

## Supplementary information

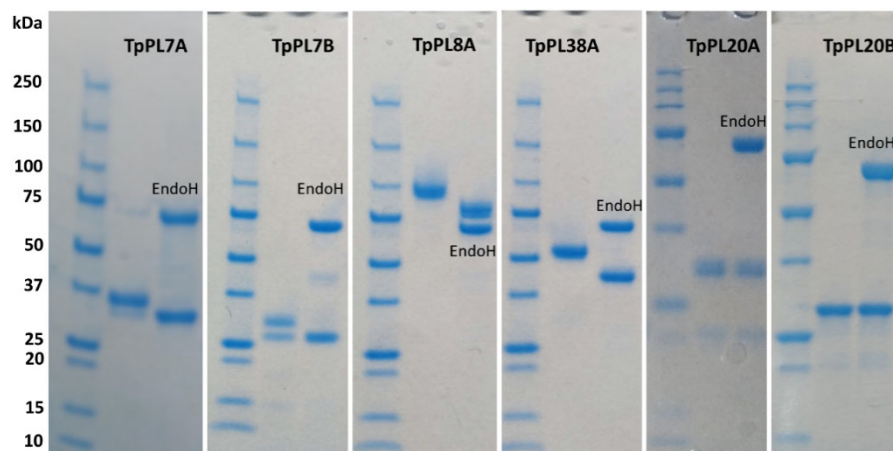
### Discovery of a novel glucuronan lyase system in *Trichoderma parareesei*

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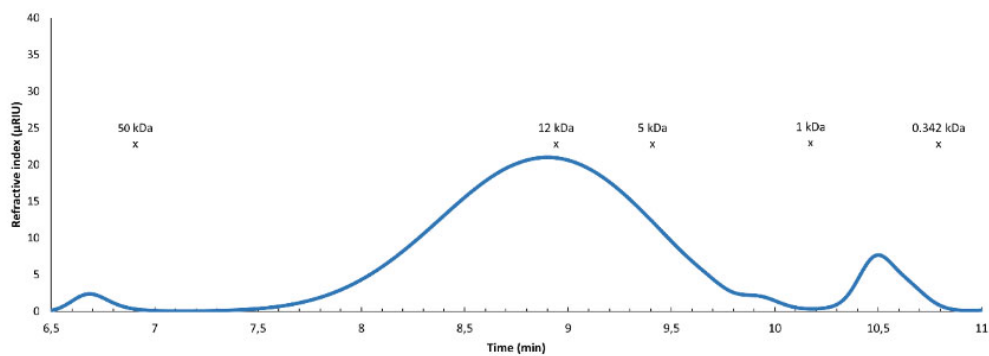
Protein Chemistry and Enzyme Technology Section, DTU Bioengineering, Department of Biotechnology and Biomedicine, Technical University of Denmark, 2800 Kgs. Lyngby, Denmark.

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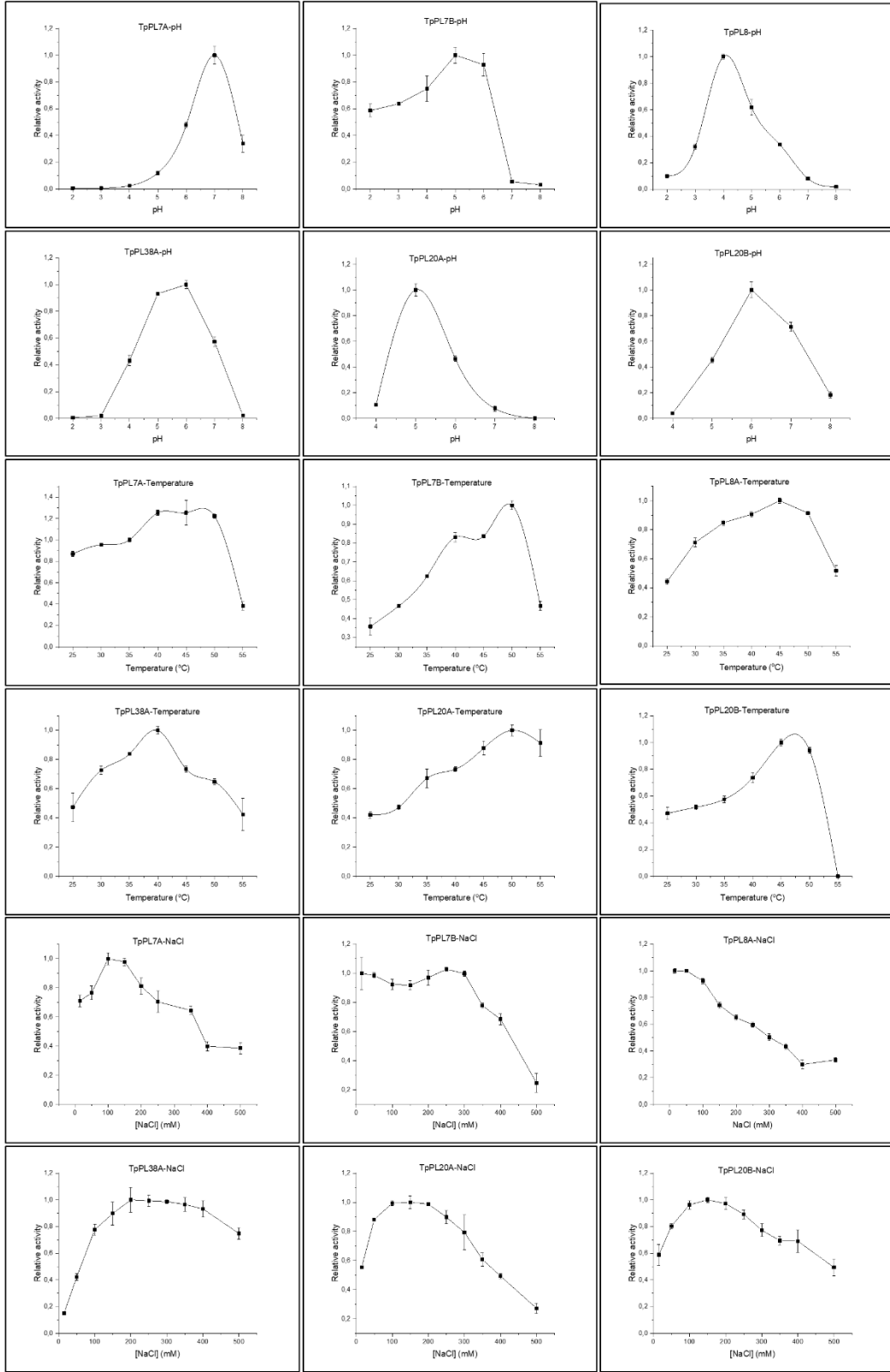
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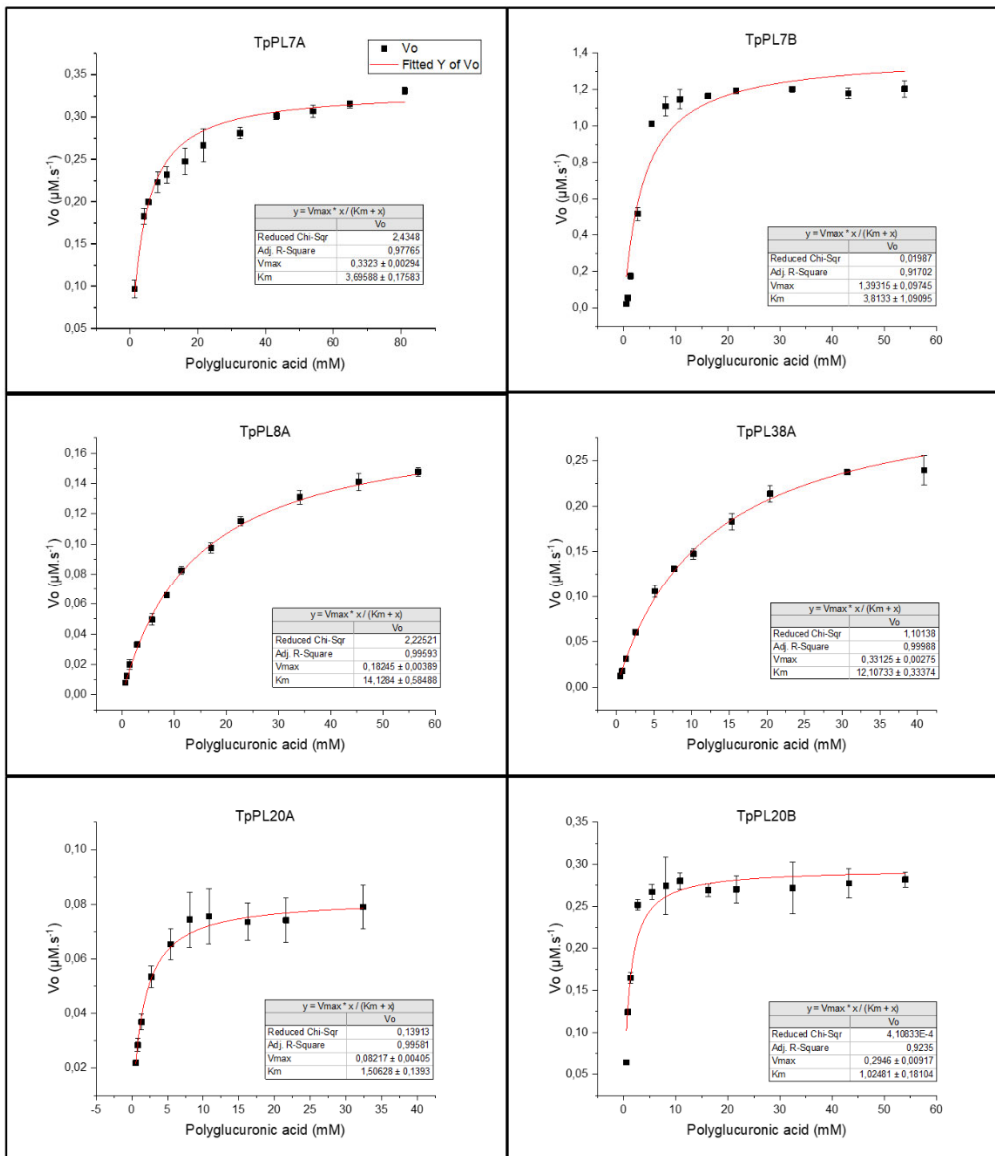
**Supplementary Figure 1.** 4-12% gradient SDS-page gels of the purified recombinant lyases before and after endoH treatment. EndoH is marked by text closest to the band representing it.



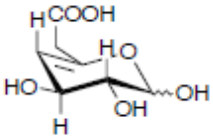
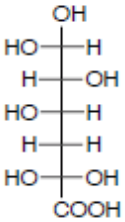
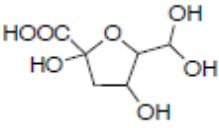
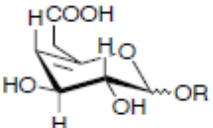
**Supplementary Figure 2.** HP-SEC of the TEMPO prepared polyglucuronic acid at a concentration of 10 g/L. The peaks of the pullulan size standards are marked by x and the size in kDa.



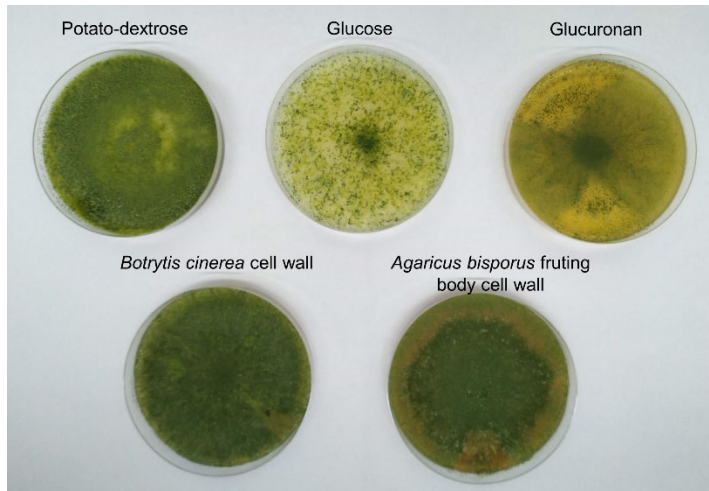
**Supplementary Figure 3.** Biochemical characterisations of the six recombinant glucuronan lyases from *T. pararesei*. Error bars represent standard deviations of triplicate experiments.



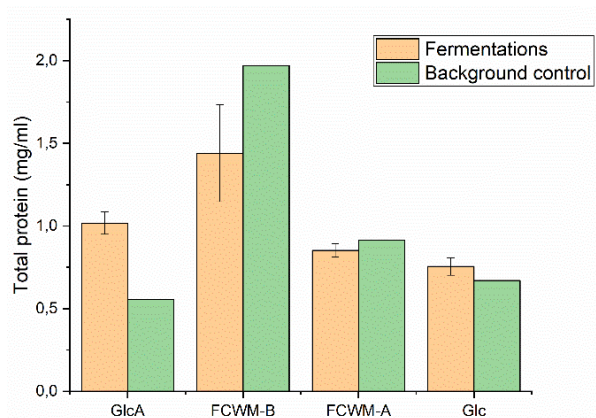
**Supplementary Figure 4.** Initial rates at optimum conditions under increasing substrate concentrations. Red lines are fitted Michaelis-Menten curves. Calculated  $V_{\text{max}}$  and  $K_m$  values are displayed in the grey boxes. Error bars represent standard deviations from triplicate experiments. The graphs and model-fittings were performed in Origin v. 2019b.

	<sup>1</sup> H	<sup>13</sup> C	
β-glucuronan	C-1	4.49	102.3
	C-2	3.35	72.9
	C-3	3.61	74.4
	C-4	3.66	80.9
	C-5	3.85	75.3
	C-6		175.1
		5.21	92.7
		3.69	70.2
		4.20	65.9
		5.71	106.2
			145.5
			169.3
		4.89	93.1
		3.28	76.5
		3.91	68.1
		2.1/1.87	39.2
			95.9
			175.6
		5.17	89.5
		4.00	85.0
		4.42	72.0
		2.38/2.32	44.3
			103.8
			177.1
		5.08	100.1
		3.85	69.4
		4.12	n.d.
		5.86	107.2
			144.9
			169.1

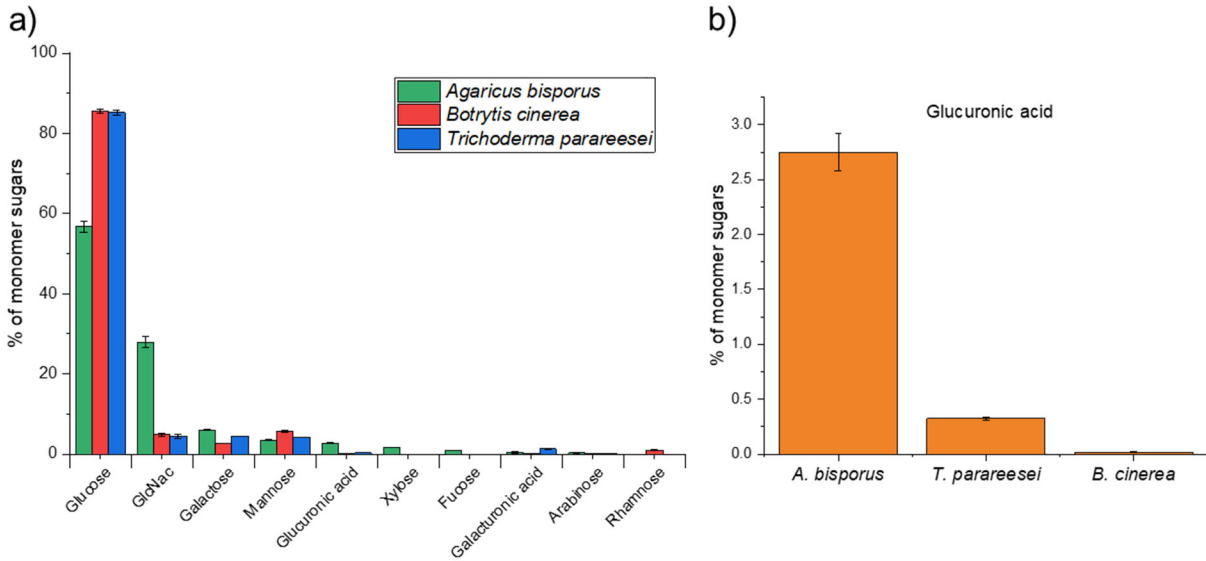
**Supplementary Figure 5.** NMR, <sup>1</sup>H and <sup>13</sup>C chemical shift assignments for the main isomers of the displayed molecules.



**Supplementary Figure 6.** Agar plates showing growth of *T. parareseei* after six days at 30°C on the various carbon sources used in this study.



**Supplementary Figure 7.** Total protein content of fermentation supernatant. Estimated by BCA with BSA as standard. Error bars represent standard deviations of triplicate experiments.

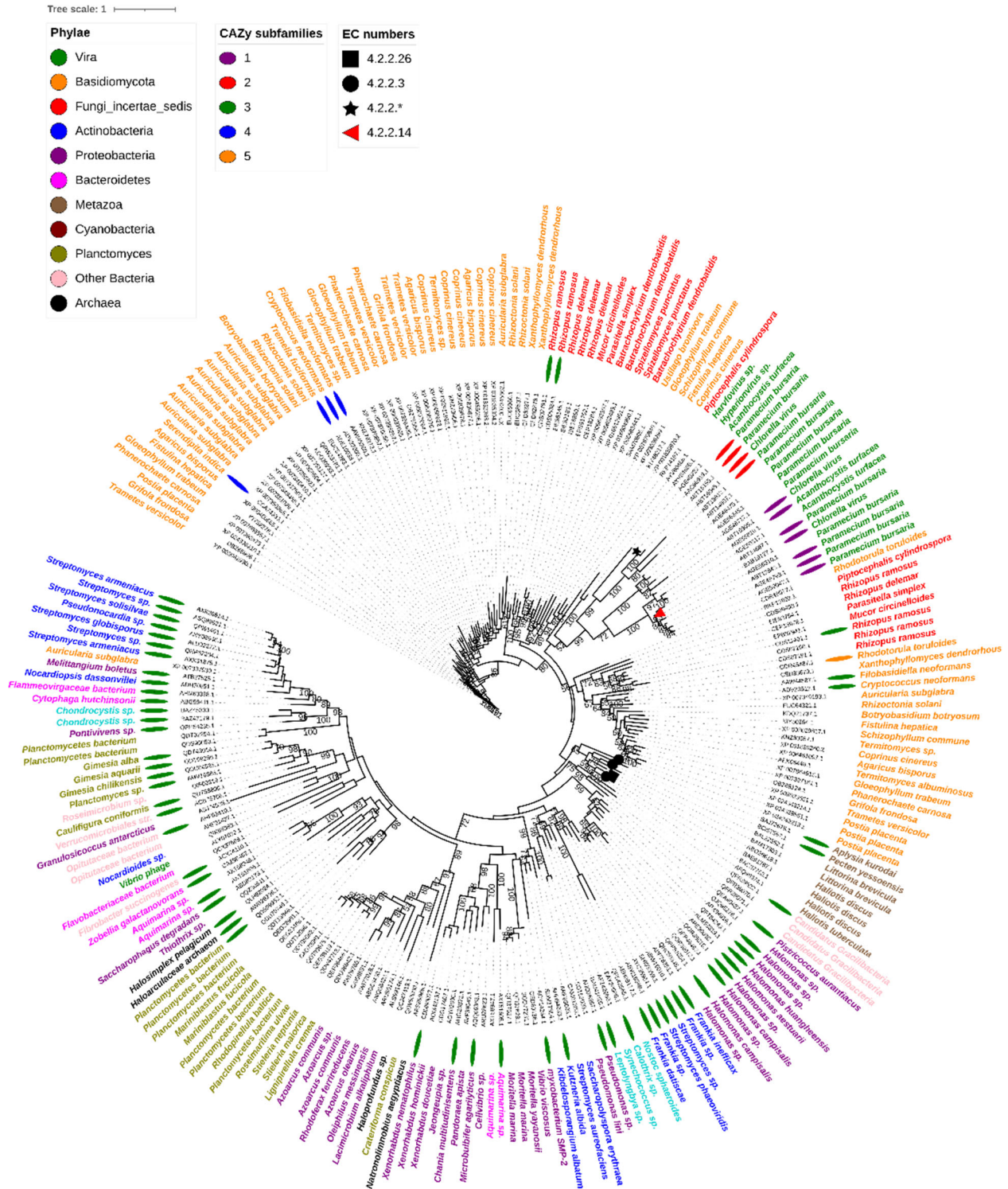


**Supplementary Figure 8.** HPAEC-PAD analysis of sugar monomer composition of fungal cell walls after acid hydrolysis. Values are normalized to total amount of detected sugars. Error bars represent standard deviation of triplicate analysis. **a)** All sugars. **b)** Subset of a) showing only glucuronic acid.

Phylum	Class	Order	Family	Organism	Major ecology	Glucuronan					GAG	Pectin								
						PL7	PL8	PL20	PL38	PL14	PL35	PL1	PL3	PL4	PL9	PL11	PL26	PL27		
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma arundinaceum</i>	Mycotrophic / generalistic	2	1	2	2	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma asperillum</i>	Mycotrophic / generalistic	3	1	2	2	0	0	1	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma atroviride</i>	Mycotrophic / generalistic	3	1	2	2	0	0	2	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma gamsii</i>	Mycotrophic / generalistic	3	1	2	2	0	0	2	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma harzianum</i>	Mycotrophic / generalistic	3	1	2	2	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma longibrachiatum</i>	Mycotrophic / generalistic	2	1	2	1	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma parareesei</i>	Mycotrophic / generalistic	2	1	2	1	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma reesei QMA</i>	Mycotrophic / generalistic	2	1	2	1	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Trichoderma virens</i>	Mycotrophic / generalistic	3	1	2	2	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Cladobotryum protusium</i>	Mycotrophic	4	1	2	2	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Hypomyces periosus</i>	Mycotrophic	2	1	1	1	0	0	0	0	0	0	0	0	0	0	1
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Hypomyces rosellus</i>	Mycotrophic	4	1	2	2	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Hypocreaceae	<i>Escovopsis weberi</i>	Mycotrophic	1	1	0	2	0	0	2	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Clavicipitaceae	<i>Metarhizium acidum</i>	Entomopathogenic	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Clavicipitaceae	<i>Metarhizium album</i>	Entomopathogenic	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Clavicipitaceae	<i>Metarhizium brunneum</i>	Entomopathogenic	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Clavicipitaceae	<i>Metarhizium robertsii</i>	Entomopathogenic	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Clavicipitaceae	<i>Pachonia chlamydosporia</i>	Nematophagous	5	1	1	2	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Cordycipitaceae	<i>Cordyceps fumosorosea</i>	Entomopathogenic	1	1	1	0	0	1	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Cordycipitaceae	<i>Cordyceps militaris</i>	Entomopathogenic	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Ophiocordycipitaceae	<i>Purpureocillium lilacinum</i>	Entomopathogenic	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Ophiocordycipitaceae	<i>Ophiocordyceps sinensis</i>	Entomopathogenic	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Bionectriaceae	<i>Clonostachys rosea</i>	Mycotrophic / generalistic	0	0	1	0	0	0	18	11	4	1	2	1	3	0	0
Ascomycota	Sordariomycetes	Hypocreales	Bionectriaceae	<i>Geosmithia morbida</i>	Phytopathogenic	0	0	1	0	0	0	2	1	1	0	0	2	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Clavicipitaceae	<i>Ustilaginoides vires</i>	Phytopathogenic	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Nectriaceae	<i>Fusarium fujikuroi</i>	Phytopathogenic	0	0	0	0	0	0	11	5	2	2	0	1	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Nectriaceae	<i>Fusarium graminearum</i>	Phytopathogenic	0	0	1	0	0	0	9	7	3	1	0	1	0	0	0
Ascomycota	Sordariomycetes	Hypocreales	Nectriaceae	<i>Fusarium oxysporum</i>	Saprobic	0	0	0	0	0	0	10	7	4	2	1	1	0	0	0
Ascomycota	Sordariomycetes	Diaporthales	Cryphonectriaceae	<i>Cryphonectria parasitica</i>	Phytopathogenic	1	0	0	0	0	0	5	1	2	0	0	0	0	0	
Ascomycota	Sordariomycetes	Diaporthales	Diaporthaceae	<i>Diaporthe citri</i>	Phytopathogenic	0	0	0	0	0	0	18	8	7	1	0	1	2	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum anigma</i>	Phytopathogenic	0	0	1	0	0	0	16	14	7	3	1	1	2	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum fructicola</i>	Phytopathogenic	0	0	1	0	0	0	18	14	7	3	2	2	2	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum gramincola</i>	Phytopathogenic	0	0	0	0	0	0	7	4	3	1	0	1	1	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum higinianum</i>	Phytopathogenic	0	0	0	0	0	0	18	16	7	4	1	1	1	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum kartsii</i>	Phytopathogenic	0	0	0	0	0	0	18	15	8	3	1	1	2	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum orchidophilum</i>	Phytopathogenic	0	0	0	0	0	0	14	9	6	3	0	2	0	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum scovillei</i>	Phytopathogenic	0	0	1	0	0	0	15	13	6	3	1	2	2	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum siamense</i>	Phytopathogenic	0	0	1	0	0	0	17	14	7	3	1	2	2	0	
Ascomycota	Sordariomycetes	Glomerellales	Glomerellaceae	<i>Colletotrichum truncatum</i>	Phytopathogenic	0	0	0	0	0	1	22	15	9	5	1	2	2	0	
Ascomycota	Sordariomycetes	Glomerellales	Plectosphaerellaceae	<i>Verticillium alfalfae</i>	Phytopathogenic	0	0	0	0	0	0	16	11	4	2	1	1	0	0	
Ascomycota	Sordariomycetes	Glomerellales	Plectosphaerellaceae	<i>Verticillium dahliae</i>	Phytopathogenic	0	0	0	0	0	0	16	11	5	2	1	1	0	0	
Ascomycota	Sordariomycetes	Magnaporthales	Magnaporthaceae	<i>Gaeumannomyces tritici</i>	Phytopathogenic	0	0	1	0	0	0	3	1	1	0	0	1	0	0	
Ascomycota	Sordariomycetes	Magnaporthales	Pyriculariaceae	<i>Pyricularia grisea</i>	Phytopathogenic	0	0	1	0	0	0	2	1	1	0	0	1	0	0	
Ascomycota	Sordariomycetes	Magnaporthales	Pyriculariaceae	<i>Pyricularia oryzae</i>	Phytopathogenic	0	0	1	0	0	0	2	1	1	0	0	1	0	0	
Ascomycota	Sordariomycetes	Magnaporthales	Pyriculariaceae	<i>Pyricularia pennisetigena</i>	Phytopathogenic	0	0	1	0	0	0	2	1	1	0	0	1	0	0	
Ascomycota	Sordariomycetes	Microascales	Microasceae	<i>Scedosporium apiospermum</i>	Saprobic	0	0	0	0	0	0	1	0	1	0	0	0	0	0	
Ascomycota	Sordariomycetes	Ophiostomatales	Ophiostomataceae	<i>Grosmanina clavigera</i>	Wood-rot	0	0	0	0	0	0	2	1	1	0	0	1	0	0	
Ascomycota	Sordariomycetes	Ophiostomatales	Ophiostomataceae	<i>Sporothrix brasiliensis</i>	Saprobic	0	0	0	1	0	0	0	0	0	0	0	0	0	0	
Ascomycota	Sordariomycetes	Ophiostomatales	Ophiostomataceae	<i>Sporothrix schenckii</i>	Saprobic	0	0	0	1	0	0	0	0	0	0	0	0	0	0	
Ascomycota	Sordariomycetes	Sordariales	Chaetomiaceae	<i>Chaetomium globosum</i>	Saprobic	0	0	1	0	0	0	7	4	2	1	0	1	0	0	
Ascomycota	Sordariomycetes	Sordariales	Chaetomiaceae	<i>Chaetomium thermophilum</i>	Saprobic	0	0	0	0	0	0	1	3	1	0	0	0	0	0	
Ascomycota	Sordariomycetes	Sordariales	Chaetomiaceae	<i>Podospora anserina</i>	Saprobic	0	0	0	0	0	1	4	2	1	0	0	0	0	0	
Ascomycota	Sordariomycetes	Sordariales	Chaetomiaceae	<i>Thermothelomyces thermophilus</i>	Saprobic	0	0	1	0	0	0	5	1	1	0	0	1	0	0	
Ascomycota	Sordariomycetes	Sordariales	Chaetomiaceae	<i>Thermothelomyces terrestris</i>	Saprobic	1	0	0	0	0	0	0	0	2	0	0	1	0	0	
Ascomycota	Sordariomycetes	Sordariales	Sordariaceae	<i>Neurospora crassa</i>	Saprobic	0	0	1	0	0	0	1	1	1	0	0	0	0	0	
Ascomycota	Sordariomycetes	Sordariales	Sordariaceae	<i>Neurospora tetrasperma</i>	Saprobic	0	0	1	0	0	0	1	1	1	0	0	0	0	0	
Ascomycota	Sordariomycetes	Sordariales	Sordariaceae	<i>Sordaria macrospora</i>	Saprobic	0	0	1	0	0	0	1	2	1	0	0	0	0	0	
Ascomycota	Sordariomycetes	Togniniales	Togniniaceae	<i>Phaeoacremonium minimum</i>	Phytopathogenic	0	0	1	0	0	0	4	3	3	1	0	0	0	0	
Ascomycota	Sordariomycetes	Xylariales	Hypoxylaceae	<i>Daldinia chlidiae</i>	Wood-rot	0	0	2	0	0	0	2	0	5	0	0	1	0	0	
Ascomycota	Sordariomycetes	Xylariales	Pseudomassariaceae	<i>Pseudomassaria vexata</i>	Saprobic	0	0	0	0	0	0	10	8	7	1	0	1	0	0	
Ascomycota	Sordariomycetes	Xylariales	Sporocadaceae	<i>Pestalotiopsis fici</i>	Phytopathogenic	0	0	0	0	0	0	19	7	7	1	1	3	1	0	
Ascomycota	Sordariomycetes	Xylariales	Cephalothecaaceae	<i>Phialemoniopsis curvata</i>	Saprobic	0	0	1	0	0	0	1	1	2	0	0	2	0	0	
Ascomycota	Eurotiomycetes	Eurotiales	Aspergillaceae	<i>Aspergillus oculatus</i>	Saprobic	1	0	0	0	0	0	6	0	3	0	0	1	0	0	
Ascomycota	Eurotiomycetes	Eurotiales	Aspergillaceae	<i>Aspergillus flavus</i>	Saprobic	1	0	2	0	0	0	12	3	3	1	0	2	1	0	
Ascomycota	Eurotiomycetes	Eurotiales	Aspergillaceae	<i>Aspergillus fumigatus</i>	Saprobic	0	0	1	1	0	0	6	3	3	1	0	1	0	0	
Ascomycota	Eurotiomycetes	Eurotiales	Aspergillaceae	<i>Aspergillus nidulans</i>	Saprobic	0	0	2	0	0	0	8	5	4	1	1	1	1	0	
Ascomycota	Eurotiomycetes	Eurotiales	Aspergillaceae	<i>Aspergillus niger</i>	Saprobic	0	0	0	0	0	0	6	0	2	0	0	0	1	0	
Ascomycota	Eurotiomycetes	Eurotiales	Aspergillaceae	<i>Penicillium chrysogenum</i>	Saprobic	0	0	1	0	0	0	5	1	2	0	0	1	0	0	
Ascomycota	Eurotiomycetes	Eurotiales	Aspergillaceae	<i>Penicillium digitatum</i>	Phytopathogenic	0	0	0												



**Supplementary Figure 9.** Genome annotation of PL genes in a broad selection of fungi covering major ecological niches and taxonomic groups. The putative substrate categories, glucuronan, GlycosAmino Glycans (GAG) and pectin are inferred by substrate specificities of experimentally characterized members of the CAZy families and phylogenetic analysis of the protein sequences.



**Supplementary Figure 10.** Maximum likelihood phylogenetic tree of protein sequences from the PL14 family. All sequences were subjected to redundancy, fragmentation and alignment checks. Characterized members are highlighted by symbols at the node ending. Subfamily annotations are highlighted by the colored ellipsis next to the species names. Branch numbers indicate bootstrap values above 50.

**Supplementary Table 1.** Experimental determination of the extinction coefficient (E) for the enzymatic 4-5 double bond formation in glucuronan. Glucuronic acid was used as quantitative standard in the reducing end assay. TpPL7B was used for producing stable double bonds in the substrate under standard assay conditions.

<b>Glucuronan (mg/mL)</b>	<b>A<sub>235</sub></b>	<b>s.d.</b>	<b>Reduced ends (mM)</b>	<b>s.d.</b>	<b>E</b>
0.8	0.221	0.0025	0.0641	0.0419	6621
1.0	0.515	0.0064	0.1650	0.0074	6013
1.5	0.322	0.0006	0.0957	0.0298	6470
<b>Mean</b>					<b>6368 ± 317</b>

**Supplementary Table 2.** Subset of the most abundant CAZymes found in the proteomics analysis. The standard deviations are based on triplicate experiments.

Accession	CAZy family	EC number	Enzyme description	Substrate	Normalized intensities							
					Glucuronan	Std. d	FCWM-B	Std. d	FCWM-A	Std. d	Glucose	Std. d
OTA07244.1	GH15	3.2.1.3	glucan 1,4- $\alpha$ -glucosidase	Starch	0.221	0.011	0.326	0.139	0.058	0.031	0.097	0.020
OTA07404.1	GH13_1	3.2.1.1	$\alpha$ -amylase	Starch	0.000	0.000	0.212	0.067	0.000	0.001	0.012	0.010
OTA02846.1	GH31	-	-	Starch	0.265	0.020	0.102	0.042	0.121	0.085	0.294	0.089
OTA01276.1	GH79	-	-	Proteoglycan	0.342	0.085	1.231	0.158	0.004	0.002	0.004	0.002
OTA07787.1	GH79	3.2.1.31	$\beta$ -glucuronidase	Proteoglycan	0.572	0.088	0.892	0.185	0.062	0.046	0.091	0.097
OTA03228.1	GH89	3.2.1.50	$\alpha$ -N-acetylglucosaminidase	Proteoglycan	0.032	0.001	0.067	0.078	0.034	0.018	0.061	0.020
OTA08831.1	GH20	3.2.1.52	$\beta$ -N-acetylhexosaminidase	Proteoglycan	0.270	0.070	0.014	0.017	0.024	0.009	0.099	0.054
OTA01106.1	GH20	3.2.1.52	$\beta$ -N-acetylhexosaminidase	Proteoglycan	0.053	0.004	0.000	0.000	0.081	0.028	0.071	0.016
OTA02666.1	AA2	-	-	Oxidase	0.064	0.022	0.220	0.044	0.101	0.016	0.144	0.062
OTA07476.1	AA7	-	-	Oxidase	1.083	0.183	0.209	0.069	4.863	1.046	2.833	0.093
OTA07907.1	AA5	-	-	Oxidase	0.380	0.041	0.073	0.036	0.923	0.174	0.382	0.068
OTA02290.1	AA14	-	-	Oxidase	0.039	0.006	0.059	0.016	0.131	0.026	0.049	0.012
OTA02426.1	AA7	-	-	Oxidase	0.267	0.014	0.049	0.017	0.231	0.098	0.080	0.066
OTA03014.1	AA2	-	-	Oxidase	0.175	0.011	0.025	0.026	0.260	0.059	0.107	0.056
OTA03614.1	AA2	1.11.1.5	cytochrome-c peroxidase	Oxidase	0.098	0.004	0.006	0.002	0.038	0.006	0.070	0.017
OTA03261.1	GH38	3.2.1.24	$\alpha$ -mannosidase	$\alpha$ -1,6-mannan	0.078	0.018	0.000	0.000	0.077	0.016	0.119	0.044
OTA05230.1	GH92	3.2.1.24	$\alpha$ -mannosidase	$\alpha$ -1,6-mannan	0.023	0.002	0.000	0.000	0.052	0.012	0.057	0.016
OTA02561.1	GH71	3.2.1.59	glucan endo-1,3- $\alpha$ -glucosidase	$\alpha$ -1,3-glucan	0.004	0.001	0.006	0.003	0.007	0.005	0.015	0.007
OTA00561.1	GH71	3.2.1.59	glucan endo-1,3- $\alpha$ -glucosidase	$\alpha$ -1,3-glucan	0.040	0.012	0.000	0.000	0.016	0.006	0.036	0.012
OSZ9986.1	CBM24	3.2.1.59	glucan endo-1,3- $\alpha$ -glucosidase	$\alpha$ -1,3-glucan	0.038	0.006	0.000	0.000	0.005	0.004	0.129	0.163
OTA05307.1	GH54	-	-	Xylan	0.163	0.019	0.467	0.109	0.272	0.214	0.399	0.125
OTA05875.1	GH115	3.2.1.131	xylan $\alpha$ -1,2-glucuronosidase	Xylan	0.066	0.013	0.000	0.000	0.023	0.002	0.067	0.031
OTA01922.1	GH18	3.2.1.96	syl-glycoprotein endo- $\beta$ -N-acetylglucosam	Glycoprotein	0.165	0.016	0.251	0.126	0.062	0.042	0.188	0.068
OTA05399.1	GH47	3.2.1.113	iannosyl-oligosaccharide 1,2- $\alpha$ -mannosidas	Glycoprotein	0.225	0.019	0.220	0.162	0.267	0.142	0.427	0.237
OTA07247.1	PL20_B	4.2.2.14	glucuronan lyase	Glucuronan	0.412	0.016	0.863	0.453	0.003	0.002	0.007	0.003
OTA00986.1	PL7_4_A	-	-	Glucuronan	0.822	0.102	0.236	0.209	0.019	0.003	0.124	0.182
OTA01262.1	PL7_4_B	-	-	Glucuronan	0.049	0.023	0.083	0.099	0.001	0.000	0.001	0.000
OTA06285.1	PL38	4.2.2.14	glucuronan lyase	Glucuronan	0.348	0.151	0.067	0.048	0.010	0.007	0.009	0.003
OTA05836.1	PL8_4	-	-	Glucuronan	0.959	0.181	0.049	0.026	0.018	0.000	0.024	0.014
OSZ99978.1	-	3.2.1.22	$\alpha$ -galactosidase	Galactan	0.141	0.038	0.000	0.000	0.014	0.003	0.024	0.012
OTA00159.1	GH2	3.2.1.146	$\beta$ -galactofuranosidase	idophosphogalactoma	0.252	0.074	0.000	0.000	0.034	0.025	0.043	0.024
OTA07992.1	CBM66	3.2.1.146	$\beta$ -galactofuranosidase	idophosphogalactoma	0.077	0.002	0.000	0.000	0.174	0.048	0.200	0.095
OTA00212.1	GH75	3.2.1.132	chitinase	Chitosan	0.423	0.087	1.053	0.229	0.015	0.004	0.034	0.006
OTA04600.1	GH18	3.2.1.14	chitinase	Chitin	0.325	0.027	0.146	0.045	0.425	0.095	0.654	0.402
OTA00069.1	GH2	3.2.1.165	exo-1,4- $\beta$ -D-glucosaminidase	Chitosan	0.631	0.114	0.043	0.027	0.142	0.094	0.045	0.021
OTA08247.1	GH18	3.2.1.14	chitinase	Chitin	0.220	0.007	0.000	0.000	0.247	0.040	0.392	0.104
OTA06834.1	GH18	3.2.1.14	chitinase	Chitin	0.107	0.015	0.000	0.000	0.194	0.058	0.229	0.128
OTA01114.1	CE4	-	-	Chitin	0.029	0.005	0.000	0.000	0.102	0.032	0.033	0.020
OTA00434.1	GH18	3.2.1.14	chitinase	Chitin	0.017	0.005	0.000	0.000	0.009	0.002	0.020	0.006
OTA04876.1	GH5_5	3.2.1.4	cellulase	$\beta$ -1,4-glucan	0.006	0.008	0.037	0.022	0.037	0.035	0.153	0.119
OTA02682.1	GH3	-	-	$\beta$ -1,4-glucan	0.003	0.000	0.005	0.006	0.002	0.001	0.000	0.000
OSZ99894.1	GH3	-	-	$\beta$ -1,4-glucan	0.016	0.002	0.000	0.000	0.015	0.005	0.017	0.004
OTA01208.1	GH30_3	3.2.1.75	glucan endo-1,6- $\beta$ -glucosidase	$\beta$ -1,6-glucan	0.147	0.007	1.567	0.488	0.045	0.004	0.064	0.010
OTA00769.1	GH55	3.2.1.58	glucan 1,3- $\beta$ -glucosidase	$\beta$ -1,3-glucan	0.875	0.203	0.335	0.165	0.228	0.097	0.488	0.225
OTA05146.1	GH55	3.2.1.39	glucan endo-1,3- $\beta$ -D-glucosidase	$\beta$ -1,3-glucan	0.443	0.042	0.238	0.124	0.051	0.014	0.016	0.009
OTA02562.1	GH64	3.2.1.39	glucan endo-1,3- $\beta$ -D-glucosidase	$\beta$ -1,3-glucan	0.661	0.060	0.229	0.067	0.828	0.566	0.869	0.328
OTA08417.1	GH128	-	-	$\beta$ -1,3-glucan	0.000	0.000	0.147	0.070	0.000	0.000	0.000	0.000
OTA08148.1	GH55	-	-	$\beta$ -1,3-glucan	2.400	0.284	0.108	0.078	0.032	0.020	0.014	0.011
OTA01266.1	GH55	3.2.1.39	glucan endo-1,3- $\beta$ -D-glucosidase	$\beta$ -1,3-glucan	0.023	0.001	0.065	0.013	0.014	0.007	0.011	0.008
OTA01439.1	GH128	-	-	$\beta$ -1,3-glucan	0.005	0.001	0.038	0.008	0.008	0.002	0.011	0.006
OTA04977.1	GH55	3.2.1.39	glucan endo-1,3- $\beta$ -D-glucosidase	$\beta$ -1,3-glucan	0.024	0.004	0.020	0.022	0.016	0.003	0.016	0.001
OTA04295.1	GH81	3.2.1.39	glucan endo-1,3- $\beta$ -D-glucosidase	$\beta$ -1,3-glucan	0.000	0.000	0.019	0.019	0.001	0.000	0.000	0.000
OTA01419.1	GH132	3.2.1.x	-	$\beta$ -1,3-glucan	0.354	0.045	0.000	0.000	0.337	0.022	0.388	0.091
OTA06148.1	GH16_3	3.2.1.6	endo-1,3(4)- $\beta$ -glucanase	$\beta$ -1,3-glucan	0.238	0.031	0.000	0.000	0.536	0.065	0.265	0.071
OTA06837.1	GH128	-	-	$\beta$ -1,3-glucan	0.121	0.071	0.000	0.000	0.121	0.024	0.130	0.093
OTA05871.1	GH81	3.2.1.39	glucan endo-1,3- $\beta$ -D-glucosidase	$\beta$ -1,3-glucan	0.011	0.002	0.000	0.000	0.006	0.002	0.010	0.003
OTA01208.1	GH30_3	3.2.1.75	glucan endo-1,6- $\beta$ -glucosidase	$\beta$ -1,6-glucan	0.147	0.007	1.567	0.488	0.045	0.004	0.064	0.010
OTA07339.1	GH5_15	3.2.1.75	glucan endo-1,6- $\beta$ -glucosidase	$\beta$ -1,6-glucan	0.004	0.000	0.000	0.000	0.015	0.005	0.017	0.004

**Supplementary Table 3.** Genomes downloaded from NCBI and used in this study.

<b>Strain name</b>	<b>NCBI assembly accession nb.</b>	<b>Strain name</b>	<b>NCBI assembly accession nb.</b>
<i>Achlya hypogyna</i>	GCA_002081595.1	<i>Pyricularia oryzae</i>	GCF_000002495.2
<i>Agaricus bisporus</i>	GCA_000300575.1_Agabi_varbisH97_2	<i>Grosmannia clavigera</i>	GCF_000143105.1
<i>Aspergillus aculeatus</i>	GCF_001890905.1_Aspac1	<i>Chaetomium globosum</i>	GCF_000143365.1
<i>Aspergillus flavus</i>	GCA_000006275.3	<i>Gaeumannomyces tritici</i>	GCF_000145635.1
<i>Aspergillus fumigatus</i>	GCF_000002655.1_ASM265v1	<i>Colletotrichum graminicola</i>	GCF_000149035.1
<i>Aspergillus nidulans</i>	GCF_000149205.2_ASM14920v2	<i>Verticillium dahliae</i>	GCF_000150675.1
<i>Aspergillus niger</i>	GCF_000002855.3_ASM285v2	<i>Verticillium alfalfae</i>	GCF_000150825.1
<i>Auricularia subglabra</i>	GCF_000265015.1_Auricularia_subglabra_SS-5_V1.0	<i>Sordaria macrospora</i>	GCF_000182805.2
<i>Batrachochytrium dendrobatidis</i>	GCF_000203795.1_v1.0	<i>Neurospora crassa</i>	GCF_000182925.2
<i>Botryobasidium botryosum</i>	GCA_000697705.1_Botbo1	<i>Metarhizium acridum</i>	GCF_000187405.1
<i>Botrytis cinerea</i>	GCF_000143535.2_ASM14353v4	<i>Metarhizium robertsii</i>	GCF_000187425.2
<i>Catenaria anguillulae</i>	GCA_002102555.1_Catan2	<i>Neurospora tetrasperma</i>	GCF_000213175.1
<i>Cladobotryum protrusum</i>	GCA_004303015.1_ASM430301v1	<i>Chaetomium thermophilum</i>	GCF_000221225.1
<i>Clonostachys rosea</i>	GCA_000963775.2_ASM96377v2	<i>Cordyceps militaris</i>	GCF_000225605.1
<i>Coprinopsis cinerea</i>	GCF_000182895.1_CC3	<i>Thermothelomyces thermophilus</i>	GCF_000226095.1
<i>Cordyceps fumosorosea</i>	GCF_001636725.1_SF_1.0	<i>Thermothielavioides terrestris</i>	GCF_000226115.1
<i>Cordyceps militaris</i>	GCF_000225605.1	<i>Podospira anserina</i>	GCF_000226545.1
<i>Cryptococcus gattii</i>	GCF_000185945.1_ASM18594v1	<i>Beauveria bassiana</i>	GCF_000228075.1
<i>Escovopsis weberi</i>	GCA_001278495.1_ASM127849v1	<i>Phaeoacremonium minimum</i>	GCF_000392275.1
<i>Fistulina hepatica</i>	GCA_000934395.1_Fishe1	<i>Pestalotiopsis fici</i>	GCF_000516985.1
<i>Fusarium fujikuroi</i>	GCF_900079805.1_Fusarium_fujikuroi_IM58289_V2	<i>Ustilagoidea vires</i>	GCF_000687475.1
<i>Fusarium graminearum</i>	GCF_000240135.3_ASM24013v3	<i>Scedosporium apiospermum</i>	GCF_000732125.1
<i>Fusarium oxysporum</i>	GCF_000149955.1_ASM14995v2	<i>Metarhizium album</i>	GCF_000804445.1
<i>Gloeophyllum trabeum</i>	GCF_000344685.1_Glotr1_1	<i>Metarhizium brunneum</i>	GCF_000814965.1
<i>Gonapodya prolifera</i>	GCA_001574975.1_Ganpr1	<i>Sporothrix brasiliensis</i>	GCF_000820605.1
<i>Grifola frondosa</i>	GCA_001683735.1_ASM168373v1	<i>Sporothrix schenckii</i>	GCF_000961545.1
<i>Hypomyces perniciosus</i>	GCA_0008477525.1_ASM847752v1	<i>Drechmeria coniospora</i>	GCF_001625195.1
<i>Hypomyces rosellus</i>	GCA_011799845.1_ASM1179984v1	<i>Cordyceps fumosorosea</i>	GCF_001636725.1
<i>Lentinus edodes</i>	GCA_002003045.1	<i>Pochonia chlamydosporia</i>	GCF_001653235.2
<i>Lichtheimia ramosa</i>	GCA_000945115.1_Lramosa_hybrid_454_Illumina	<i>Purpureocillium lilacinum</i>	GCF_001653265.1
<i>Lichtheimia ramosa</i>	GCA_000945115.1	<i>Colletotrichum higginsianum</i>	GCF_001672515.1
<i>Moesziomyces antarcticus</i>	GCF_000747765.1_ASM74776v1	<i>Colletotrichum orchidophilum</i>	GCF_001831195.1
<i>Mucor circinelloides</i>	GCA_000401635.1_Muco_sp_1006Ph_V1	<i>Pseudomassariella vexata</i>	GCF_002105095.1
<i>Ophiocordyceps sinensis</i>	GCA_012934285.1_ASM1293428v1	<i>Sodiomyces alkalinus</i>	GCF_003711515.1
<i>Paradendryphiella salina</i>	GCA_900634815.1	<i>Verticillium nonalfalfae</i>	GCF_003724135.2
<i>Parasitella parasitica</i>	GCA_000938895.1_PPv1	<i>Pyricularia pennisetigena</i>	GCF_004337985.1
<i>Penicillium chrysogenum</i>	GCA_000710275.1_ASM71027v1	<i>Phialemoniopsis curvata</i>	GCF_004353045.1
<i>Penicillium digitatum</i>	GCF_000315645.1_PdigPd1_v1	<i>Pyricularia grisea</i>	GCF_004355905.1
<i>Penicillium expansum</i>	GCF_000769745.1_ASM76974v1	<i>Daldinia childiae</i>	GCF_008694065.1
<i>Penicillium roqueforti</i>	GCA_000513255.1_FROQFM164_20130607	<i>Colletotrichum fructicola</i>	GCF_009771025.1
<i>Phanerochaete carnososa</i>	GCF_000300595.1_Phanerochaete_carnososa_HH1B-10118-Sp_v1.0	<i>Colletotrichum scovillei</i>	GCF_011075155.1
<i>Piptocephalis cylindrospora</i>	GCA_000361445.1_Ppicy3_1	<i>Cryphonectria parasitica</i>	GCF_011745365.1
<i>Pochonia chlamydosporia</i>	GCF_001653235.2_ASM165323v2	<i>Colletotrichum karsti</i>	GCF_011947395.1
<i>Podospira anserina</i>	GCF_000226545.1	<i>Geosmithia morbida</i>	GCF_012550715.1
<i>Postia placenta</i>	GCF_002117355.1_PospIRSB12_1	<i>Colletotrichum aenigma</i>	GCF_013390185.1
<i>Pseudozyma flocculosa</i>	GCF_000417875.1_Pfloc1_1.0	<i>Colletotrichum siamense</i>	GCF_013390195.1
<i>Puccinia striiformis</i>	GCA_001191645.1	<i>Colletotrichum truncatum</i>	GCF_014235925.1
<i>Puccinia triticina</i>	GCA_000151525.2_P_triticina_1_1_V2	<i>Diaporthe citri</i>	GCF_014595645.1
<i>Rhizoctonia solani</i>	GCA_000524645.1_Rhizoctonia_solani_AG-3	<i>Pseudogymnoascus verrucosus</i>	GCA_001662655.1
<i>Rhizophlyctis rosea</i>	GCA_002214945.1		
<i>Rhizopus delemar</i>	GCA_000149305.1_RO3		
<i>Schizophyllum commune</i>	GCF_000143185.1_v1.0		
<i>Sclerotinia sclerotiorum</i>	GCF_000146945.2_ASM14694v2		
<i>Spizellomyces punctatus</i>	GCF_000182565.1_S_punctatus_V1		
<i>Syncephalis pseudoplumigaleata</i>	GCA_003614755.1_Synps1		
<i>Talaromyces marneffei</i>	GCF_000001985.1		
<i>Talaromyces verruculosus</i>	GCA_001305275.1		
<i>Termitomyces</i>	GCA_001263195.1_TerJ1321.0		
<i>Trametes versicolor</i>	GCF_000271585.1_Trametes_versicolor_v1.0		
<i>Trichoderma asperellum</i>	GCF_003025105.1_Trias_v_1.0		
<i>Trichoderma atroviride</i>	GCF_000171015.1_TRIAT_v2.0		
<i>Trichoderma gamsii</i>	GCF_001481775.2_TGAM01v2		
<i>Trichoderma harzianum</i>	GCF_003025095.1_Triha_v1.0		
<i>Trichoderma longibrachiatum</i>	GCA_003025155.1_TriLo3		
<i>Trichoderma parareesei</i>	GCA_001050175.1_Tpara_v1.0		
<i>Trichoderma reesei QMA</i>	GCF_000167675.1_v2.0		
<i>Trichoderma virens</i>	GCF_000170995.1_TRV1_v2.0		
<i>Ustilago maydis</i>	GCF_000328475.2_Umaydis521_2.0		
<i>Verticillium alfalfae</i>	GCF_000150825.1_ASM15082v1		
<i>Verticillium dahliae</i>	GCF_000150675.1_ASM15067v2		