

# Supporting Information

## **Bis-rhodamines bridged with a diazoketone linker: synthesis, structure and photolysis**

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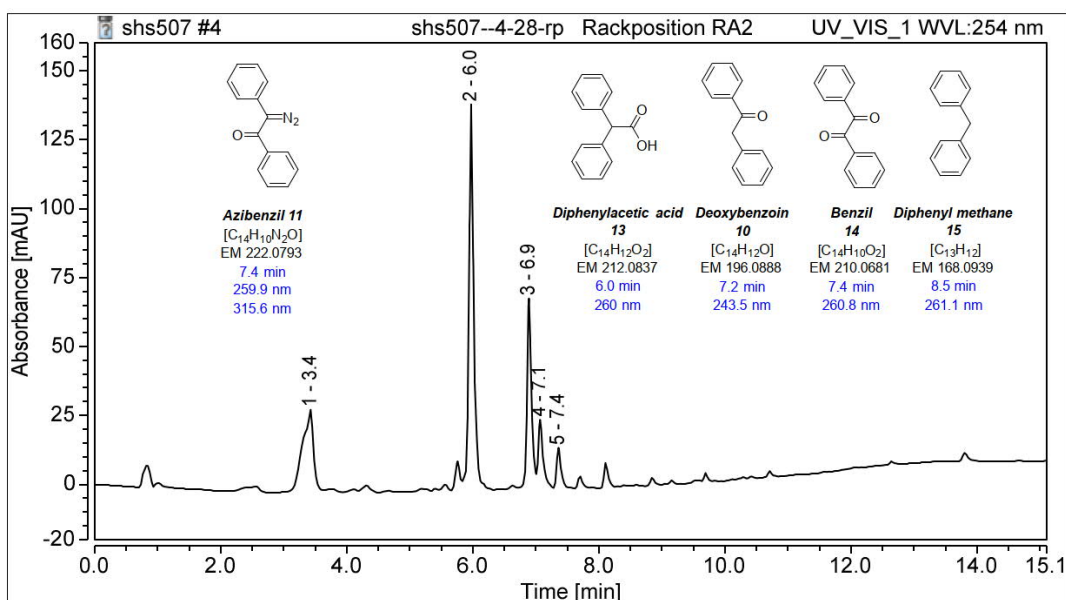
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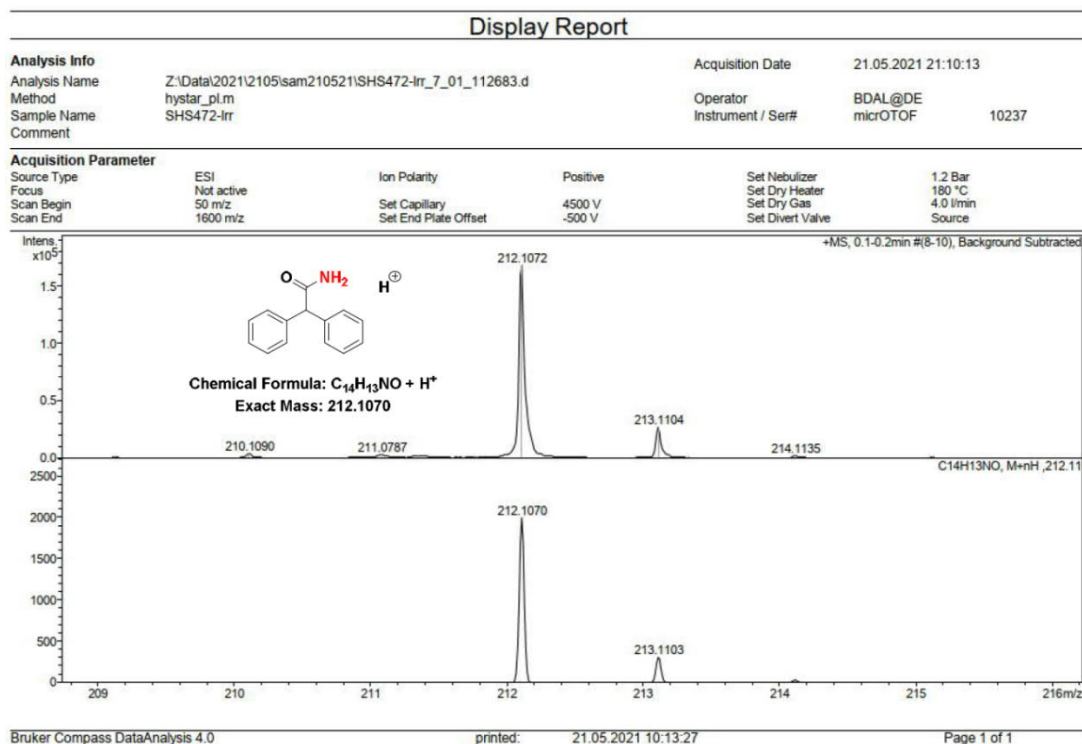
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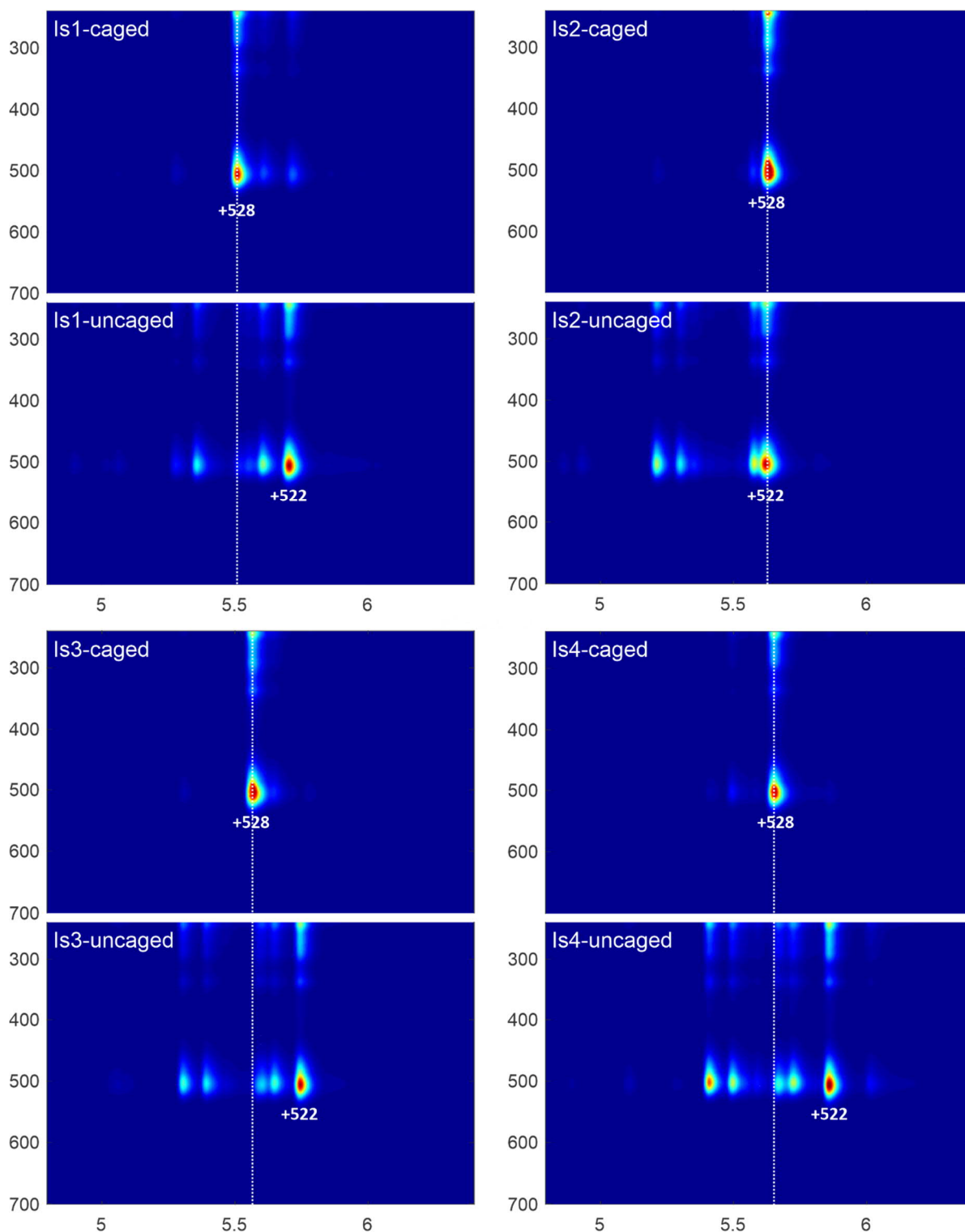
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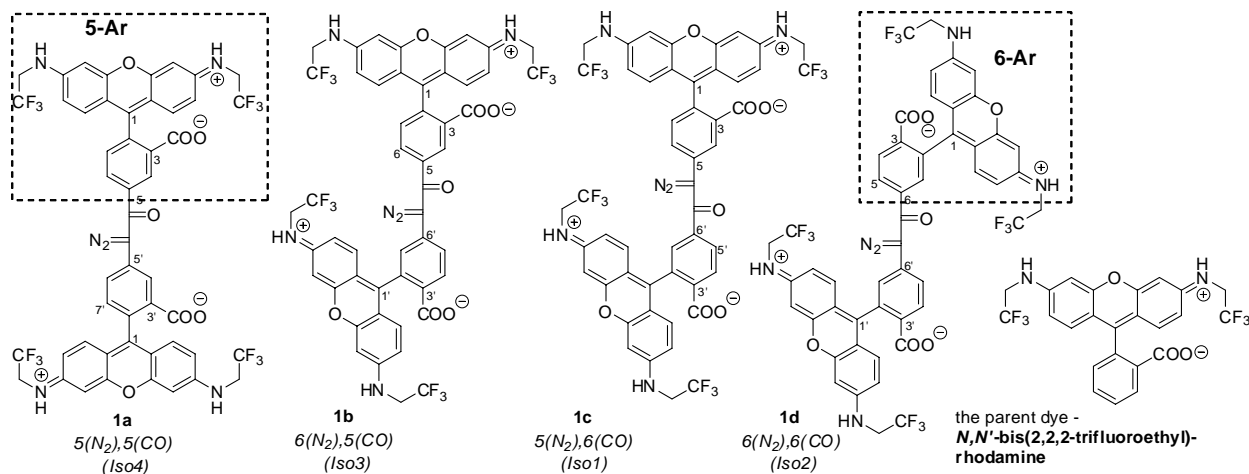
**Figure S1.** HPLC analysis of the reaction mixture: azibenzil (**11**) (0.1 mM) in acetonitrile (80% v/v) and aqueous HEPES buffer (20%, v/v) at pH 6.5 was irradiated with 365 nm light providing full conversion of the starting material.



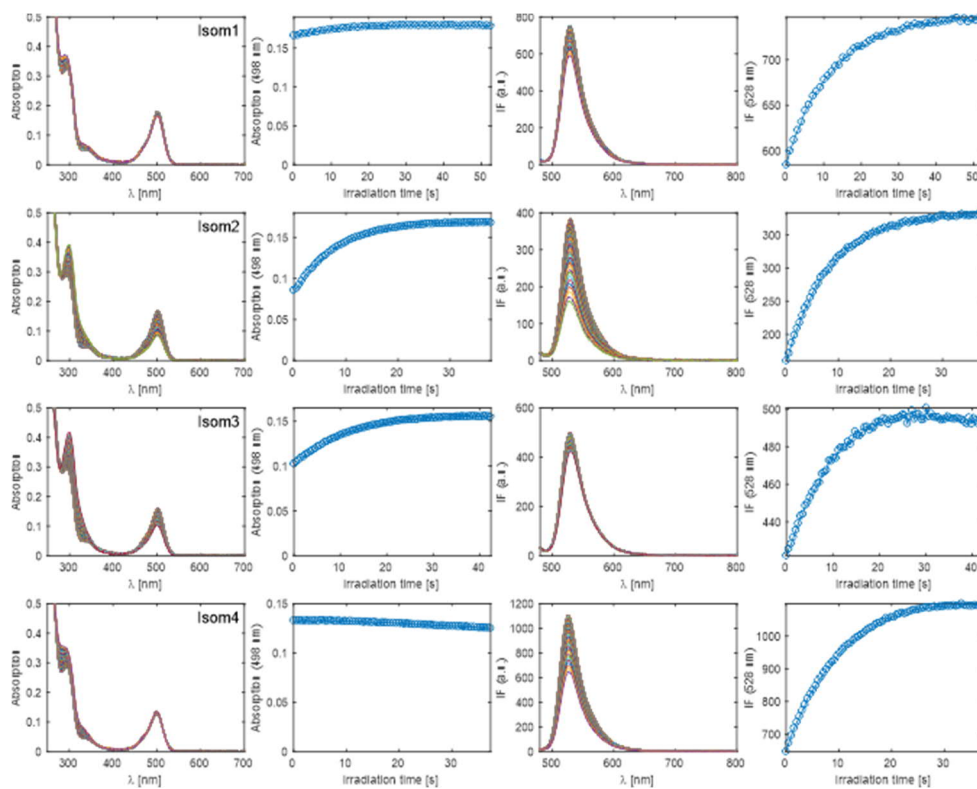
**Figure S2.** HR-MS of the reaction solution obtained after photolysis of azibenzil (**11**, PhCOCN<sub>2</sub>Ph) solutions in aqueous acetonitrile (80% acetonitrile, 20% water, v/v) with HCOONH<sub>4</sub> buffer (pH 7.3). The full conversion to a new substance with the same retention time, but without absorption maximum at 319 nm, was observed (see Figure 2 in the main text).



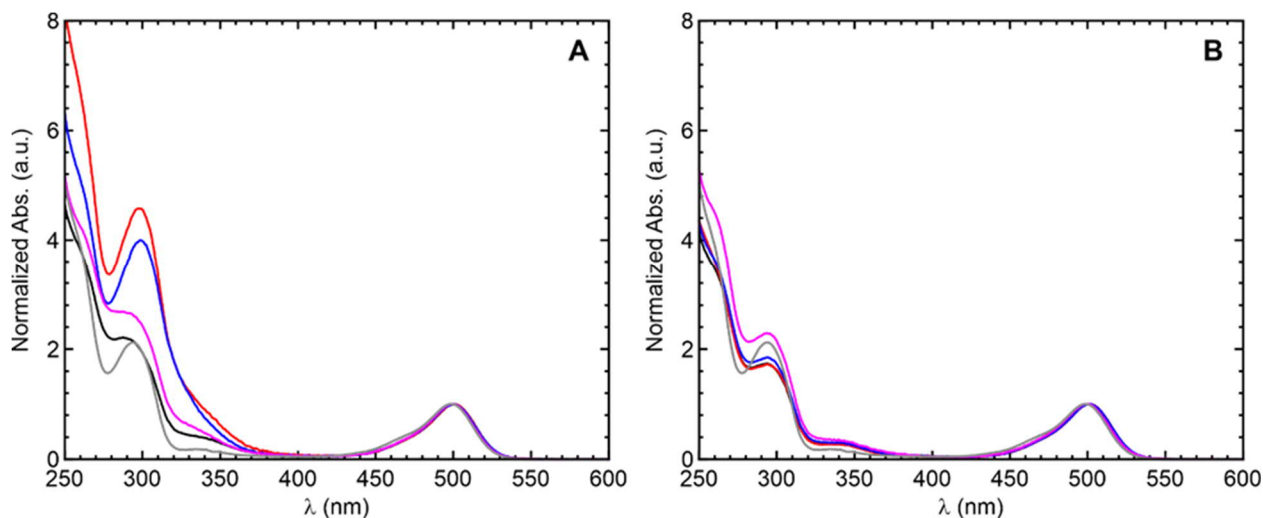
**Figure S3a.** 2D HPLC maps of the solutions of isomers 1-4 (for structures, see Figure S3b), before and after irradiation in MeCN (80% v/v) and aqueous 10 mM ammonium formate buffer (20% v/v; pH = 7.4), with 365 nm light, upon complete conversion. The  $m/z$  values of the main peaks  $[M+2H]^{2+}$  are given for the starting materials ( $m/z$  528) and the main reaction products ( $m/z$  522).



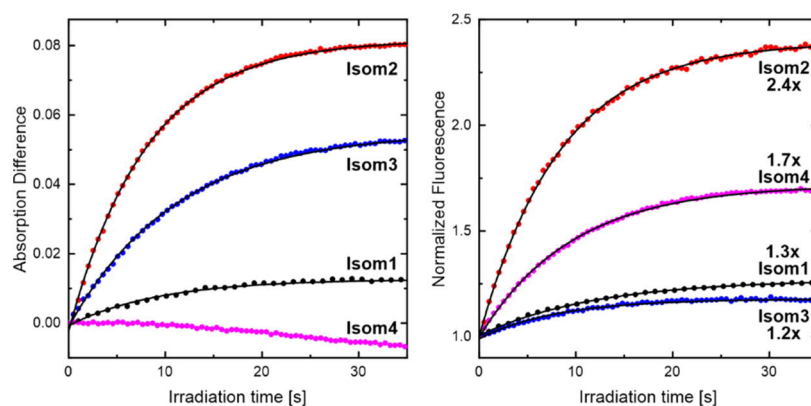
**Figure S3b.** Structures of compounds **1a**, **1b**, **1c** and **1d** (isomers 4, 3, 1 and 2, respectively), and the parent rhodamine dye.



**Figure S4a.** Irradiation of isomers 1-4 (12  $\mu$ M, for structures, see Figure S3b) in MeCN (80% v/v) and aqueous 10 mM ammonium formate buffer, pH = 7.4 (20% v/v). The samples were continuously stirred and kept at 20°C during irradiation with 365 nm light. Absorption at 498 nm and emission at 528 nm ( $\lambda_{EX} = 470$  nm) were monitored and read out at fixed time intervals.



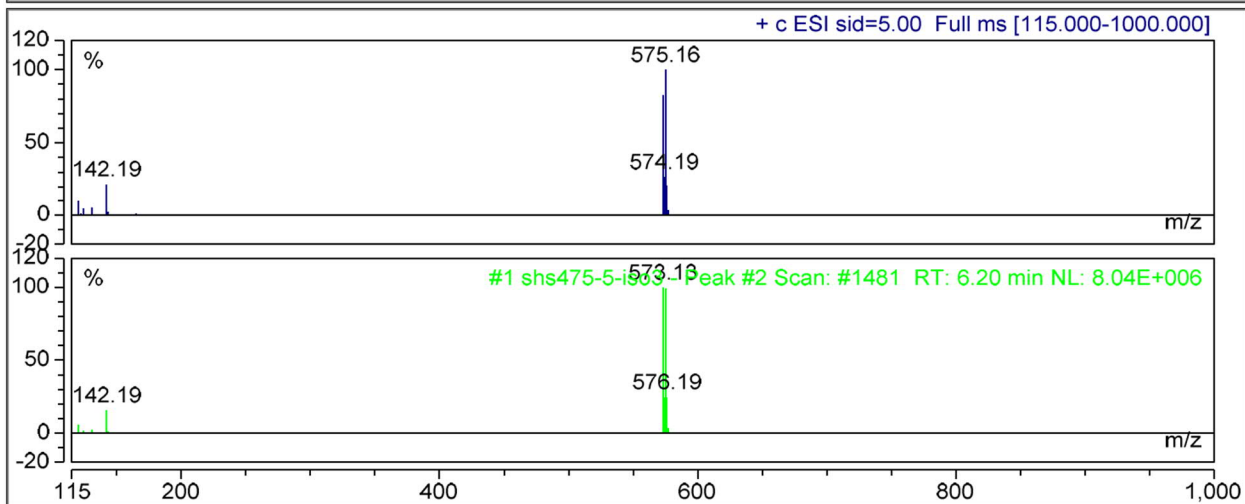
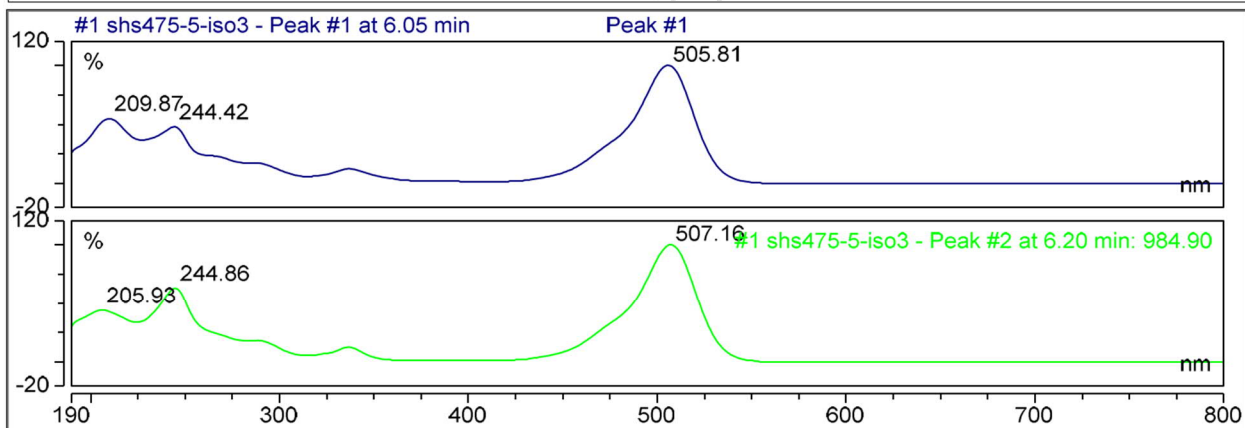
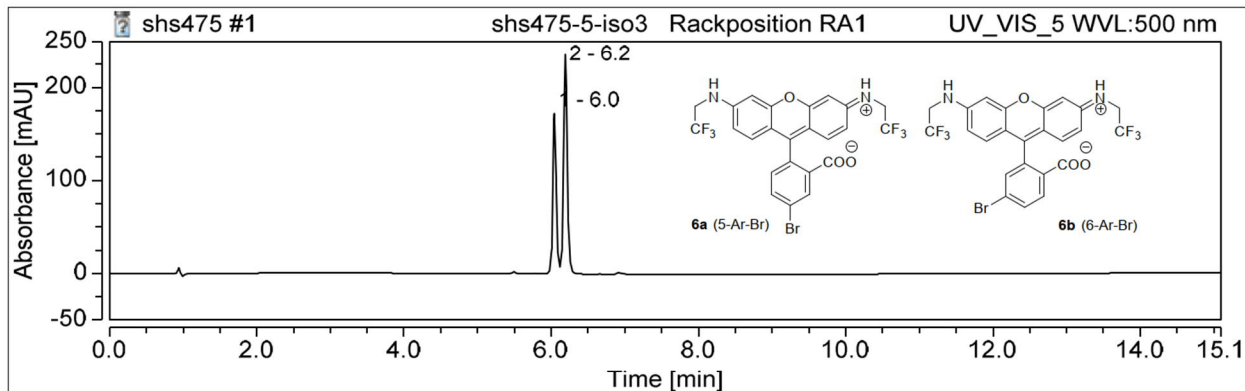
**Figure S4b.** Absorption spectra (normalized to the intensity of the band with the maximum at 498 nm) of compounds **1a** (purple), **1b** (blue), **1c** (black), **1d** (red) and the parent *N,N'*-bis(2,2,2-trifluoroethyl)rhodamine (grey) before (A) and after (B) photoconversion by irradiation with 365 nm source. For structures, see Figure S3b. Solvent: aqueous acetonitrile (80% acetonitrile, 20% water, v/v) with addition of HCOONH<sub>4</sub> buffer (pH 7.3).



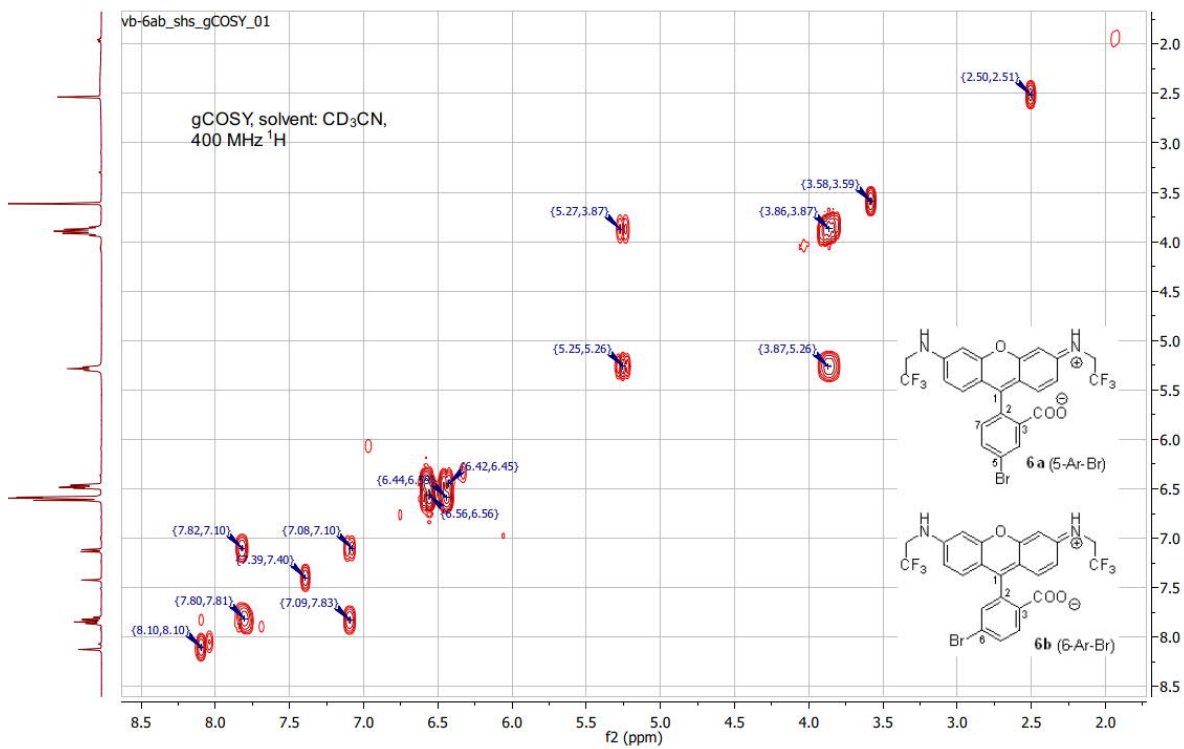
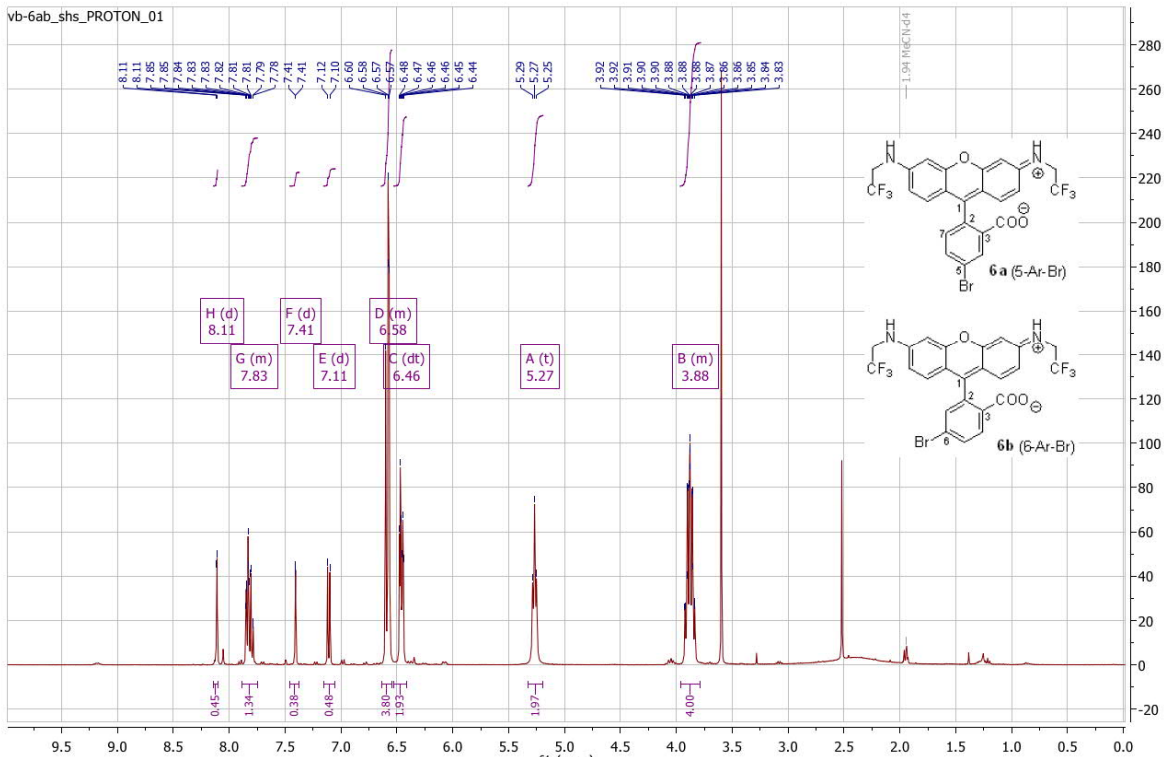
**Figure S5.** Normalized fluorescence (right) and absorption (left) transients during irradiation of isomers 1-4 (**1a-d**, 12  $\mu$ M) with 365 nm light, as solutions in MeCN (80%) and aqueous 10 mM ammonium formate buffer; pH = 7.4 (20% v/v). Samples were continuously stirred and kept at 20°C during irradiation. Absorption at 498 nm and emission at 528 nm ( $\lambda_{\text{EX}} = 470$  nm) were monitored and read out at fixed time intervals. The emission intensities were divided (normalized) by the corresponding initial value for each compound, and the initial absorbance was subtracted for each sample.

Bromorhodamines **6a** (5'-Br) and **6b** (6'-Br)

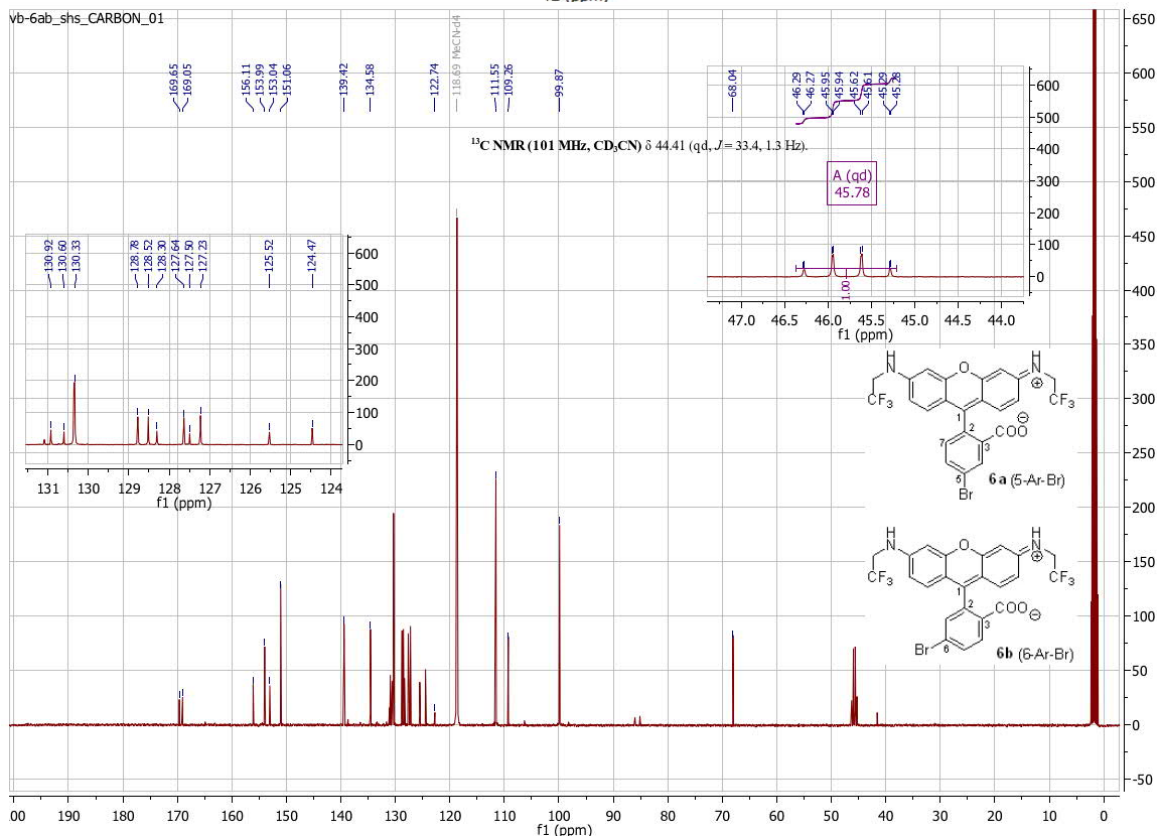
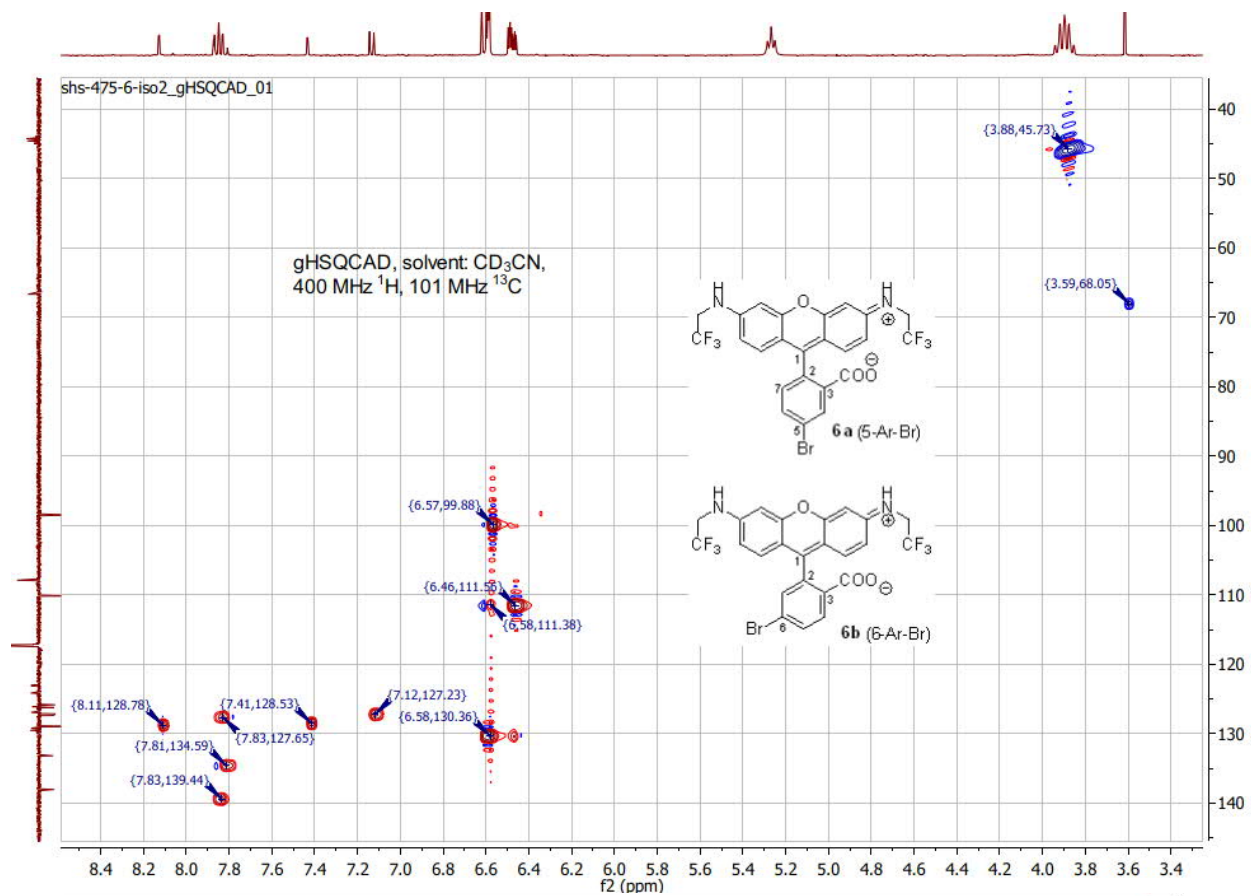
<b>Probe :</b>	shs475-5-iso3		
<b>Lösungsmittel :</b>	MeCN/H2O	<b>Aufgabemenge:</b>	3.0 µl
<b>Säule:</b>	Phenomenex Kinetex C18 2.6 µm	<b>Länge:</b>	75 mm iO : 3.0 mm
<b>Fluß (ml / Min) :</b>	0.5	<b>Temperatur :</b>	25.0
<b>Detektor:</b>	DAD-3000	<b>Pumpe:</b>	HPG-3200SD
<b>Laufmittel:</b>	A = Acetonitril 0.1% FA		<b>Sampler:</b> WPS-3000
<b>Gradient:</b>	A 20.0 % B 80.0 % ---->	A 100.0 % B 0.0 %	T = 10 Min.

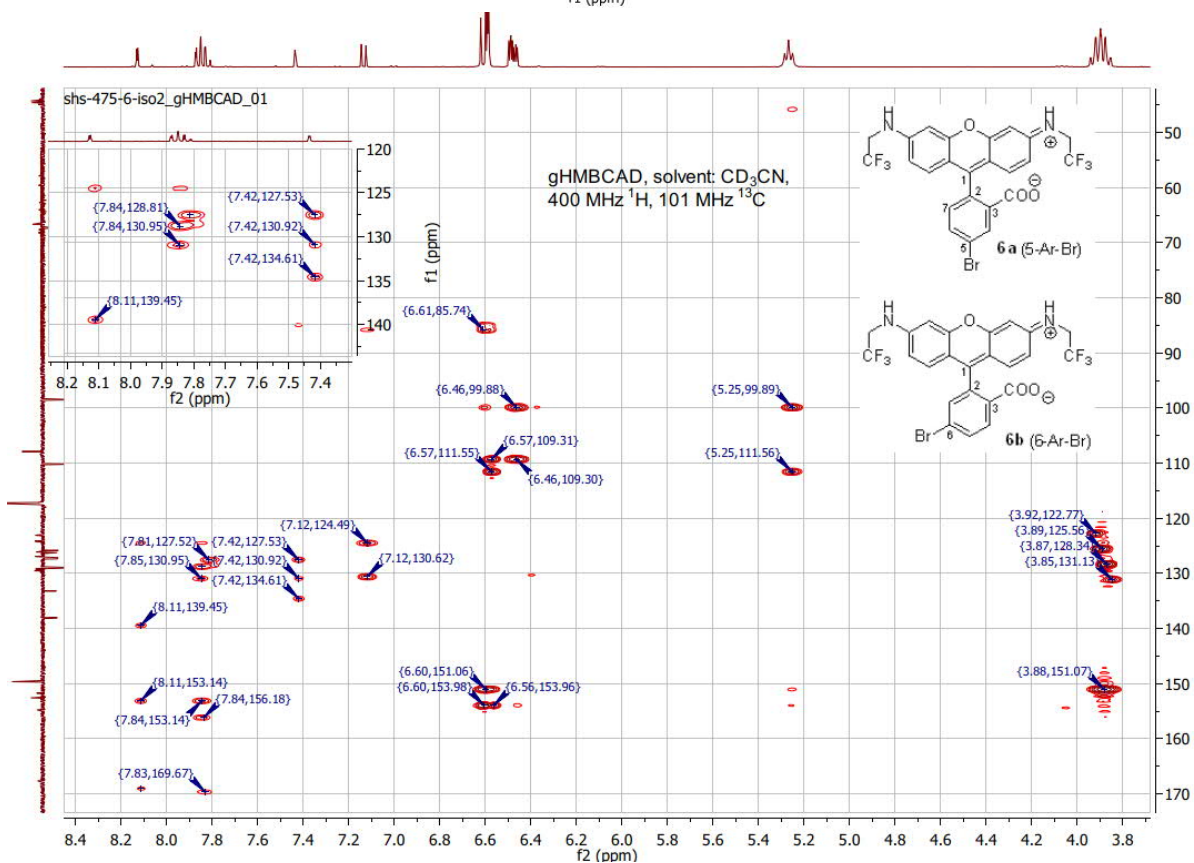
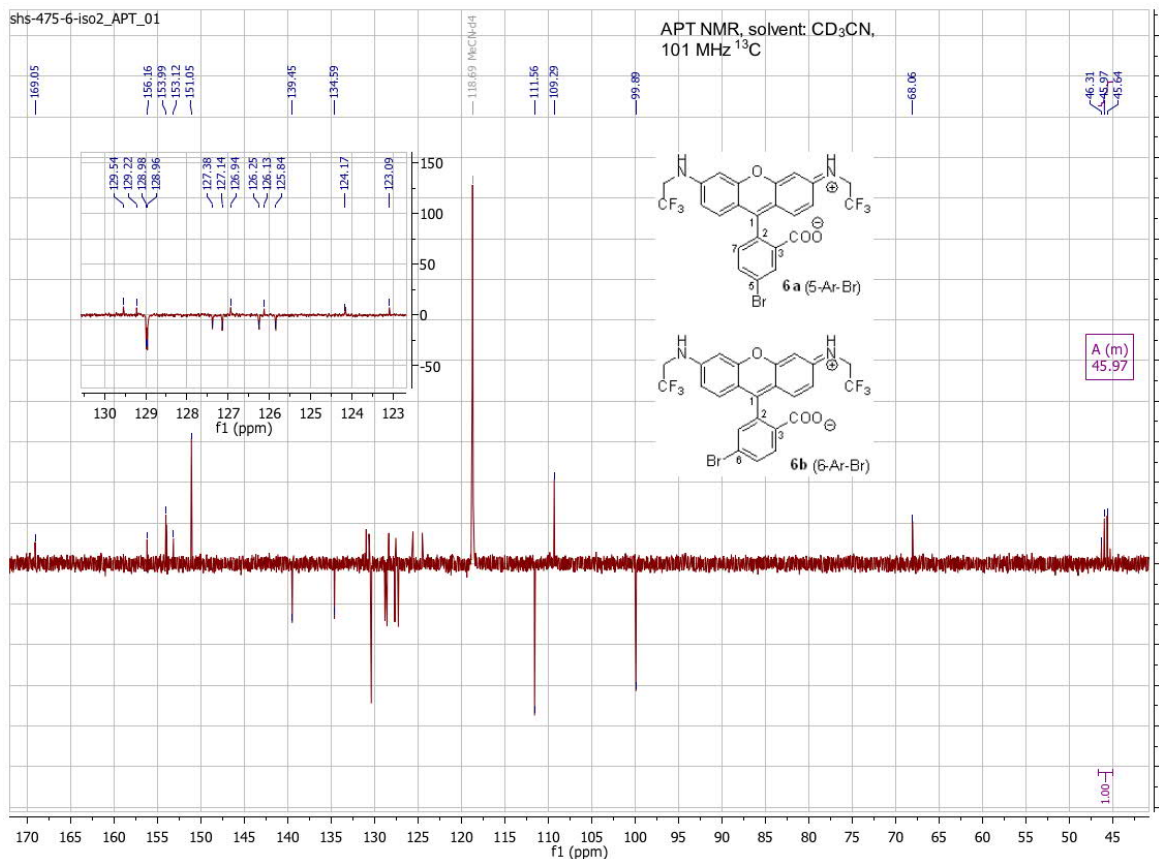


<sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN) δ 8.11 (d, *J* = 1.8 Hz, 0H), 7.89 – 7.75 (m, 1H), 7.41 (d, *J* = 1.6 Hz, 0H), 7.11 (d, *J* = 8.2 Hz, 0H), 6.64 – 6.55 (m, 4H), 6.46 (dt, *J* = 8.7, 2.8 Hz, 2H), 5.27 (t, *J* = 6.9 Hz, 2H), 3.96 – 3.78 (m, 4H).

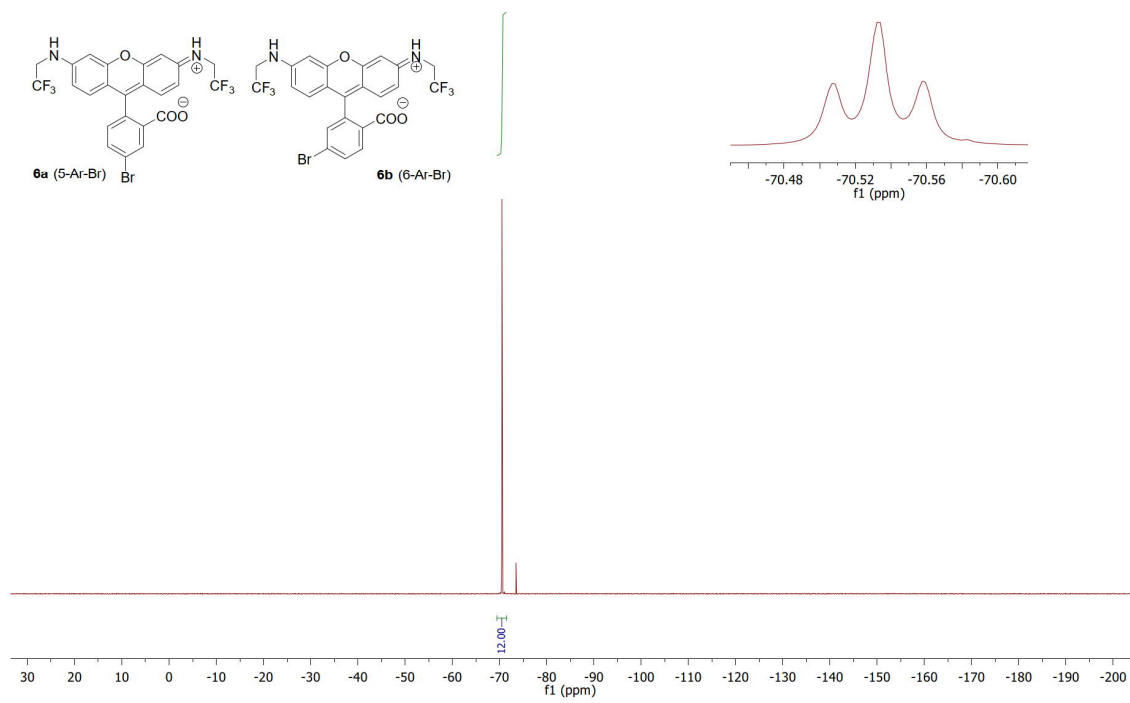






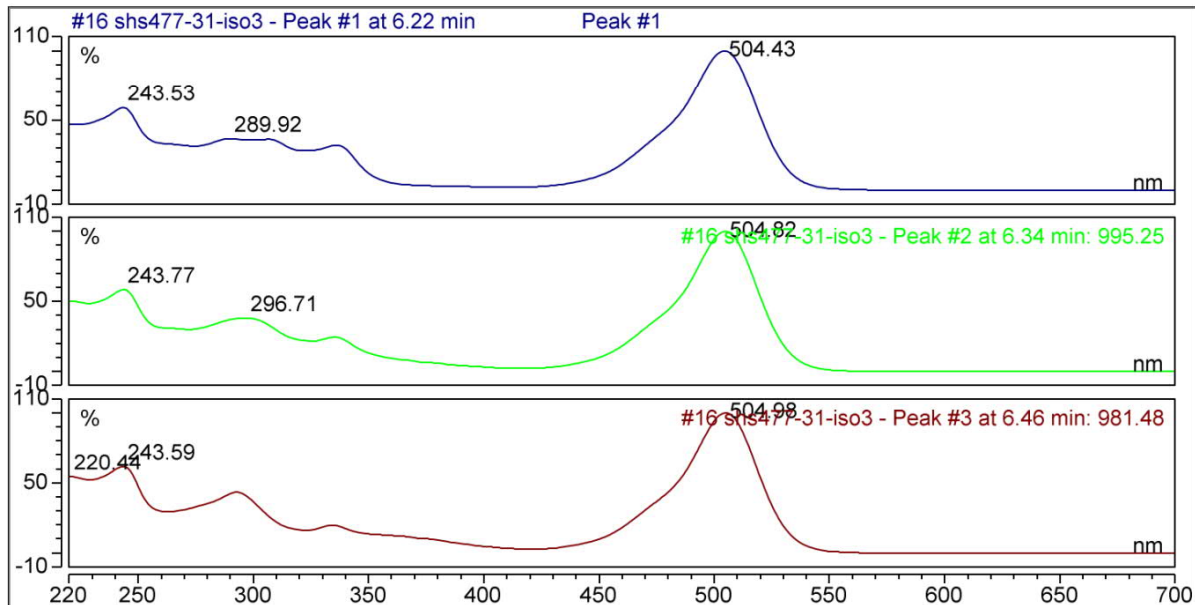
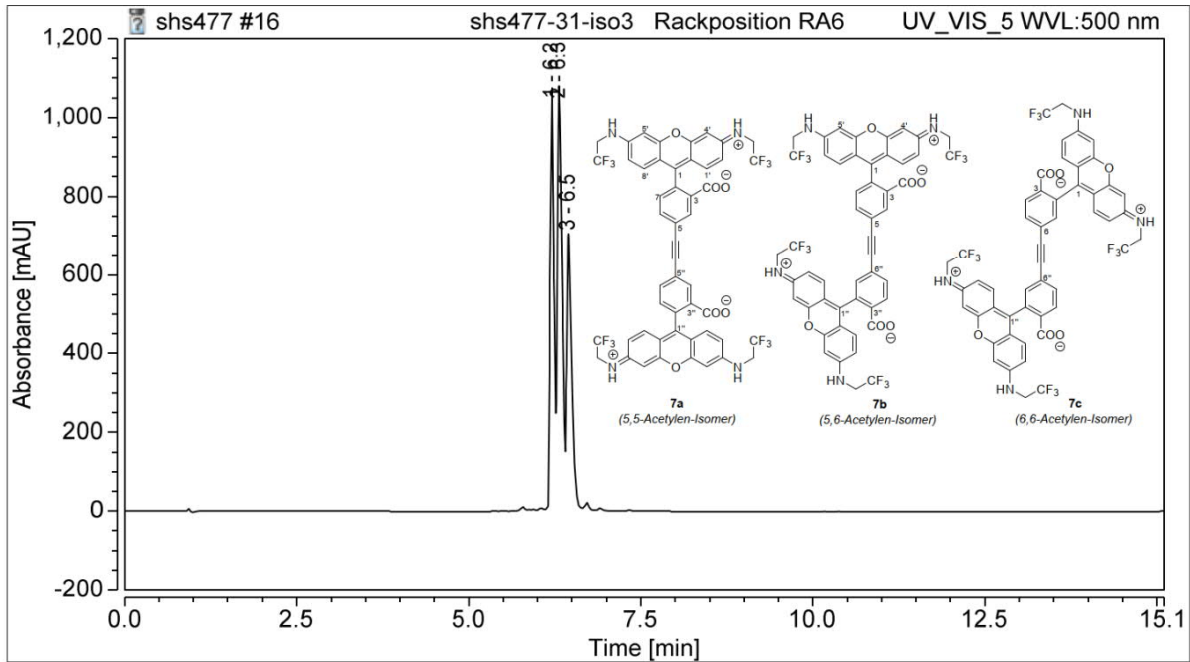


$^{19}\text{F}$  NMR (376 MHz,  $\text{DMSO-}d_6$ )  $\delta$  -70.53 (t,  $J$  = 9.6 Hz, 12F) ppm.  
shs475-1-iso-2a\_FLUORINE\_01



Bis(rhodamine) acetylenes **7a** (peak 3), **7b** (peak 2), and **7c** (peak 3) are 5,5'-, 5,6'- and 6,6'- isomers, respectively.

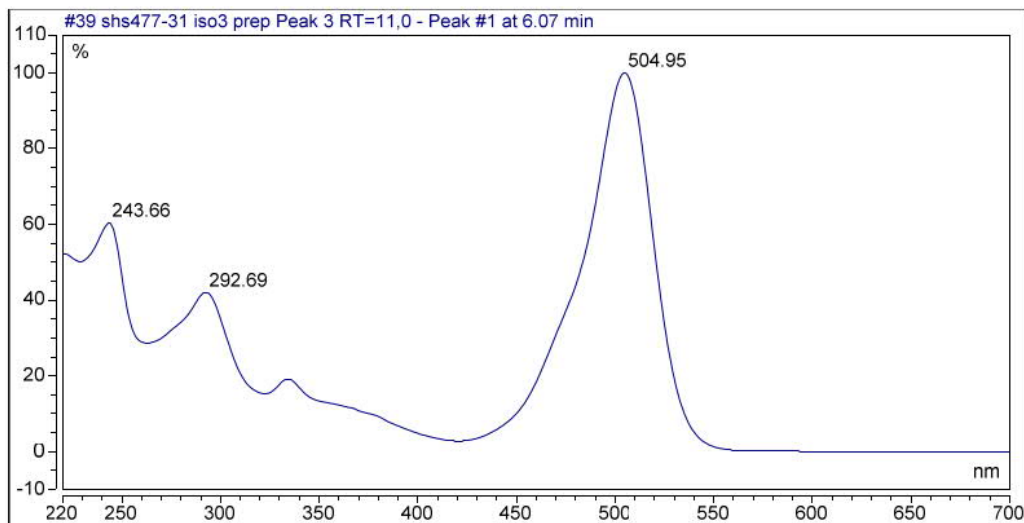
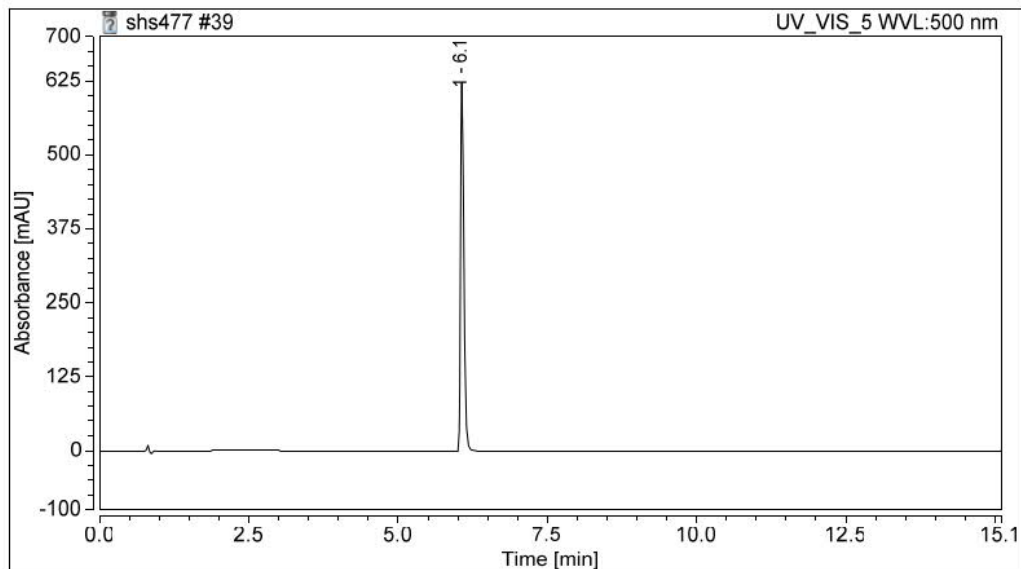
<b>Probe :</b> shs477-31-iso3			<b>Aufgabemenge:</b>	3.0 $\mu$ l
<b>Lösungsmittel :</b> MeCN/H <sub>2</sub> O			<b>Länge:</b>	75 mm
<b>Säule:</b> Phenomenex Kinetex C18	2.6 $\mu$ m		<b>iO :</b>	3.0 mm
<b>Fluß (ml / Min) :</b> 0.5	<b>Temperatur :</b>		25.0	
<b>Detektor:</b> DAD-3000	<b>Pumpe:</b> HPG-3200SD	<b>Sampler:</b> WPS-3000		
<b>Laufmittel:</b>	<b>A = Acetonitril 0.1% FA</b>		<b>B = Wasser 0.1% FA</b>	
<b>Gradient:</b> A 20.0 %	B 80.0 %	---->	A 100.0 %	B 0.0 %
T = 10 Min.				



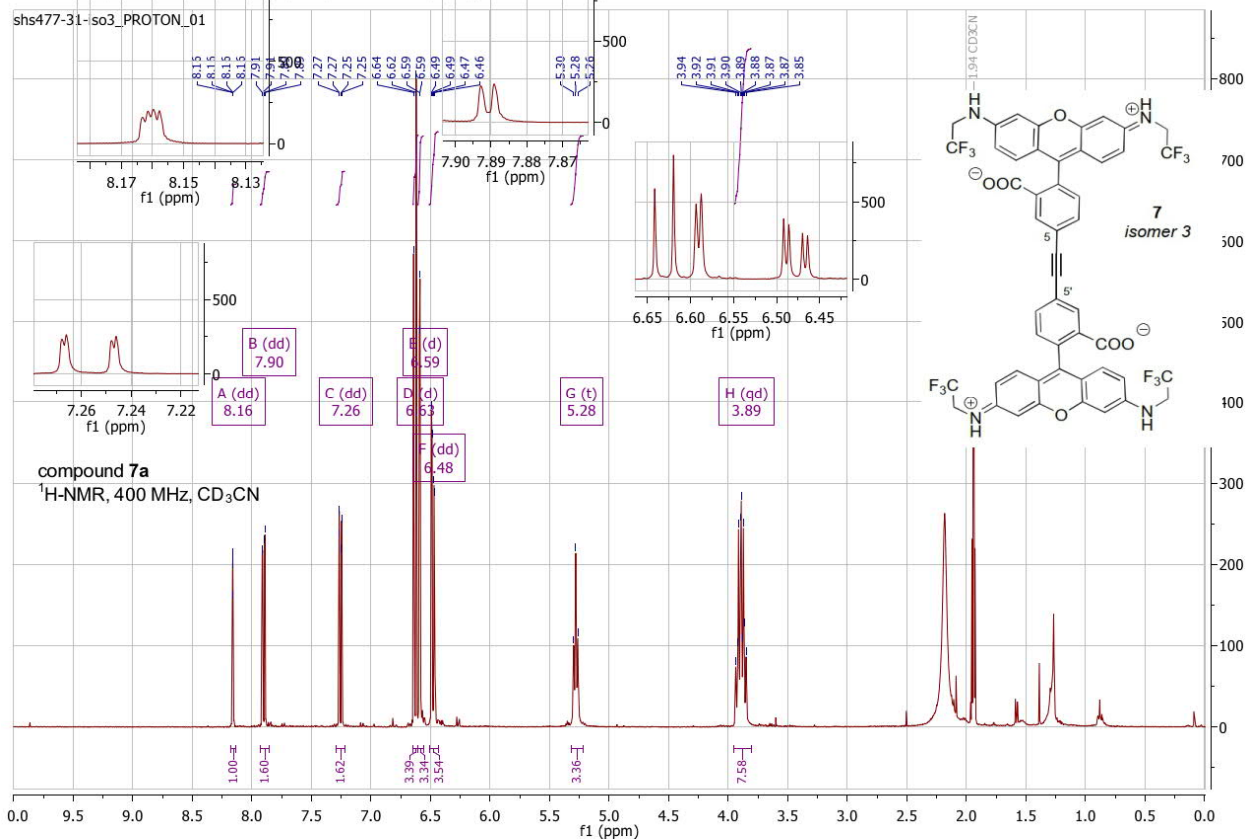
Analytical HPLC traces and NMR spectra of the individual regioisomers **7a** (peak 3), **7b** (peak 2) and **7c** (peak 1) isolated by means of preparative HPLC

Compound **7a**

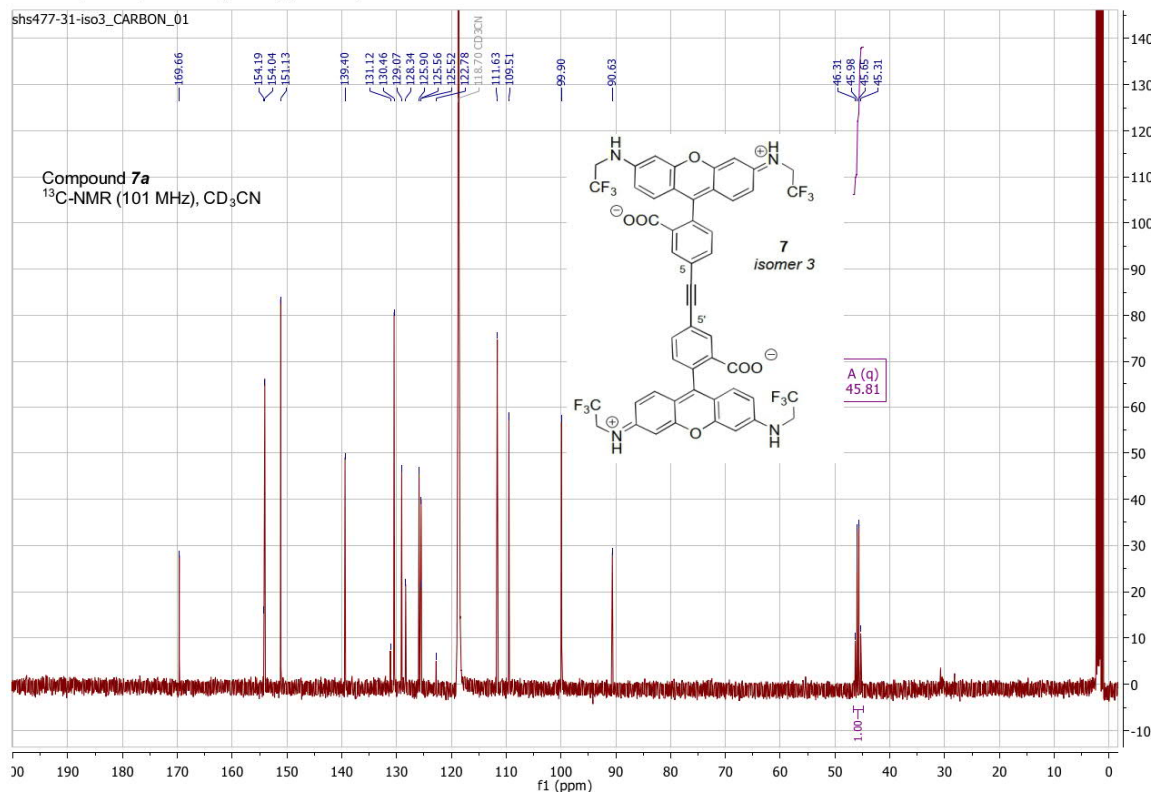
Probe :	shs477-31 iso3 prep Peak 3 RT=11,0		
Lösungsmittel :	MeCN/H <sub>2</sub> O	Aufgabemenge:	3.0 µl
Säule:	Phenomenex 2.6 µm	Länge:	75 mm iO : 3.0 mm
Fluß (ml / Min) :	0.5	Temperatur :	25.0
Detektor:	DAD-3000	Pumpe:	HPG-3200SD Sampler: WPS-3000
Laufmittel:	A = Acetonitril 0.1% FA		B = Wasser 0.1% FA
Gradient:	A 20.0 % B 80.0 %	---->	A 100.0 % B 0.0 % T = 10 Min.



<sup>1</sup>H NMR (400 MHz, Acetonitrile-*d*<sub>3</sub>) δ 8.16 (dd, *J* = 1.5, 0.8 Hz, 1H), 7.90 (dd, *J* = 8.0, 1.5 Hz, 2H), 7.26 (dd, *J* = 8.0, 0.9 Hz, 2H), 6.63 (d, *J* = 8.7 Hz, 3H), 6.59 (d, *J* = 2.4 Hz, 3H), 6.48 (dd, *J* = 8.7, 2.4 Hz, 4H), 5.28 (t, *J* = 7.0 Hz, 3H), 3.89 (qd, *J* = 9.3, 6.8 Hz, 3H).

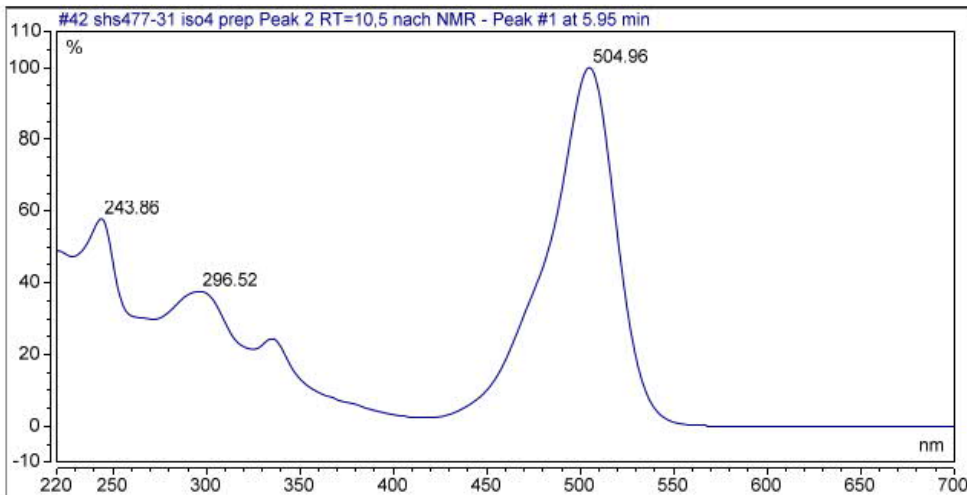
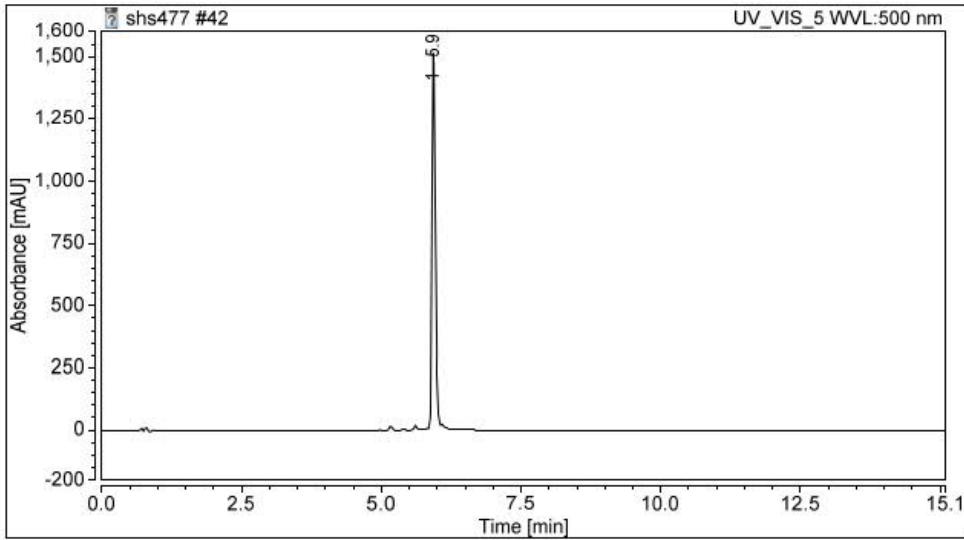


<sup>13</sup>C NMR (101 MHz, Acetonitrile-*d*<sub>3</sub>) δ 45.81 (q, *J* = 33.4 Hz).



## Compound 7b

Probe : **shs477-31 iso4 prep Peak 2 RT=10,5 nach**  
Lösungsmittel : **MeCN/H<sub>2</sub>O** Aufgabemenge: **3.0 µl**  
Säule: Phenomenex 2.6 µm Länge: 75 mm iO : 3.0 mm  
Fluß (ml / Min) : 0.5 Temperatur : 25.0  
Detektor: DAD-3000 Pumpe: HPG-3200SD Sampler: WPS-3000  
Laufmittel: **A = Acetonitril 0.1% FA** **B = Wasser 0.1% FA**  
Gradient: A 20.0 % B 80.0 % ----> A 100.0 % B 0.0 % T = 10 Min.

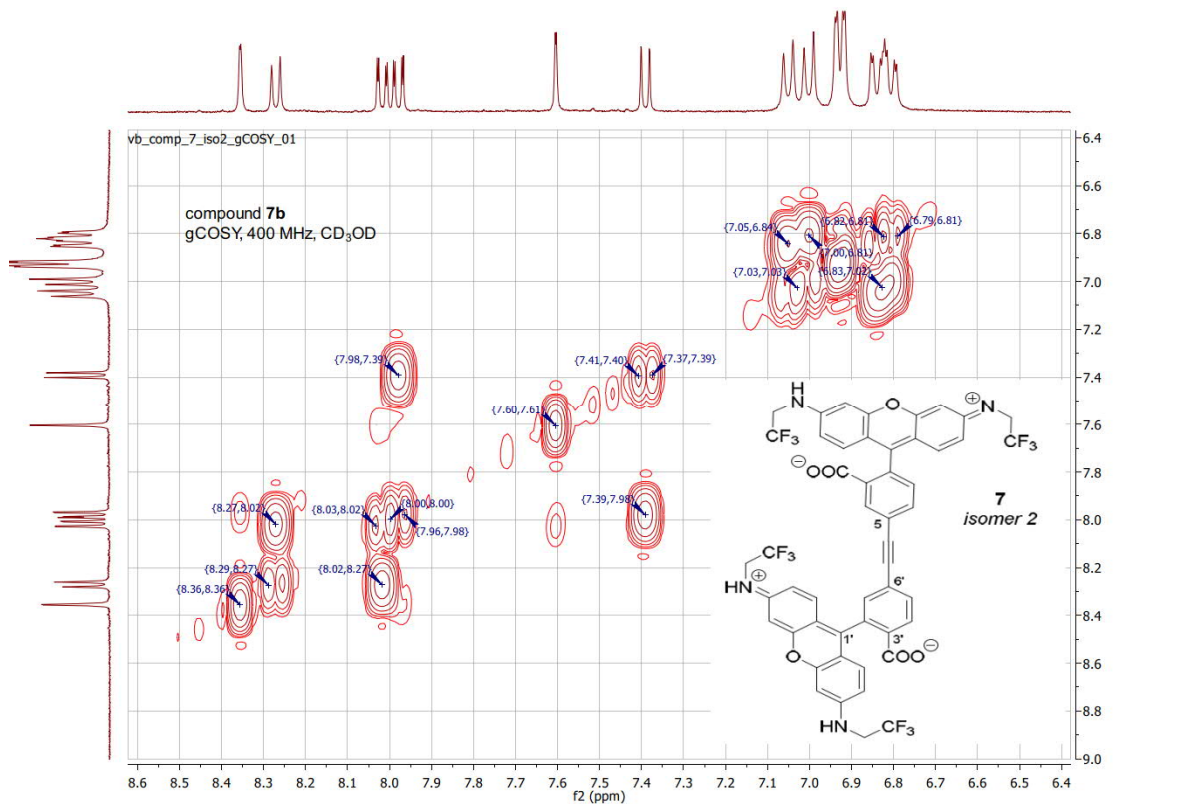
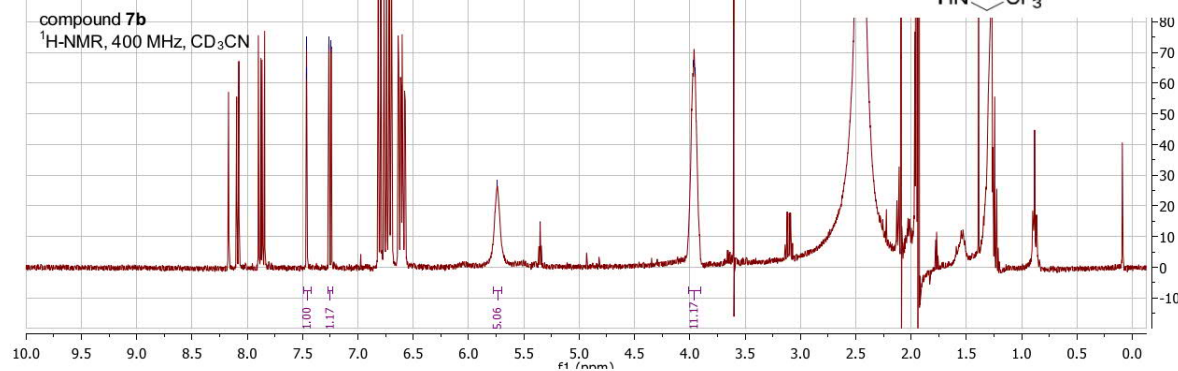
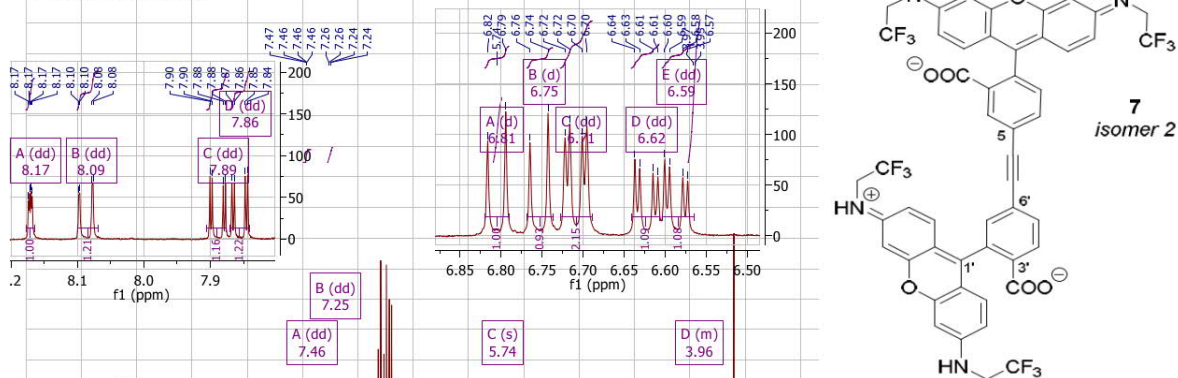


<sup>1</sup>H-NMR (400 MHz, Acetonitrile-d<sub>3</sub>) δ 7.46 (dd, *J* = 1.4, 0.7 Hz, 1H), 7.25 (dd, *J* = 8.0, 0.7 Hz, 1H), 5.74 (s, 5H), 4.01 – 3.91 (m, 11H).

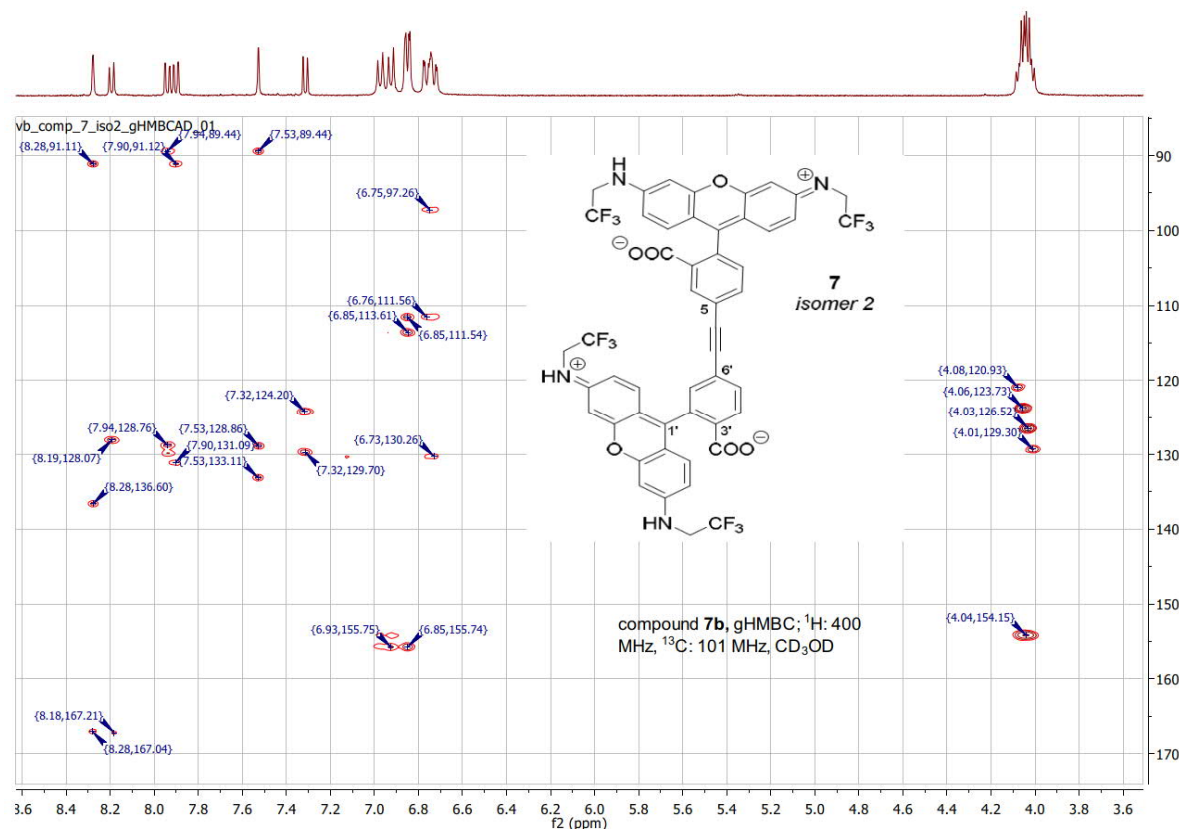
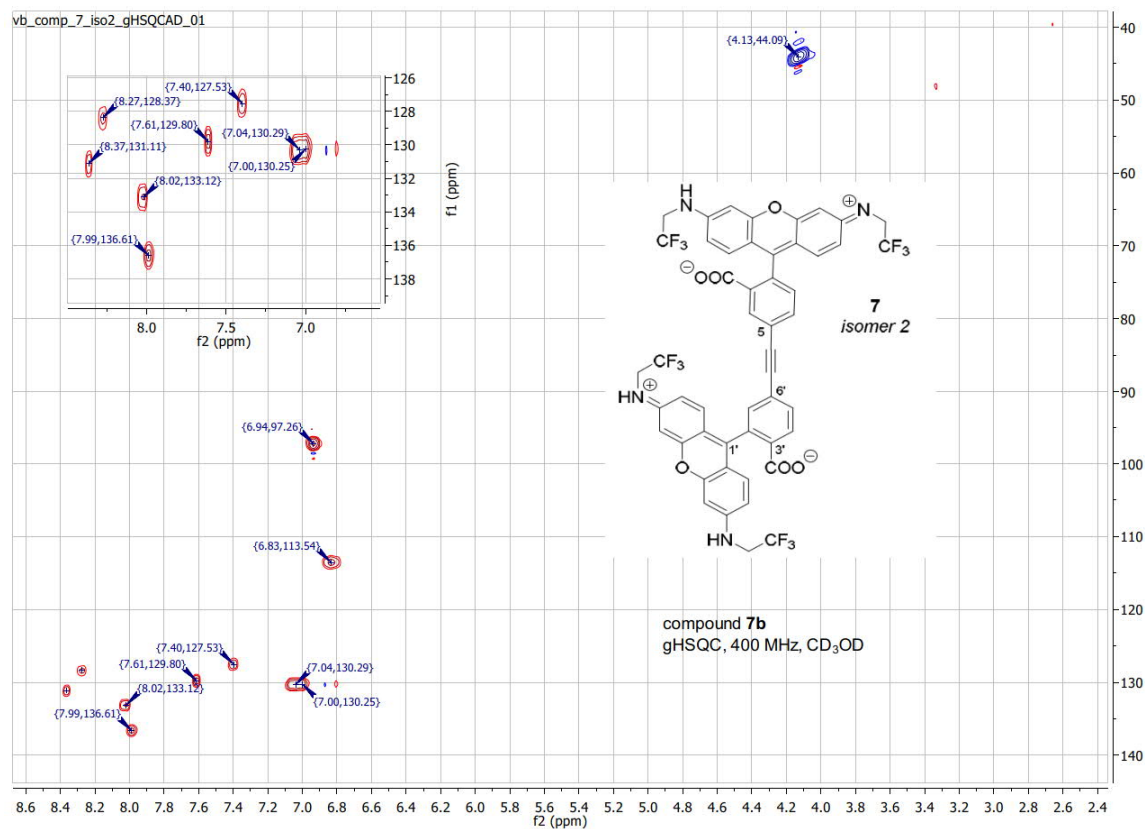
δ 8.17 (dd, *J* = 1.6, 0.7 Hz, 1H), 8.09 (dd, *J* = 8.0, 0.7 Hz, 1H), 7.89 (dd, *J* = 8.0, 1.4 Hz, 1H), 7.86 (dd, *J* = 8.0, 1.5 Hz, 1H).

δ 6.81 (d, *J* = 8.8 Hz, 1H), 6.75 (d, *J* = 8.8 Hz, 1H), 6.71 (dd, *J* = 8.4, 2.4 Hz, 2H), 6.62 (dd, *J* = 8.8, 2.4 Hz, 1H), 6.59 (dd, *J* = 8.8, 2.4 Hz, 1H).

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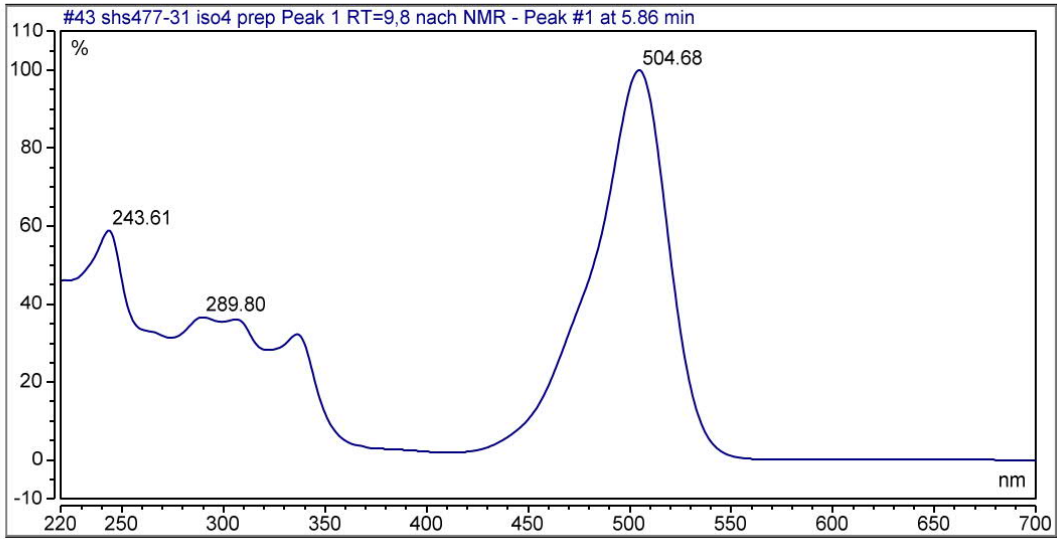
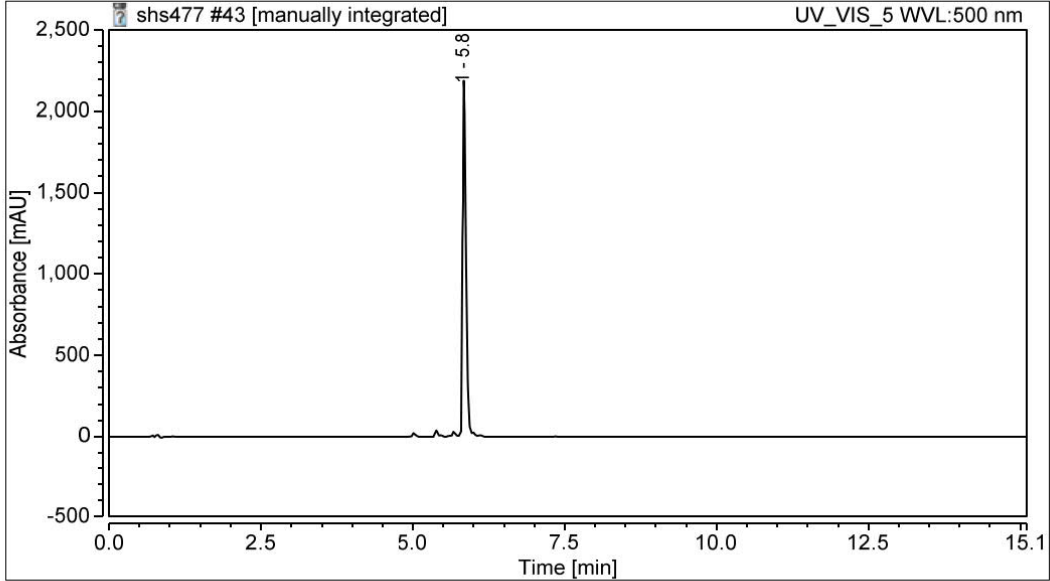






# Compound 7c

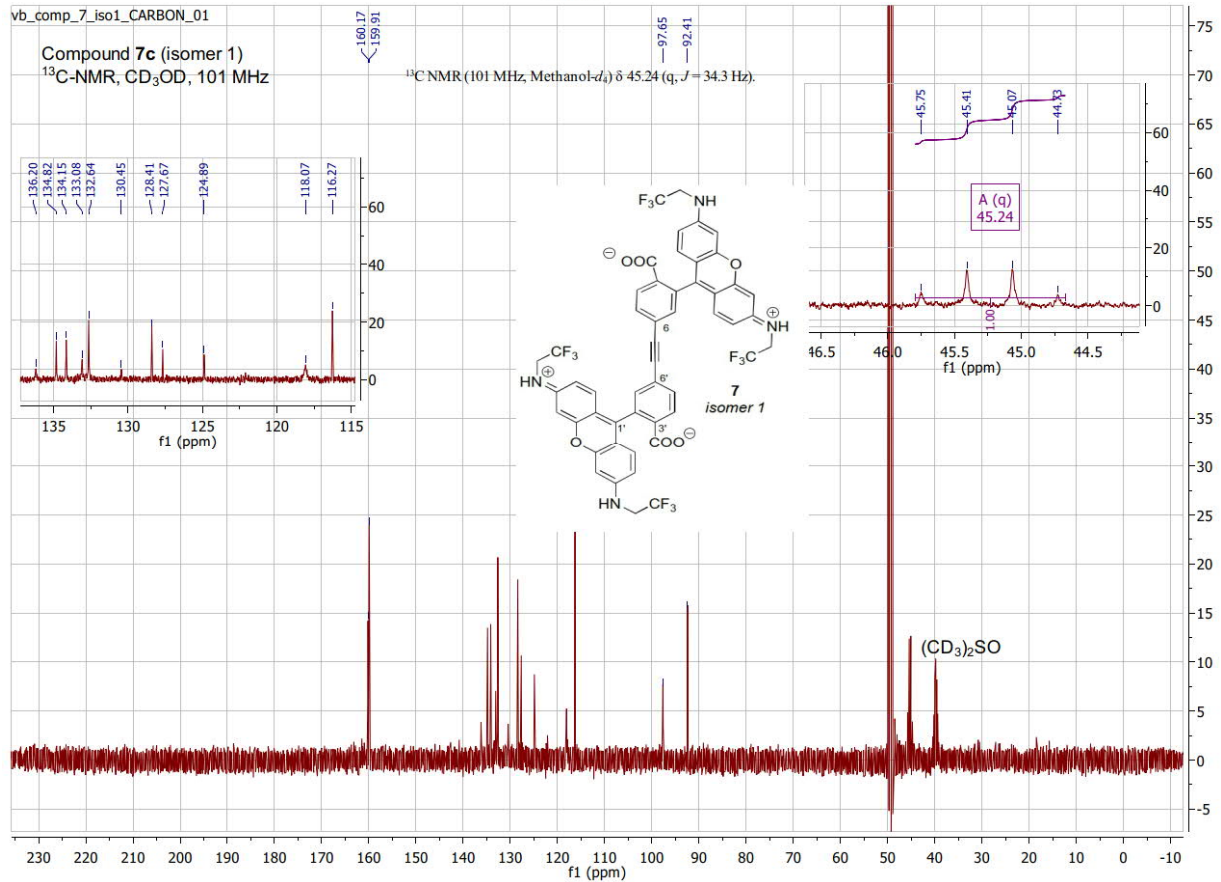
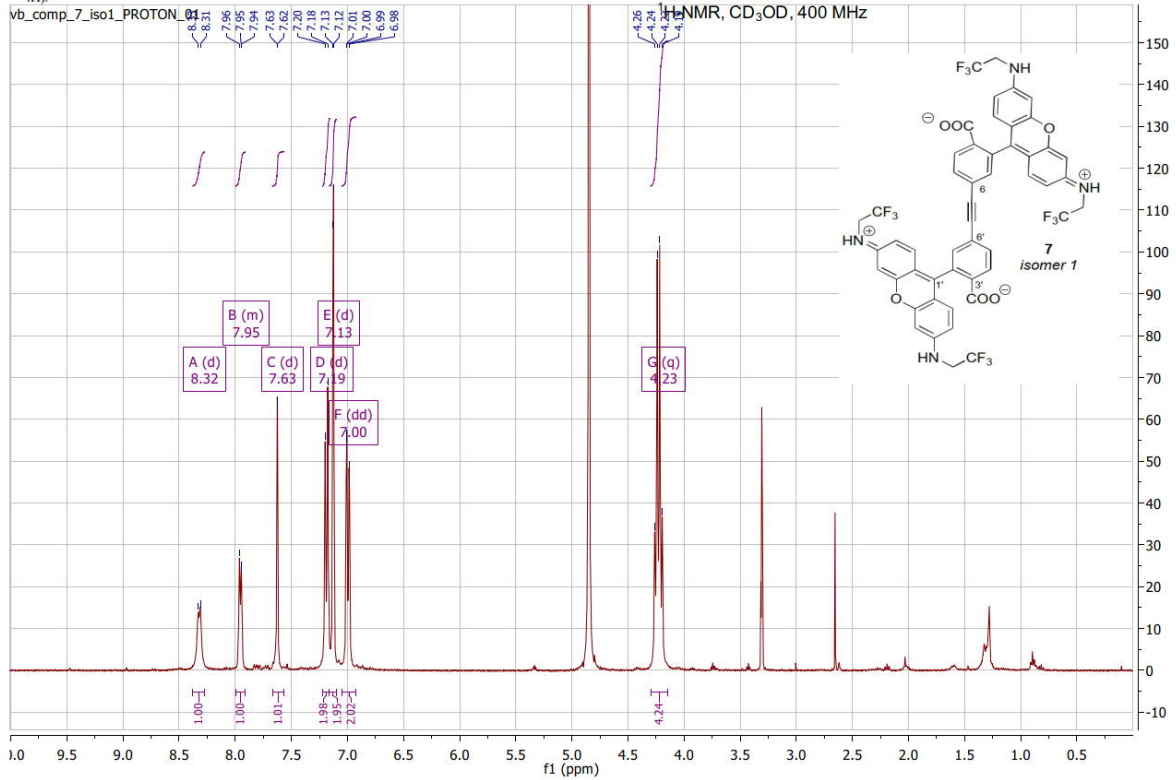
Probe : **shs477-31 iso4 prep Peak 1 RT=9,8 nach N**  
Lösungsmittel : **MeCN/H<sub>2</sub>O**      Aufgabemenge: **3.0 µl**  
Säule: Phenomenex      2.6 µm      Länge: 75 mm      iO : 3.0 mm  
Fluß (ml / Min) : **0.5**      Temperatur : **25.0**  
Detektor: DAD-3000      Pumpe: HPG-3200SD      Sampler: WPS-3000  
Laufmittel: **A = Acetonitril 0.1% FA**      **B = Wasser 0.1% FA**  
Gradient: A 20.0 %      B 80.0 %      ---->      A 100.0 %      B 0.0 %      T = 10 Min.



<sup>1</sup>H NMR (400 MHz, Methanol-*d*<sub>4</sub>) δ 8.32 (d, *J* = 7.9 Hz, 1H), 8.00 – 7.91 (m, 1H), 7.63 (d, *J* = 1.4 Hz, 1H), 7.19 (d, *J* = 9.2 Hz, 2H), 7.13 (d, *J* = 2.1 Hz, 2H), 7.00 (dd, *J* = 9.1, 2.2 Hz, 2H), 4.23 (q, *J* = 9.0 Hz, 4H).

Compound **7c** (isomer 1)

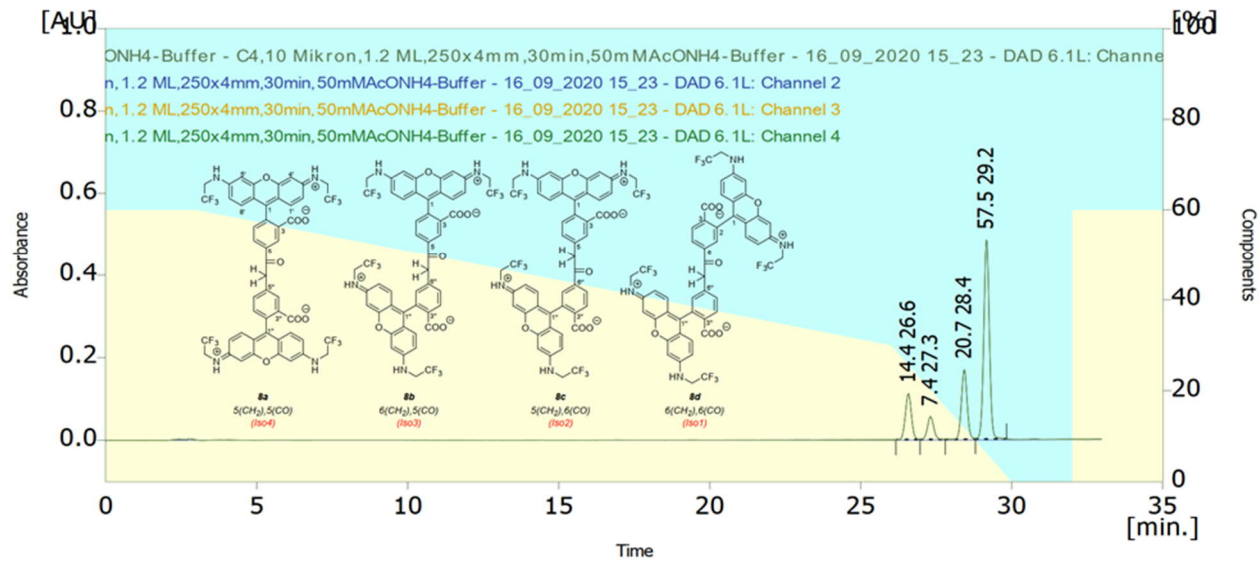
<sup>1</sup>H NMR, CD<sub>3</sub>OD, 400 MHz



Bis-rhodamine ethanones **8a** [5(CH<sub>2</sub>),5(CO)] (peak 4), **8b** [6(CH<sub>2</sub>),5(CO)] (peak 3), **8c** [5(CH<sub>2</sub>),6(CO)] (peak 2) and **8d** [6(CH<sub>2</sub>),6(CO)] (peak 1)

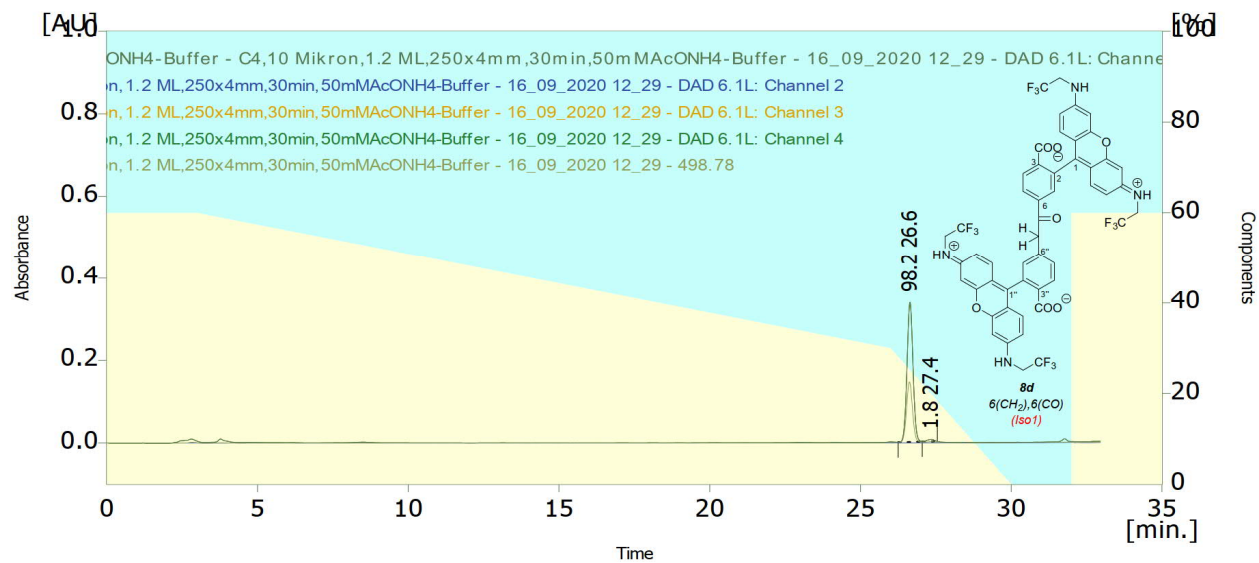
### HPLC trace of the mixture of regioisomers

Chromatogram C:\ClarityChrom\WORK1\Heydar\SHS479-31-Iso1234-ACN-H2O+50mM-AcONH4-Buffer-40-60-70-30-254-630nm C4,10 Mikron,1.2 ML,250x4mm,30min,

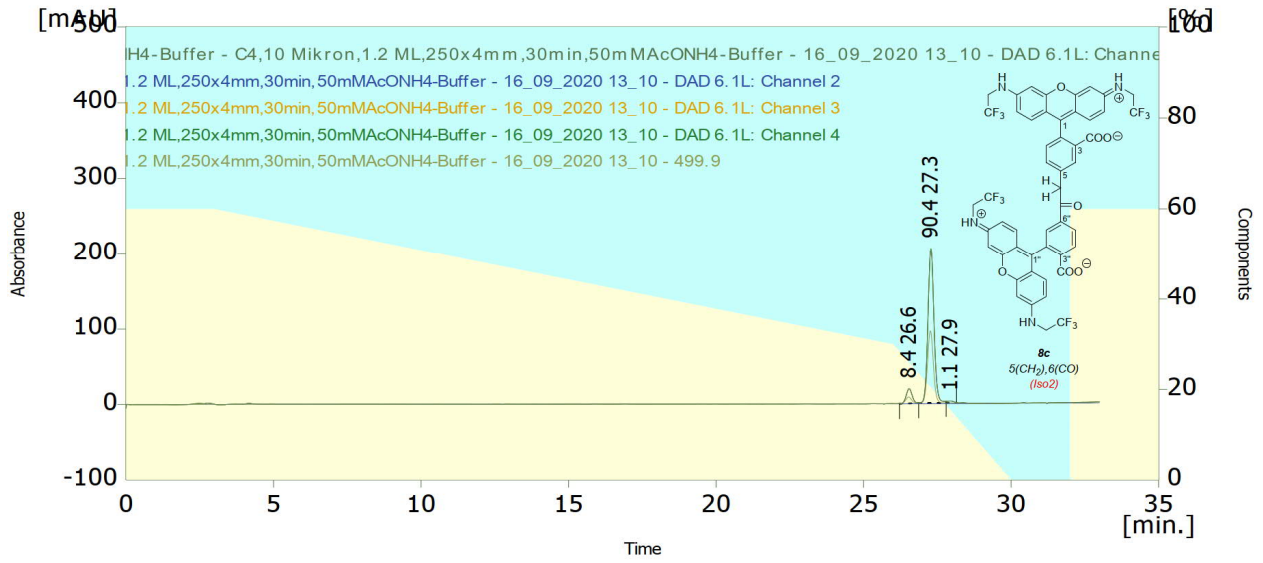


### HPLC traces and NMR spectra of the isolated individual regioisomers **8a-d**

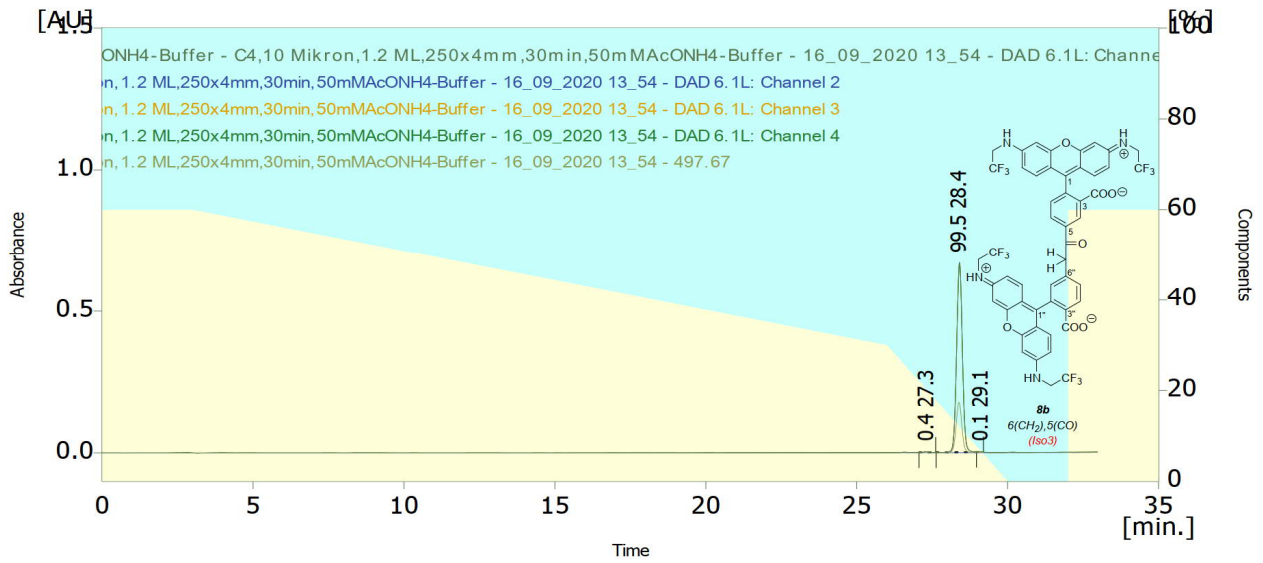
Chromatogram C:\ClarityChrom\WORK1\Heydar\SHS479-31-Iso1-ACN-H2O+50mM-AcONH4-Buffer-40-60-70-30-254-630nm C4,10 Mikron,1.2 ML,250x4mm,30min,5

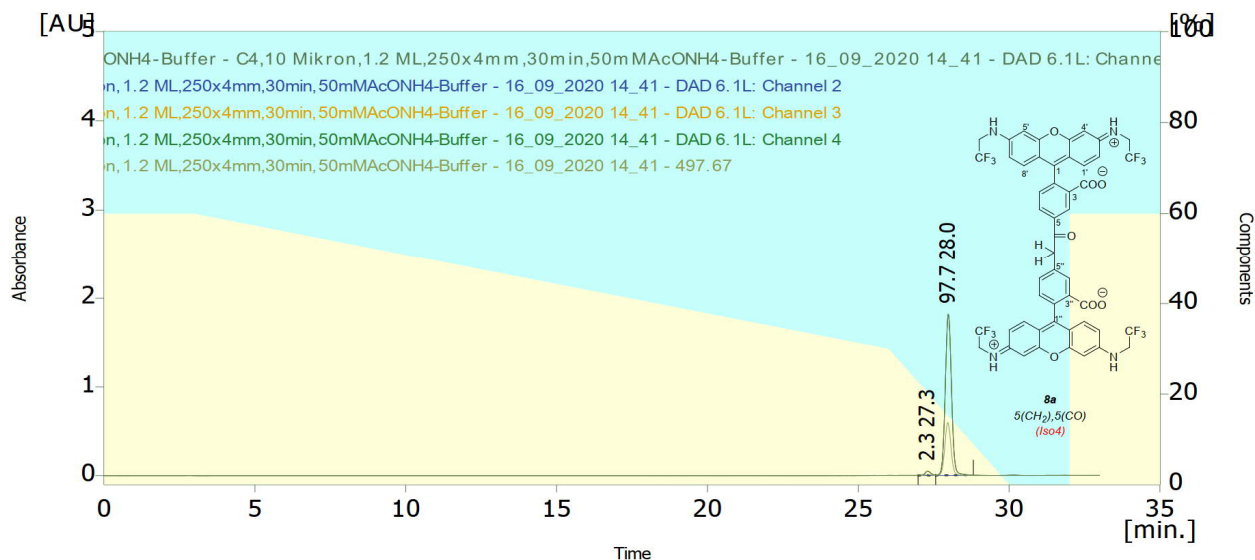


Chromatogram C:\ClarityChrom\WORK1\Heydar\SHS479-31-Iso2-ACN-H2O+50mM-AcONH4-Buffer-40-60-70-30-254-630nm C4,10 Mikron,1.2 ML,250x4mm,30min,5



Chromatogram C:\ClarityChrom\WORK1\Heydar\SHS479-31-Iso3-ACN-H2O+50mM-AcONH4-Buffer-40-60-70-30-254-630nm C4,10 Mikron,1.2 ML,250x4mm,30min,5

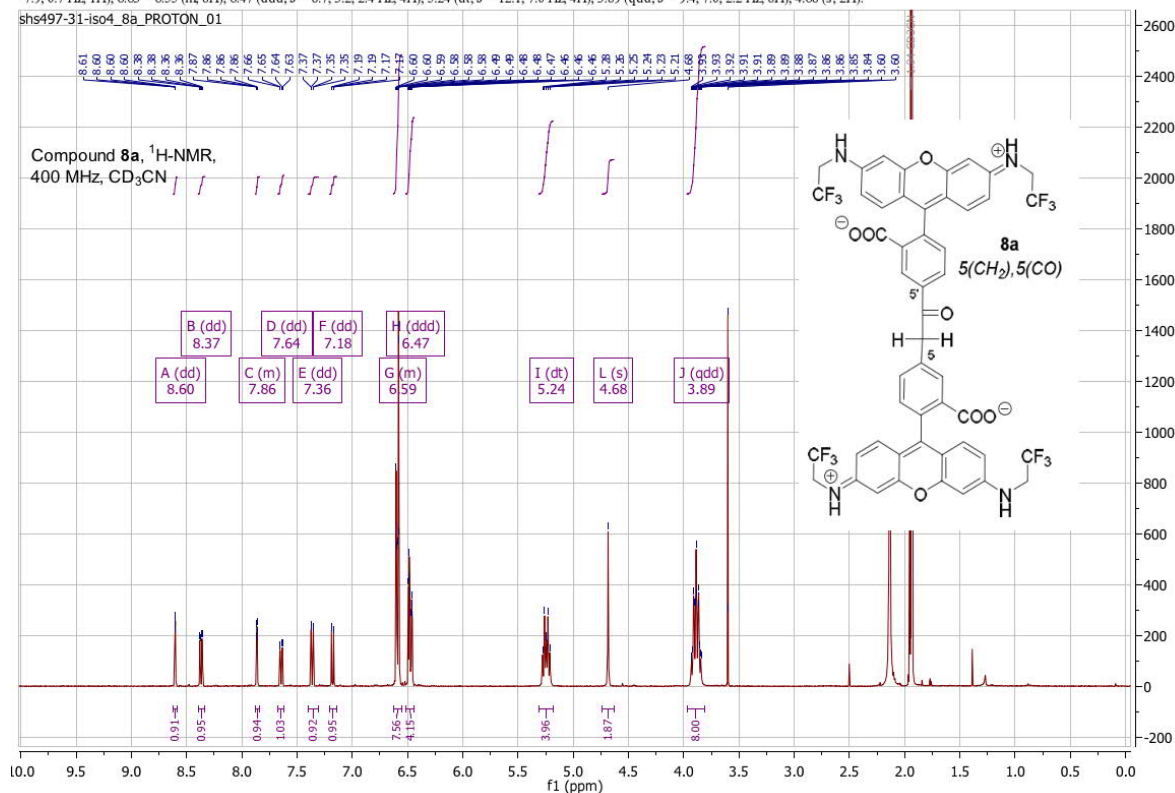


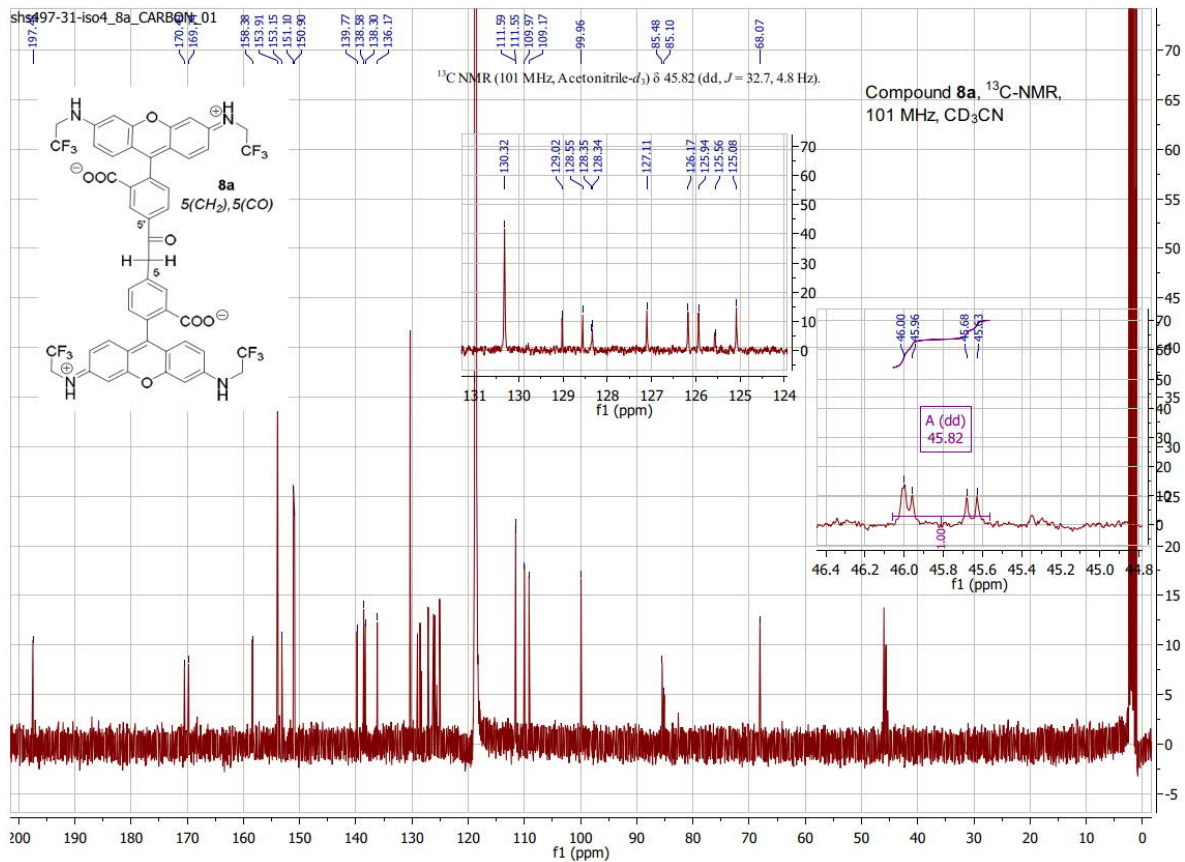


NMR spectra of compound **8a** [5(CH<sub>2</sub>)-6(CO)]: <sup>1</sup>H (400 MHz) and <sup>13</sup>C{<sup>1</sup>H} (101 MHz)

<sup>1</sup>H NMR (400 MHz, Acetonitrile-d<sub>3</sub>) δ 8.60 (dd, J = 1.6, 0.7 Hz, 1H), 8.37 (dd, J = 8.1, 1.6 Hz, 1H), 7.87 – 7.84 (m, 1H), 7.64 (dd, J = 7.9, 1.6 Hz, 1H), 7.36 (dd, J = 8.0, 0.7 Hz, 1H), 7.18 (dd, J = 7.9, 0.7 Hz, 1H), 6.63 – 6.55 (m, 8H), 6.47 (ddd, J = 8.7, 3.2, 2.4 Hz, 4H), 5.24 (dt, J = 12.1, 7.0 Hz, 4H), 3.89 (qdd, J = 9.4, 7.0, 2.2 Hz, 8H), 4.68 (s, 2H).

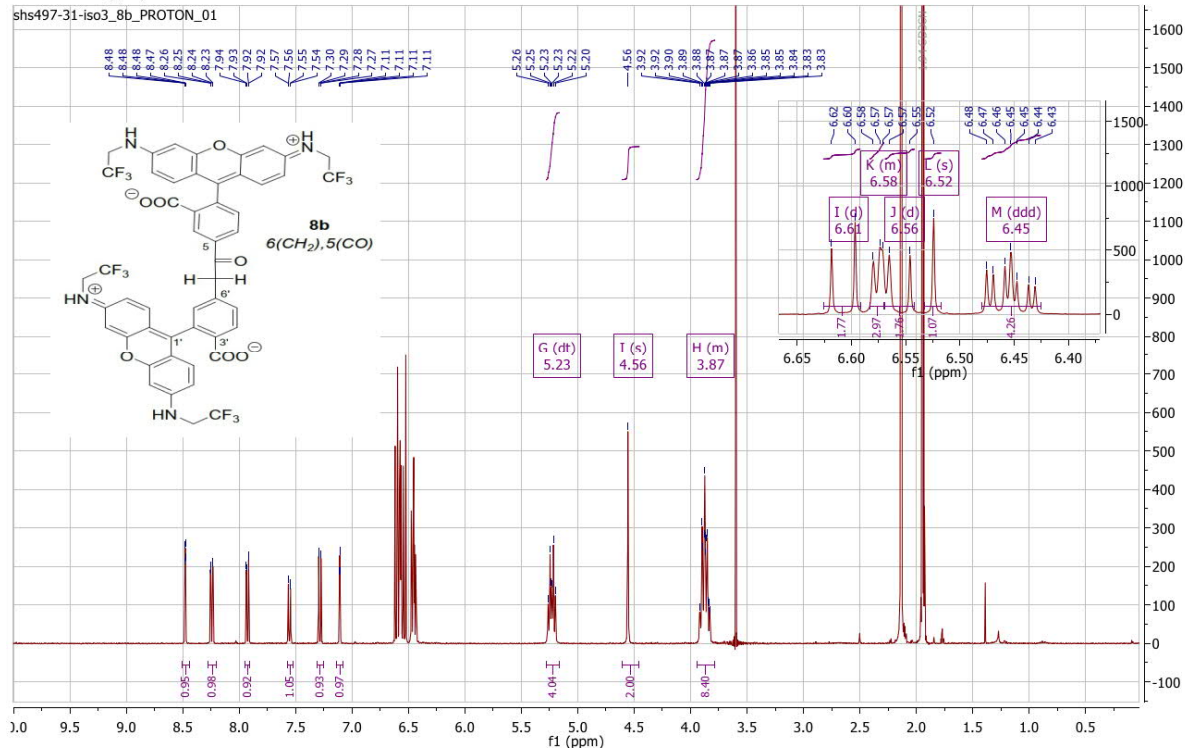
shs497-31-iso4\_8a\_PROTON\_01

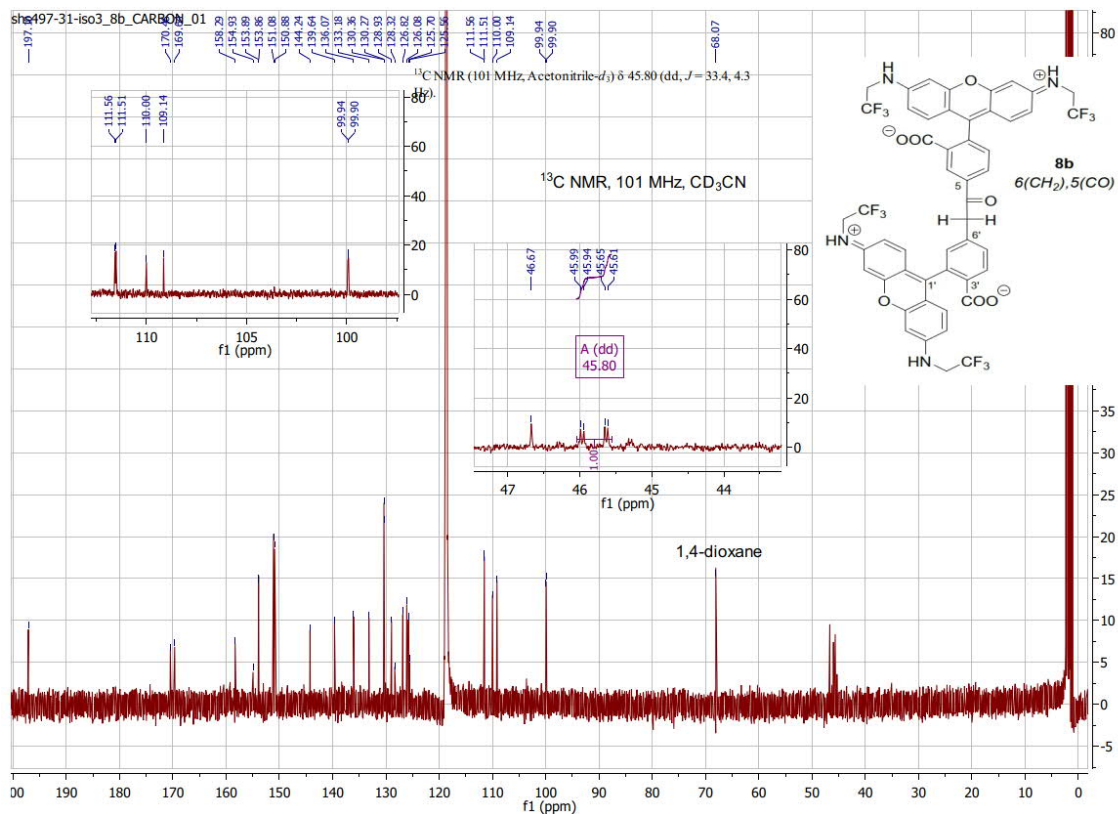




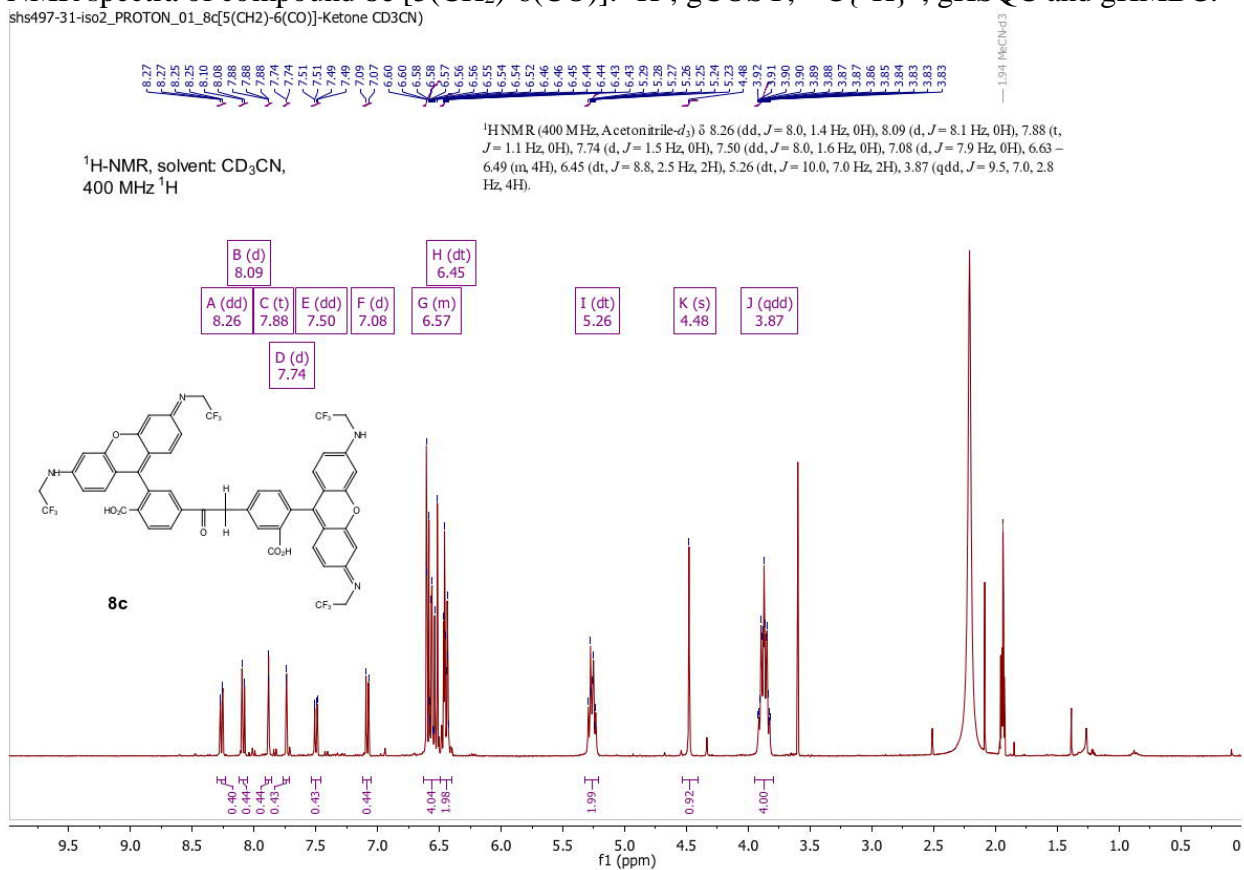
**NMR spectra of compound 8b [6(CH<sub>2</sub>)-5(CO)]:  $^1\text{H}$  (400 MHz) and  $^{13}\text{C}\{^1\text{H}\}$  (101 MHz)**

$^1\text{H}$ NMR (400 MHz,  $\text{CD}_3\text{CN}$ )  $\delta$  8.48 (dd,  $J = 1.6, 0.8$  Hz, 1H), 8.24 (dd,  $J = 8.1, 1.6$  Hz, 1H), 7.93 (dd,  $J = 7.9, 0.7$  Hz, 1H), 7.56 (dd,  $J = 7.9, 1.4$  Hz, 1H), 7.28 (dd,  $J = 8.1, 0.7$  Hz, 1H), 7.11 (dd,  $J = 1.4, 0.7$  Hz, 1H), 6.61 (d,  $J = 8.7$  Hz, 2H), 6.58 – 6.57 (m, 3H), 6.56 (d,  $J = 7.6$  Hz, 2H), 6.52 (s, 1H), 6.45 (ddd,  $J = 8.9, 6.7, 2.4$  Hz, 4H), 5.23 (dt,  $J = 11.7, 7.0$  Hz, 4H), 3.95 – 3.79 (m, 8H) 4.56 (s, 2H).

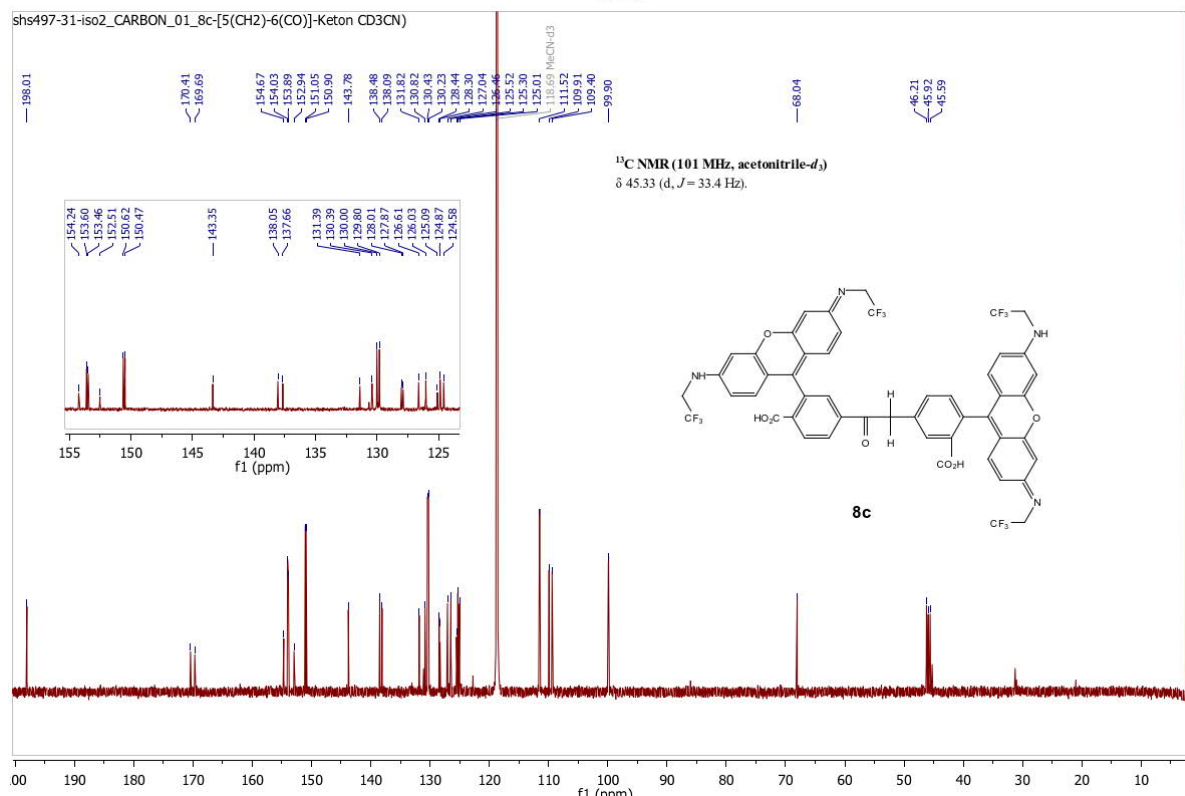
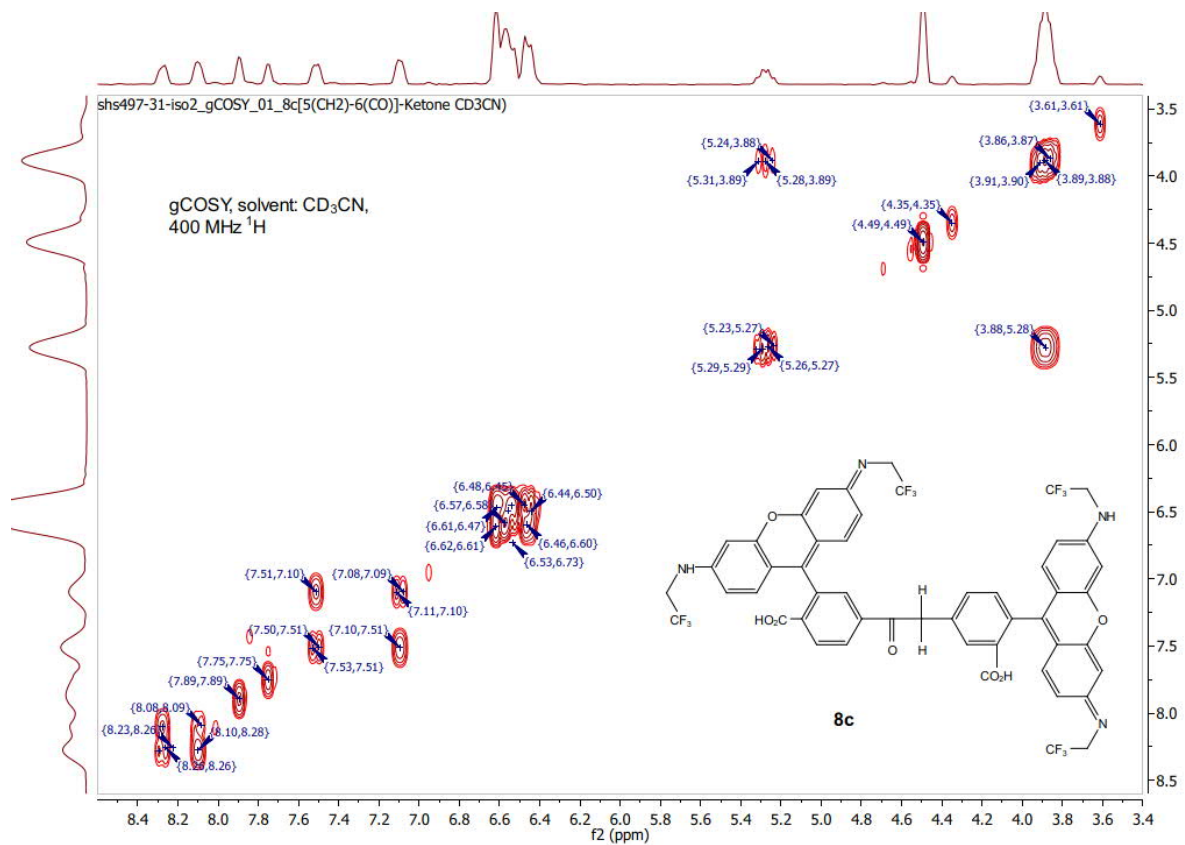


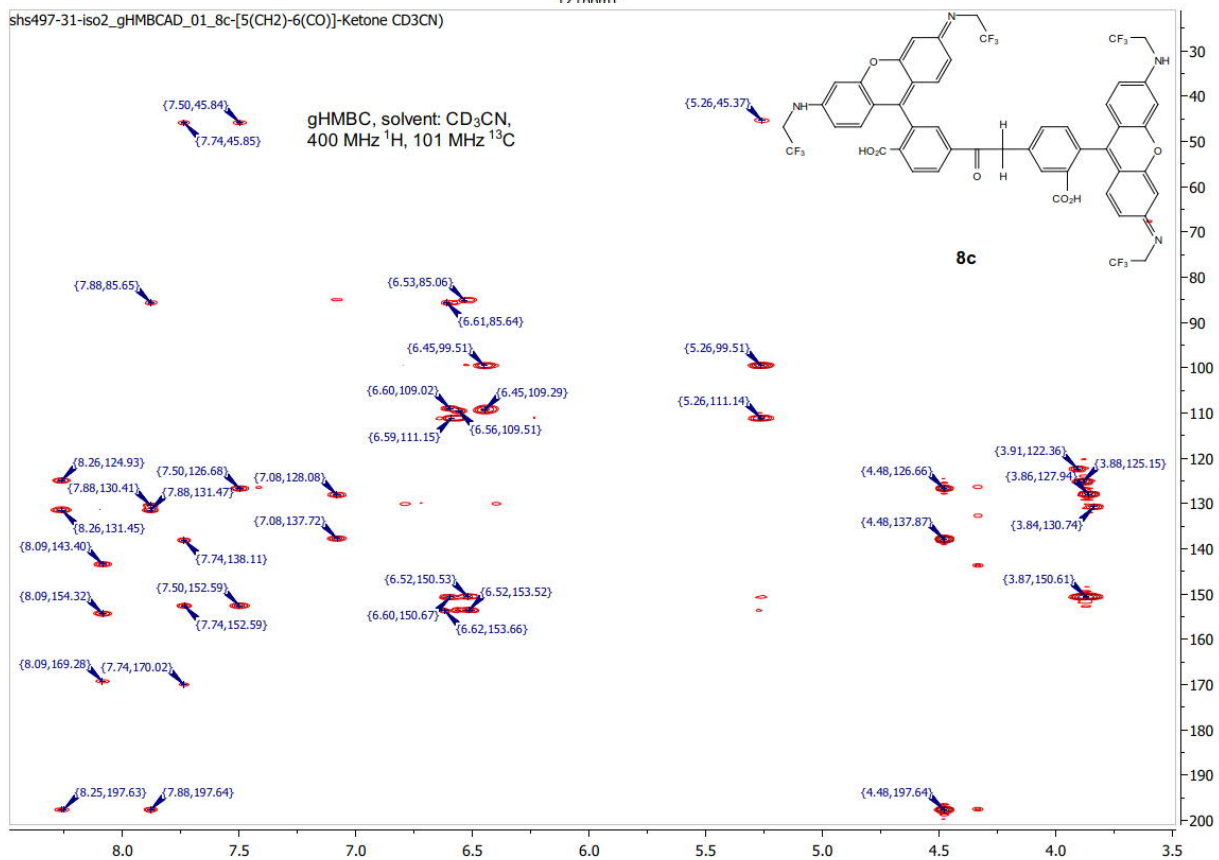
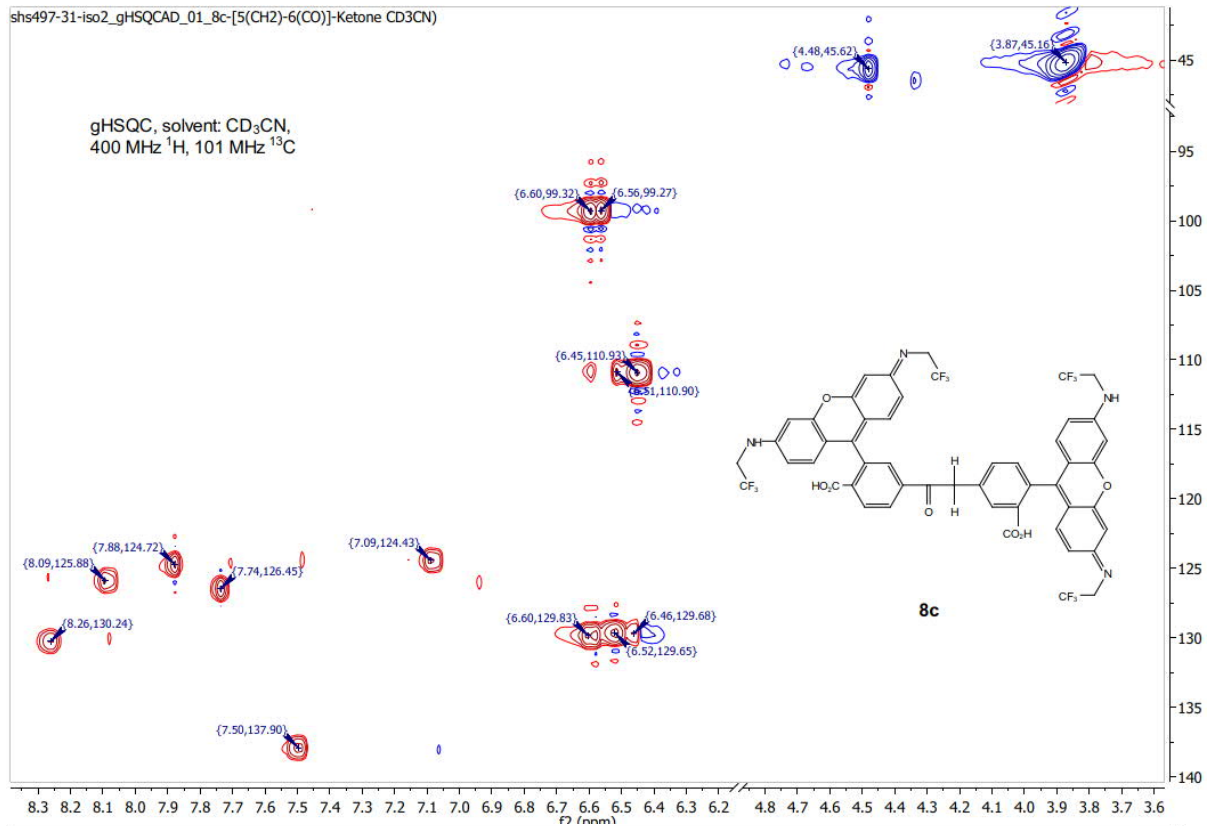


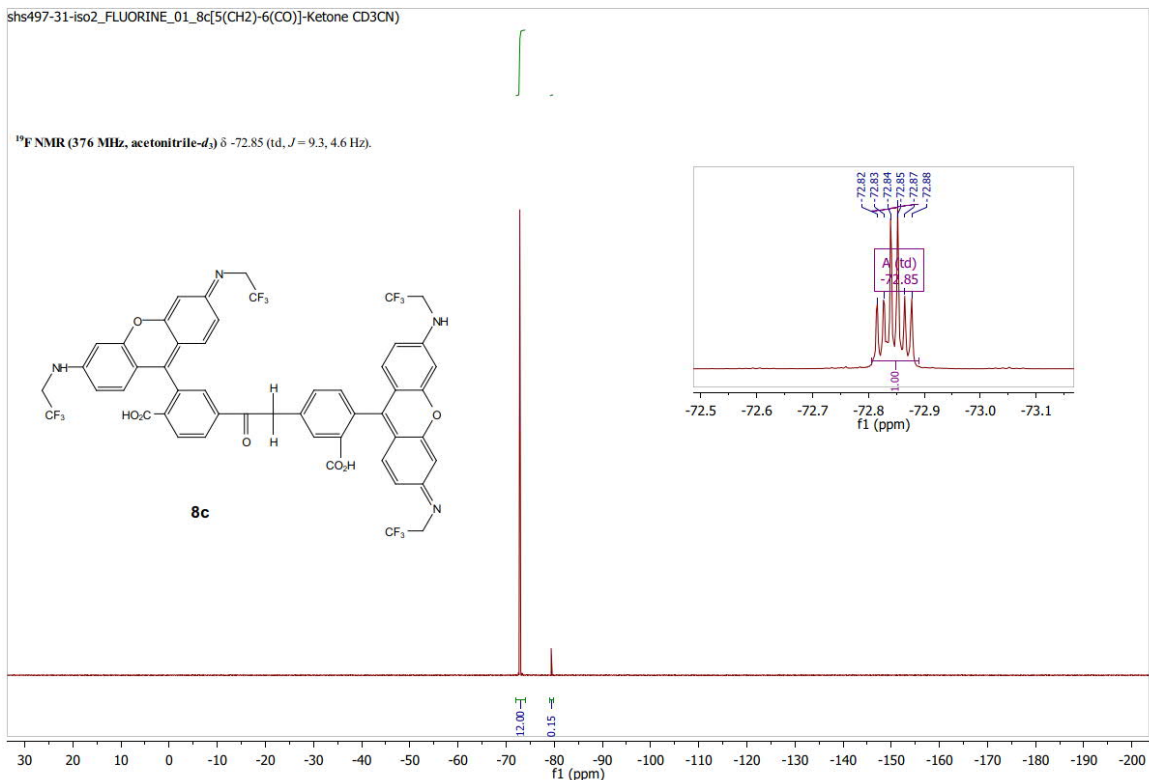
NMR spectra of compound **8c** [5( $\text{CH}_2$ )-6(CO)]:  $^1\text{H}$ -, gCOSY,  $^{13}\text{C}\{^1\text{H}\}$ -, gHSQC and gHMBC.



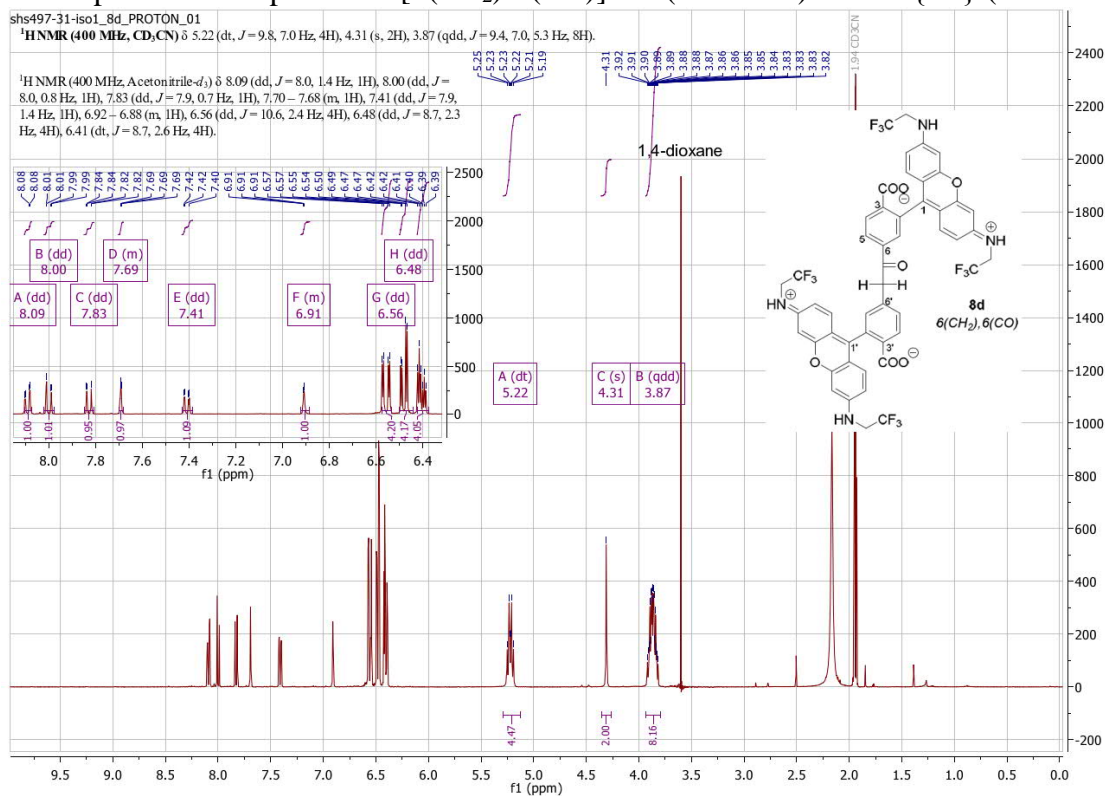


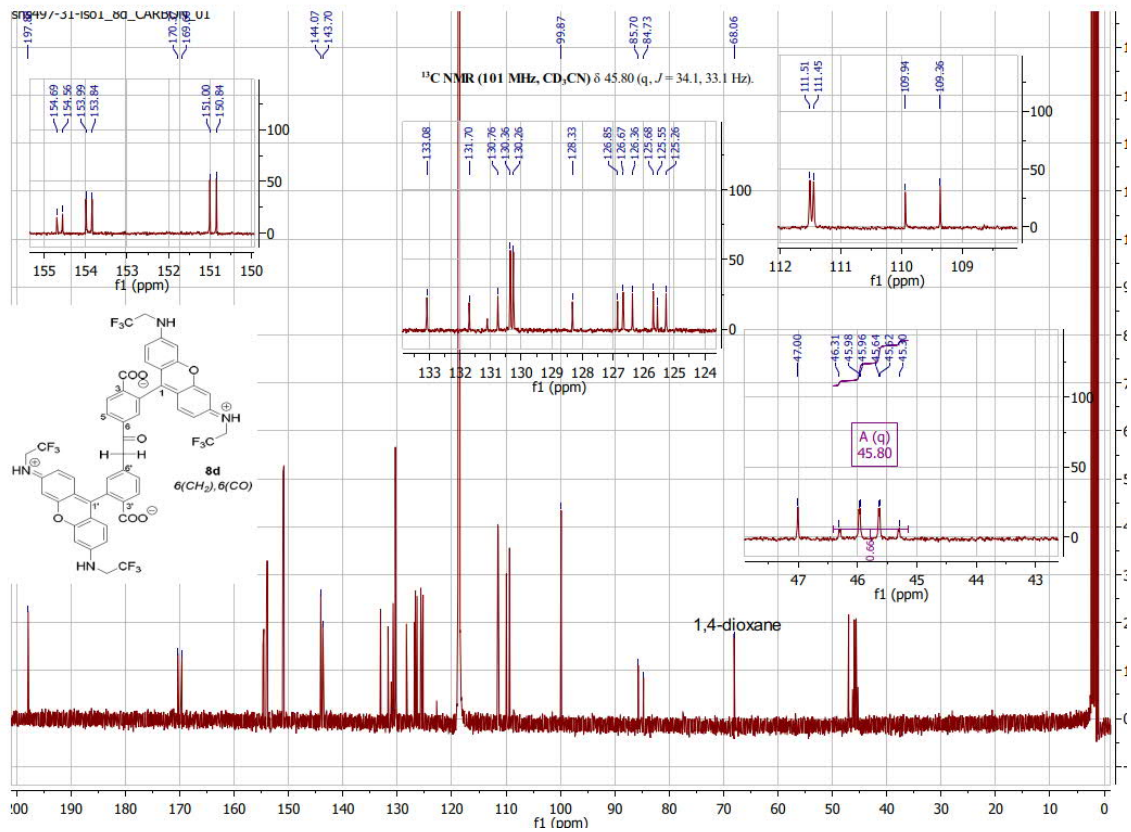






NMR spectra of compound **8d** [6(CH<sub>2</sub>)-6(CO)]: <sup>1</sup>H (400 MHz) and <sup>13</sup>C{<sup>1</sup>H} (101 MHz)

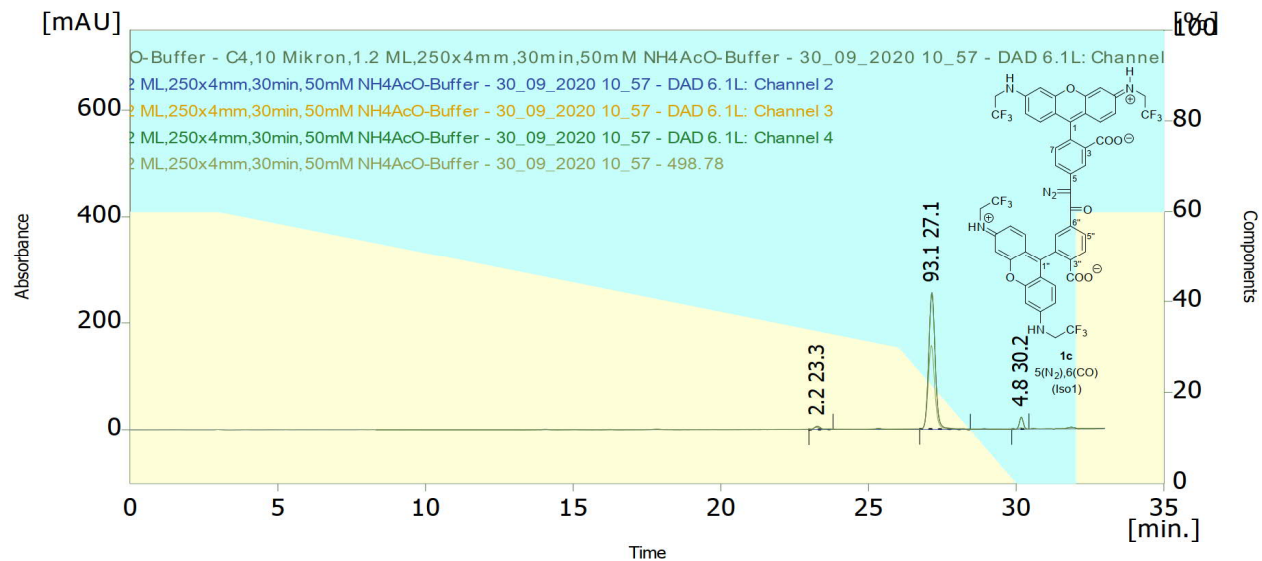




Bis-rhodamine diazoketones **1a** [5(N<sub>2</sub>),5(CO)] (isomer 4), **1b** [6(N<sub>2</sub>),5(CO)] (isomer 3), **1c** [5(N<sub>2</sub>),6(CO)] (isomer 1) and **1d** [6(N<sub>2</sub>),6(CO)] (isomer 2)

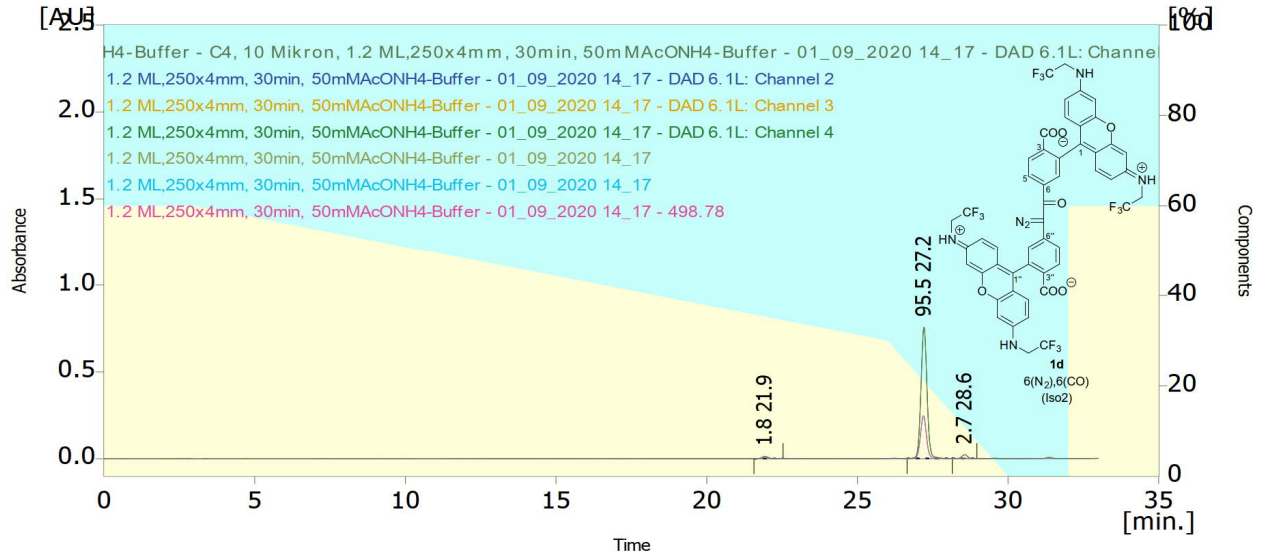
HPLC trace, isomer 1, **1c** [5(N<sub>2</sub>),6(CO)]

Chromatogram C:\ClarityChrom\WORK1\Heydar\SHS505-13-Iso1-ACN-H2O+50mMNH4AcO-40-60-70-30-30min-254-630nm C4,10 Mikron,1.2 ML,250x4mm,30min,50



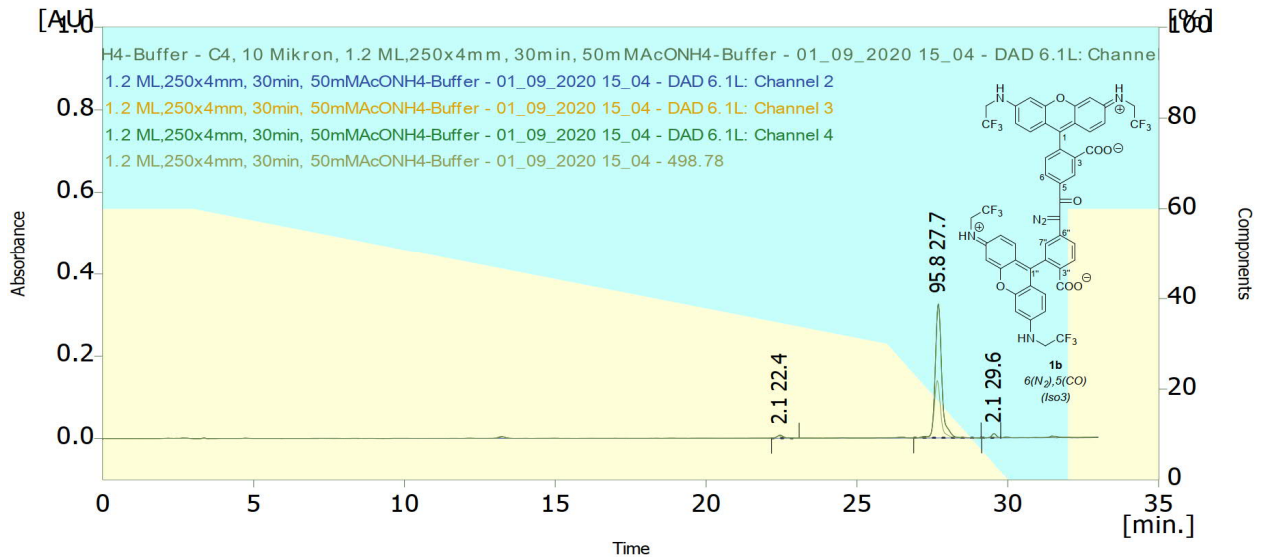
### HPLC trace, isomer 2, **1d** [6(N<sub>2</sub>),6(CO)]

Chromatogram C:\ClarityChrom\WORK1\Heydar\SHS505-11-Iso2-ACN-ACN-H2O+50mM-AcONH4-Buf-40-60-70-30-254-630nm C4, 10 Mikron, 1.2 ML,250x4mm, 30



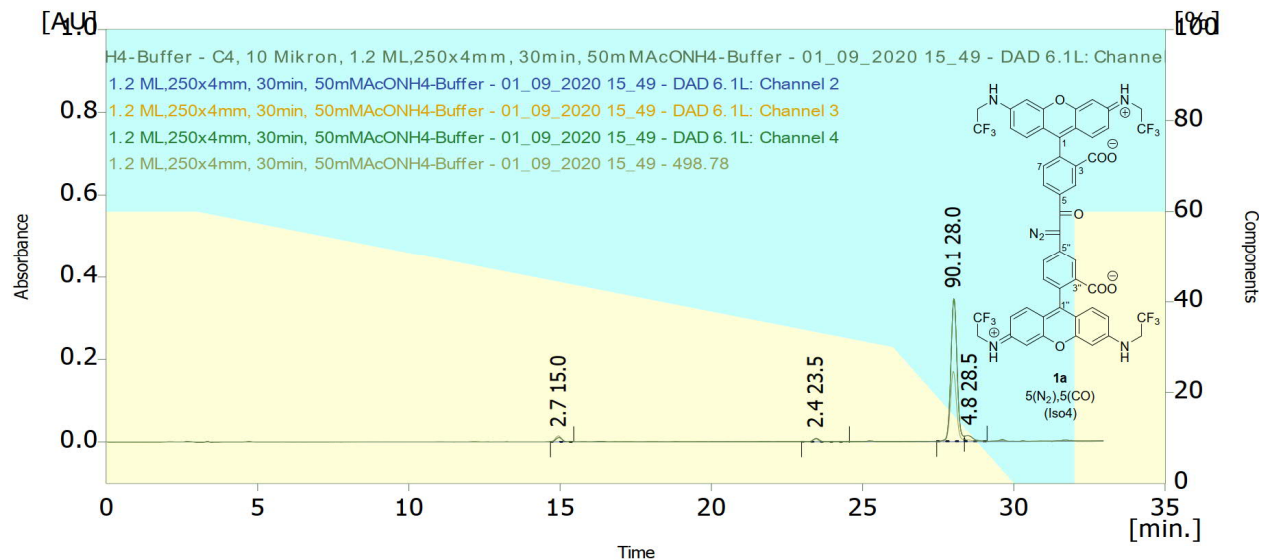
### HPLC trace, isomer 3, **1b** [6(N<sub>2</sub>),5(CO)]

Chromatogram C:\ClarityChrom\WORK1\Heydar\SHS505-11-Iso3-ACN-ACN-H2O+50mM-AcONH4-Buf-40-60-70-30-254-630nm C4, 10 Mikron, 1.2 ML,250x4mm, 30



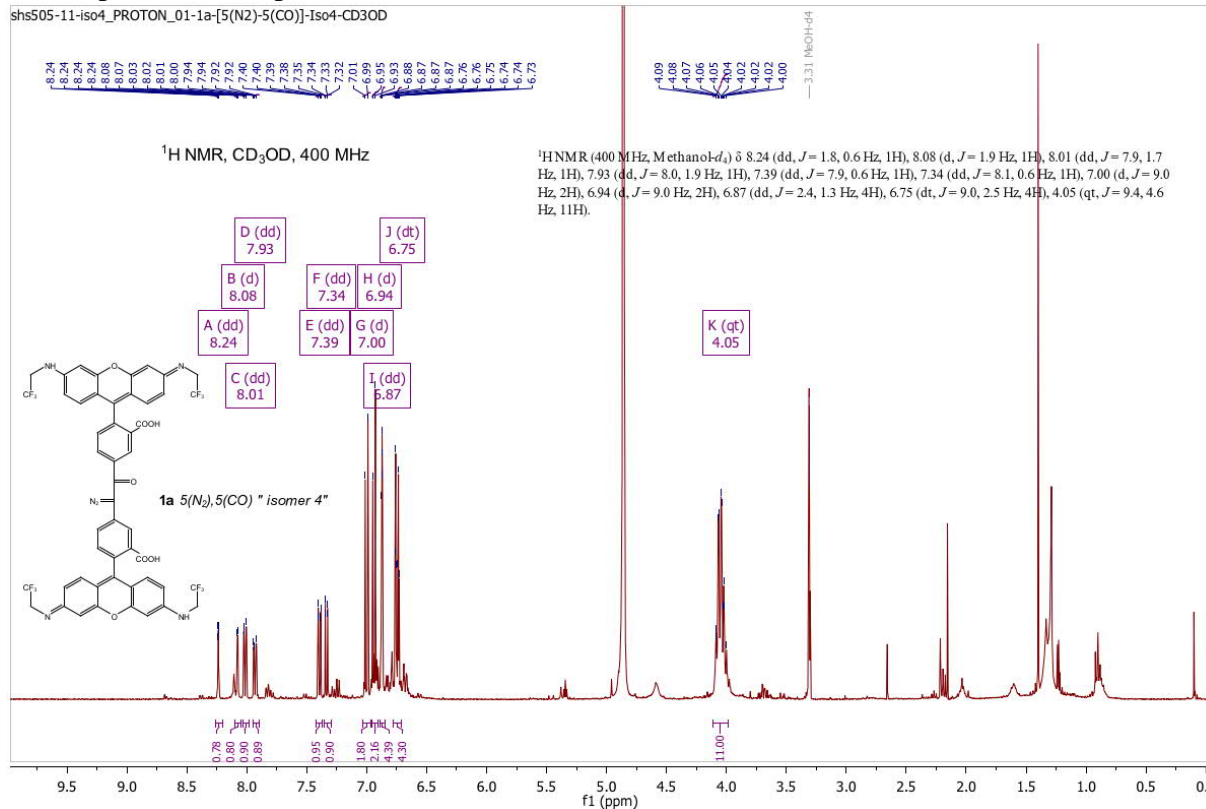
## HPLC trace, isomer 4, **1a** [5(N<sub>2</sub>),5(CO)]

Chromatogram C:\ClarityChrom\WORK1\Heydar\SHS505-11-Iso4-ACN-ACN-H2O+50mM-AcONH4-Buf-40-60-70-30-254-630nm C4, 10 Mikron, 1.2 ML,250x4mm, 30

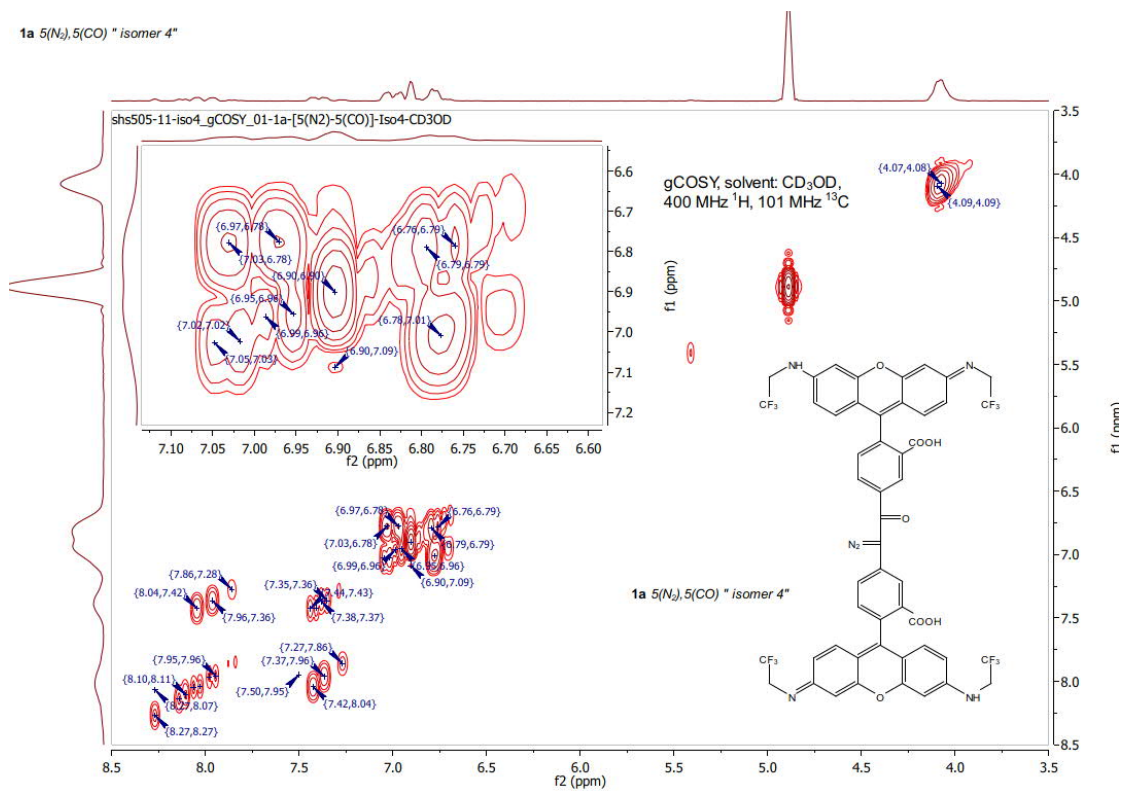


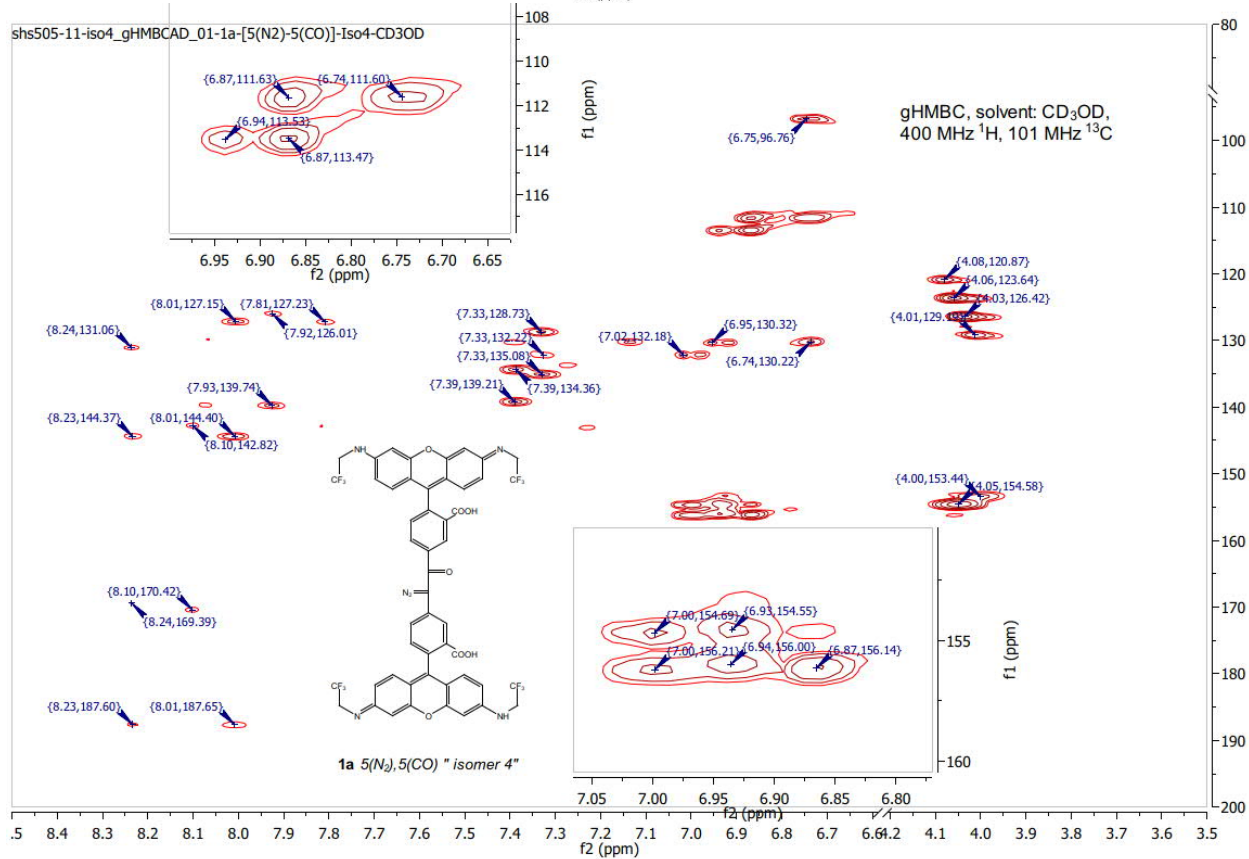
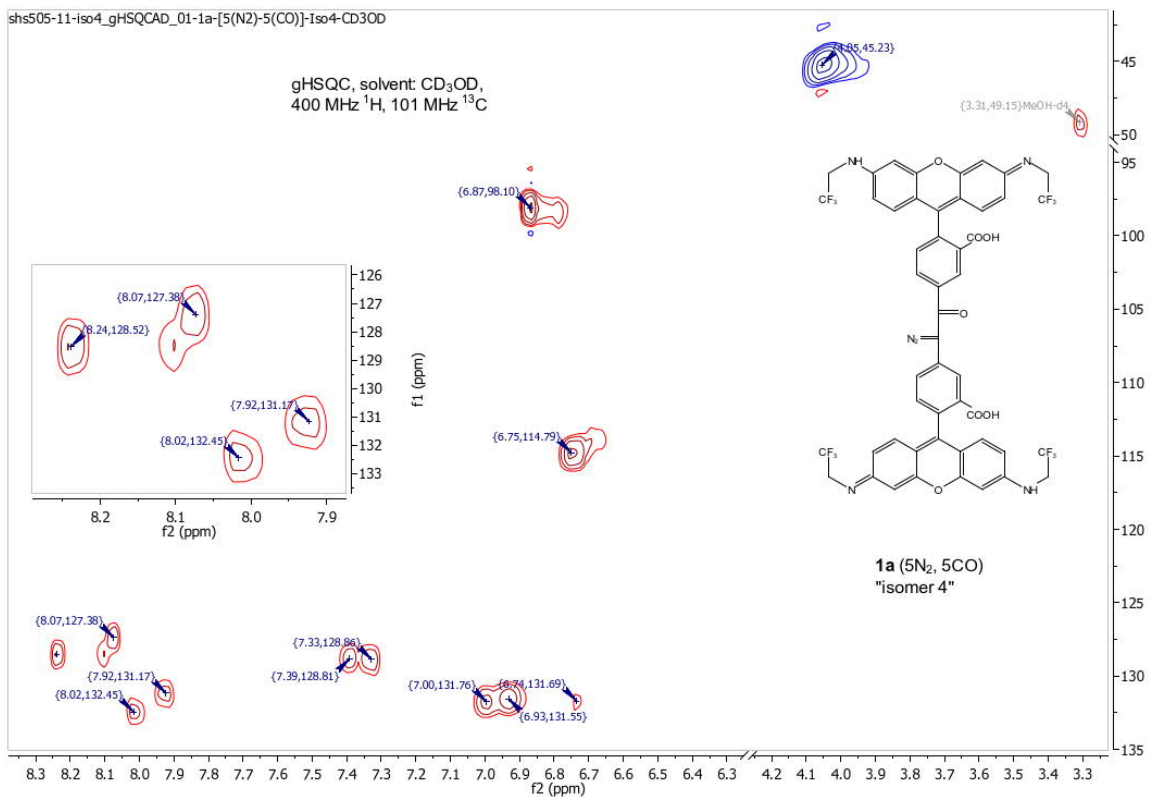
## NMR spectra of compound **1a** [5(N<sub>2</sub>), 5(CO)] (isomer 4)

shs505-11-iso4\_PROTON\_01-1a-[5(N2)-5(CO)]-Iso4-CD3OD



1a 5(N<sub>2</sub>),5(CO) " isomer 4"

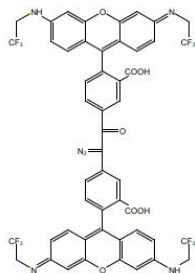




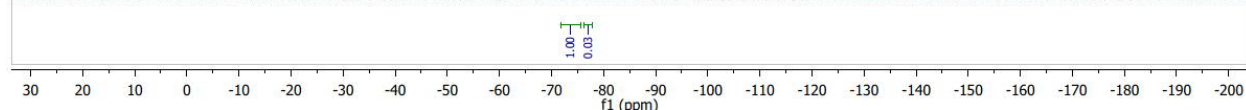
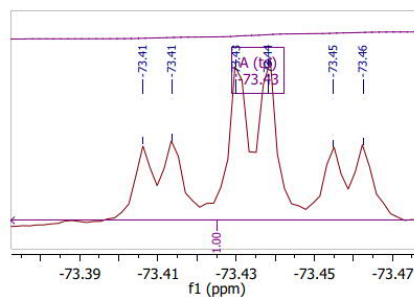


shs505-11-iso4\_FLUORINE\_01-1a-[5(N<sub>2</sub>)-5(CO)]-Iso4-CD3OD  
STANDARD FLUORINE PARAMETERS

<sup>19</sup>F NMR (376 MHz, CD<sub>3</sub>OD) δ -73.43 (td, J = 9.1, 2.8 Hz) ppm



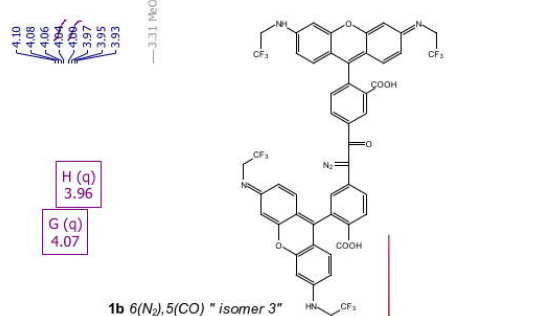
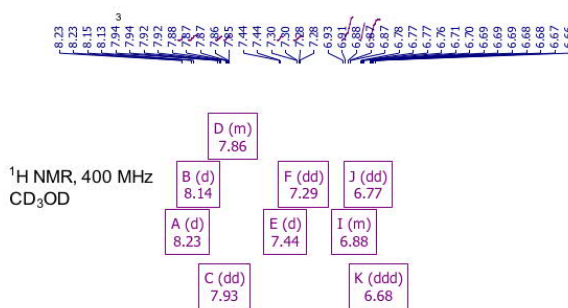
1a 5(N<sub>2</sub>),5(CO) " isomer 4"



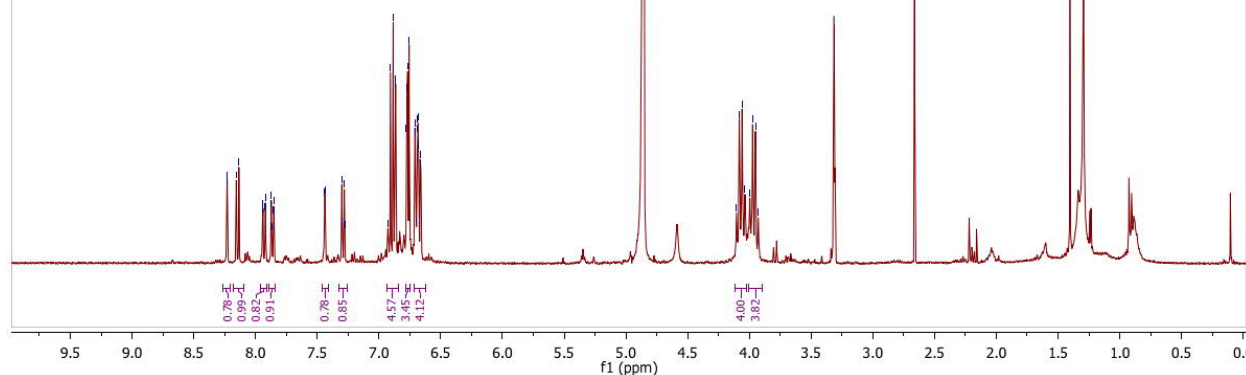
### NMR spectra of compound 1b [6(N<sub>2</sub>),5(CO)] (isomer 3)

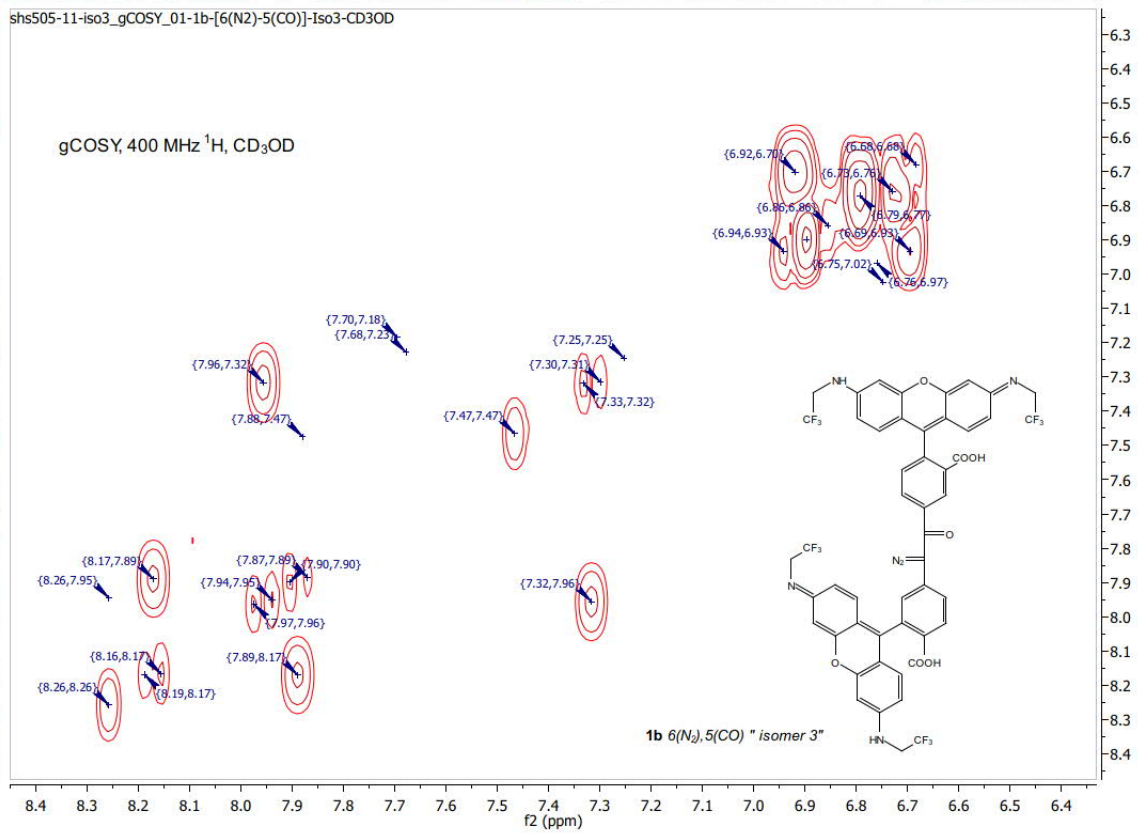
shs505-11-iso3\_PROTON\_01-1b-[6(N<sub>2</sub>)-5(CO)]-Iso3-CD3OD

<sup>1</sup>H NMR (400 MHz, Methanol-d<sub>4</sub>) δ 8.23 (d, J = 1.6 Hz, 1H), 8.14 (d, J = 8.2 Hz, 1H), 7.93 (dd, J = 7.9, 1.6 Hz, 1H), 7.89 – 7.84 (m, 1H), 7.44 (d, J = 1.7 Hz, 1H), 7.29 (dd, J = 7.9, 1.1 Hz, 1H), 6.93 – 6.85 (m, 5H), 6.77 (dd, J = 5.7, 3.3 Hz, 3H), 6.68 (ddd, J = 9.0, 7.8, 2.3 Hz, 4H), 4.07 (q, J = 9.1 Hz, 4H), 3.96 (q, J = 9.1 Hz, 4H).

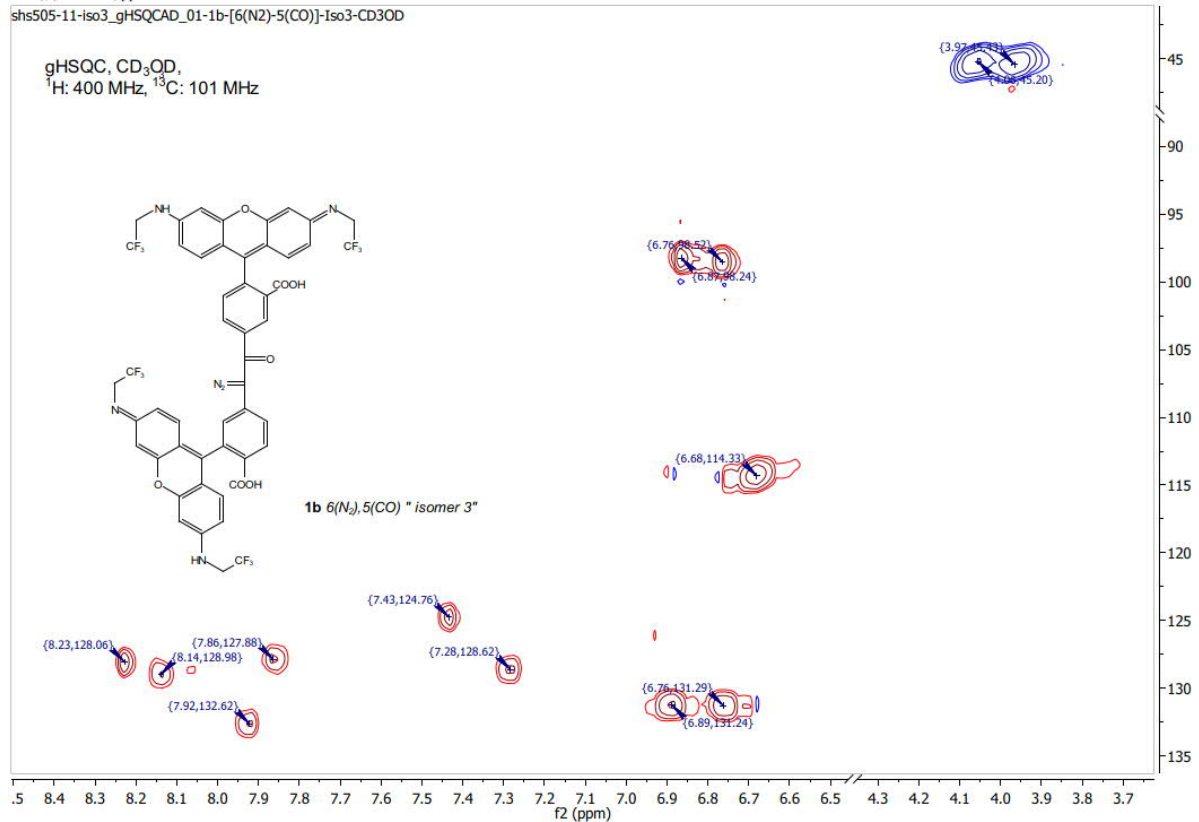


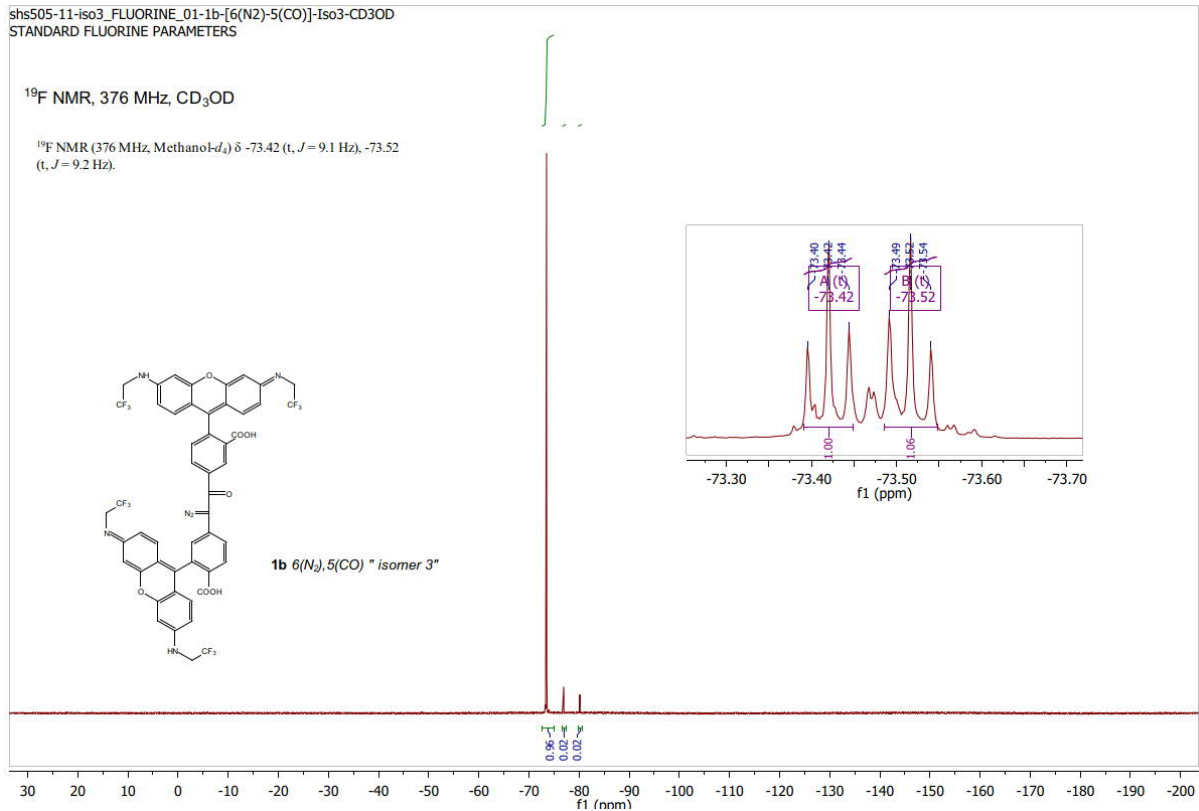
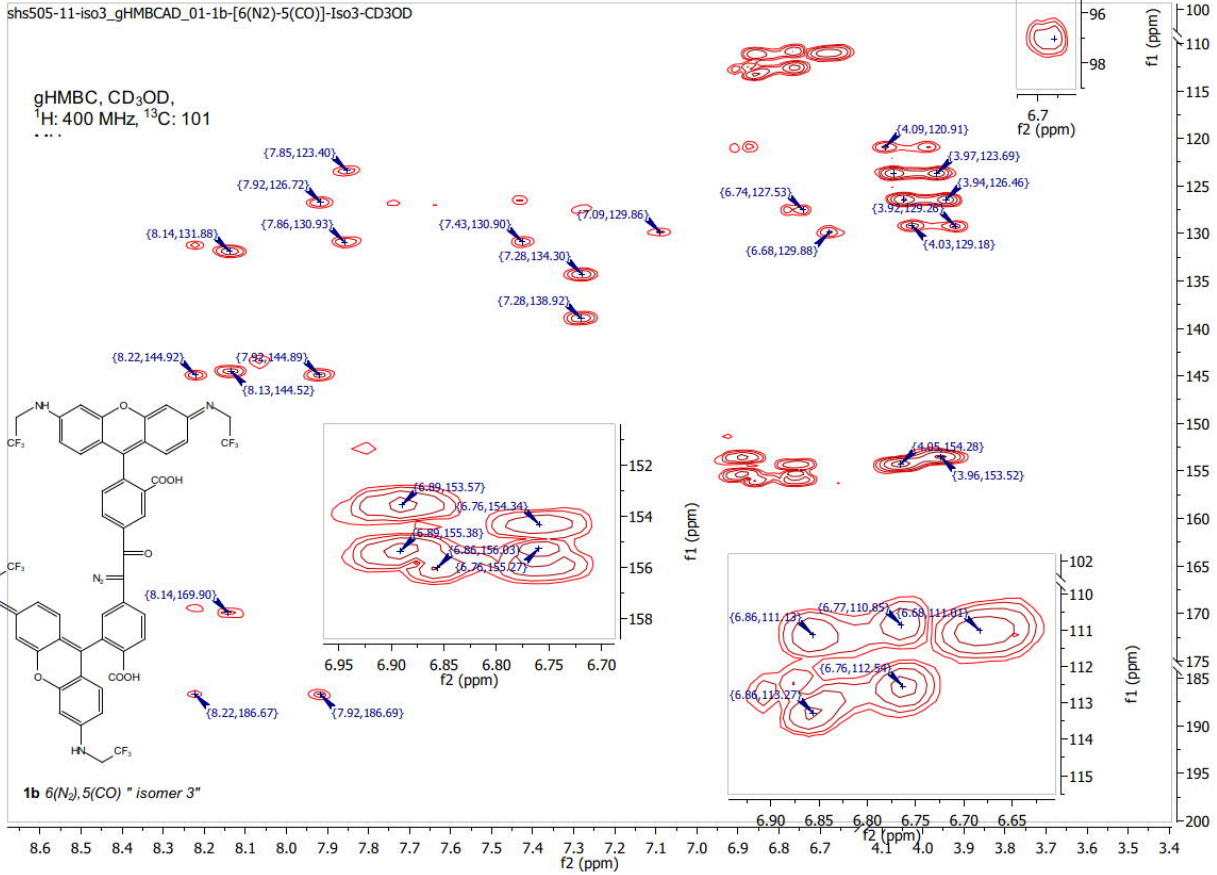
1b 6(N<sub>2</sub>),5(CO) " isomer 3"



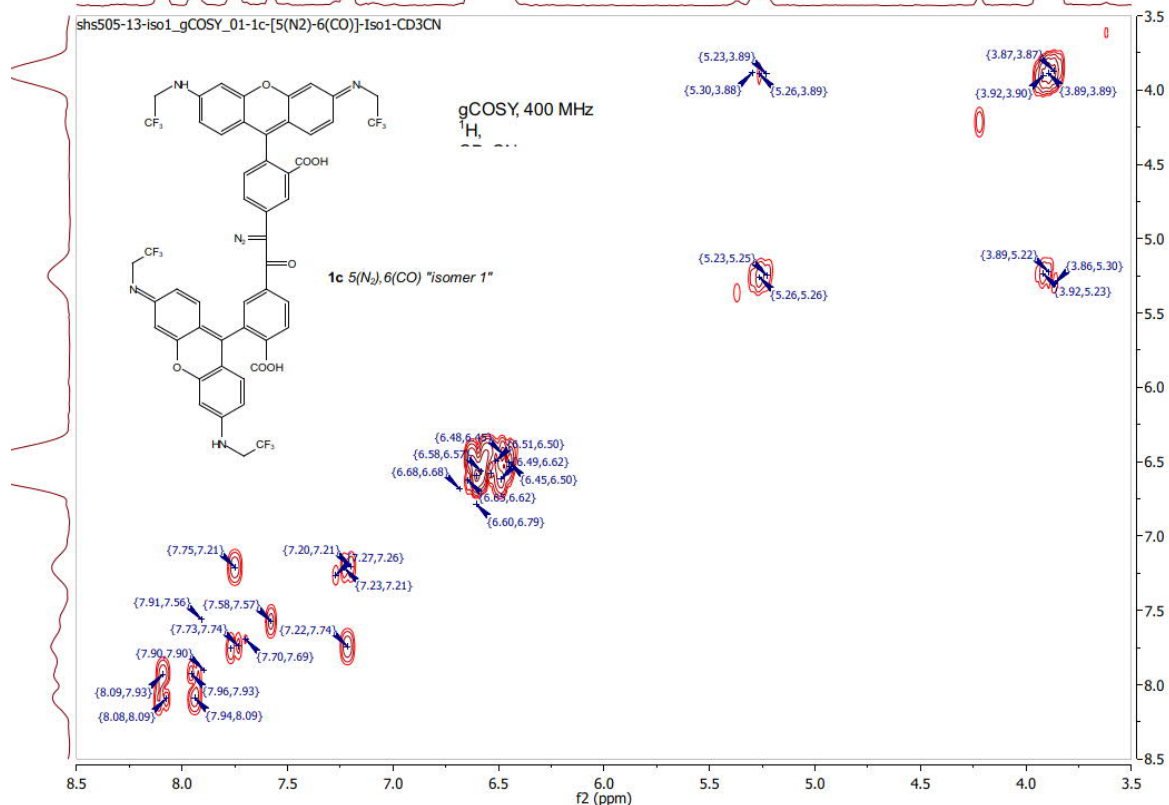
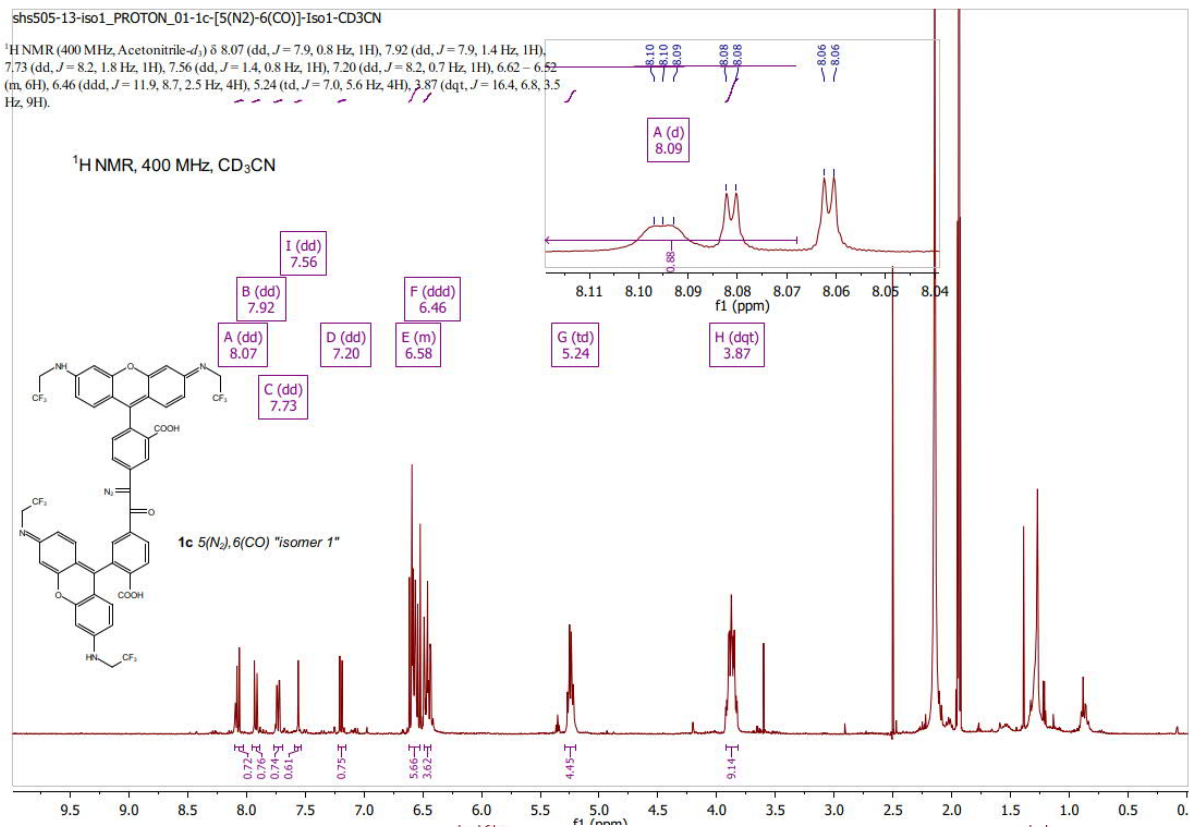


<sup>1</sup>H-<sup>13</sup>C NMR ((400, 101) MHz, cd<sub>3</sub>od) δ (8.26 126.72), (8.17 127.60), (7.95 131.24), (7.89 126.44), (7.46 123.38), (7.31 127.24), (6.92 129.86), (6.90 96.86), (6.79 97.13), (6.79 129.91), (6.71 112.85), (4.09 43.82), (4.00 44.05) ppm.

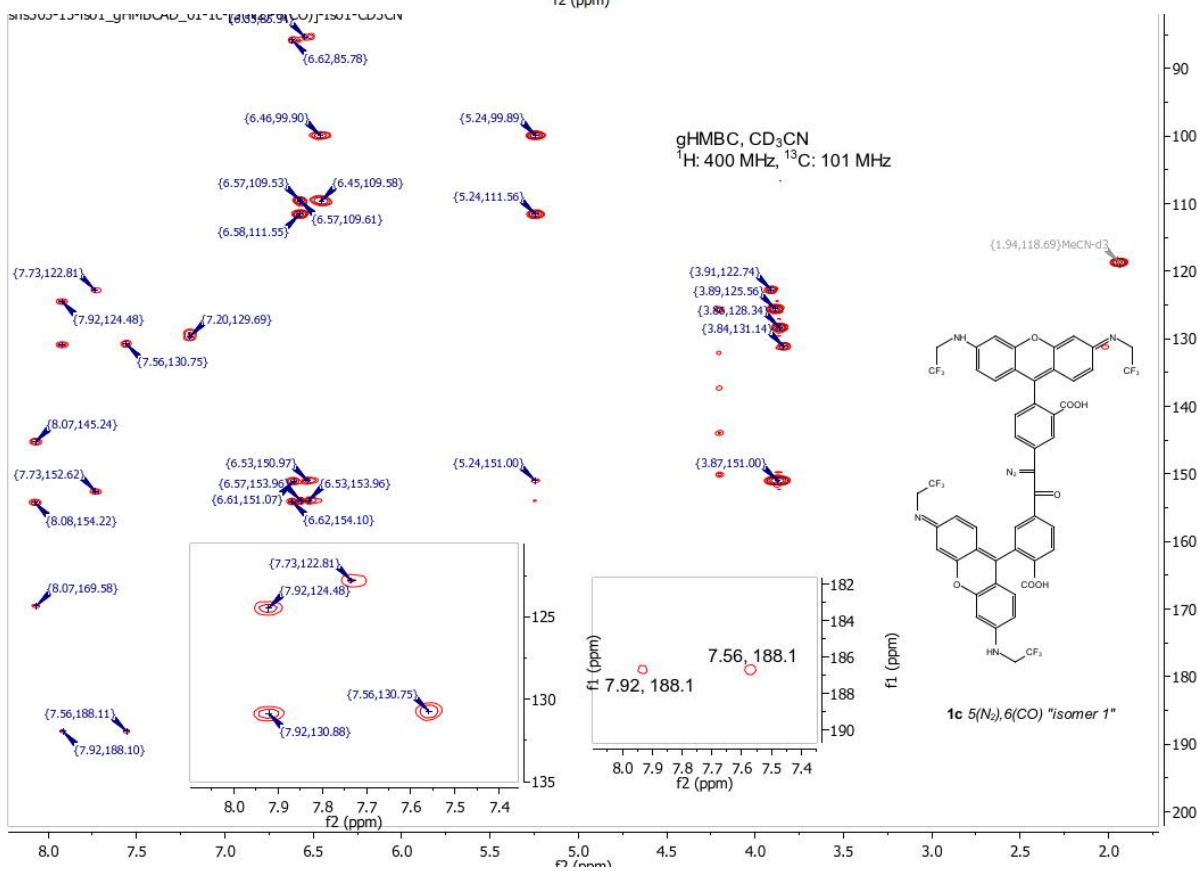
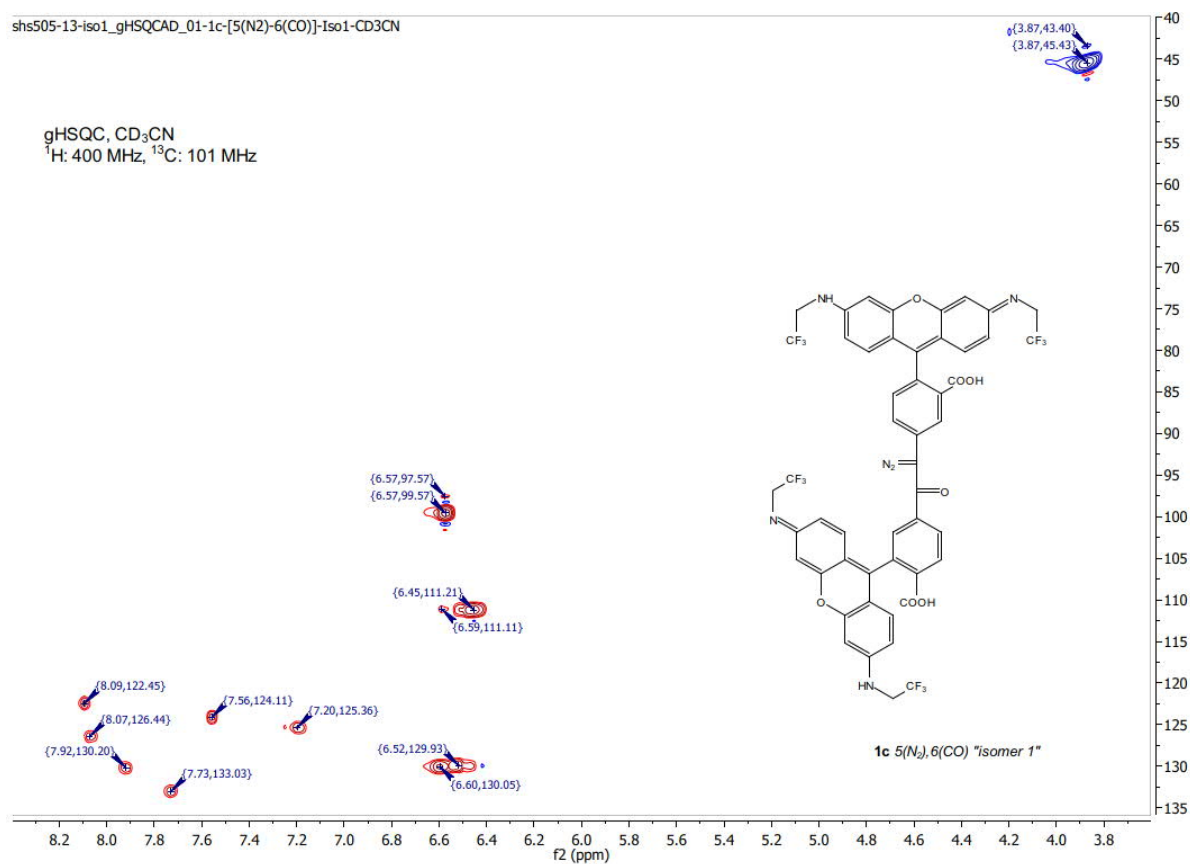




# Compound 1c [5(N<sub>2</sub>),6(CO)] (isomer 1)

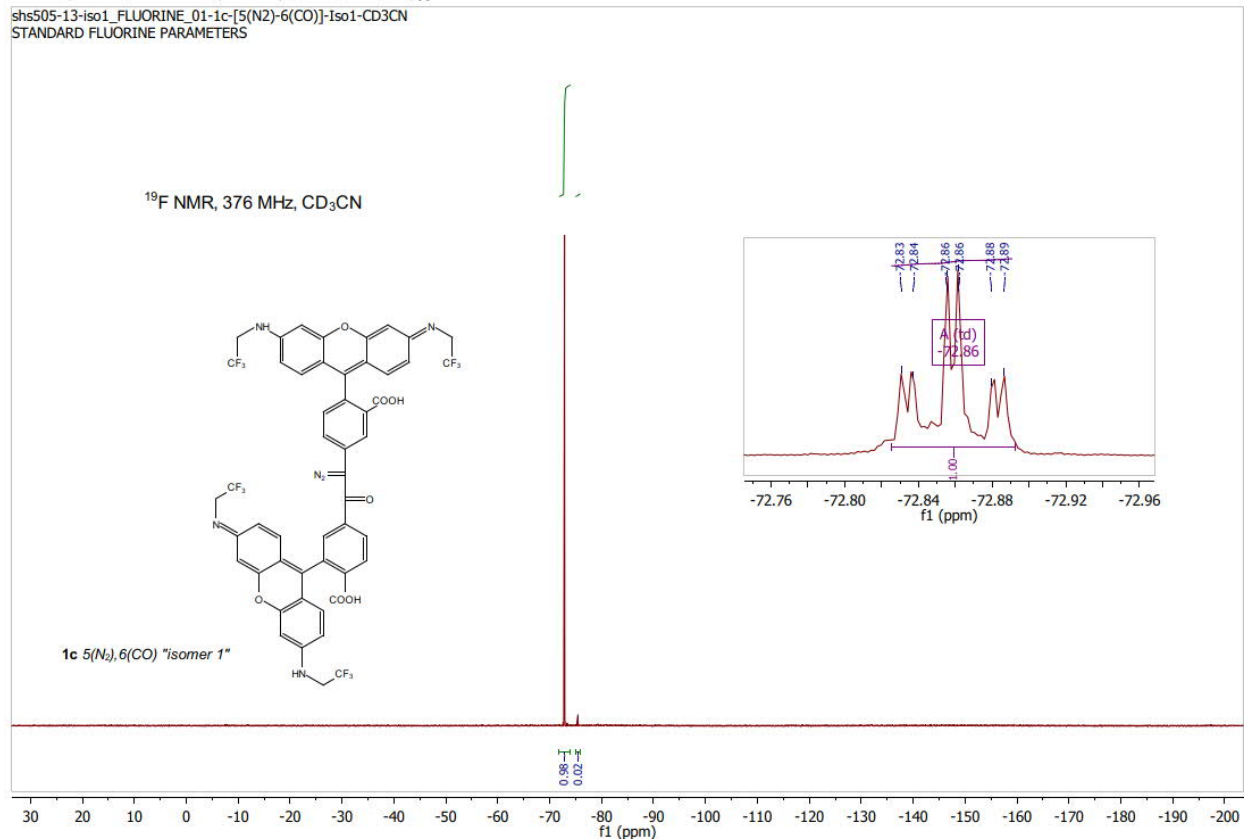


gHSQC, CD<sub>3</sub>CN  
<sup>1</sup>H: 400 MHz, <sup>13</sup>C: 101 MHz



$^{19}\text{F}$  NMR (376 MHz, Acetonitrile- $d_3$ )  $\delta$  -72.86 (td,  $J = 9.4, 2.1$  Hz, 12F) ppm.

shs505-13-iso1\_FLUORINE\_01-1c-[5(N<sub>2</sub>)-6(CO)]-Iso1-CD<sub>3</sub>CN  
STANDARD FLUORINE PARAMETERS

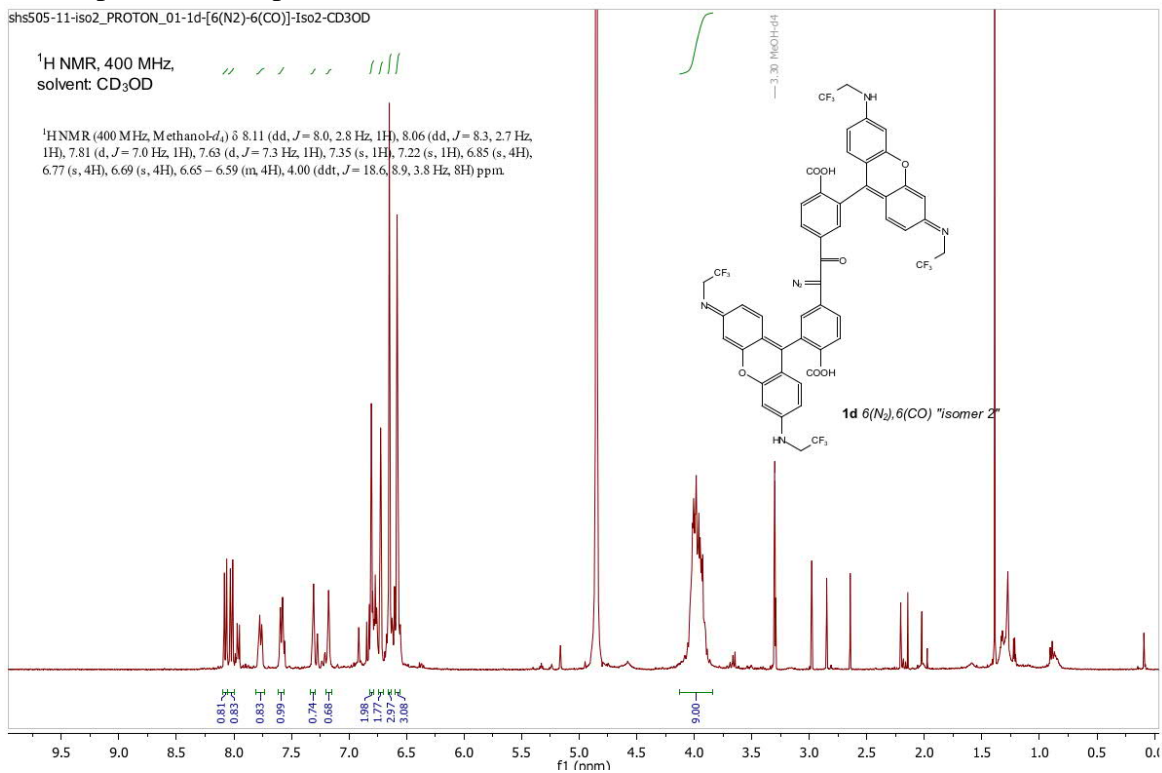


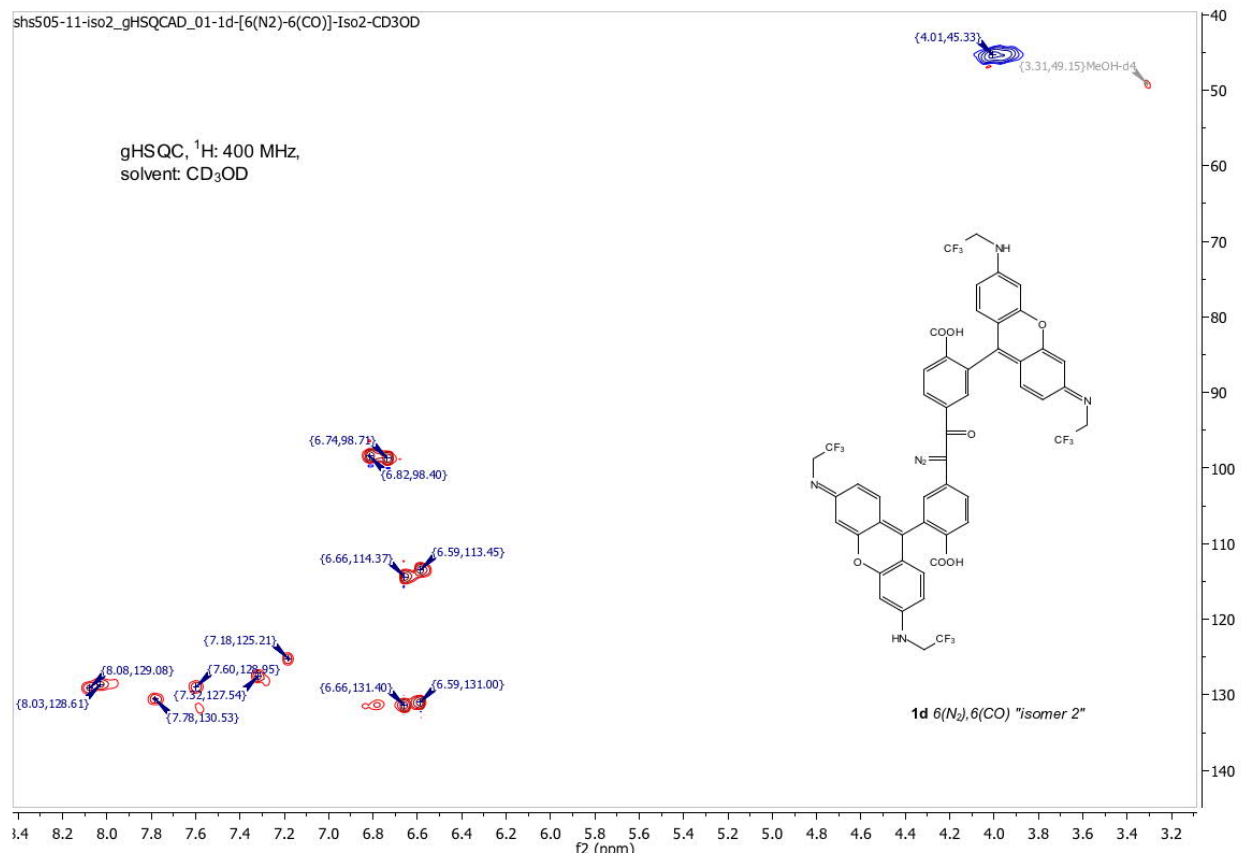
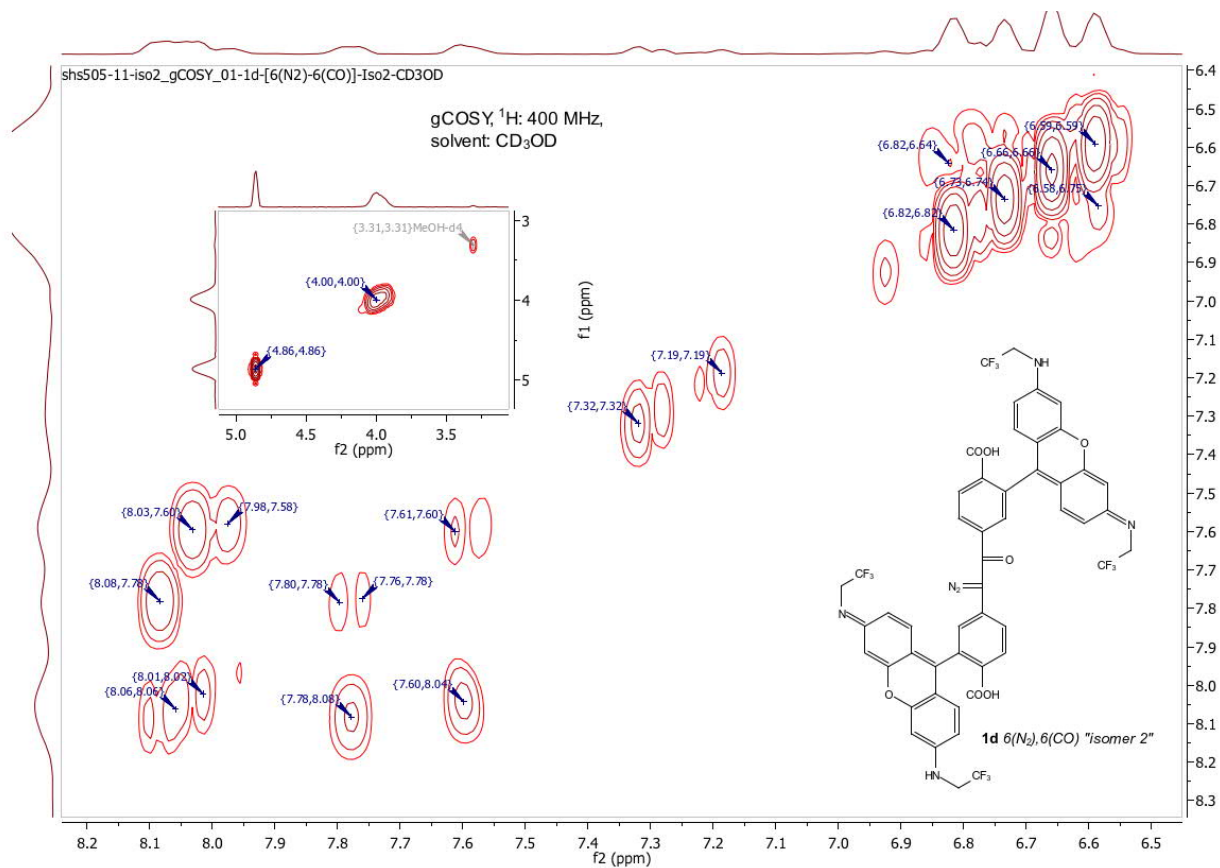
## NMR spectra of compound **1d** [6(N<sub>2</sub>),6(CO)] (isomer 2)

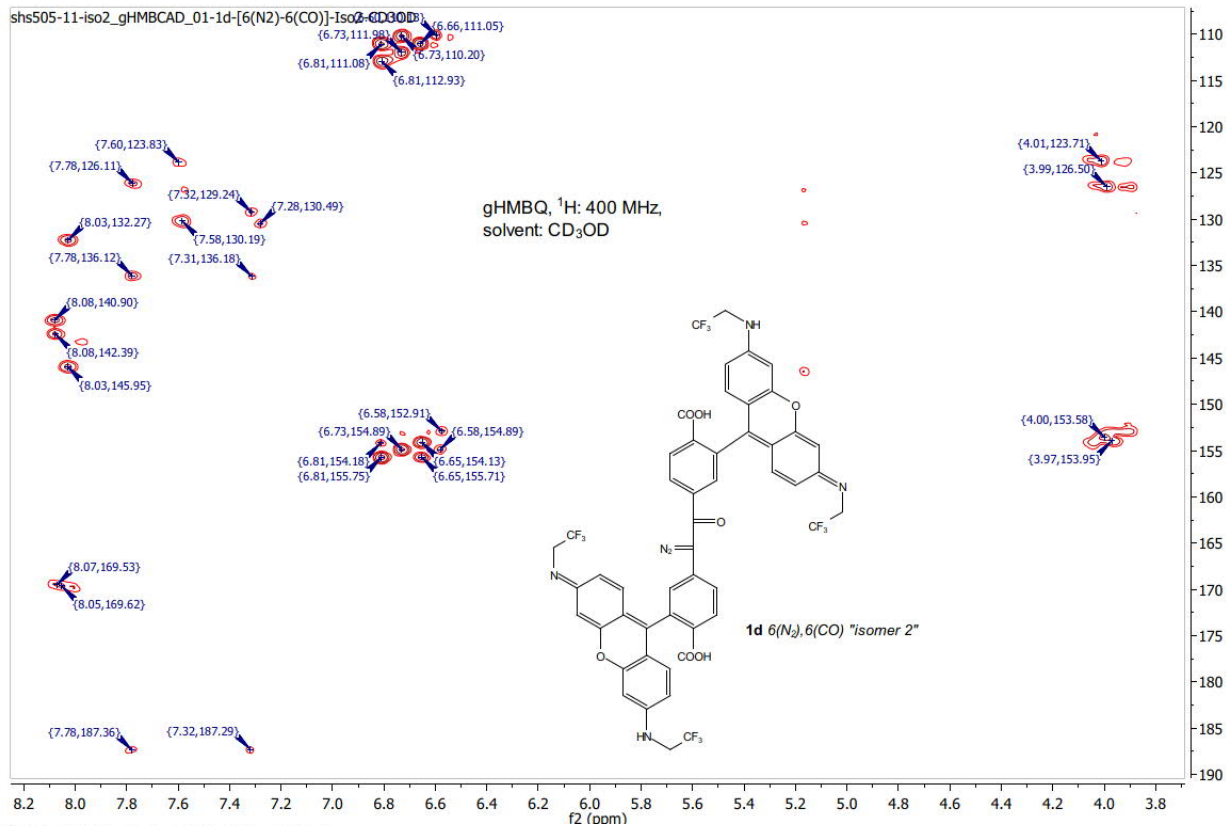
shs505-11-iso2\_PROTON\_01-1d-[6(N<sub>2</sub>)-6(CO)]-Iso2-CD<sub>3</sub>OD

$^1\text{H}$  NMR, 400 MHz,  
solvent: CD<sub>3</sub>OD

$^1\text{H}$  NMR (400 MHz, Methanol- $d_4$ )  $\delta$  8.11 (dd,  $J = 8.0, 2.8$  Hz, 1H), 8.06 (dd,  $J = 8.3, 2.7$  Hz, 1H), 7.81 (d,  $J = 7.0$  Hz, 1H), 7.63 (d,  $J = 7.3$  Hz, 1H), 7.35 (s, 1H), 7.22 (s, 1H), 6.85 (s, 4H), 6.77 (s, 4H), 6.69 (s, 4H), 6.65 – 6.59 (m, 4H), 4.00 (ddt,  $J = 18.6, 8.9, 3.8$  Hz, 8H) ppm.







$^{19}\text{F}$  NMR (376 MHz, Methanol- $d_4$ )  $\delta$  -71.80 – -74.71 (m) ppm

shs505-11-iso2\_FLUORINE\_01-1d-[6(N2)-6(CO)]-Iso2-CD3OD  
STANDARD FLUORINE PARAMETERS

