD promotes the release of KGDHC and subsequent increases succinylation.  
 441

514

- 515 • The authors should visit larger proteomic datasets for more accurate steady-state  
 516 protein level quantification in AD cohorts that can fully describe ADAM/SIRT family  
 517 levels in AD as compared with controls.

518

519 In the paper by (Seyfried, Dammer et al. 2017), a total of 2745 proteins in two regions  
 520 (dorsolateral prefrontal cortex (FC, Brodmann Area 9) and precuneus (PC, Brodmann  
 521 Area 7) were quantified. The number that overlapped was about 2332 proteins  
 522 (85.3%) compared with our data (4442 proteins from 10 controls and 10 AD,  
 523 Brodmann area 44/45).

524 The four ADAM family members identified in that paper were also identified in our  
 525 proteome. The protein level of ADAM 10, 22, 23 did not change in that paper nor our  
 526 data, while ADAM11 showed a similar decrease in the two cases. The paper only  
 527 identified SIRT2 and SIRT5. However, the SIRT family levels did not vary in the two  
 528 cases. Furthermore, protein levels of SIRT5 do not necessarily reflect activity, which  
 529 are often regulated by substrates and post-translational modifications.

530

531

Symbol	Unique ID	Frontal Cortex (FC)			Precuneus (PC)		
		p value	Tukey's	log <sub>2</sub> (FC)	p value	Tukey's	log <sub>2</sub> (FC)
		ANOVA	AD-CTL	FC (AD-CT)	ANOVA	AD-CTL	PC (AD-CT)
ADAM10	O14672	0.051817	0.623137	0.234946	0.613604	0.691346	0.147285
ADAM23	E7EWD3	0.054739	0.194228	0.1514	0.242464	0.230508	0.198556

ADAM22	Q9P0K1	0.598328	0.589506	0.065565	0.098855	0.110043	0.142058
ADAM11	B4DKD2	0.016906	0.043947	-0.27101	0.000878	0.035282	-0.24924
SIRT2	Q8IXJ6	0.168041	0.868595	0.116332	0.722619	0.782255	0.088465
SIRT5	Q9NXA8	0.295005	0.294575	-0.16512	0.413197	0.465075	0.136716

532

533

The following has added to the manuscript lines 318-330

318 bond, and missense mutation at K612N produces early onset AD<sup>37</sup>. Furthermore, global proteomics  
 319 showed an increase in  $\beta$ -secretase (BACE1) abundance of 31% in AD brains compared to controls  
 320 (Supplementary Data Table 6), while no changes occurred for either  $\alpha$ -secretase or the sirtuins (SIRT)  
 321 family (Extended Data Figure 2c). Seyfried et al., quantified a total of 2,745 proteins in two regions  
 322 (dorsolateral prefrontal cortex (FC, Brodmann Area 9) and precuneus (PC, Brodmann Area 7)  
 323 were quantified. The number that overlapped was about 2,332 proteins (85.3%) compared with  
 324 our data (4442 proteins from 10 controls and 10 AD, Brodmann area 44/45).

325

326 The four ADAM family members identified in that paper were also identified in our proteome.  
 327 The protein level of ADAM 10, 22, 23 did not change in that paper nor our data, while ADAM11  
 328 showed a similar decrease in the two cases. SIRT2 and SIRT5 levels did not vary<sup>38</sup>. Further,  
 329 protein levels of SIRTs do not necessarily reflect activity, which are often regulated by substrates  
 330 and post-translational modifications.

534

535

536

537

538

539

540

- Can the authors isolate AD brain PHF tau aggregates then perform immunogold labeling with the pan lysine succinylation antibody? Also for Abeta plaques? Then with a non-AD tauopathy control (ex: Corticobasal degeneration with prominent K311 Acetylation?)

541 It would be interesting do measure succinylation by mass spec within the plaques and  
 542 tangles by mass spec and compare with the tau and APP not in the plaques and  
 543 tangles. Such experiments are beyond the scope of the current manuscript. We feel  
 544 that are mass spec studies already show succinylation definitively. We even know the  
 545 precise site of succinylation.

546

547

548

549

Describing that your in vitro tau fibrils look like AD brain fibrils doesn't stand up. This would be an independent validation of your mass spectrometry experiments

550 We agree that we overstated the conclusion. We changed the text to indicate  
 551 that we increased aggregation. The formation PHF is complicated process that  
 552 cannot be mimicked in a one protein system.

553 Please see lines 388-393 in the manuscript.

388 protein (Figure 6e). Longer incubation time (24 h) with PHF6, S-PHF6, and a 90%/10% mixture was  
 389 visualized by EM (Figure 6f-h). All the reactions exhibited fibrils with a typical paired helical filament  
 390 appearance. However, the succinylated peptide formed abundant, short filaments, a feature of brain-  
 391 derived Alzheimer PHFs<sup>49-51</sup>, while unmodified PHF6 filaments are longer and sparser, morphologies  
 392 more typical of recombinant tau peptide fibers (Figure 6i and 6j). Thus, both ThS and EM results support  
 393 an important role of succinylation in promoting pathological tau aggregation.

554

555

556

We observed each individual fiber in normal PHF6 showed ~15 nm in width

557 and ~107 nm of crossover repeat, while a width of ~12 nm and periodically  
558 appearing twists every ~86 nm are found in the succinylated PHF6 and mixture.

559 Morphologically speaking, the normal PHF6 fibers are much longer than  
560 these succinylated ones. Compared with the reported electron micrographs of  
561 PHFs from AD brain or assembled from recombinant tau peptides  
562 (10.1021/bi0357006), the normal PHF6 is just like these recombinant tau peptides  
563 with morphology, while the succinylated one is much like brain-derived  
564 Alzheimer PHFs characteristics, short and in a mess. However, the mixture  
565 seems much more like the normal PHF6 but it does have some small parts mixed  
566 among the main fibers. See the following text in the manuscript lines 384-393.

567

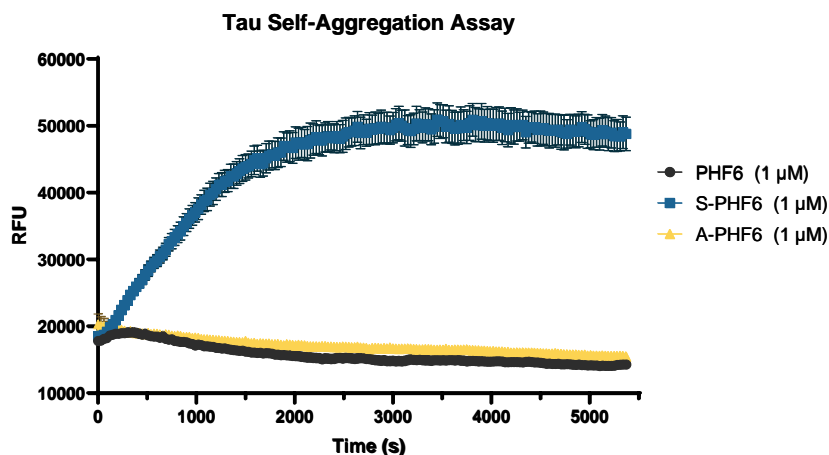
384 In contrast, PHF6 and K311-succinylated PHF6 (S-PHF6) fibrillated by 20 min and 80 min, respectively  
385 (Figure 6e). The aggregation of PHF6 was remarkably accelerated by the K311 succinylation. A  
386 substantial enhancement of PHF6-induced aggregation occurred even with a mixture containing 90%  
387 PHF6 and only 10% S-PHF6, suggesting that succinylated tau can promote aggregation of unmodified  
388 protein (Figure 6e). Longer incubation time (24 h) with PHF6, S-PHF6, and a 90%/10% mixture was  
389 visualized by EM (Figure 6f-h). All the reactions exhibited fibrils with a typical paired helical filament  
390 appearance. However, the succinylated peptide formed abundant, short filaments, a feature of brain-  
391 derived Alzheimer PHFs<sup>49-51</sup>, while unmodified PHF6 filaments are longer and sparser, morphologies  
392 more typical of recombinant tau peptide fibers (Figure 6i and 6j). Thus, both ThS and EM results support  
393 an important role of succinylation in promoting pathological tau aggregation.

568

569  
570  
571 The authors should include PHF6-K311Acetyl as a positive control to compare the  
572 succinylated peptide for the Tau Self-aggregation assay.

573

574 We purchased the requested peptides from GenScript. At peptide concentration of 1  
575  $\mu\text{M}$  in the presence of 2 nM heparin, neither PHF6 nor A-PHF6 fibrillated during a  
576 90-min incubation period. Since there is no data to support/report that K311  
577 acetylation can promote tau self-aggregation. We think the unmodified PHF6 can  
578 serve as an adequate positive control instead of A-PHF6, as it is well documented to  
579 aggregate under these conditions in the main test. Thus, we did not add these  
580 experiments to the text.



581

582 Fig. Tau peptides concentrations were 1  $\mu$ M in presence of 2 nM heparin: PHF6 (●),  
583 S-PHF6 (■), A-PHF6 (▲). Experiments were performed in quadruplicate and  
584 repeated three times with similar results. All values in the present graph were  
585 expressed as mean  $\pm$  SEM.

586

587

588 **Minor Comments:**

- 589 • Fig. 2A: change “succinlyated” to “succinylated”

590

591 We modified the manuscript. Please see line 18 in the Figures and Tables.

592 18 a. Subcellular distribution of succinylated-K proteins identified by ~~Cytoscape~~ and ~~stringAPP~~

592

- 593 • Fig. 2B: change “succinlated” to “succinylated”

594

595 We modified the manuscript. Please see line 20 in the Figures and Tables.

596 20 b. Overlap of succinylated-K proteins located in the mitochondrion, nucleus, cytosol and plasma

597

- 598 • Change APP770 to APP695 (as well as accompanying notations), as APP695 is the  
599 major neuronal species (Fig. 5)

600

601 These have all been corrected in the manuscript, Figure 5 and extended data figures.

602 See lines 78 and 79 in the figures and tables.

603 78 a. Location and identity of succinylation K612 near the A $\beta$  region. Residues are numbered

604 79 according to APP695 sequence. Purple amino acids refer to  $\alpha$ - or  $\beta$ - or  $\gamma$ - cleavage sites. The red

605

- 605 • Specify the mass shift specific to lysine succinylation in the methods

606 We add the following to the methods lines 91-93

607 91 For label-free SuccK peptides analysis, one MS survey scan was followed by 3 second “Top

608 92 Speed” data-dependent CID ion trap MS/MS scans with normalized collision energy of 30%. Dynamic

609 93 exclusion parameters were set at 1 within 45 s exclusion duration with  $\pm 10$  ppm exclusion mass width.

610

611

- 610 • There are some very minor fold changes between the control and AD brain sample  
611 proteins in Extended Data Figure 2.

612

613 Is this due to MS/MS spectral interference and compression. Can the authors discuss.

614

615 Yes, that is correct. This is a common issue for TMT labeled quantitative proteomics based on  
616 MS/MS fragmentation. It is well known that the co-isolation of near isobaric-labeled  
617 background ions causes spectral interference resulting in ratio distortion or ratio compression.

618 The main text was modified lines 261-264

619

261 states<sup>17</sup>. Thus, changes in protein levels and succinylation may be important in AD. Relatively small fold  
262 changes found between control and AD brain samples, were probably due to a well-known ratio  
263 compression caused by the co-isolation of isobaric-labeled background ions in MS2-based TMT  
264 quantitative proteomics.

620

621

622 • Page 11, Line 252-253: The Tg19959 line contains three mutations:

623

624 Please see the following in the [methods lines 398-403](#)

398 All the experiments were carried out in four and ten-month-old transgenic mouse models of AD.  
399 Tg19959 mice (that overexpress a double mutant form of the human amyloid precursor protein) were  
400 obtained from Dr. George Carlson (McLaughlin Research Institute, Great Falls, MT, USA). Tg19959  
401 mice were constructed by injecting FVB X 129S6 F1 embryos with a cosmid insert containing  
402 human APP<sub>695</sub> with 2 familial AD mutations (KM670/671NL and V717F), under the control of the  
403 hamster PrP promoter.<sup>25</sup> P301S (PS19, that overexpress the human tau gene harboring the P301S

625

12

626

627 • Page 9, Lines 205-206. What was the correlation between change in succinylation  
628 and change in protein abundance? Please illustrate/state

629

630 We have modified this section in the manuscript [lines 250-264](#).

631

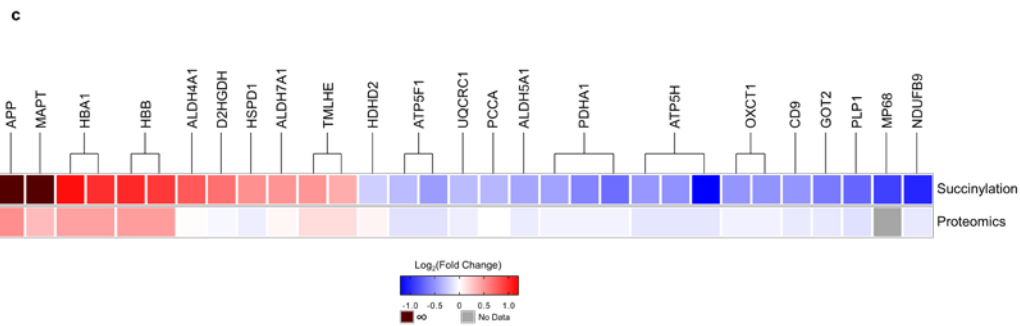
250 between AD and control subjects (Figures 3a, b). Ten succinylated peptides were increased while  
251 succinylation of 19 peptides declined in AD. Proteomic analysis of 20 samples in two cohorts (Figure 1c)  
252 showed that of the 4,678 identified proteins, 4,442 common proteins were quantifiable in both AD and  
253 controls (Figure 1d and Extended Data Figure 2a, b). A comparison of the succinylome with the  
254 proteome demonstrated little AD-related changes in protein levels of those succinylated proteins, and  
255 therefore the succinylation variations are most likely independent from the changes of the corresponding  
256 protein abundance (Figure 3c). The proteomic analysis showed that 81 proteins changed significantly  
257 (two-tailed Student's *t*-test,  $p < 0.05$  and  $|\log_2FC| > 0.25$ ). Eight proteins decreased in brains from AD  
258 patients while 73 protein levels were increased (Extended Data Figure 2a). In a recent large-scale  
259 proteomic scan, the protein abundance of PDHA, PDHB, and DLD were all decreased in AD, which is  
260 consistent with our finding, representing a decreased abundance of proteins in impaired mitochondrial

8

261 states<sup>17</sup>. Thus, changes in protein levels and succinylation may be important in AD. Relatively small fold  
262 changes found between control and AD brain samples, were probably due to a well-known ratio  
263 compression caused by the co-isolation of isobaric-labeled background ions in MS2-based TMT  
264 quantitative proteomics.

632

633



634

635

636 • Has acetylation been observed at A $\beta$  K16?

637

638 No one has reported this. We did not examine the acetylation since we focused on  
 639 succinylation. We anticipate acetylation is probably similar to succinylation with relatively  
 640 low occupancy rate. Therefore, an enrichment step using anti-acetylated Lysine antibody prior  
 641 to LC-MS/MS analysis would be required to confidently identify acetylation sites.

642

643 • The authors should provide High-resolution images for Extended Data Figure 3a-b?

644

645 We agree. Figures 3a and 3b have been replaced with high resolution figures. have  
 646 modified [this](#). Please see Extended Data

647

648 • Please provide a representative image for cleaved A $\beta$  6-29 fragment precursor ion  
 649 peaks in Extended Data Figure 3. The plot in Extended Figure 3f-g is confusing

650

651

652 **Table: MRM parameters with their retention time of targeted peptides and their**  
 653 **fragments:**

Compounds	RT (min)	Precursor ion m/z; (z)	Compounds parameter		Fragment ion m/z; b/y ions
			DP (V)	CE (V)	
Peptide 1	5.64	675.826; (4)	50	30	876.406; (y9)
Peptide 2	5.71	700.830; (4)	50	30	815.884; (b13 <sup>2+</sup> )
Fragment 1	1.2	668.805; (2)	50	40	788.321; (b7)
Fragment 2	6.21	691.851; (2)	50	30	876.406; (y9)
Fragment 3	3.87	479.545; (3)	50	25	560.210; (b5)

654 **Peptide 1: HDSGYEVHHQKLVFFAEDVGSNKG**

655 **Peptide 2: HDSGYEVHHQK(100.016)LVFFAEDVGSNKG**

656 **Fragment 1: HDSGYEVHHQK**

657 **Fragment 2: LVFFAEDVGSNKG**

658 **Fragment 3: HDSGYEVHHQK(100.016)**

659 **RT: retention time, DP: declustering potential, CE: Collision energy**

660 **Here, we performed both A $\beta$ 6-29 and cleavage fragments measurement. We analyzed**  
 661 **changes in these peptides incubated with rhADAM10 samples relative to the control**

662 peptides (without rhADAM10). For the fragments measurement, we set the results of  
663 fragments from the 24 hrs incubation with rhADAM10 as 100%.

664

665 The figure was also changed to make a new and more clear representation figure as Fig 3g  
666 where only cleaved A $\beta$  6-29 fragment precursor ion changes were shown with incubation  
667 time.

668 We modified the legend to figure 3 in extended data lines 25-26

25 f. Multiple Reaction Monitoring (MRM) parameters used in assay for quantitation with their retention time of  
26 targeted peptides and their fragments.

669

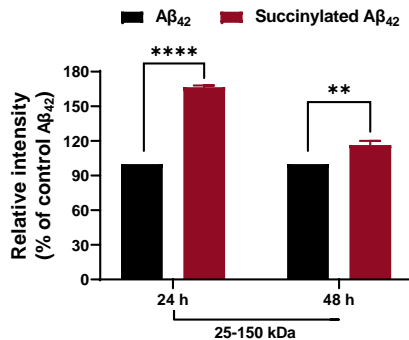
670

671 • What is “Percentage Change (%)” quantified in Figure 5f?

672

673 We modified Figure 5F to clarify the percentage change

674 The percent is simply the ratio with and without succinyl-lysine



675

676

677 • Page 13, Line 300: effects of what?

678

679 We have modified this. Please see Line 363-366 in the manuscript.

680

362 Previous studies indicate the removal of residue K311 in PHF6 abrogates fibril formation<sup>45</sup>, but the  
363 structural and functional implications of K311 succinylation are unknown. Thus, exploring the influence  
364

681 365 of tau succinylation on K311 may be important as we seek to develop a comprehensive understanding of  
682 366 its biological functions.

681

682

683

684 • Page 13, Line 302: “IF staining to compare the presence or abeta of succinylation



685 with that of tau” – What does this mean

686

687 We have modified this. Please see **Lines 363-376** in the manuscript.

368 **To characterize tau** succinylation in a transgenic mouse model of tangle formation, we used  
369 immunofluorescence staining to compare the presence of lysine succinylation within tau oligomers (T-22)  
370 <sup>46</sup> and phospho-tau (AT8) in the hippocampus from wild type and TgP301S mice. No phosphorylated tau  
371 and few tau oligomers were present in the brain of wild type mice (**Figure 6c, d and Extended Data**  
372 **Figure 4a, b**). The immunofluorescence signal of tau oligomers and phospho-tau was significantly  
373 augmented in the hippocampal region of TgP301S mice<sup>47</sup>. A parallel increase in lysine succinylation and  
374 oligomeric tau T-22 (green) and phospho-tau (green) was observed in tau mice compared to wild type  
375 animals (**Figure 6c, d and Extended Data Figure 4a, b**). Thus, tau succinylation associates with tau  
376 aggregates in a transgenic mouse model of tauopathy.  
377

688

689

690 • Page 13, Line 304: “but in 4-month-old TgP301S mice”.

691 o What occurred in 4-month-old TgP301S mice?

692

693 We have modified this. Please see **Lines 363-376** in the manuscript.

368 **To characterize tau** succinylation in a transgenic mouse model of tangle formation, we used  
369 immunofluorescence staining to compare the presence of lysine succinylation within tau oligomers (T-22)  
370 <sup>46</sup> and phospho-tau (AT8) in the hippocampus from wild type and TgP301S mice. No phosphorylated tau  
371 and few tau oligomers were present in the brain of wild type mice (**Figure 6c, d and Extended Data**  
372 **Figure 4a, b**). The immunofluorescence signal of tau oligomers and phospho-tau was significantly  
373 augmented in the hippocampal region of TgP301S mice<sup>47</sup>. A parallel increase in lysine succinylation and  
374 oligomeric tau T-22 (green) and phospho-tau (green) was observed in tau mice compared to wild type  
375 animals (**Figure 6c, d and Extended Data Figure 4a, b**). Thus, tau succinylation associates with tau  
376 aggregates in a transgenic mouse model of tauopathy.  
377

694

695

696

697

698

699 • Figure 6d: “d” is covered up by panel c

700

701 Corrected

702 • Please label Fig.6f-h with the peptide at hand (f=PHF6, g=S-PHF6, h=9:1 mix)

703

704 We have modified the legends Figure 6f-6h in the “figures and tables” (**lines 148-**

705 **150**)

148 **f-h.** Negative stain electron microscopy of *in vitro* polymerized PHFs after 24 hrs incubation. f: 50  
149  $\mu$ M PHF6; g: 50  $\mu$ M S-PHF6; h: 50  $\mu$ M mixture (PHF6:S-PHF6=9:1). White arrows denote paired  
150 helical filaments. Scale bar is 100 nm.

706

707

708 • Page 17, Line 394: “bot amyloidosis”?

709

710 We have modified this. Please see Line **461-463** in the manuscript.

461 Overall, our data represent the first report of the human brain succinylome and its implications for  
462 mitochondrial function as well as for the molecular pathogenesis of amyloidosis and tauopathy, two of the  
463 cardinal features of AD. We provide a rich resource for functional analyses of lysine succinylation, and

711  
712

713 • Page 8 Line 179: “Since no specific motifs for lysine succinylation in human cells  
714 have been reported” – They have been reported in HeLa cells in Weinert et al., Cell  
715 Reports (2013) <http://dx.doi.org/10.1016/j.celrep.2013.07.024>. Since they have  
716 reported previously, the authors should compare Motifs in AD brain to theirs, which  
717 upon first glance, don’t exactly match.

718

719 We modified this **Lines 223-226** in the manuscript.

223 Since no specific motifs for lysine succinylation in human tissues have been reported, a succinylation  
224 motif analysis of all 1908 succinylated peptides using Motif-X was performed. A total of five conserved  
225 motifs were identified (**Figure 2d**) in which non-polar, aliphatic residues including alanine, valine and

7

---

720 226 isoleucine surround the acceptor succinylated lysines. The succinylated lysine site analysis indicated a

721

722

723

724 • Page 16, Line 375: fix spelling errors

725

726 We removed this sentence based on another reviewer's comments.

727

728 **Reviewer #2 (Remarks to the Author)**

729

730 The manuscript “Succinylation Links Metabolic Reductions to Amyloid and Tau  
731 Pathology” compares succinylation in AD versus controls and surprisingly identifies  
732 AB and tau as targets that succinylated exclusively in AD. As a potential rationale for  
733 why these proteins may be succinylated in AD, the paper shows that mitochondrial  
734 dysfunction in cells leads to escape of proteins from the mitochondria that may be  
735 functioning in the succinylation of pathological AB and tau. In addition, the authors  
736 perform a series of biochemical experiments which suggest a way that succinylation  
737 may facilitate pathological AB and tau.

738

739 In the end, I think they can say that this paper demonstrates that a new protein  
740 modification is found on APP and tau that may correlate with AD status. In addition,  
741 some nice biochemical experiments raise a potential way in which this modification

742 can potentially influence the aggregation of these proteins. There is no evidence that it  
743 is actually functionally doing so in vivo, nor is there any evidence that is correlates  
744 with progression in humans, and the correlation in mice is the weakest part of the  
745 paper. It remains possible that the modification is simply a consequence of  
746 mitochondrial dysfunction in AD patients and the biochemistry not actually relevant  
747 to what is functionally occurring in vivo- perhaps because the succinylation moiety is  
748 rather large.

749 We think the large size of the succinylation suggests that it is biologically important.  
750 We have shown that succinylation alters the activity of TCA cycle enzymes, APP  
751 cleavage and tau function.

752

753 Nevertheless, even in this case, succinylation could serve as a marker, so it is still  
754 potentially relevant. Also, there is sufficient data to warrant following up the work.  
755 Thus, overall the findings are interesting.

756 However, the level of over-interpretation and over-blown claims are reckless and  
757 unwarranted, so the text needs major revisions. There are also some experimental  
758 concerns.

759 We have corrected all of the over-interpretation and over-blown claims.

760

761 **Major comments:**

762 The introduction is really short for an unknown topic and the Nature Communication  
763 format. A lot more needs to be added for the reader to understand succinylation,  
764 mitochondrial dysfunction, and AD pathology.

765

766

767

768

769 We redid the introduction according to reviewer's request **Lines 153-187** in the  
770 manuscript.

153 **Introduction**

154 Misfolded deposits of the amyloid beta peptide (A $\beta$ )<sup>1,2</sup> and the microtubule-associated protein tau  
155 (MAPT)<sup>3</sup> are pivotal pathological features in Alzheimer's disease (AD), wherein reduced brain regional  
156 glucose metabolism and synaptic density are correlated with the development of clinical cognitive  
157 dysfunction<sup>4</sup>. Preclinical research studies show that reduced glucose metabolism exacerbates learning  
158 and memory deficits concurrent with the accumulation of A $\beta$  oligomers and plaques<sup>5</sup>, and misfolded  
159 hyperphosphorylated tau<sup>6,7</sup>. However, the interrelationship(s) linking these keys but apparently disparate  
160 pathological processes remains unknown. While pro-amyloidogenic and/or immune-inflammatory genetic  
161 factors have played prominent roles in advancing our understanding of AD, more recent formulations  
162 have expanded the scope of molecular underpinnings of the disease<sup>8,9</sup>. Sims and colleagues coined the  
163 term "multiplex hypothesis of AD" to highlight the increasingly recognized shortcomings of the "amyloid  
164 hypothesis of AD"<sup>9</sup>.

166 Post-translational modifications (PTMs) of proteins provide an efficient and rapid biological regulatory  
167 mechanism that links metabolism to protein and cell functions. PTMs contribute to the functional  
168 diversity of proteomes without the formation of new proteins or a change in their abundance by covalent  
169 addition of functional groups that can alter protein charge, structure, and their interactions. Protein PTMs  
170 play a central role in the pathology of neurological diseases. The function of tau can be altered via its  
171 phosphorylation<sup>10</sup>, acetylation<sup>11</sup>, methylation<sup>12</sup> and O-GlcNAcylation<sup>13</sup>. Protein succinylation of lysine  
172 residues is a relatively novel PTM and changes the charge from positive to negative. The interactions of  
173 lysine succinylation and acetylation play an important role in metabolic pathways<sup>14</sup>. However,  
174 succinylation is poorly studied in the nervous system; our previous work demonstrated that lysine  
175 succinylation functionally modifies enzymes of energy metabolism<sup>15</sup>.

177 There is an increasing interest in defining the precise metabolic pathways involved in the pathogenesis of  
178 AD<sup>9,16-19</sup>. A significant correlation between reduced brain regional glucose metabolism and decreased  $\alpha$ -  
179 ketoglutarate dehydrogenase complex (KGDHC)<sup>20,21</sup> has been described in AD. Inhibition of KGDHC  
180 activity leads to a wide-spread reduction in regional brain post-translational lysine succinylation, for  
181 which the succinyl donor is presumably succinyl-CoA, both in yeast<sup>22</sup> and cultured neurons<sup>15,23,24</sup>. Studies  
182 of organisms deficient in NAD<sup>+</sup>-dependent desuccinylase sirtuin 5 (SIRT5)<sup>25</sup> provide evidence of the  
183 regulatory importance of succinylation in metabolic processes<sup>26-30</sup>. However, the role of succinylation in  
184 metabolic pathways of the human nervous system or in neurodegenerative diseases is unknown. Our  
185 report represents the first investigation of the human brain succinylome and its changes in AD. The  
186 results suggest that succinylation may link AD-related metabolic deficits to structural, functional, and  
187 pathological alterations involving APP and tau.

188

771

772

773 In Figure 3C, demonstrating that the change in succinylation is not just due to changes  
774 in protein levels is a critical point. The correlation that is shown, though weak, is a bit  
775 troubling. There is a new paper (Johnson et al, Nature Medicine 2020) focusing on  
776 proteomics in neurodegenerative diseases. I think it would be important to compare  
777 the changes in succinylated proteins to the changes in proteins presented in an  
778 independent paper, such as this one to make this point more convincing.

779

780 **The goal of our proteomics dataset is to serve as benchmark demonstrating if in**  
781 **very same 10 controls and 10 AD the abundance change of succinylated**  
782 **peptides/sites found in AD is due to change of protein abundance or not.**

783 **We took 22 significantly changed succinylated proteins from Johnson's paper**  
784 **and listed as below. Total protein levels vary by less than 12% (|Log<sub>2</sub>(FC)|<0.12)**

785  
786

**in Johnson's paper, and we think this also support our view that changes in succinylation levels are not based on changes in protein levels.**

Our paper					Johnson's paper			
UniProtKB ID	Gene name	Modifications	Succinylome (log <sub>2</sub> FC)	Proteome (log <sub>2</sub> FC)	Protein		Significance (Tukey P values)	Volcanoes log <sub>2</sub> (FC)
					Gene Name	ID	Control-AD	Control-AD
P6995	HBA1	1xSuccinyl [K4]	1.127	0.441	HBA2	P69905	0.075	-0.119
		1xSuccinyl [K5]	0.978					
P68871	HBB	1xSuccinyl [K]	1.013	0.460	HBB	P68871	0.208	-0.087
		1xSuccinyl [K9]	0.933					
P338	ALDH4A1	1xSuccinyl [K19]	0.783	0.010	ALDH4A1	P30038	0.963	-0.007
Q8N465	D2HGDH	1xSuccinyl [K6]	0.670	-0.044	D2HGDH	Q8N465	0.011	0.298
P189	HSPD1	1xSuccinyl [K3]	0.519	-0.075	HSPD1	P10809	0.014	0.038
P49419	ALDH7A1	1xSuccinyl [K2]	0.496	0.043	ALDH7A1	P49419-2	0.306	0.038
Q9NVH6	TMLHE	1xSuccinyl [K10]	0.492	0.161	TMLHE	Q9NVH6-8	0.909	0.037
		1xSuccinyl [K8]	0.391					
Q9HR4	HDHD2	1xSuccinyl [K7]	-0.225	0.054	HDHD2	Q9H0R4-2	-	-
P24539	ATP5F1	1xSuccinyl [K8]	-0.315	-0.144	ALDH5A1	P51649	0.086	0.037
		1xSuccinyl [K11]	-0.469					
P3193	UQCRC1	2xCarbamidomethyl [C3;C11];1xSuccinyl [K5]	-0.323	-0.079	UQCRC1	P31930	0.000	0.093
P5165	PCCA	1xSuccinyl [K6]	-0.343	-0.002	PCCA	P05165	0.452	-0.028
P51649	ALDH5A1	1xAcetyl [K];1xSuccinyl [K]	-0.420	-0.090	ATP5F1	P24539	0.178	0.034
P8559	PDHA1	1xSuccinyl [K9]	0.439	-0.061	PDHA1	P08559	0.000	0.114
		1xSuccinyl [K1]	-0.582					
		1xOxidation [M1];1xSuccinyl [K4]	-0.691					
O75947	ATP5H	1xSuccinyl [K]	-0.490	-0.120	ATP5H	O75947	0.012	0.058
	ATP5H	1xSuccinyl [K2]	-0.516					
	ATP5H	1xAcetyl [K5];1xSuccinyl [K]	-1.334					
P5589	OXCT1	1xSuccinyl [K3]	-0.501	-0.069	OXCT1	P55809	0.056	0.038
		1xSuccinyl [K5]	-0.507					
P21926	CD9	1xSuccinyl [K7]	-0.503	-0.101	CD9	P21926	0.858	-0.031
P55	GOT2	1xAcetyl [K];1xCarbamidomethyl [C8];1xSuccinyl [K]	-0.627	-0.105	GOT2	P00505	0.996	-0.004
P621	PLP1	2xCarbamidomethyl [C15;C23];1xSuccinyl [K13]	-0.719	-0.152	PLP1	P60201	0.203	0.097
P56378	MP68	1xSuccinyl [K7]	-0.902	NA		P56378	-	-

Q9Y6M9	NDUFB9	1xSuccinyl [K3]	-1.026	-0.110	NDUFB9	Q9Y6M9	0.000	0.114
--------	--------	-----------------	--------	--------	--------	--------	-------	-------

787

788

We inserted the following into the text (Manuscript lines 258-264)

258 patients while 73 protein levels were increased (Extended Data Figure 2a). In a recent large-scale  
 259 proteomic scan, the protein abundance of PDHA, PDHB, and DLD were all decreased in AD, which is  
 260 consistent with our finding, representing a decreased abundance of proteins in impaired mitochondrial

8

261 states<sup>17</sup>. Thus, changes in protein levels and succinylation may be important in AD. Relatively small fold  
 262 changes found between control and AD brain samples, were probably due to a well-known ratio  
 263 compression caused by the co-isolation of isobaric-labeled background ions in MS2-based TMT  
 264 quantitative proteomics.

789

790

791

792

793

794

795

796

797

798

799

800

801

802

803

804

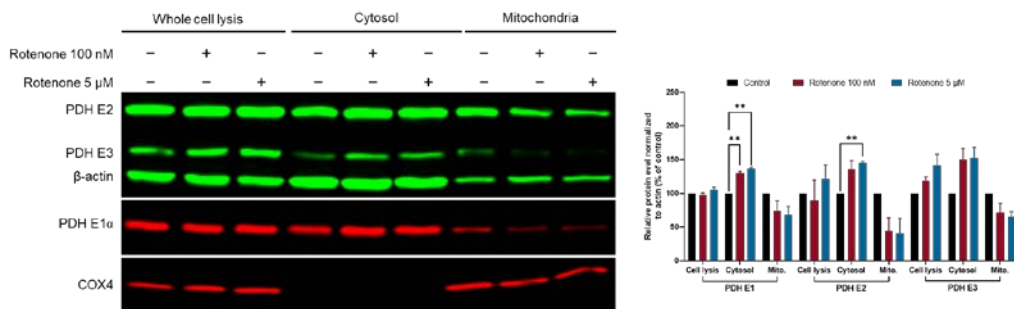
805

806

807

Figure 4A, without a control for protein loading, the overall change in levels of succinylation are meaningless. This is somewhat mitigated by the B-actin control for the individual proteins in B. However, is the B-actin from 4B also being used to normalize 4C? The B-actin should be shown on the same blot in C. Also, there probably should be controls showing that the fractions have been sorted intact.

**Figure 4A. The low cellular abundance of succinylation necessitates that we pull down the succinylated proteins with the anti-succinylation antibody. The IP only pulls down the succinylated proteins so we lose much of the beta actin which is not succinylated. No data suggests that the rotenone treatment will not affect the succinylation of beta-actin, so we think that beta-actin cannot be used as a loading control. In the Figure 4b we ran the two gels at the same time, so the loading samples were same, and we used the beta-actin from 4b to normalize 4c. We have repeated the whole assay and present the new results in the Figure 4c. See lines 65-68 in Figures and Tables.**



808

809

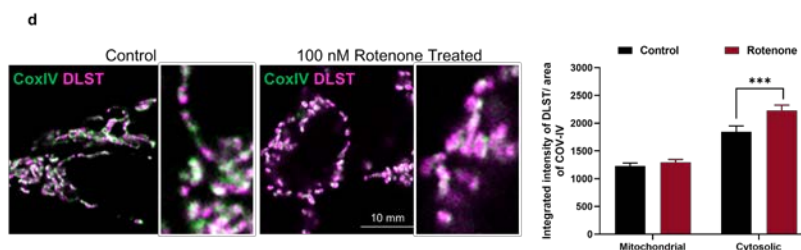
810  
811

65 c. The effects of rotenone (100 nM, 5  $\mu$ M/20 min) on the distribution of PDHC protein between  
66 mitochondria and non-mitochondrial fractions. The data from three different replicate experiments  
67 were expressed as the mean with error bars from SEM (n = 3, \*\*:  $p < 0.01$ , \*:  $p < 0.05$ , two-way  
68 ANOVA followed by Tukey's multiple comparisons test).

812  
813

In Figure 4D, it looks like there is less colocalization in the Rotenone treatment. Are the images reversed? In either case, the resolution of the images is too low to comment on the localization. Also, why was only 100nm Rotenone shown? What about 5uM? Overall, I probably buy the interpretation, but the data could be cleaned up.

819 Figure 4d is correct. But we revised it and inserted magnified regions on the right.  
820 Rotenone induces release of DLST into cytoplasm. In the control conditions, DLST  
821 (magenta) was concentrated inside mitochondria defined by COX-IV labeling (green).  
822 So, co-localization should maintain at the maximum level. After 1h of 100 nM  
823 Rotenone treatment, additional DLST labeling was out of the mitochondria and spread  
824 into the cytoplasm.



825  
826

Lines 69-73 in Figures and tables legend

69 d. Rotenone induces release of DLST into cytoplasm. In the control conditions, DLST (magenta)  
70 was concentrated inside mitochondria defined by COX-IV labeling (green). After 1h of 100 nM  
71 Rotenone treatment, additional DLST labeling was found in the cytoplasm. Inserts on the right are  
72 magnified regions. Magenta: DLST; Green: CoxIV; Error bars represent SEM deviation from the  
73 mean (n = 98 fields from 19 dishes, \*\*\*:  $p < 0.001$ , Tukey's multiple comparisons test).

827  
828

829  
830

831  
832

In Figure 5C and 5D, it would be nice to have an unaffected staining control to show that the change succinylation is specific.

834

835  
836

We apologize, but we are not sure that we have understood the question. Wild type mice are unaffected. Succinylation is a part of normal brain metabolism in the same way as acetylation or phosphorylation. The succinyloomics data show that hundreds of proteins are succinylated in normals. These results are the first visualization in the

838

839 brain of AD and tau mice.

840

841 We expect to see succinylation not only in A $\beta$  plaques or tangles, but also other  
842 proteins. Indeed, the hypersuccinylation in AD brains was in APP and tau. We have  
843 now discussed it in [lines 442-446 in the manuscript](#)

844 442 We show that transgenic mouse strains of either tauopathy or amyloidosis phenotype including plaques  
443 exhibit widespread increases in lysine succinylation, which is not exclusive to tau and APP, but parallels  
444 the formation of pathological species. While proteins in addition to tau or APP maybe altered, APP and  
445 tau are only succinylated in brains from AD patients, which suggests that increased tau and APP  
446 succinylation may play a role in the development of AD pathology. Thus, the modification of metabolism

845

846 More importantly, in WT there is a dramatic decrease in succinylation between 4  
847 months and 10 months. Why is this? Could this be due to mitochondrial changes in  
848 normal aging. This should probably be commented on in the discussion.

849

850 As discussed in detail above, the goal of this paper is to look at the association of  
851 succinylation to tau and APP. The effects of aging and the maturation of the pathology  
852 are beyond this paper and the ten-month data has been omitted.

853 However, for the review we offer the following speculative answer. This is a hard  
854 question to resolve because this is the first study investigating succinylation in the  
855 brain of APP or tau mice. Succinylation is a post-translational modification and  
856 several factors can regulate the balance between succinylation and desuccinylation.  
857 Our findings show that  $\alpha$ -ketoglutarate dehydrogenase (KGDHC) is a major  
858 succinylase in neurons. We have shown that brain KGDHC is not altered with age out  
859 to 30 months (Freeman, Nielsen et al. 1987) suggesting the change is not a reduction  
860 in succinylation. Whether aging may alter KGDHC migration to the cytosol has never  
861 been studied. The desuccinylases in brain remain unknown. A prominent paper by two  
862 of our co-authors have shown that sirtuin 5 (SIRT5) plays a central role in modulating  
863 heart metabolism and function (Sadhukhan, Liu et al. 2016).

864 SIRT5 is localized in the mitochondria and shows a weak deacetylase activity but  
865 a potent desuccinylase activity on lysine residues both *in vitro* and *in vivo* (Park, J  
866 2013 Mol. Cell; Du et al., 2011; Peng et al., 2011). The catalytic reaction involves the  
867 removal of a succinyl group from the lysine side chain of protein substrates, a process  
868 that consumes NAD<sup>+</sup> as a co-substrate and generates nicotinamide (NAM) and 2'-O-  
869 succinyl-ADP-ribose (Rardin MJ 2013 Cell Metabol). SIRT5 KO mouse embryonic  
870 fibroblasts display an increase in lysine succinylation but not acetylation (Du et al.  
871 2011 Science). We have used SIRT5 to desuccinylate enzymes such as the pyruvate  
872 dehydrogenase complex. We have also looked at succinylation in SIRT5 KO mice,  
873 which show a significant increase in succinylation levels in the liver while trivial  
874 changes were found in the brain. Liver succinylation, but not that in brain responds to  
875 fasting. Furthermore, the data from the AD samples suggest that different  
876 desuccinylases are likely important in the cytosol and mitochondria. We believe that  
877 our current results justify further studies on the regulation of succinylation in the brain  
878 during aging and in neurodegenerative diseases.



879           However, no studies reveal the relationship between aging and succinylation.  
880 Since aging is one of the most important risk factors for the development of AD, we  
881 think a separate deeper research need to investigate the age-dependent decrease in  
882 succinylation in future studies. We believe that 4-month-old mice clearly show an  
883 association between increased succinylation and disease progression (pathology).

884           The large increase in succinylation in 4-month-old Tg19959 mice agrees with our  
885 hypothesis: abnormal mitochondrial function promotes the release of KGDHC, which  
886 in turn, increases succinylation.

887

888 In the Tg mouse, there a lot more succinylation. However, there is also a decrease in  
889 succinylation from 4 to 10 months that is similar to WT, despite the fact that AB is  
890 definitely increasing in the Tg mice over this time period. Thus the two do not seem to  
891 be particularly well correlated. Also, is the level of succinylation increase in the Tg  
892 mice prior to appearance of AB? This should be checked because they already observe  
893 a dramatic increase in succinylation at 4 months when AB is first forming. Thus, it is  
894 possible that succinylation is changing in the Tg model well before this. Overall,  
895 though it is clear that succinylation is responding to the Tg, it is not at all clear that it  
896 correlates with the build up of AB. The exact same thing is true for tau in Figure 6.

897

898           The results clearly demonstrate that succinylation is clearly responding to the  
899 transgenic in both mouse models. This is consistent and suggest that increased  
900 succinylation at early stages of pathology development may be involved in the  
901 succinylation of multiple proteins. Howearly this occurs is of great interest to us but  
902 beyond the scope of this manuscript.

903           We used a pan succinylation antibody. We know a few hundred proteins are  
904 succinylated with multiple largely unknown functions. Our data suggest that  
905 succinylation could be involved in early stages of plaque or tangle formation to  
906 promote overproduction of A $\beta$  and inability of tau to bind to microtubules.  
907 We agree with the reviewer that it would be great to know the role of succinylation  
908 from development to old age, as well as well as pathology. Interestingly, the activity  
909 of KGDHC peaks at about 30 days post-natal, when the cortex reaches final  
910 maturation. We hope that this paper will encourage examination of succinylation  
911 under all these conditions, but we feel it is beyond the scope of this paper.

912

913 The fact that the same phenomenon is true in both AB and Tau models is perhaps even  
914 more disconcerting, because it suggests that the phenomenon is not specific. That is to  
915 say, the overall increase in succinylation seems to occur regardless of the pathological  
916 insult, which does not cause mitochondrial dysfunction exactly the same way in the  
917 two models, and the timing of that pathological insult, which is not exactly the same  
918 in the two mouse models. This should at least be discussed. In particular, it would be  
919 nice to know how the changes in succinylation relate to the changes in mitochondrial  
920 dysfunction in the two Tg models in their hands. Overall, the data supports a change  
921 in succinylation that occurs in the two pathological models, but does not support the  
922 conclusion that the change in succinylation truly correlates with pathology. At a

923 minimum, this should be clearly noted in the results and discussion.

924

925 The reviewer clearly states an important issue that speaks to the essence of the  
926 whole paper. Since the extensive data required to answer is not available, we have  
927 now discussed it in the revised version, as suggested by the Reviewer. As indicated by  
928 the reviewer, the data shows and increase in succinylation in two models of pathology.

929 One question is whether precise changes in mitochondria are required to alter  
930 succinylation of tau and APP. The precise changes in mitochondria from AD patients  
931 or in animal models of pathology are unknown, so it hard to exactly model the precise  
932 mitochondrial changes. The data on relation of the mitochondrial changes to  
933 succinylation in brain is limited to our published studies, which show that  
934 mitochondrial succinylation decreases rapidly in response to many altered metabolic  
935 states and increases in response to others. In addition, the relation to cytosolic  
936 succinylation has never been explored. The results in this manuscript suggest new  
937 experiments to test the relation of select metabolic insults to the release of KGDHC  
938 from the mitochondria and succinylation of specific cytosolic proteins.

939 The fascinating question of specificity may not be only related to the kind of  
940 inhibition but to the severity and duration of the inhibition. The current studies  
941 provide justification for exploring cytosolic succinylation in response to a variety of  
942 insults. Future experiments could also determine which metabolic insults lead to  
943 release of KGDHC and the succinylation of specific cytosolic proteins. The coupling  
944 between succinylation due to unique modifications of the mitochondria or the severity  
945 of the sick mitochondria must be different in diverse regions of the brain or maybe,  
946 the sensitivity is different.

947

948 We have modified the discussion by adding the following paragraph Please see Lines  
949 452-459 in the manuscript.

452 Lysines are highly modified residues. Understanding the relationship of succinylation to the other PTM is  
453 critical to a complete understanding of its role in AD pathology. A direct comparison is practically  
454 difficult because each PTM requires a different enrichment strategy. Lysine311 has been associated with  
455 ubiquitination, dimethylation, and acetylation in transgenic mice<sup>59</sup>, but not human brains. Our  
456 experiments raise the question of whether precise changes in mitochondria are required to alter  
457 modification of specific proteins. Succinylation appears directly linked to KGDHC and mitochondria.  
458 Whether other post-translational modifications are also linked to mitochondrial dysfunction remains to be  
459 determined.

950

460

951

952 There is a slight concern that the peptide used for the tubulin polymerization assay is  
953 succinylated throughout, when they only detected succinylation at K311 in AD.

954 Nevertheless, the loss of Tau polymerization function is impressive. This should be  
955 more clearly stated and used to qualify the interpretation. This is particularly true  
956 because the succinylation moiety is quite large. The authors should definitely discuss  
957 how such a large modification could affect proteins and the biochemical assays that  
958 they perform on them. The decrease in Tau-tubulin interactions is more convincing,  
959 particularly because they also performed this assay with Tau only succinylated at

960 K311.

961

962 Performing the site-specific succinylation of the lysine 311 in vitro is very  
963 difficult, in the present case of the K19 peptide has more than one lysine. It is  
964 also quite difficult to synthesize a 99-aa length peptide by using peptide  
965 synthesis techniques. Finally, we decided that we first use a pan-succinylation  
966 on K19 peptide and tested its ability on tubulin polymerization assay. Then  
967 we tested if the succinylation of K311 is sufficient to specifically decrease  
968 tau-tubulin interactions by 1H saturation transfer difference (STD) NMR.  
969 We added the following comment to the discussion in the manuscript **lines**  
970 **425-428**

971

425 dysfunction<sup>54-57</sup>, thereby contributing to progression of amyloidosis and tauopathy. It is not perhaps  
426 surprising that succinylation would have such an effect since it increases the size of the lysine side chain  
427 considerable and could lead to steric clashes, as well as reversing the charge of the side chain (Extended  
428 Data Fig. 4j).

972

429

973

974

975

976 Overall, it is possible that succinylation is simply a consequence of mitochondrial  
977 dysfunction, and not necessarily functional in AD. To mitigate this, I think it might be  
978 good to provide some additional negative controls if possible. ie do AB and tau  
979 accumulate any other post-translational modifications that might just be due to  
980 disruption of mitochondria or the abnormal appearance of the pathological versions of  
981 these proteins in the cytosol.

982

983 As indicated, the data suggests that the succinylation is a consequence of  
984 mitochondrial dysfunction. We show succinylation is altered in AD at very specific  
985 sites. We show that succinylation causes pathologically important changes in tau and  
986 APP. How the succinylation interacts with other modification is critical but beyond  
987 this study. Our studies show that the link of mitochondria and acetylation is very  
988 different than the link mitochondria and acetylation. We added the following to the  
989 **manuscript (Lines 455-459)**

455 ubiquitination, dimethylation, and acetylation in transgenic mice<sup>59</sup>, but not human brains. Our  
456 experiments raise the question of whether precise changes in mitochondria are required to alter  
457 modification of specific proteins. Succinylation appears directly linked to KGDHC and mitochondria.  
458 Whether other post-translational modifications are also linked to mitochondrial dysfunction remains to be  
459 determined.

990

460

991

992

993 There is a lot of wild speculation:

994

995 Example: The last sentence of the abstract is wildly overexaggerated- While there is a  
996 possibility that succinylation could contribute to pathologies in AD, the data presented

997 in the manuscript certainly are not by themselves enough to even raise the possibility  
998 that succinylation must be addressed therapeutically for meaningful clinical benefit  
999

1000 Example: The last sentence of the introduction is overstated and unnecessary

1001  
1002 We have completely re-written the abstract (manuscript lines 118-129) and  
1003 introduction (manuscript lines 153-187). We avoided wild speculations.  
1004

1005 Example: line 311- this reflected a potential existence of succinylation-  
1006 phosphorylation switch as in the case of acetylation- the paper provides absolutely no  
1007 evidence for this. This could be speculated about in the discussion, but is completely  
1008 inappropriate for the results section.  
1009

1010 We modified the manuscript in two places

1011 Lines 423-425

1012

423 altered cerebral metabolic function in AD as the disease progresses. Other PTMs, such as ubiquitination,  
424 acetylation and phosphorylation, have been recently shown to affect amyloid degradation<sup>54,55</sup> and tau  
1013 425 dysfunction<sup>54,57</sup>, thereby contributing to progression of amyloidosis and tauopathy. It is not perhaps  
1014

1014

1015 Lines 461-466

461 Overall, our data represent the first report of the human brain succinylome and its implications for  
462 mitochondrial function as well as for the molecular pathogenesis of amyloidosis and tauopathy, two of the  
463 cardinal features of AD. We provide a rich resource for functional analyses of lysine succinylation, and  
464 facilitate the dissection of metabolic networks in AD. The current studies also lay the foundation for  
465 future investigation into the crosstalk between different PTMs, including acetylation, phosphorylation,  
1016 466 ubiquitination and succinylation associated with AD and tau pathology. The discovery that succinylation  
1017

1017

1018 Example: line 283- taken together the accumulated data strongly suggest that  
1019 succinylation of K678 might lead to an early-onset enhanced generation,  
1020 oligomerization and plaque biogenesis, consistent with the effects of known genetic  
1021 disease mutations at this site. While the data suggest a potential was for succinylation  
1022 to affect A $\beta$  cleavage, there is no functional evidence that it does so.  
1023

1023 We have modified this. Please see Line 347-350 in the manuscript.

347 at t = 24 or 48 hrs (Figure 5g). These data demonstrate that succinylation of K612 of APP is a key  
348 molecular event that promotes A $\beta$  oligomerization. Taken together, our findings suggest that  
349 succinylation of K678 might lead to early-onset and/or enhanced generation, oligomerization and plaque  
350 deposition of A $\beta$ , consistent with the effects of known genetic disease mutations at this site<sup>37,42</sup>.  
1024 351

1024

1025

1026

1027

1028

1029 Example line 369: Notably, these results demonstrate for the first time that

1030 succinylation is the key link between the signature metabolic reductions and amyloid

1031 plaques and neurofibrillary tangles in AD.- Again, although this is possible, there is  
1032 absolutely no evidence for this in the manuscript

1033

1034 We have modified this. Please [see Line 419-425](#) in the manuscript.

1035

419 Our study provides a system level view of the human brain succinylome in metabolic process, particularly  
420 in mitochondria, and reveals a dramatic alteration on succinylation in AD. Our results demonstrate for the  
421 first time that succinylation is a key link between the signature metabolic reductions and A $\beta$  plaques as  
422 well as NFTs in AD. We show that changes in protein succinylation, as a molecular signal, correlate with  
423 altered cerebral metabolic function in AD as the disease progresses. Other PTMs, such as ubiquitination,  
424 acetylation and phosphorylation, have been recently shown to affect amyloid degradation<sup>54,55</sup> and tau  
425 dysfunction<sup>54,57</sup>, thereby contributing to progression of amyloidosis and tauopathy. It is not perhaps

1036

1037

1038 Example line 371: The current results reveal that varied in protein succinylation, as a  
1039 molecular signal, correlates with altered cerebral metabolic function in AD as the  
1040 disease progresses.- While there may be a small amount of evidence of this in the  
1041 mice data (if you compare to previous analysis of the mice strains employed and you  
1042 ignore the fact that the mouse data don't truly correlate), they do not show this on  
1043 their own and they certainly did not examine succinylation across the progression of  
1044 the disease in human cases, so this statement is not warranted.

1045

1046 This phrase has been deleted in the red-write of the section.

1047

1048 Minor comments:

1049

1050 I really don't understand the math in figure 1, and there is no description of what the  
1051 29 proteins in B are? Are they differential between AD vs control, as in D? And what  
1052 percentages are up vs down? An effort should be made to make the numbers more  
1053 clear.

1054

1055 We provide Fig.1 as a schematic workflow for a better understanding of the  
1056 data analysis. Since we did independent cohorts, we would like to explain that  
1057 we have high reproducibility in two cohorts (both succinylome and proteome).  
1058 29 proteins in B are those succinylated proteins having the sites that have  $\log_2 |\text{fold change}| > 0.25$   
1059 and p value  $< 0.05$  found in succinylome analysis of human brain. Yes, they are differential between  
1060 AD vs control. Out of 29 succinylated proteins, the abundance of 12 succinylated sites of the  
1061 proteins is increased while the abundance of the rest 17 sites is decreased. The fold change of  
1062 the succinylome and proteome were provided in Supplementary Table 4 and  
1063 6, also they have been visualized in Fig 3a and Extended Data Figure 2.

1064

1065

1066 The antibody used in extended data figure 3B needs to be clearly shown in the figure.

1067

1068 The antibody names are on the figure, so we are not sure of the question. We did  
1069 improve the resolution.

1070 All antibodies are in the methods section.

1071

1072 In extended figure 3F and G, why is the full length protein so different when the  
1073 production of the cleavage products remains the same? Perhaps I don't quite  
1074 understand the assay, but this should be clarified.

1075

1076 We performed both A $\beta$ 6-29 and cleavage fragments measurement. The cleavage  
1077 products along with remaining A $\beta$ 6-29 were quantitatively determined by LC-MS/MS  
1078 analysis. Specifically, an MRM (multiple reaction monitoring) method was used in  
1079 the mass spectrometric assay to measure the amount of normal and succinylated  
1080 peptides of A $\beta$ 6-29 changes with time after incubated with rhADAM10. The extended  
1081 Fig 3g and 3h was changed into a new and clear Fig 3g where the changes with time  
1082 was shown only for the peptides, not for the fragments produced per reviewer's  
1083 suggestion.

1084 The label of the graph in 5F needs to be clarified as percentage change from 0 hrs.

1085

1086 During the 0 hrs, the intensity will be 0, and it can't be the denominator.

1087

1088 Line 309 refers to succinylated tau. However, it should refer to succinylation in the  
1089 Tau Tg mice. There is a big difference.

1090

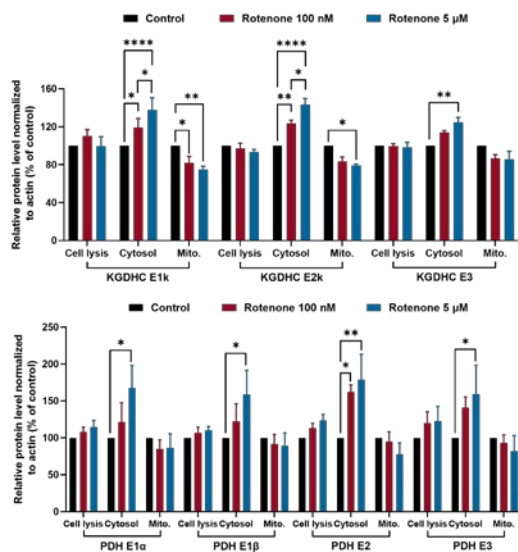
1091 This phrase has been deleted.

1092

1093 The Y-axis in Figure 4B says evel instead of level

1094

1095 We corrected the figure. We also modified the legend to Figure 4b in Figures and  
1096 Tables line 61-64.



1097

61 b. The effects of rotenone (100 nM, 5  $\mu$ M/20 min) on the distribution of KGDHC protein between  
62 mitochondria and non-mitochondrial fractions. The data from three different replicate experiments  
63 were expressed as the mean with error bars from SEM (n = 3, \*\*\*\*:  $p < 0.0001$ , \*\*:  $p < 0.01$ , \*:  $p$   
64 < 0.05, two-way ANOVA followed by Tukey's multiple comparisons test).

1098

1099

1100

The Tg19959 and P301S mouse models need to be defined for the reader.

1101

We added more detail to [the methods section lines 498-506](#) The references were also included in the main text.

1102

398 All the experiments were carried out in four and ten-month-old transgenic mouse models of AD.  
399 Tg19959 mice (that overexpress a double mutant form of the human amyloid precursor protein) were  
400 obtained from Dr. George Carlson (McLaughlin Research Institute, Great Falls, MT, USA). Tg19959  
401 mice were constructed by injecting FVB X 129S6 F1 embryos with a [cosmid](#) insert containing  
402 human APP<sub>695</sub> with 2 familial AD mutations (KM670/671NL and V717F), under the control of the  
403 hamster [PrP](#) promoter.<sup>25</sup> P301S (PS19, that overexpress the human tau gene harboring the P301S

12

404 mutation) transgenic mice were purchased from The Jackson Laboratory (Bar Harbor, ME, USA). The  
405 transgenic mice used in this study express the human pathogenic mutation P301L of tau together with the  
406 longest human brain tau isoform (htau40) under control of the neuron-specific mThy1.2 promoter.<sup>26</sup>

1103

1104

1105

Line 194 should say from rather than form

1106

We have modified [Lines 235-238](#) of the manuscript.

235 Completion of the human brain [succinylome](#) and global proteome analyses allowed direct comparison  
236 between brains from controls and AD patients. Without enrichment of [succinylated peptide](#) in global  
237 proteome data, the number of [succinylated peptides](#) identified is 0.13% total peptides for cohort 1 and  
238 0.28% for cohort 2. The notable difference in ratio of [succinylated peptides](#) over total peptides between

1107

1108

1109

PDHA1 is mentioned with no context

1110

We modified the text lines [205-208](#)

205 73% (229/314) of the [succinylated proteins](#) were mitochondrial (**Figure 2b**). The pyruvate dehydrogenase  
206 complex E1 component subunit alpha (PDHA1), which links glycolysis to the TCA cycle, was  
207 significantly [succinylated](#). The eight enzymes of the TCA cycle located within the mitochondrial matrix  
208 and their multiple subunits were also extensively [succinylated](#). Furthermore, succinylation of proteins was

1111

1112

1113

1114

Line 257: paralleled should be parallel

1115

We have modified the text on lines 306-310

306 which carries the human APP with two mutations (Figure 5c and Extended Data Figure 3a). Double  
307 immunofluorescence staining with antibodies to pan-lysine-succinylation and to A $\beta$  oligomers (NU-4)<sup>35</sup>  
308 or to A $\beta$  plaque ( $\beta$ -Amyloid, D3D2N) revealed an early increase in lysine succinylation that appeared to  
309 parallel oligomer formation and subsequent plaque deposition in the hippocampus. These findings suggest  
310 that the APP succinylation might be involved in A $\beta$  oligomerization and plaque formation *in vivo*.

1116

1117 The logic in line 266-267 needs to be better spelled out for the reader with regard to  
1118 the competition of the two enzymes. Likewise in line 271, ADAM10 needs to be  
1119 introduced as a secretase for the reader.

1120

1121

A: We have modified Lines 314-330 in the manuscript

314 The generation of the A $\beta$  is a highly regulated process by the secretases.  $\beta$ -secretase initiates the  
315 amyloidogenic pathway, while  $\alpha$ -secretase is part of the non-amyloidogenic pathway bisection the A $\beta$   
316 domain and thereby inhibiting the formation of A $\beta$ . In subsequent experiments, we tested the relationship  
317 between succinylation and APP processing by the secretase enzymes. K612-L613 is the APP  $\alpha$ -secretase  
318 bond, and missense mutation at K612N produces early onset AD<sup>37</sup>. Furthermore, global proteomics  
319 showed an increase in  $\beta$ -secretase (BACE1) abundance of 31% in AD brains compared to controls  
320 (Supplementary Data Table 6), while no changes occurred for either  $\alpha$ -secretase or the sirtuins (SIRT)  
321 family (Extended Data Figure 2c). Seyfried et al., quantified a total of 2,745 proteins in two regions  
322 (dorsolateral prefrontal cortex (FC, Brodmann Area 9) and precuneus (PC, Brodmann Area 7)  
323 were quantified. The number that overlapped was about 2,332 proteins (85.3%) compared with  
324 our data (4442 proteins from 10 controls and 10 AD, Brodmann area 44/45).

325

326 The four ADAM family members identified in that paper were also identified in our proteome.  
327 The protein level of ADAM 10, 22, 23 did not change in that paper nor our data, while ADAM11  
328 showed a similar decrease in the two cases. SIRT2 and SIRT5 levels did not vary<sup>38</sup>. Further,  
329 protein levels of SIRTs do not necessarily reflect activity, which are often regulated by substrates  
330 and post-translational modifications.

1122

1123

1124 The S and C labels in 5C need to be defined in the main figure.

1125 We presume the reviewer means the label in Figure 5e. We have modified the text in  
1126 Figures and Legends lines 100-108

100 e. Succinylation blocks  $\alpha$ -cleavage. Peptides were incubated for 24 h with or without rhADAM10.

101 Peak area ratio values were calculated and are shown relative to corresponding controls without

102 rhADAM10. Each sample was run in triplicate and data were expressed as the mean with SEM

103 (\*\*:  $p < 0.01$ , two-way ANOVA followed by Bonferroni's multiple comparisons test; except for

104 one sample from the group of succinylated peptide without rhADAM10 was damaged).

105 f. Western blot analysis of succinylated and control A $\beta$ <sub>42</sub> from aggregation assay showed that the

106 succinylation generates more oligomerized A $\beta$  even after a long incubation. The data from two

107 different replicate experiments were expressed as the mean with error bars from SEM (\*\*\*\*:  $p <$

108 0.0001, \*\*:  $p < 0.01$ , two-way ANOVA followed by Bonferroni's multiple comparisons test).

1127

1128

1129

1130



1131 **Line 302: abeta should be absence?**

1132 We modified the manuscript **Lines 368-370**

368 To characterize tau succinylation in a transgenic mouse model of tangle formation, we used  
369 immunofluorescence staining to compare the presence of lysine succinylation within tau oligomers (T-22)  
370 <sup>46</sup> and phospho-tau (AT8) in the hippocampus from wild type and TgP301S mice. No phosphorylated tau

1133

1134

1135 **Line 371: varied should be variation**

1136 We modified the manuscript lines **Lines 419-422**

419 Our study provides a system level view of the human brain succinylome in metabolic process, particularly  
420 in mitochondria, and reveals a dramatic alteration on succinylation in AD. Our results demonstrate for the  
421 first time that succinylation is a key link between the signature metabolic reductions and A $\beta$  plaques as  
422 well as NFTs in AD. We show that changes in protein succinylation, as a molecular signal, correlate with

1137

1138

1139 **Line 375: involvon should be involved in?**

1140 We removed this sentence based on another reviewer's comments.

1141

1142 **Line 394: bot should be both?**

1143 We have modified the text lines **430-432**.

430 The mechanisms and control of both non-enzymatic and enzymatic succinylation by cellular  
431 succinyltransferases and desuccinylases are unknown. Our data clearly demonstrate that impairing  
432 mitochondrial function decreases mitochondrial succinylation and promotes succinylation of specific non-

1144

1145

1146 **Line 385: The decline in succinylation of mitochondrial proteins suggests that**  
1147 **activation of descuccinylases- The alternative, that there could be a failure to maintain**  
1148 **succinylation levels, should be mentioned.**

1149 We modified **the text lines 427-437**

430 The mechanisms and control of both non-enzymatic and enzymatic succinylation by cellular  
431 succinyltransferases and desuccinylases are unknown. Our data clearly demonstrate that impairing  
432 mitochondrial function decreases mitochondrial succinylation and promotes succinylation of specific non-  
433 mitochondrial proteins by altering the distribution of succinyltransferases from the mitochondria to  
434 cytosol. Precedent for this concept is provided by results showing that translocation of the DLST subunit  
435 of KGDHC to the nucleus increases histone succinylation<sup>35</sup>. Rotenone induces translocation of PDHC

13

---

436 from mitochondria to other cellular compartments<sup>38</sup>. The decline in succinylation of mitochondrial  
437 proteins, appears due to a failure in maintaining succinylation levels, and may suggest that activation of  
438 desuccinylases (e.g., Sirtuins) or general increases in NAD<sup>+</sup> should be reconsidered. The large increase  
439 in succinylation in 4-month-old Tg19959 mice agrees with our hypothesis, in which abnormal  
440 mitochondrial function in AD promotes the release of KGDHC and subsequent increases succinylation.  
441

1150

1151

1152

1153 **The manuscript should also be edited for grammar.**

1154 Examination of the marked copy shows that we carefully changed the manuscript  
1155 including changes in grammar.

1156

1157 **Reviewer #3 (Remarks to the Author)**

1158 The authors have investigated the potential role of succinylation and Amyloid and Tau  
1159 pathology using brain tissue from AD cases and controls. They analysed brain tissue  
1160 cell lysate proteomes using 10 plex TMT. They also analyzed the same 10 controls  
1161 and 10 AD patients' brain samples Succinylome using Cell Signaling Tech IP-MS kit  
1162 and ran LCMS of the PTM enriched tryptic peptides.

1163

1164 **Comments:**

1165

1166 **1- In addition to bioinformatics analysis of succinylome IP-MS data it would be**  
1167 **useful to analyze and show the biological significance of whole tissue lysate 10- plex**  
1168 **TMT data as well. It would be useful to cover the global proteome analysis done**  
1169 **which may be relevant to disease pathogenesis in addition to the succinylome targeted**  
1170 **concept.**

1171 **We agree that the whole proteome analysis relevant to disease progression is**  
1172 **important to find out some potential biomarkers in AD. It would be too much to add**  
1173 **to this paper since the primary focus of this manuscript is the succinylation. We had**  
1174 **added the global proteome data to highlight the changes in succinylation**  
1175 **peptides/sites of the protein are unrelated to any changes in protein abundance in AD**

1176 compared with healthy samples.  
1177 We have added references to recent papers that report the complete proteome.  
1178 The whole proteome is posted online if the reviewer wants to see but it would change  
1179 the focus from this paper. We have added several mentions of the Johnson et al paper  
1180 and the focus of that paper is proteomics.

1181

1182 2- It is important to pinpoint sites of protein succinylation. Succinylome localization  
1183 shown in Figure 1a is not clear.

1184

1185

1186 In Fig 1a, we only present a schematic workflow for succinylome studies where it is  
1187 shown that succinylation on lysine residues were immune- enriched prior to nano  
1188 scale LC-MS/MS analysis.

1189

1190 3- The authors considered impaired mitochondrial function resulted in succinylome  
1191 localization to be pushed out of mitochondria to cytosol by leakage (Figure 4, Line  
1192 244-247), however, whole tissue lysate mass spec succinylome data suggested an  
1193 overall decrease in AD. These findings need to be reconciled

1194

1195 Please remember the majority of succinylated proteins in AD brains are in the  
1196 mitochondria. Mitochondrial damage results in impaired enzyme activity at the total  
1197 succinylation level. So, our succinylome supports this idea. The leakage of  
1198 mitochondrial enzymes, this will result in unexpected modifications on some proteins  
1199 (APP or Tau) that have no or less access to these succinyl transferases.

1200

1201

1202 4- The K687 site in the middle of APP is the interaction site of  $\alpha$ -secretase and  
1203 the cleavage was inhibited when the K was succinylated in vitro (Figure 5e).

1204 However, it cannot be assumed that succinylated A $\beta$  has more aggregation property  
1205 since the comparison shown in Figure 5f did not show a significant difference. The  
1206 AD patient succinylome mass spec identified succinylated K687 peptide (S Table 5).  
1207 It is not clear whether the succinylome IP conditions favored solubilizing Abeta  
1208 plaque and protofibril oligomers.

1209

1210

1211 We think Figure 5f shows a significant difference. Compared with the intensity, we  
1212 found succinylation generates more oligomerized A $\beta$ . During the MS protocol, we  
1213 broke the protein complex molecule into peptides, and then we did the IP. So, during  
1214 the IP processing, there is no big complex (no plaque nor tangle), so it is unlikely  
1215 there will be such a preference. The anti-Succ-K antibody recognizes the exposed  
1216 succinylated lysine. As described in the methods, all of the exposed succinyl-lysines is  
1217 expected to be effectively pull down by the antibody in our enrichment steps.

1218 However, if this antibody pull down would favor certain sequence dependent species  
1219 of AB or tau is unknown. Since we really compare the same succinylated

1220 peptides/sites among all samples, the possible bias introduced by variable peptide  
1221 sequence would not affect our quantitative results between AD and health cases.  
1222

1223 **5- APOE4 mutation is a risk factor of AD. Does the proteomics data reveal mutation**  
1224 **in the 20 patient brains analyzed?**  
1225

1226 No, the global proteomic data cannot reveal the mutation sites of APOE4. In shotgun-  
1227 based mass spectrometry analysis, we were only able to identify proteins by a series  
1228 of tryptic peptides. Normally, these detected tryptic peptides cannot cover the whole  
1229 protein sequence. When the software searches acquired spectra against a database  
1230 containing a particular protein, it can only match the exact same sequences of the  
1231 tryptic peptide of the native protein in the database. If mutations occur at certain  
1232 sites, the database searching software cannot recognize these peptides unless the  
1233 mutated protein sequence was added to the database prior to database search. We have  
1234 verified the APP data. The missing succinylation of APP in only one case was not due  
1235 to a mutation.  
1236

1237 Freeman, G. B., P. E. Nielsen and G. E. Gibson (1987). "Effect of age on behavioral and enzymatic  
1238 changes during thiamin deficiency." Neurobiology of Aging **8**(5): 429-434.

1239 Kaden, D., A. Harmeier, C. Weise, L. M. Munter, V. Althoff, B. R. Rost, P. W. Hildebrand, D. Schmitz,  
1240 M. Schaefer, R. Lurz, S. Skodda, R. Yamamoto, S. Arlt, U. Finckh and G. Multhaup (2012). "Novel  
1241 APP/A $\beta$  mutation K16N produces highly toxic heteromeric A $\beta$  oligomers." EMBO Molecular Medicine  
1242 **4**(7): 647-659.

1243 Morris, M., G. M. Knudsen, S. Maeda, J. C. Trinidad, A. Ioanoviciu, A. L. Burlingame and L. Mucke  
1244 (2015). "Tau post-translational modifications in wild-type and human amyloid precursor protein  
1245 transgenic mice." Nature Neuroscience **18**(8): 1183-1189.

1246 Sadhukhan, S., X. Liu, D. Ryu, O. D. Nelson, J. A. Stupinski, Z. Li, W. Chen, S. Zhang, R. S. Weiss, J.  
1247 W. Locasale, J. Auwerx and H. Lin (2016). "Metabolomics-assisted proteomics identifies succinylation  
1248 and SIRT5 as important regulators of cardiac function." Proceedings of the National Academy of  
1249 Sciences **113**(16): 4320.

1250 Seyfried, N. T., E. B. Dammer, V. Swarup, D. Nandakumar, D. M. Duong, L. Yin, Q. Deng, T. Nguyen,  
1251 C. M. Hales, T. Wingo, J. Glass, M. Gearing, M. Thambisetty, J. C. Troncoso, D. H. Geschwind, J. J. Lah  
1252 and A. I. Levey (2017). "A Multi-network Approach Identifies Protein-Specific Co-expression in  
1253 Asymptomatic and Symptomatic Alzheimer's Disease." Cell Systems **4**(1): 60-72.e64.

1254 Tracy, T., K. C. Claiborn and L. Gan (2019). Regulation of Tau Homeostasis and Toxicity by Acetylation.  
1255 Tau Biology. A. Takashima, B. Wolozin and L. Buee. Singapore, Springer Singapore: 47-55.  
1256

## REVIEWER COMMENTS

Reviewer #1 (Remarks to the Author):

The authors have done an excellent job addressing the previous critiques.

Reviewer #2 (Remarks to the Author):

Overall, the manuscript is improved. However, the following points still need to be addressed:

In 5C, the authors need to perform dual IF with another protein that is unchanged in the same image that they are staining for succinylation, so that we can be sure that the changes are not just due the one sample staining a bit better, and therefore being brighter. The other Ab that is used currently is for AB oligomers is nice for co-localization, but is uninformative for comparing levels of succinylation. This is standard control when trying to quantify changes in levels from staining.

I think that removing the 10 month time point from figure 5 is not appropriate, as it risks just hiding data that potentially does not fit their model. I think the authors need to put the 10 month time point back and add a potential explanation. Personally, I don't find the argument that they provide in the rebuttal to be that compelling, but that is for the reader to judge. Removing the data completely does not provide the reader the opportunity to do so, and is therefore inappropriate.

Along these lines, the authors response to the critique that the changes in succinylation in the transgenic mouse model don't correlate very well with the build up of AB is insufficient. While it is clearly true that succinylation is responding to the transgene, the fact that succinylation doesn't correlate well with the build up of AB provides a major caveat to the interpretation. If the authors do not wish to perform additional experiments to address why this might not be the case, they need to at least acknowledge the caveat and discuss it. As is, the authors seem to be saying that because the transgene causes a change in succinylation, it doesn't matter that that change does not seem to correlate well with the build up of pathological AB. When actually, this disconnect raises the clear possibility that AB build up and succinylation are not directly related. This doesn't mean that their results are not potentially interesting or important, but it does need to be acknowledged and discussed so that the reader is not left with an incorrect view of the data.

Similarly, the discussion that has been added to supposedly address the review point about the changes occurring in models of both AB and Tau is also not sufficient. The authors need to specifically discuss why succinylation may be changing in both AB and Tau models, despite the models being different. The new discussion that was added does not address this directly at all, and so only serves to further muddy the interpretation.

The above points are especially important, considering the revised abstract that states "Our results reveal a tight relationship linking lysine succinylation status and AD-associated proteinopathies" As discussed above, this relationship is not necessarily tight, so this seems inappropriate. It is fine for the authors to state that they find a relationship in both human cases and mouse models. This certainly raises the possibility that there is a mechanistic relationship, which is interesting. However, there are major caveats to this relationship, and the actual mechanistic relationship is certainly unclear from their data. So, I don't understand why the authors keep insisting on such language.

I think the authors need to add a sentence to the results clearly stating that the peptide used for the tubulin polymerization assay is succinylated throughout, so that it is very clear for the reader.

The authors also continue to have language that is too strong for their data. For example, line 347 should say something like "may be a key molecular event that contributes" rather than "is a key molecular event that promotes." In line 421 it could say something like "our results raise the possibility that succinylation may provide a link" rather than our results "demonstrate that succinylation is a key

link." The data are compelling. There is no need to over sell it.

Reviewer #3 (Remarks to the Author):

The authors investigated lysine succinylation changes in the brain associated with Alzheimer's disease comparing lysine succinylomes and proteomes from AD and control brains. They found in AD brains, succinylation declined for multiple mitochondrial proteins, and increased for a smaller number of cytosolic proteins, among which the amyloid precursor (APP) and tau exhibited the largest increases. In transgenic mice models of AD, they also found elevated succinylation of soluble and insoluble APP and tau. They examined the effect of succinylation and observed disrupted normal secretase processing of APP and A $\beta$  accumulation. Succinylation of tau also promoted its aggregation and impaired microtubule assembly.

PTMs have been investigated in connection with neurodegenerative diseases. Tau proteins are known to undergo PTMs While succinylation is a well known modification that causes significant changes in proteins the studies presented by the authors are novel and expand our knowledge of the pathology in AD.

The authors have largely addressed prior reviewer concerns related to mass spectrometry. A concern about the prior submission was the exclusive focus on succinylation without comparison to other PTMS. The authors response seems to acknowledge this issue by expanding the introduction to cover other PTMs in AD as background but no additional data is provided. The response of the authors to other issues such as independent validation of data being beyond the scope of the paper is understandable.

Overall the quality of the revised paper is much improved and the findings are of interest to the field

We appreciate the insightful consideration and valuable comments of reviewer #2. We have addressed each of the constructive comments on an item-by-item basis, and provide responses addressing each of them (we insert the line numbers of the marked documents). The changes are highlighted in yellow in the marked manuscript and methods section and our answers are inserted in blue immediately after each reviewer's question.

## **REVIEWER COMMENTS**

Reviewer #1 (Remarks to the Author):

The authors have done an excellent job addressing the previous critiques.

Reviewer #2 (Remarks to the Author):

Overall, the manuscript is improved. However, the following points still need to be addressed:

In 5C, the authors need to perform dual IF with another protein that is unchanged in the same image that they are staining for succinylation, so that we can be sure that the changes are not just due the one sample staining a bit better, and therefore being brighter. The other Ab that is used currently is for AB oligomers is nice for co-localization, but is uninformative for comparing levels of succinylation. This is standard control when trying to quantify changes in levels from staining.

### **Methods lines 411-415.**

We thank you the reviewer for raising this point. All sections were stained at the same time under the same conditions (solutions, washing, temperature, etc.) and analyzed under identical experimental settings. Our results are expressed as the mean with SEM representative of the average of ~900-1000 pyramidal neurons comprised in 3-4 different brain sections per animal (n = 4 mice per each group).

### **Manuscript lines 379-381.**

Similar to what we observed found in A $\beta$  deposits, 10-month-old wild-type and transgenic tau mice displayed a significant reduction in the levels of succinyl-lysine in comparison to 4-month-old mice, thereby leading to an attenuated colocalization between succinylation epitopes and tauopathy epitopes.

### **Manuscript lines 468-490.**

While proteins in addition to tau or APP are succinylated, APP and tau succinylation status increase in brains from AD patients, which suggests

that increased tau and APP succinylation may play a role in the development of AD pathology. Intriguingly, lysine succinylation levels decrease in 10-month-old mice over 4-month mice, while both amyloid aggregation and tauopathy continued to increase. This may reflect either de-succinylation processes, or sequestration of succinylated sites away from labeling antibodies. Notably, both K16 in A $\beta$  and K311 in tau are buried in the structured core of their respective aggregated forms(Goedert *et al.* 2021),(Zhang *et al.* 2019). The decrease in the association between succinylation and pathology at 10 months may be due to results at least in part incorporation of succinylated sites inside aggregated species, preventing detection by immunohistochemistry. However, based on the current data, it is not possible to rule out alternative explanations, including potential changes in metabolism leading to de-succinylation reactions that may be related, or unrelated, to the progression of pathology and disease. Importantly, some precedent is provided by reports in which tau acetylation at residue K280 also peaks and decays during the course of tangle formation and cell death, leading to the suggestion that this epitope is either masked in paired helical filaments (PHFs) or else is subjected to deacetylation in later stages of aggregate maturation(Irwin *et al.* 2012). An adequate explanation requires a complete accounting of which proteins are involved (i.e., a complete mouse brain succinylome at multiple ages) and knowing which proteins are incorporated into deposits in both humans and mouse models. The mitochondrial succinylome in human brain tissues was significantly reduced in AD while succinylation of APP and tau was increased. Despite these remaining questions, our results suggest that the modification of metabolism in disease may lead to critical succinyl-mediated modifications of extramitochondrial proteins including APP and tau leading to aggregation and deposition. Preventing APP and tau succinylation and/or increasing mitochondrial succinylation may provide novel therapeutic targets for the prevention and/or treatment of AD and associated pathologies(Yang & Gibson 2019).

I think that removing the 10 month time point from figure 5 is not appropriate, as it risks just hiding data that potentially does not fit their model. I think the authors need to put the 10 month time point back and add a potential explanation. Personally, I don't find the argument that they provide in the rebuttal to be that compelling, but that is for the reader to judge. Removing the data completely does not provide the reader the opportunity to do so, and is therefore inappropriate.

We thank the reviewer for the suggestion. We have now restored the 10-month age data. While we cannot provide a conclusive explanation for the



decrease in succinylation immunoreactivity at 10 months, we propose that succinylated sites become masked in fibrillar species, pointing out that both the A $\beta$  and tau succinylation sites (that we have discovered) are buried in the core of the respective fibrillar forms of these proteins. At the same time, we acknowledge that our data do not state directly to this possibility and that it is also possible that the decrease may be due to a de-succinylation process, which can occur as a consequence of metabolic alterations related to pathology or disease. Sorting out these different possibilities would require a way to assess the presence of inaccessible succinylation sites at different time points in the mouse brains. This could potentially be addressed by further mass spectrometry experiments but would be a major effort that is beyond the scope of the current study.

**Manuscript lines 312-319. Figure 5d and Extended Data Figure 3b.**

However, the immunoreactivity of lysine succinylation was significantly decreased in 10-month-old wild-type and transgenic mice relative to 4-month-old mice, which results in a reduced colocalization between lysine succinylation and A $\beta$  plaque accumulation (**Figure 5d** and **Extended Data Figure 3b**). This could result either from a decrease in lysine succinylations or from their sequestration into a context (e.g., perhaps in the form of A $\beta$  plaques) that prevents Succ-K antibody from access to possibly buried succinylation sites. These findings suggest that APP succinylation might be involved in early A $\beta$  aggregation events *in vivo*, while its role and mechanism in later events leading to subsequent plaque development remain to be further explored.

Along these lines, the authors response to the critique that the changes in succinylation in the transgenic mouse model don't correlate very well with the build up of AB is insufficient. While it is clearly true that succinylation is responding to the transgene, the fact that succinylation doesn't correlate well with the build up of AB provides a major caveat to the interpretation. As is, the authors seem to be saying that because the transgene causes a change in succinylation, it doesn't matter that that change does not seem to correlate well with the build up of pathological AB. When actually, this disconnect raises the clear possibility that AB build up and succinylation are not directly related. This doesn't mean that their results are not potentially interesting or important, but it does need to be acknowledged and discussed so that the reader is not left with an incorrect view of the data.

**Manuscript lines 312-319. Figure 5d and Extended Data Figure 3b.**

However, the immunoreactivity of lysine succinylation was significantly decreased in 10-month-old wild-type and transgenic mice relative to 4-month-old mice, which results in a reduced colocalization between lysine

succinylation and A $\beta$  plaque accumulation (**Figure 5d** and **Extended Data Figure 3b**). This could result either from a decrease in lysine succinylations or from their sequestration into a context (e.g., perhaps in the form of A $\beta$  plaques) that prevents Succi-K antibody from access to possibly buried succinylation sites. These findings suggest that APP succinylation might be involved in early A $\beta$  aggregation events *in vivo*, while its role and mechanism in later events leading to subsequent plaque development remain to be further explored.

**Manuscript lines 379-381. Figure 6c,d and extended data 4a,b.**

Similar to what we observed found in A $\beta$  deposits, 10-month-old wild-type and transgenic tau mice displayed a significant reduction in the levels of succinyl-lysine in comparison to 4-month-old mice, thereby leading to an attenuated colocalization between succinylation epitopes and tauopathy epitopes.

Similarly, the discussion that has been added to supposedly address the review point about the changes occurring in models of both AB and Tau is also not sufficient. The authors need to specifically discuss why succinylation may changing in both AB and Tau models, despite the models being different.

We agree with the reviewer that this should be discussed more thoroughly. Although we do not know the exact answer, it is tempting to speculate that this is a consequence of the complex interplay between metabolic/mitochondrial dysfunction and pathological aggregation. We agree that data mapping the progression of mitochondrial dysfunction to changes in succinylation in the two mouse models would help to establish the relationship between these events and protein aggregation, as well as to reveal any commonalities between the two mouse models. This represents an important set of experiments that would not be practicable to incorporate into the current study.

**Manuscript lines 448-459**

We showed that transgenic mouse strains of either tauopathy or amyloidosis phenotype, exhibit widespread increases in lysine succinylation at 4 months of age, which is not exclusive to tau and APP but parallels the early appearance of these proteinopathies. This suggests that each transgene is altering common processes (e.g., mitochondria/metabolism) in addition to tau or APP processing. Metabolism is altered even in embryonic cultures of mouse models of AD(Trushina *et al.* 2012). The data in Figure 4 demonstrate that disrupted mitochondrial function increases succinyl transferase in the cytosol. Indeed, the widespread succinylation in both models provides further evidence of that possibility. Interestingly, a pharmacological increase in vitamin B1 (a key vitamin in metabolism) significantly reduces A $\beta$  burden(Pan *et al.* 2010) and tauopathy(Tapias *et al.*

2018) in mice and also showed encouraging results in AD patients(Gibson *et al.* 2020), suggesting these fundamental processes are critical even in mice genetically engineered to create the pathologies. A more precise interpretation requires knowing which proteins are succinylated since the human brain succinylome probably involves hundreds of succinylated proteins.

The above points are especially important, considering the revised abstract that states It is fine for the authors to state that they find a relationship in both human cases and mouse models. This certainly raises the possibility that there is a mechanistic relationship, which is interesting. However, there are major caveats to this relationship, and the actual mechanistic relationship is certainly unclear from their data. So, I don't understand why the authors keep insisting on such language.

As requested, we have changed the abstract to improve our findings, indicating that our results suggest a potential link (rather than establish a tight relationship) between succinylation and AD/proteinopathy.

**Manuscript lines 129-131.**

Our results suggest the potential existence of a link between lysine succinylation and AD-associated proteinopathies and that aberrant succinylation may be involved in the initiation and/or progression of AD.

I think the authors need to add a sentence to the results clearly stating that the peptide used for the tubulin polymerization assay is succinylated throughout, so that it is very clear for the reader.

This has been amended in the results section.

**Manuscript 400-405.**

To understand the role of succinylation in tau function, tubulin polymerization was assessed using the tau K19 peptide, a 99-residue 3-repeat tau microtubule-binding domain (MBD) fragment (MQ244-E372), and succinylated K19 (**Extended Data Figure 4d-f**). Native tau K19 promoted tubulin assembly as determined by increased light scattering at 350 nm, as previously reported (Cohen *et al.* 2011; Lu *et al.* 1999). Nevertheless, succinyl-CoA treated K19, which is succinylated at multiple lysine residues including Lys311, showed a complete suppression of tubulin assembly activity (**Figure 6k**).

The authors also continue to have language that is too strong for their data. For example, line 347 should say something like "may be a key molecular event that contributes" rather than "is a a key molecular even that promotes."

In line 421 it could say something like “our results raise the possibility that succinylation may provide a link” rather than our results “demonstrate that succinylation is a key link.” The data are compelling. There is no need to over sell it.

As requested, we have changed the abstract to improve our findings, indicating that our results suggest a potential link (rather than establish a tight relationship) between succinylation and AD/proteinopathy.

**Manuscript lines 129-131.**

Our results suggest the potential existence of a link between lysine succinylation and AD-associated proteinopathies and that aberrant succinylation may be involved in the initiation and/or progression of AD.

**Reviewer #3 (Remarks to the Author):**

The authors investigated lysine succinylation changes in the brain associated with Alzheimer’s disease comparing lysine succinylomes and proteomes from AD and control brains. They found in AD brains, succinylation declined for multiple mitochondrial proteins, and increased for a smaller number of cytosolic proteins, among which the amyloid precursor (APP) and tau exhibited the largest increases. In transgenic mice models of AD, they also found elevated succinylation of soluble and insoluble APP and tau. They examined the effect of succinylation and observed disrupted normal secretase processing of APP and A $\beta$  accumulation. Succinylation of tau also promoted its aggregation and impaired microtubule assembly.

PTMs have been investigated in connection with neurodegenerative diseases. Tau proteins are known to undergo PTMs While succinylation is a well known modification that causes significant changes in proteins the studies presented by the authors are novel and expand our knowledge of the pathology in AD.

The authors have largely addressed prior reviewer concerns related to mass spectrometry. A concern about the prior submission was the exclusive focus on succinylation without comparison to other PTMS. The authors response seems to acknowledge this issue by expanding the introduction to cover other PTMs in AD as background but no additional data is provided. The response of the authors to other issues such as independent validation of data being beyond the scope of the paper is understandable.

Overall the quality of the revised paper is much improved and the findings are of interest to the field

Cohen, T. J., Guo, J. L., Hurtado, D. E., Kwong, L. K., Mills, I. P., Trojanowski, J. Q. and Lee, V. M. (2011) The acetylation of tau inhibits its function and promotes pathological tau aggregation. *Nature communications* **2**, 252.

- Gibson, G. E., Luchsinger, J. A., Cirio, R. et al. (2020) Benfotiamine and Cognitive Decline in Alzheimer's Disease: Results of a Randomized Placebo-Controlled Phase IIa Clinical Trial. *Journal of Alzheimer's Disease* **78**, 989-1010.
- Goedert, M., Spillantini, M. G., Falcon, B., Zhang, W., Newell, K. L., Hasegawa, M., Scheres, S. H. W. and Ghetti, B. (2021) Tau Protein and Frontotemporal Dementias. In: *Frontotemporal Dementias : Emerging Milestones of the 21st Century*, (B. Ghetti, E. Buratti, B. Boeve and R. Rademakers eds.), pp. 177-199. Springer International Publishing, Cham.
- Irwin, D. J., Cohen, T. J., Grossman, M., Arnold, S. E., Xie, S. X., Lee, V. M. Y. and Trojanowski, J. Q. (2012) Acetylated tau, a novel pathological signature in Alzheimer's disease and other tauopathies. *Brain* **135**, 807-818.
- Lu, P.-J., Wulf, G., Zhou, X. Z., Davies, P. and Lu, K. P. (1999) The prolyl isomerase Pin1 restores the function of Alzheimer-associated phosphorylated tau protein. *Nature* **399**, 784.
- Pan, X., Gong, N., Zhao, J. et al. (2010) Powerful beneficial effects of benfotiamine on cognitive impairment and  $\beta$ -amyloid deposition in amyloid precursor protein/presenilin-1 transgenic mice. *Brain* **133**, 1342-1351.
- Tapias, V., Jainuddin, S., Ahuja, M. et al. (2018) Benfotiamine treatment activates the Nrf2/ARE pathway and is neuroprotective in a transgenic mouse model of tauopathy. *Human Molecular Genetics* **27**, 2874-2892.
- Trushina, E., Nemutlu, E., Zhang, S. et al. (2012) Defects in Mitochondrial Dynamics and Metabolomic Signatures of Evolving Energetic Stress in Mouse Models of Familial Alzheimer's Disease. *PLOS ONE* **7**, e32737.
- Yang, Y. and Gibson, G. E. (2019) Succinylation Links Metabolism to Protein Functions. *Neurochemical Research* **44**, 2346-2359.
- Zhang, W., Falcon, B., Murzin, A. G., Fan, J., Crowther, R. A., Goedert, M. and Scheres, S. H. W. (2019) Heparin-induced tau filaments are polymorphic and differ from those in Alzheimer's and Pick's diseases. *eLife* **8**, e43584.

## REVIEWER COMMENTS

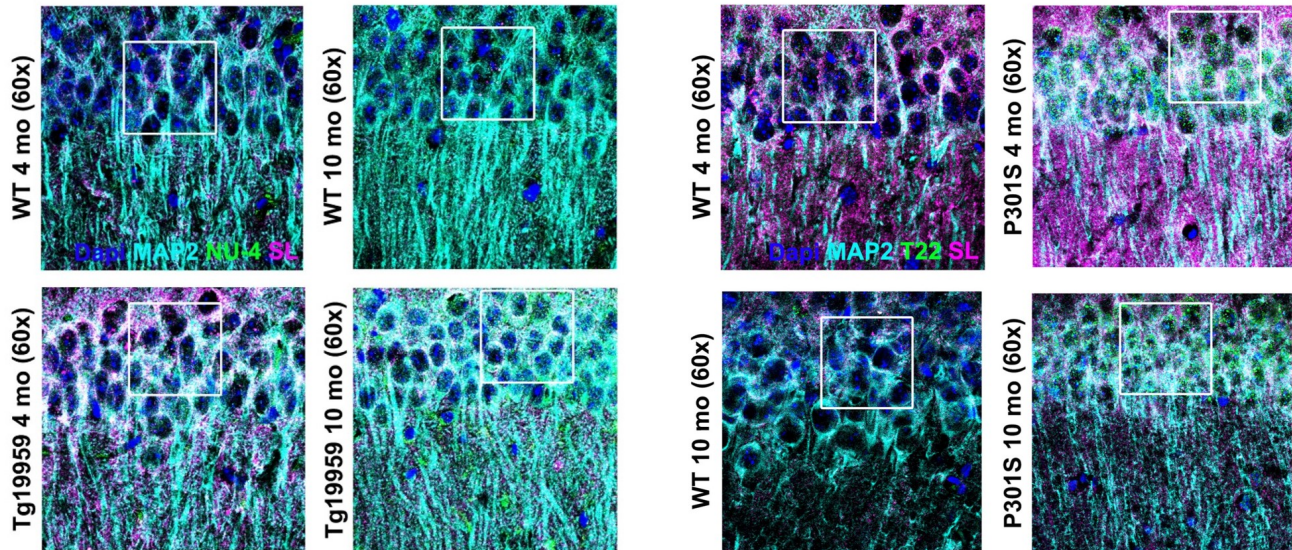
Reviewer #2 (Remarks to the Author):

Overall, the manuscript is substantially improved and almost all of my comments have been addressed. However, in figure 5, the authors still have not provided dual IF with a control antibody that is unchanged. While I understand that performing quantification of imaging on multiple sections and multiple mice somewhat mitigates the concern, it seems like it is really not that difficult to perform this standard control. The authors should have some tissue remaining from the original experiments to do this quickly. Thus, this control should be provided or a compelling reason stated why this control cannot be provided.

Reviewer #2 (Remarks to the Author):

Overall, the manuscript is substantially improved and almost all of my comments have been addressed. However, in figure 5, the authors still have not provided dual IF with a control antibody that is unchanged. While I understand that performing quantification of imaging on multiple sections and multiple mice somewhat mitigates the concern, it seems like it is really not that difficult to perform this standard control. The authors should have some tissue remaining from the original experiments to do this quickly. Thus, this control should be provided or a compelling reason stated why this control cannot be provided.

We agree with the reviewer about the importance of the concern. We believe that our extended data, in particular Extended Data Figures 3 and 4, already answer the Reviewer's question. We have shown that the immunoreactivity of MAP2 (a neuronal marker, cyan) does not change during aging, either in WT or A $\beta$ /tau TG mice. We have created a collage from the first column of panel a in those figures, which demonstrates that the decline in succinylation at 10 months is not an artifact.



We added the following comment to address this concern (Lines 471-476) of the marked manuscript.

This change is not likely to be a technical artifact. All sections were stained at the same time under the same conditions (solutions, washing, temperature, antibody preparation, etc.) and analyzed under identical experimental settings. In addition, perusal of the first column of panel in the Extended Data Figures 3 and 4 show that the immunoreactivity of MAP2 (a neuronal marker, cyan) does not change during aging, either in WT or A $\beta$ /tau TG mice. The decline in succinylation may reflect either de-succinylation processes, or sequestration of succinylated sites away from labeling antibodies.

## **REVIEWER COMMENTS**

Reviewer #2 (Remarks to the Author):

The authors have now addressed my main concerns.