## Environ Health Perspect

## DOI: 10.1289/EHP9373

**Note to readers with disabilities:** *EHP* strives to ensure that all journal content is accessible to all readers. However, some figures and Supplemental Material published in *EHP* articles may not conform to <u>508 standards</u> due to the complexity of the information being presented. If you need assistance accessing journal content, please contact <u>ehp508@niehs.nih.gov</u>. Our staff will work with you to assess and meet your accessibility needs within 3 working days.

#### **Supplemental Material**

# Role of Hepatocyte- and Macrophage-Specific PPAR $\gamma$ in Hepatotoxicity Induced by Diethylhexyl Phthalate in Mice

Miao Xu, Yongning Li, Xiaohong Wang, Qiannan Zhang, Lei Wang, Xin Zhang, Wenming Cui, Xiaomin Han, Ning Ma, Haishan Li, Hongyun Fang, Song Tang, Jingguang Li, Zhaoping Liu, Hui Yang, and Xudong Jia

## **Table of Contents**

Table S1. Key reagents and materials.

**Table S2.** Liver weight and liver index in DEHP treated mice [mean  $\pm$  SD; n=10/group].

**Table S3.** Concentration of MEHP in plasma and liver of DEHP treated mice [mean  $\pm$  SD (n)].

**Table S4.** Clinical chemistry panel of DEHP treated mice [mean  $\pm$  SD; n=5/group].

**Table S5.** Concentration of hepatic TG in DEHP treated mice [mean  $\pm$  SD; n=5/group].

**Table S6.** Hepatic pathological severity and oil red staining evaluation in DEHP treated mice [evaluating scores; mean  $\pm$  SD; n=6/group].

**Table S7.** Fatty acid uptake and lipid accumulation evaluation in MEHP treated HepG2 cells [Mean stain area ( $\mu$ m<sup>2</sup>) (% of control); mean ± SD; n=4/treatment].

**Table S8.** Data for Figure 2A: Binding affinity analysis of PPARα [Resonance units (RU); n=2006-2146; provided as Excel Table S8].

**Table S9.** Data for Figure 2A: Binding affinity analysis of PPAR $\gamma$  [Resonance units (RU); n=1583-2131; provided as <u>Excel Table S9</u>].

**Table S10.** PPRE and CD36 response in PPRE-THP-1 cells derived macrophages [positive cells (%); mean±SD; n=6 sites/treatment].

**Table S11.** Dose response curves of PPRE in PPRE-THP-1 cells derived macrophages [mean stain area ( $\mu$ m<sup>2</sup>); mean; n=3/treatment].

**Table S12.** Dose response curve of CD36 in PPRE-THP-1 cells derived macrophages [mean stain area ( $\mu$ m<sup>2</sup>); mean, n=3/treatment)].

**Table S13.** Dose response curve of PPRE and CD36 in MEHP treated macrophages with/without PPAR $\alpha/\gamma$  antagonists [mean stain area ( $\mu$ m<sup>2</sup>); mean, n=3/treatment].

**Table S14.** Liver weight and liver index in WT and Hep-KO mice [mean  $\pm$  SD (n)].

**Table S15.** Clinical chemistry panel in WT and Hep-KO mice [mean  $\pm$  SD (n)].

**Table S16.** Concentration of hepatic TG in WT and Hep-KO mice [ $\mu$ g/mg tissue; mean  $\pm$  SD (n)].

**Table S17.** Hepatic pathological severity evaluation in WT and Hep-KO mice [evaluating scores; mean  $\pm$  SD (n)].

**Table S18.** Macrophages evaluation in liver sections from WT and Hep-KO mice [cell number per site; mean  $\pm$  SD (n)].

**Table S19.** Liver weight and liver index in WT and Mac-KO mice [mean  $\pm$  SD; n=5/group].

**Table S20.** Concentration of hepatic TG in WT and Mac-KO mice [ $\mu$ g/mg tissue; mean  $\pm$  SD; n=5/group].

**Table S21.** Hepatic pathological severity and oil red staining evaluation in WT and Mac-KO mice [evaluating scores; mean  $\pm$  SD; n=5/group].

**Table S22.** M2 macrophages analysis in liver of WT and Mac-KO mice using flow cytometry [% of CD45<sup>+</sup> cells; mean  $\pm$  SD; n=5/group].

**Table S23.** Macrophages evaluation in liver sections from WT and Mac-KO mice [cell number per site; mean  $\pm$  SD; n=5/group].

**Table S24.** Monocyte evaluation in liver from WT and Mac-KO mice [% of CD45<sup>+</sup>cells; mean  $\pm$  SD; n=5/group].

**Table S25.** M2 macrophages polarization evaluation in BMDM from WT and Mac-KO mice  $[CD206^+ \text{ cells } (\%); \text{ mean } \pm \text{ SD}; n=15 \text{ technical replicates/treatment}].$ 

**Table S26.** Relative expression of DEGs associated with M2 macrophages in MEHP treated macrophages [Z scores; n=3/treatment; provided as <u>Excel Table S26</u>].

**Table S27.** Relative expression of DEGs associated with PPAR pathway in MEHP treated macrophages [Z scores; n=3/treatment; provided as <u>Excel Table S27</u>].

**Table S28.** Characteristics of PPRE-THP-1 cells derived M2 macrophages [Mean stain area ( $\mu$ m2); mean  $\pm$  SD; n=2/subtype].

**Table S29.** Dose response curves of PPRE/CD209/LD in DEHP/MEHP treated M2 macrophages [% of control; mean, n=3/treatment].

**Table S30.** Data for Figure 5G: Dose response curves of CD209 in PPAR $\alpha/\gamma$  agonists treated M2 macrophages [Mean stain area ( $\mu$ m<sup>2</sup>); mean; n=3/treatment].

**Table S31.** Data for Figure 5H: Dose response curves of CD209 in PPAR $\alpha/\gamma$  antagonists treated M2 macrophages [Mean stain area ( $\mu$ m<sup>2</sup>); mean, n=3/treatment].

**Table S32.** Response of CD209/CD36/LD in MEHP treated M2 macrophages with/without PPAR $\alpha/\gamma$  antagonists [Mean stain area ( $\mu$ m<sup>2</sup>); mean ± SD (n)].

**Table S33.** Data for Figure 6A: Transcriptomic profiling of livers from different mice models [DEGs; provided as <u>Excel Table S33</u>].

**Table S34.** Data for Figure 6B: Relative expression of DEGs associated with lipid metabolism in liver from different mice models [Z scores; n=4-6/group; provided as <u>Excel Table S34</u>].

**Table S35.** Pathway enrichment analysis of DEGs in liver from different mice models[enrichment scores].

**Table S36.** Data for Figure 6D: GO analysis of DEGs in livers from different mice models [provided as <u>Excel Table S36</u>].

**Table S37.** Superclass constitution of lipid metabolites in liver from different mice models [lipid counts; average ratio (DEHP/CT ratio)].

**Table S38.** Data for Figure 6F: Class constitution of lipid metabolites in livers from different mice models [lipid counts; average ratio (DEHP/CT ratio); provided as <u>Excel Table S38</u>].

**Table S39.** Evaluation of mitochondrial OXPHOS, fatty acid uptake and LD in HepG2 cells treated with MEHP with/without specific inhibitors [Mean stain area ( $\mu$ m<sup>2</sup>); mean±SD; n=5/treatment].

**Table S40.** Evaluation of mitochondrial OXPHOS and LD in HepG2 cells treated with MEHP with/without M2 macrophages [Mean stain area ( $\mu$ m2); mean  $\pm$  SD; n=5/treatment].

**Table S41.** Relative expression of DEGs associated with OXPHOS in HepG2 cells treated with MEHP with/without M2 macrophages [Z scores; n=2/treatment].

**Table S42.** Relative expression of DEGs associated with glycolysis in HepG2 cells treated with MEHP with/without M2 macrophages [Z scores; n=3/treatment].

**Table S43.** Relative expression of DEGs associated with lipid metabolism in HepG2 cells treated with MEHP with/without M2 macrophages [Z scores; n=3/treatment].

**Table S44.** Body weight fluctuation in DEHP treated mice [g; mean ± SD; n=10/group].

**Table S45.** Data for Figure S2B: Enriched KEGG pathways (level 2) in DEHP treated mice [number of genes; provided as <u>Excel Table S45</u>].

**Table S46.** Relative expression of DEGs associated with lipid metabolism in HepG2 cells treated with MEHP [Z scores; n=3/treatment].

**Table S47.** Relative expression of DEGs associated with PPAR pathway in DEHP treated mice [Z scores; n=6/group; provided as Excel Table S47].

**Table S48.** Pathway enrichment analysis of DEGs in HepG2 cells treated with MEHP [n=3/treatment].

**Table S49.** Clinical chemistry panel in WT and Mac-KO mice [mean  $\pm$  SD (n)].

**Table S50.** Macrophages and neutrophils analysis in liver of WT and Mac-KO mice using flow cytometry [% of CD45<sup>+</sup> cells; mean  $\pm$  SD (n)].

**Table S51.** Macrophages featured cytokines analysis in liver of WT and Mac-KO mice using flow cytometry [pg/mg protein; mean  $\pm$  SD; n=5].

**Table S52.** CD69<sup>+</sup> M1 macrophages evaluation in BMDM from WT and Mac-KO mice [mean stain area ( $\mu$ m2); mean±SD (n)].

**Table S53.** Relative expression of DEGs associated with M1 macrophages in MEHP treated macrophages [Z scores; n=3/treatment; provided as Excel Table S53].

**Table S54.** Macrophages lipid metabolism analysis in liver of WT and Mac-KO mice using flow cytometry [% of CD45<sup>+</sup> cells; mean  $\pm$  SD; n=5/treatment].

Figure S1. Gating strategy for flow cytometry of mouse liver.

Figure S2. The effect of DEHP on hepatic lipid metabolism.

Figure S3. Activation of PPAR isoforms by DEHP and MEHP at the molecular level.

**Figure S4.** The function of PPAR $\gamma$  in hepatic macrophages.

Figure S5. The effects of DEHP and MEHP on Macrophages polarization.

Additional File- Excel Document

Table S1 Key reagents and materials

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
PE anti-human CCR7 (clone#G043H7, FACS: 2 µl /10 <sup>6</sup> cells)	Biolegend	Cat#353204,
		RRID: AB_10913813
Alexa Fluor® 647 anti-human CCR7 (clone#G043H7, FACS:	Biolegend	Cat#353218;
$2 \mu l /10^6 \text{cells})$		RRID: AB_10917385
PE anti-human CD209 (clone#9E9A8, FACS: 2 μl /10 <sup>6</sup> cells)	Biolegend	Cat#330106;
		RRID:AB_1134052
FITC anti-human CD209 (clone#9E9A8, FACS: 2 µl /10 <sup>6</sup> cells)	Biolegend	Cat#330104;
		RRID:AB_1134048
PE anti-human CD36 antibody(clone#AC106, FACS: 2 µl /10 <sup>6</sup>	Miltenyi	Cat#130-095-472;
cells)		RRID:AB_10827705
PE anti-mouse CD206 (clone#C068C2, FACS: 0.5 µg /10 <sup>6</sup>	Biolegend	Cat#141706;
cells)		RRID:AB_10895754
APC anti-mouse CD206 (clone#C068C2, FACS: 0.5 µg /10 <sup>6</sup>	Biolegend	Cat#141708;
cells)		RRID:AB_10900231
FITC anti-mouse CD206 (clone#C068C2, FACS: 0.5 µg /10 <sup>6</sup>	Biolegend	Cat#141704;
cells)		RRID:AB_10901166
Alexa Fluor® 647 anti-mouse CD69 (clone#H1.2F3, FACS:	Biolegend	Cat#104518;
$0.25 \ \mu g \ /10^6 \ cells)$		RRID:AB_492847
APC anti-mouse CD36 (clone#HM36, FACS: 0.25 µg /106	Biolegend	Cat#102612;
cells)		RRID:AB_2072639
PerCP/Cyanine5.5 anti-CD11b (clone#M1/70, FACS: 0.25	Biolegend	Cat#123126;
$\mu$ g /10 <sup>6</sup> cells)		RRID:AB_893483
PE anti-mouse F4/80(Clone#BM8, FACS: 0.5 µg /10 <sup>6</sup> cells)	Biolegend	Cat#123110;
		RRID:AB_893486
PerCP anti-mouse F4/80(Clone#BM8, FACS: 0.5 µg /10 <sup>6</sup> cells)	Biolegend	Cat#123126;
		RRID:AB_893483
PerCP/Cyanine5.5 anti-mouse CD45 (clone#30-F11, FACS:	Biolegend	Cat#103132;
$0.25 \ \mu g \ / 10^6 \ cells)$		RRID:AB_893340
FITC anti-mouse CD45 (clone#30-F11, FACS: 0.25 $\mu g$ /106	Biolegend	Cat#103108;
cells)		RRID:AB_312973
APC Anti-mouse CD45 (clone#30-F11, FACS: 0.25 µg /106	Biolegend	Cat#103112;
cells)		RRID:AB_312977
PE anti-mouse CCR2 (clone# SA203G11, FACS: 0.5 $\mu g$ /10 $^6$	Biolegend	Cat#150610;
cells)		RRID:AB_2616982
Alexa Fluor® 647 anti-mouse CCR2 (clone# SA203G11,	Biolegend	Cat#150604;
FACS: 0.5 µg /10 <sup>6</sup> cells)		RRID:AB_2566140
APC anti-mouse LY6C (clone#HK1.4, FACS: $0.1 \mu g / 10^6$ cells)	Biolegend	Cat# 128016;
		RRID:AB_1732076
PPARγ rabbit mAb (clone# C26H12, IHC:1/200)	Cell Signaling	Cat#2435S;
	Technology	RRID:AB_2166051

Ki-67 rabbit mAb (clone# D3B5, IHC:1/200)	Cell Signaling	Cat#12202;	
	Technology	RRID:AB_2620142	
F4/80 rabbit mAb (clone# D4C8V, IHC:1/200)	Cell Signaling	Cat#30325;	
	Technology	RRID:AB_2798990	
PPAR alpha polyclonal antibody(IHC:1/200)	Invitrogen	Cat#PA1-822A;	
		RRID:AB_2165595	
Mouse CLEC4F/CLECSF13 antibody(IHC:1/200)	R&D systems	Cat#AF2784;	
		RRID:AB_2081339	
Mouse IL-1ra/IL-1F3 antibody (clone# 962802, IHC: 10	R&D systems	Cat#MAB4802;	
µg/ml)		N/A	
COX7A2L Rabbit Polyclonal antibody(IHC: 1/100)	Proteintech	Cat#11416-1-AP;	
		RRID:AB_2245402	
Rabbit anti-rat IgG H&L, HRP (IHC: 1/1000)	Abcam	Cat#ab6734;	
		RRID:AB_955450	
Donkey anti-goat IgG H&L, HRP (IHC:1/1000)	Santa Cruz	Cat#SC-2033;	
		RRID:AB_631729	
Chemicals, Peptides, and Recombinant Proteins			
Recombinant Human IL-4	Peprotech	Cat#200-04	
Recombinant Human IL-13	Peprotech	Cat#AF-200-13	
Recombinant Human IFNy	Peprotech	Cat#300-02	
Recombinant Murine M-CSF	Peprotech	Cat#315-02	
Recombinant Murine IL-4	Peprotech	Cat#214-14	
Recombinant Murine IL-13	Peprotech	Cat#210-13	
Recombinant Murine IFNy	Peprotech	Cat#315-05	
Recombinant human IL1RA	R&D systems	Cat#280-RA-050	
Human PPARy protein	Sino Biological	Cat#12019-H20B	
Human PPARa protein	Abnova	Cat#H00005465-P01	
Human PPARo protein	Abnova	Cat#H00005467-P01	
Lipopolysaccharides (LPS)	Sigma Aldrich	Cat#L6529	
Phorbol 12-myristate 13-acetate (PMA)	Sigma Aldrich	Cat#P8139	
DEHP	Sigma Aldrich	Cat# 67261	
МЕНР	MedChemExpress	Cat# HY-W018392	
WY14643	R&D systems	Cat#1312	
Rosiglitazone	Cayman	Cat#71740	
T0070907	R&D systems	Cat#2301	
GW9662	Cayman	Cat#70785	
GW6471	MedChemExpress	Cat# HY-15372	
Critical Commercial Assays			
Foxp3 / Transcription Factor Staining Buffer Set	Invitrogen	Cat#00-5523-00	
LEGENDplex Human Macrophage/Microglia Panel	Biolegend	Cat#740502	
LEGENDplex Mouse Macrophage/Microglia Panel	Biolegend	Cat#740845	
RNeasy Mini Kit	QIAGEN	Cat#74104	
Opal multiplex IHC research kit	Perkin Elmer	Cat# NEL810001KT	

DAB substrate kit	Cell Signaling	Cat#8095S
Deposited Data	Technology	
Pow and analyzed data	This paper and	http://dy.doi.org/10.17
Kaw and anaryzed data	Mandalay Data	632/zd6pt6hb7y 1
PNA sequencing data	This paper	GEO: GSE150120
RNA-sequencing data	This paper	GEO: GSE159120
RNA-sequencing data	This paper	GEO: GSE160373:
RNA-sequencing data	This paper	GEO: GSE160375,
Experimental Models: Cell Lines		GEO. G3E100820
THP_1	American Type Culture	Cat#TIB_202
1111-1	Collection	RRID:CVCL_0006
THP-1 with PPRE-eGFP	This paper	N/A
HenG?	Cell Bank of Type	Cat#SCSP-510: N/A
11902	Culture Collection	
	Committee of the	
	Chinese Academy of	
	Sciences	
Experimental Models: Organisms/Strains		
Mouse: C57BL/6	In house breeding	N/A
Mouse: PPARγ <sup>loxP</sup> :B6.129- <i>Pparg</i> <sup>tm2Rev</sup> /J	Jackson Laboratory	Cat# JAX:004584,
		RRID:IMSR_JAX:004
		584
Mouse: LysM <sup>Cre</sup> : B6.129P2-Lyz2 <sup>tm1(cre)Ifo</sup> /J	Jackson Laboratory	Cat# JAX:004781,
		RRID:IMSR_JAX:004
		781
Mouse: Alb <sup>Cre</sup> : B6.Cg- <i>Speer6-ps1</i> <sup>Tg(Alb-cre)21Mgn</sup> /J	Jackson Laboratory	Cat# JAX:003574,
		RRID:IMSR_JAX:003
		574
Recombinant DNA		
Plasmid: PPRE-H2B-eGFP	Addgene	Cat#84393;
		RRID:Addgene_84393
Software and Algorithms		-
GraphPad prism (v 6.01)	GraphPad Software	https://www.graphpad.
		com/
ImageXpress(v6.0)	Molecular Device	https://www.molecular
		devices.com
Other		
Human TruStain FcX	Biolegend	Cat#422302
TruStain FcX (anti-mouse)	Biolegend	Cat#101320
Fetal Bovine Serum (FBS)	Gibco	Cat#10099141C
Fixation and Permeabilization Solution	BD Bioscience	Cat#554722
FluoroBrite TM DMEM	Gibco	Cat#A1896702

Hoechst 33342	Solarbio	Cat#C0031
Perm/Wash buffer	BD Bioscience	Cat#554723
RPMI 1640 (ATCC modification)	Gibco	Cat#A1049101
2-mercaptoethanol	Sigma Aldrich	Cat#M3148
BODIPY 493/503	Invitrogen	Cat#D3922
HCS LipidTOX <sup>™</sup> Red Neutral Lipid Stain, for cellular	Invitrogen	Cat#34476
imaging		
TMRE	Abcam	Cat# ab113852
MitoSOX <sup>TM</sup> Red Mitochondrial Superoxide Indicator	Invitrogen	Cat# M36008
CellROX deep red	Invitrogen	Cat#C10491

**Table S2** Liver weight and liver index in DEHP treated mice [mean  $\pm$  SD; n=10/group].

Table 52 Erver weight and river index in DETT reduced inice [inean + 5D, if 10/group].					
Dose group	0 mg/kg BW	625 mg/kg BW	1250 mg/kg BW	2500 mg/kg BW	
Liver weight(g)	$1.08 \pm 0.13$	1.29±0.05***	1.28±0.11***	1.49±0.08***	
Liver index(%BW)	4.68±0.37	5.38±0.30***	5.35±0.39***	6.50±0.29***	

Note: Correspond to Figure 1A. BW, body weight; DEHP, di(2-Ethylhexyl) phthalate; SD, standard deviation. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Dunnett's multiple comparisons test].

Table S3 (	Concentration	of MEHP ir	ı plasma	and liver	of DEHP	treated mice	$[mean \pm SD]$	(n)].
------------	---------------	------------	----------	-----------	---------	--------------	-----------------	-------

Dose group	0 mg/kg BW	625 mg/kg BW	1250 mg/kg BW	2500 mg/kg BW
For Figure 1B:	$0.40{\pm}0.44$	16.36±5.71*	25.34±4.54**	53.26±18.61***
Plasma(ng/ml)	(6)	(6)	(6)	(6)
For Figure 1C:	24.45±12.69	273.64±32.97*	335.77±80.03*	659.85±277.48***
Liver(ng/g tissue)	(5)	(5)	(5)	(5)

Note: Correspond to Figure 1B and 1C. BW, body weight; DEHP, di(2-Ethylhexyl) phthalate; SD, standard deviation; MEHP, mono-2-ethylhexyl phthalate. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Dunnett's multiple comparisons test].

Table 54 Chillean	enemisary panel of L		$\lim_{n \to \infty} \frac{1}{2} \int \frac{1}$	oupj
Dose group	0 mg/kg BW	625 mg/kg BW	1250 mg/kg BW	2500 mg/kg BW
TG(mM)	$0.74{\pm}0.11$	$0.58{\pm}0.07{**}$	0.53±0.04***	0.55±0.03**
CHO(mM)	$1.55 \pm 0.14$	$1.66{\pm}0.08$	1.86±0.07**	2.12±0.18***
LDLC(mM)	$0.25 \pm 0.04$	$0.26{\pm}0.02$	$0.27{\pm}0.04$	0.32±0.03**
HDLC(mM)	$1.04{\pm}0.13$	$1.15 \pm 0.05$	1.19±0.10	1.37±0.10***
ALP(U/L)	73.96±3.05	79.60±7.16	90.00±7.04**	106.40±5.27***

Table S4 Clinical chemistry panel of DEHP treated mice [mean ± SD; n=5/group]

Note: Correspond to Figure 1D. ALP, alkaline phosphatase; BW, body weight; CHO, cholesterol; DEHP, di(2-Ethylhexyl) phthalate; HDLC, high-density lipoprotein cholesterol; LDLC, low-density lipoprotein cholesterol; SD, standard deviation; TG, triglycerides; U/L, units per liter. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Dunnett's multiple comparisons test].

**Table S5** Concentration of hepatic TG in DEHP treated mice [mean  $\pm$  SD; n=5/group].

Dose group	0 mg/kg BW	625 mg/kg BW	1250 mg/kg BW	2500 mg/kg BW	
Liver(µg/mg tissue	)1.02±0.25	2.48±0.57**	4.59±1.04***	6.25±0.55***	

Note: Correspond to Figure 1E. BW, body weight; DEHP, di(2-Ethylhexyl) phthalate; SD, standard deviation; TG, triglycerides. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Dunnett's multiple comparisons test].

**Table S6** Hepatic pathological severity and oil red staining evaluation in DEHP treated mice [evaluating scores; mean  $\pm$  SD; n=6/group].

Dose group	0 mg/kg BW	625 mg/kg BW	1250 mg/kg BW	2500 mg/kg BW
Hepatic pathological severity	v1.33±0.52	6.00±2.45**	11.17±3.37***	14.17±2.99***
Oil red staining intensity	$1.17 \pm 0.41$	3.33±1.51	10.00±3.52***	12.17±3.37***

Note: Correspond to Figure 1F. BW, body weight; DEHP, di(2-Ethylhexyl) phthalate; SD, standard deviation. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Dunnett's multiple comparisons test].

Dose(log10, µM)	BODIPY FL C12 uptake	LD lipid accumulation
2.30	386.06±20.77***	/
2.00	385.18±68.99***	/
1.41	309.85±72.84***	/
1.13	265.58±67.09***	/
0.86	202.41±21.33*	/
0.62	166.26±29.19	/
0.41	151.71±15.03	/
0.00(control)	$100.00 \pm 0.00$	/
2.30	/	145.91±12.61***
2.00	/	155.81±5.08***
1.71	/	135.88±2.01***
1.41	/	134.99±10.78***
1.13	/	118.89±9.69*
0.62	/	104.89±5.42
0.41	/	103.87±9.74
0.00(control)	/	100.00±0.00

**Table S7** Fatty acid uptake and lipid accumulation evaluation in MEHP treated HepG2 cells [Mean stain area ( $\mu$ m<sup>2</sup>) (% of control); mean ± SD; n=4/treatment].

Note: Correspond to Figure 1G. BODIPY FL C16, fluorescence probe for C16 fatty acids; LD, lipid droplet; MEHP, mono-2-ethylhexyl phthalate; SD, standard deviation. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Dunnett's multiple comparisons test].

**Table S8** Data for Figure 2A: Binding affinity analysis of PPARα [Resonance units (RU); n=2006-2146; provided as Excel Table S8].

**Table S9** Data for Figure 2A: Binding affinity analysis of PPARγ [Resonance units (RU); n=1583-2131; provided as Excel Table S9].

Table S10PPRE and CD36 response in PPRE-THP-1 cells derived macrophages [positive cells(%); mean±SD; n=6 sites/treatment].

Treatment	CT(DMSO)	WY	RGZ	T0070907	DEHP	MEHP
PPRE	22.84±4.63	29.54±4.64*	37.01±4.59***	<sup>•</sup> 24.21±2.22	28.83±1.67*	29.45±3.28*
CD36	35.11±4.02	47.14±6.56*	62.91±3.34***	29.48±3.07*	46.92±3.87*	52.04±6.81**
Note: Correspond to Figure 2C. CT, vehicle control; DMSO, dimethyl sulfoxide; DEHP, di(2-						
Ethylhexyl)	Ethylhexyl) phthalate; MEHP, mono-2-ethylhexyl phthalate; WY, WY14643, PPARα agonist; RGZ,					
rosiglitazon	rosiglitazone, PPAR $\gamma$ agonist; T0070907, PPAR $\gamma$ antagonist; SD, standard deviation. * $p$ <0.05,					
**p<0.01, ***p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with						
post hoc multiple comparison correction using Dunnett's multiple comparisons test].						

Concentration(l	og10, μM) WY	RGZ	GW6471	GW9662	T0070907
2.30	126.34	162.38	/	/	/
2.00	135.06	162.51	/	/	/
1.70	112.00	162.51	/	/	/
1.40	88.56	155.70	/	/	/
1.10	73.14	145.73	/	/	/
0.80	75.43	143.78	/	/	/
0.49	82.50	131.59	/	/	/
0.19	68.15	120.50	/	/	/
-6.00	73.26	73.26	/	/	/
1.30	/	/	58.37	83.08	44.42
1.00	/	/	67.38	86.19	48.07
0.70	/	/	66.57	82.34	44.93
0.40	/	/	74.31	93.06	43.01
0.10	/	/	74.07	94.15	48.01
-0.20	/	/	73.38	82.57	54.98
-0.51	/	/	62.74	75.32	58.90
-0.81	/	/	73.26	73.26	73.26

Table S11Dose response curves of PPRE in PPRE-THP-1 cells derived macrophages [mean<br/>stain area ( $\mu m^2$ ); mean; n=3/treatment].

Note: Correspond to Figure 2D. WY, WY14643, PPARα agonist; GW6471, PPARα antagonist; GW9662, PPARγ antagonist; RGZ, rosiglitazone, PPARγ agonist; T0070907, PPARγ antagonist and inverse agonist.

Concentration(le	og10, μM) WY	RGZ	GW6471	GW9662	T0070907
2.30	248.23	274.00	/	/	/
2.00	247.48	274.90	/	/	/
1.70	204.21	272.40	/	/	/
1.40	155.63	267.38	/	/	/
1.10	127.37	250.40	/	/	/
0.80	131.57	239.42	/	/	/
0.49	136.57	220.35	/	/	/
0.19	116.47	239.65	/	/	/
-1.20	118.64	118.64	118.64	118.64	118.64
1.30	/	/	91.82	142.00	32.87
1.00	/	/	104.29	142.11	38.05
0.70	/	/	106.56	144.74	49.87
0.40	/	/	111.02	146.70	44.93
0.10	/	/	120.55	156.97	44.53
-0.20	/	/	129.32	159.01	53.98
-0.51	/	/	123.97	140.34	61.82
-0.81	/	/	114.27	130.74	56.55

Table S12Dose response curve of CD36 in PPRE-THP-1 cells derived macrophages [mean<br/>stain area ( $\mu m^2$ ); mean, n=3/treatment)].

Note: Correspond to Figure 2D. WY, WY14643, PPARα agonist; GW6471, PPARα antagonist; GW9662, PPARγ antagonist; RGZ, rosiglitazone, PPARγ agonist; T0070907, PPARγ antagonist and inverse agonist.

<u> </u>	-	IID						0.500.5
Concentration	ME	/HP	M+G	W64/1	M+G	W9662	M+10	070907
(log10, µM)	PPRE	CD36	PPRE	CD36	PPRE	CD36	PPRE	CD36
2.30	135.76	274.05	83.60	198.29	101.69	208.05	72.75	156.07
2.00	131.30	251.43	65.56	153.60	113.41	200.50	87.55	155.45
1.70	117.13	206.65	61.20	122.91	113.01	208.83	79.08	125.57
1.40	101.23	182.55	51.40	98.85	91.39	172.65	69.94	94.16
1.10	91.51	152.13	58.30	105.67	92.26	173.16	45.27	58.25
0.80	85.18	143.89	49.34	103.64	93.83	151.63	51.89	56.31
0.49	80.87	153.89	53.96	103.54	82.93	145.05	46.54	44.71
0.19	72.38	120.15	50.40	96.44	77.36	138.10	38.66	41.72
2.30	135.76	274.05	83.60	198.29	101.69	208.05	72.75	156.07
2.00	131.30	251.43	65.56	153.60	113.41	200.50	87.55	155.45
1.70	117.13	206.65	61.20	122.91	113.01	208.83	79.08	125.57
1.40	101.23	182.55	51.40	98.85	91.39	172.65	69.94	94.16
1.10	91.51	152.13	58.30	105.67	92.26	173.16	45.27	58.25
0.80	85.18	143.89	49.34	103.64	93.83	151.63	51.89	56.31
0.49	80.87	153.89	53.96	103.54	82.93	145.05	46.54	44.71
0.19	72.38	120.15	50.40	96.44	77.36	138.10	38.66	41.72

**Table S13**Dose response curve of PPRE and CD36 in MEHP treated macrophages with/withoutPPAR $\alpha/\gamma$  antagonists [mean stain area ( $\mu$ m<sup>2</sup>); mean, n=3/treatment].

Note: Correspond to Figure 2E. GW6471, PPAR $\alpha$  antagonist; GW9662, PPAR $\gamma$  antagonist; M, MEHP, mono-2-ethylhexyl phthalate; T0070907, PPAR $\gamma$  antagonist and inverse agonist.

	U	1	L ()]	
	Treatment	СМС	DEHP	
Liver weight(g)	WT	1.24±0.13 (7)	1.65±0.15 (7)**	
	Нер-КО	1.16±0.17 (5)	1.60±0.08 (5)**	
Liver index(%BV	V) WT	4.69±0.48 (7)	6.34±0.37 (7)***	
	Hep-KO	4.45±0.52 (5)	6.23±0.14 (5)***	

**Table S14**Liver weight and liver index in WT and Hep-KO mice [mean  $\pm$  SD (n)]

Note: Correspond to Figure 3A. BW, body weight; CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Hep-KO, hepatocyte-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\* p<0.001, relative to vehicle control [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

Measurements	Treatment	СМС	DEHP
TG(mM)	WT	0.64±0.06 (7)	0.49±0.07 (7)***
	Нер-КО	0.64±0.05 (5)	0.49±0.05 (5)**
CHO(mM)	WT	1.62±0.41 (7)	2.30±0.26 (7)***
	Нер-КО	1.71±0.35 (5)	2.36±0.06 (5)**
CHE(mM)	WT	2.63±0.21 (7)	3.95±0.50 (7)***
	Нер-КО	2.74±0.16 (4)	3.89±0.31 (5)***
ALP(U/L)	WT	72.67±5.05 (6)	93.00±11.25 (7)**
	Нер-КО	70.20±8.07 (5)	96.40±14.84 (5)**
HDLC(mM)	WT	1.22±0.13 (7)	1.57±0.12 (6)***
	Нер-КО	1.19±0.21 (5)	1.58±0.05 (5)***
LDLC(mM)	WT	0.25±0.03 (5)	0.35±0.04 (6)**
	Нер-КО	0.27±0.06 (5)	0.42±0.04 (5)***

Table S15Clinical chemistry panel in WT and Hep-KO mice  $[mean \pm SD (n)].$ 

Note: Correspond to Figure 3B. ALP, alkaline phosphatase; BW, body weight; CHE, cholinesterase; CHO, cholesterol; CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; HDLC, high-density lipoprotein cholesterol; Hep-KO, hepatocyte-specific PPAR $\gamma$  knockout; LDLC, low-density lipoprotein cholesterol; SD, standard deviation; TG, triglycerides; U/L, units per liter; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

Table S16Concentration of hepatic TG in WT and Hep-KO mice  $[\mu g/mg \text{ tissue; mean } \pm \text{ SD} ]$ (n)].

Treatment	СМС	DEHP
WT	1.05±0.13 (7)	2.64±0.20 (7)***
Нер-КО	0.88±0.23 (5)	2.71±0.25 (5)***

Note: Correspond to Figure 3C. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Hep-KO, hepatocyte-specific PPAR $\gamma$  knockout; SD, standard deviation; TG, triglycerides; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

Table S17Hepatic pathological severity evaluation in WT and Hep-KO mice [evaluating scores;mean  $\pm$  SD (n)].

······································				
Treatment	СМС	DEHP		
WT	1.29±0.49 (7)	11.29±3.86 (7)***		
Hep-KO	1.20±0.45 (5)	10.60±3.44 (5)***		

Note: Correspond to Figure 3D. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Hep-KO, hepatocyte-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

= $=$ $=$ $=$ $=$ $=$ $=$ $=$ $=$ $=$				
	Treatment	СМС	DEHP	
CLEC4F <sup>+</sup> KCs	WT	102.43±9.85 (7)	50.71±10.05 (7)***	
	Hep-KO	98.60±10.01 (5)	46.00±18.73 (5)***	
F4/80 <sup>+</sup>	WT	286.14±26.33 (7)	306.43±26.15 (7)	
macrophages	Hep-KO	290.60±41.32 (5)	293.20±24.99 (5)	

Table S18Macrophages evaluation in liver sections from WT and Hep-KO mice [cell numberper site; mean  $\pm$  SD (n)].

Note: Correspond to Figure 3F. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Hep-KO, hepatocyte-specific PPAR $\gamma$  knockout; KCs, kupffer cells; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

	•		-	
	Treatment	СМС	DEHP	
Liver weight(g)	WT	1.36±0.30	1.87±0.20***	
	Mac-KO	1.50±0.09	1.70±0.10	
Liver index(%BV	W) WT	4.60±0.71	6.39±0.16***	
	Mac-KO	4.94±0.27	5.55±0.20ª	

Table S19Liver weight and liver index in WT and Mac-KO mice [mean  $\pm$  SD; n=5/group]

Note: Correspond to Figure 4A. BW, body weight; CMC, sodium carboxymethylcellulose,vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control; ap<0.05, bp<0.01, cp<0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

Table S20Concentration of hepatic TG in WT and Mac-KO mice [ $\mu$ g/mg tissue; mean  $\pm$  SD;n=5/group].

Treatment	СМС	DEHP
WT	0.99±0.18	3.09±0.36***
Mac-KO	0.94±0.34	1.55±0.36*,c

Note: Correspond to Figure 4B. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; TG, triglycerides; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control;  ${}^{a}p$ <0.05,  ${}^{b}p$ <0.01,  ${}^{c}p$ <0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

	Treatment	СМС	DEHP	
Pathological	WT	1.40±0.55	12.20±2.49***	
severity	Mac-KO	1.60±0.55	6.60±1.95**,c	
Oil red stain	ning WT	1.20±0.45	11.40±3.13***	
intensity	Mac-KO	$1.60\pm0.89$	5.60±1.67*,c	

Table S21Hepatic pathological severity and oil red staining evaluation in WT and Mac-KOmice [evaluating scores; mean  $\pm$  SD; n=5/group].

Note: Correspond to Figure 4C. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control;  $a_p$ <0.05,  $b_p$ <0.01,  $c_p$ <0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

Table S22M2 macrophages analysis in liver of WT and Mac-KO mice using flow cytometry [%of CD45<sup>+</sup> cells; mean  $\pm$  SD; n=5/group].

	Treatment	СМС	DEHP	
F4/80 <sup>+</sup> CD206 <sup>+</sup>	WT	16.30±3.26	14.08±2.33**	
M2 Macrophages	Mac-KO	16.96±2.01	$22.60\pm2.62^{\circ}$	

Note: Correspond to Figure 4D. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control; ap<0.05, bp<0.01, cp<0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

Table S23Macrophages evaluation in liver sections from WT and Mac-KO mice [cell numberper site; mean  $\pm$  SD; n=5/group].

	Treatment	СМС	DEHP	
CLEC4F+	WT	101.60±10.88	46.80±13.16***	
Kupffer Cells	Mac-KO	101.60±11.61	117.80±13.74°	

Note: Correspond to Figure 4E. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control; ap<0.05, bp<0.01, cp<0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

	Treatment	СМС	DEHP	
CD11b <sup>+</sup> CCR2 <sup>+</sup> Ly6c <sup>high</sup>	WT	$1.90{\pm}0.67$	3.88±0.83***	
Inflammatory monocyte	Mac-KO	1.15±0.30	$2.26 \pm 0.53^{b}$	
CD11b <sup>+</sup> CCR2 <sup>+</sup> Ly6c <sup>low</sup>	WT	11.45±2.41	12.27±1.84	
Patrolling monocyte	Mac-KO	12.81±1.22	$17.47 \pm 2.92^{b}$	

Table S24Monocyte evaluation in liver from WT and Mac-KO mice [% of CD45+cells; mean± SD; n=5/group]

Note: Correspond to Figure 4F. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control; ap<0.05, bp<0.01, cp<0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

Table S25M2 macrophages polarization evaluation in BMDM from WT and Mac-KO mice $[CD206^+ cells (\%); mean \pm SD; n=15$  technical replicates/treatment].

Treatment	CT(DMSO)	DEHP	MEHP
WT	90.13±2.00	68.14±4.50***	73.78±4.59***
Mac-KO	92.55±2.43	76.18±3.88°	84.04±4.46°

Note: Correspond to Figure 5A. BMDM, bone marrow derived macrophages; CT, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; DMSO, dimethyl sulfoxide; MEHP, mono-2-ethylhexyl phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control.  ${}^{a}p<0.05$ ,  ${}^{b}p<0.01$ ,  ${}^{c}p<0.001$ , relative to WT DEHP or WT MEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

Table S26Relative expression of DEGs associated with M2 macrophages in MEHP treatedmacrophages [Z scores; n=3/treatment; provided as <a href="mailto:ExcelTable S26">Excel Table S26</a>].

Table S27Relative expression of DEGs associated with PPAR pathway in MEHP treatedmacrophages [Z scores; n=3/treatment; provided as Excel Table S27].

$(\mu m2)$ ; mean $\pm$ SD; n=2/subtype].							
Treatment	CD209	PPRE	<b>CD36</b>	LD			
M0	21.90±0.14	37.09±5.29	115.42±25.90	116.87±24.68			

104.42±15.16

204.92±0.64\*\*

129.52±32.37

 $215.70{\pm}4.13{**}$ 

53.16±8.15

88.42±6.98\*\*

M1

M2

 $64.16 \pm 2.28*$ 

346.11±5.79\*\*

Table S28 Characteristics of PPRE-THP-1 cells derived M2 macrophages [Mean stain area

Note: Correspond to Figure 5E. LD, lipid droplet; PPRE, peroxisome proliferator-activated receptor response elements; SD, standard deviation. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

Concentration	DEHP				MEHI	1EHP	
(log10, μM)	CD209	PPRE	LD	CD209	PPRE	LD	
2.30	49.23	73.13	32.05	39.04	84.41	43.07	
2.00	67.16	/	39.74	52.23	81.93	61.16	
1.70	/	82.40	26.68	65.51	82.90	46.08	
1.40	56.72	/	/	73.63	77.84	57.08	
1.10	71.68	79.55	42.93	66.98	/	63.00	
0.80	64.71	86.07	50.59	82.05	/	78.72	
0.49	85.89	85.18	/	94.20	86.56	89.05	
0.19	78.34	92.57	71.80	/	86.99	/	
0.00	100.00	100.00	100.00	100.00	100.00	100.00	

Table S29Dose response curves of PPRE/CD209/LD in DEHP/MEHP treated M2macrophages [% of control; mean, n=3/treatment].

Note: Correspond to Figure 5F. DEHP, di(2-Ethylhexyl) phthalate; LD, lipid droplet; MEHP, mono-2-ethylhexyl phthalate; PPRE,peroxisome proliferator-activated receptor response elements.

Concentration	WW	DC7	MEIID	CW7647
(log10, μM)	VV X	KGZ	MEHP	GW/04/
1.40	116.19	26.99	37.79	/
1.10	100.02	50.91	49.02	/
0.80	119.79	60.79	56.73	/
0.49	95.07	61.34	73.97	/
0.19	93.99	58.64	76.44	/
-0.11	69.17	61.09	76.12	/
-0.41	71.35	71.35	71.35	/
1.30	/	/	/	202.88
1.00	/	/	/	195.04
0.70	/	/	/	193.10
0.40	/	/	/	180.27
0.10	/	/	/	122.67
-0.20	/	/	/	/
-0.51	/	/	/	95.27
-0.81	/	/	/	71.35

Table S30Data for Figure 5G: Dose response curves of CD209 in PPAR $\alpha/\gamma$  agonists treated M2macrophages [Mean stain area ( $\mu$ m<sup>2</sup>); mean; n=3/treatment].

Note: Correspond to Figure 5G. GW7647, PPARα agonist; MEHP, mono-2-ethylhexyl phthalate; RGZ, rosiglitazone, PPARγ agonist; WY, WY14643, PPARα agonist.

fielded W12 maerophages	s [mean stain area (µm );	mean, ii 5/ iieaimeni	J	
Concentration (log10, μM)	GW6471	GW9662	<b>T0070907</b>	
0.10	62.84	83.37	93.42	
-0.20	68.33	82.01	91.64	
-0.51	76.97	82.45	90.49	
-0.81	77.89	80.13	83.53	
-1.11	75.14	69.83	78.47	

**Table S31**Data for Figure 5H: Dose response curves of CD209 in PPAR $\alpha/\gamma$  antagoniststreated M2 macrophages [Mean stain area ( $\mu$ m<sup>2</sup>); mean, n=3/treatment]

Note: Correspond to Figure 5H. GW6471, PPARα antagonist; GW9662, PPARγ antagonist; T0070907, PPARγ antagonist and inverse agonist.

Table S32	Response	of CD2	09/CD36/LD	in MEF	IP treated	1 M2	macrophages	with/witho	ut
PPAR $\alpha/\gamma$ antag	gonists [Me	ean stain	area (µm <sup>2</sup> ); n	$mean \pm S$	O (n)].				

Treatment	CT(DMSO)	MEHP	M+GW6471	M+GW9662
CD209	153.98±29.47 (4)	98.39±26.21 (3)*	45.29±4.79 (3)***,3	111.46±14.36 (3)
CD36	52.09±2.50 (4)	35.10±3.04 (5)**	18.19±4.29 (5)***,2	41.96±9.78 (5)
LD	347.32±13.57 (4)	296.67±39.66 (4)*	233.82±15.60 (4)**,	$^{1}374.59 \pm 13.60 \ (4)^{2}$

Note: Correspond to Figure 5I. CT, vehicle control; DMSO, dimethyl sulfoxide; GW6471, PPAR $\alpha$  antagonist; GW9662, PPAR $\gamma$  antagonist; MEHP, mono-2-ethylhexyl phthalate; SD, standard deviation. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control;  $^{1}p$ <0.05,  $^{2}p$ <0.01,  $^{3}p$ <0.001, relative to Vehicle control;  $^{1}p$ <0.05,  $^{2}p$ <0.01,  $^{3}p$ <0.001, relative to MEHP treatment [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

Table S33Data for Figure 6A: Transcriptomic profiling of livers from different mice models[DEGs; provided as Excel Table S33].

**Table S34**Data for Figure 6B: Relative expression of DEGs associated with lipid metabolismin liver from different mice models [Z scores; n=4-6/group; provided as Excel Table S34].

Pathways	WT	Нер-КО	Mac-KO
PPAR signaling pathway	11.52	11.30	9.46
Fatty acid degradation	10.75	10.90	9.17
Retinol metabolism	7.59	7.05	8.18
Biosynthesis of unsaturated fatty acids	6.65	2.74	5.87
Chemical carcinogenesis	6.45	3.75	3.05
Oxidative phosphorylation	5.24	4.98	0.00
Fatty acid metabolism	4.68	1.96	3.87
Valine, leucine and isoleucine degradation	4.46	2.82	3.74
Steroid hormone biosynthesis	3.99	0.00	4.17
Arachidonic acid metabolism	3.92	4.84	6.48

**Table S35**Pathway enrichment analysis of DEGs in liver from different mice models[enrichment scores].

Note: Correspond to Figure 6C. DEGs, differentially expressed genes; Hep-KO, hepatocyte-specific PPARγ knockout; Mac-KO, macrophage-specific PPARγ knockout; WT, wild type.

Table S36Data for Figure 6D: GO analysis of DEGs in livers from different mice models[provided as Excel Table S36].

Mice model	WT		Hep	Нер-КО		Mac-KO	
NEG	LC	AR	LC	AR	LC	AR	
Glycerophospholipids[GP]	6635	1.89	5947	2.03	4579	1.39	
Glycerolipids[GL]	529	7.01	514	4.94	301	2.72	
Fatty Acyls[FA]	1794	2.89	2127	1.95	3089	3.48	
Sphingolipids[SP]	324	1.44	291	1.54	225	1.64	
Sterol Lipids[ST]	124	3.37	205	1.89	209	2.40	
Polyketides[PK]	373	1.08	218	1.43	307	1.60	
Prenol Lipids[PR]	162	3.28	156	6.81	120	4.32	
Saccharolipids[SL]	3	3.03	2	1.69	3	2.73	
POS							
Glycerophospholipids[GP]	7389	1.91	6040	1.39	6102	1.91	
Glycerolipids[GL]	3708	0.84	2917	1.03	4750	0.82	
Fatty Acyls[FA]	1656	1.07	1146	1.41	800	1.03	
Sphingolipids[SP]	593	1.74	686	2.80	313	1.23	
Sterol Lipids[ST]	399	1.90	311	1.74	215	1.61	
Polyketides[PK]	249	1.32	233	1.36	177	1.01	
Prenol Lipids[PR]	180	1.38	382	0.99	164	0.88	
Saccharolipids[SL]	9	1.82	7	1.60	9	1.10	

Table S37Superclass constitution of lipid metabolites in liver from different mice models [lipidcounts; average ratio (DEHP/CT ratio)].

Note: Correspond to Figure 6E. AR, average ratio (DEHP/CT ratio); DEHP, di(2-Ethylhexyl) phthalate; Hep-KO, hepatocyte-specific PPARγ knockout; LC, lipid counts; Mac-KO, macrophage-specific PPARγ knockout; NEG, negative ion mode; POS, postive ion mode.

**Table S38**Data for Figure 6F: Class constitution of lipid metabolites in livers from differentmice models [lipid counts; average ratio (DEHP/CT ratio); provided as Excel Table S38].

ii S/tiedtilientj.				
Determination	TMRE	<b>BODIPY FL C12</b>	LD	
CT(DMSO)	$20.96 \pm 5.60$	25.51±2.65	14.53±3.85	
MEHP	59.46±6.39***	49.27±6.79***	33.93±5.52***	
MEHP+IACS	$11.90 \pm 2.42^3$	$20.53 \pm 4.90^3$	$20.89 \pm 2.51^2$	
MEHP+VLX	$31.93 \pm 9.49^3$	$28.90 \pm 3.68^3$	$22.05 \pm 4.35^{1}$	
MEHP+3-NP	$33.53 \pm 8.19^3$	$25.03 \pm 3.73^3$	$15.63 \pm 7.11^3$	
MEHP+IL-1RA	39.07±5.34**,3	$26.12\pm6.19^{3}$	$18.45 \pm 7.00^{2}$	

Table S39Evaluation of mitochondrial OXPHOS, fatty acid uptake and LD in HepG2 cellstreated with MEHP with/without specific inhibitors [Mean stain area ( $\mu$ m<sup>2</sup>); mean±SD;n=5/treatment].

Note: Correspond to Figure 7A, 7B. 3-NP, 3-Nitropropanoic acid (2 mM); CT, vehicle control; DMSO, dimethyl sulfoxide; IACS, IACS-10759 (10 nM); IL-1RA (200 ng/ml); LD, lipid droplet; MEHP, mono-2-ethylhexyl phthalate (200  $\mu$ M); OXPHOS, oxidative phosphorylation; TMRE, tetramethylrhodamine ethyl ester, mitochondrial membrane potential indicator; SD, standard deviation; VLX, VLX600 (1  $\mu$ M). \**p*<0.05, \*\**p*<0.01, \*\*\**p*<0.001, relative to vehicle control; <sup>1</sup>*p*<0.05, <sup>2</sup>*p*<0.01, <sup>3</sup>*p*<0.001, relative to MEHP treatment [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

	1 0 1	Q //	/	
With/without M2	Treatment	TMRE	MitoSOX	LD
HepG2	CT(DMSO)	$145.98 \pm 3.62$	51.79±3.00	60.98±4.32
(without M2)	MEHP	163.29±3.05**	56.49±1.79***	117.28±17.67***
Co-M2	CT(DMSO)	$129.66 \pm 9.46$	37.76±2.30	$59.38 {\pm} 7.07$
(HepG2 with M2)	MEHP	140.69±11.12 <sup>3</sup>	$37.27 \pm 4.53^3$	$88.42 \pm 4.45^3$

Table S40Evaluation of mitochondrial OXPHOS and LD in HepG2 cells treated with MEHPwith/without M2 macrophages [Mean stain area ( $\mu$ m2); mean  $\pm$  SD; n=5/treatment].

Note: Correspond to Figure 7C. CT, vehicle control; DMSO, dimethyl sulfoxide; LD, lipid droplet; MEHP, mono-2-ethylhexyl phthalate (200  $\mu$ M); mitoSOX, mitochondrial superoxide, mitochondrial reactive oxygen species indicator; OXPHOS, oxidative phosphorylation; TMRE, tetramethylrhodamine ethyl ester, mitochondrial membrane potential indicator; SD, standard deviation. \**p*<0.05, \*\**p*<0.01, \*\*\**p*<0.001, relative to vehicle control; <sup>1</sup>*p*<0.05, <sup>2</sup>*p*<0.01, <sup>3</sup>*p*<0.001, relative to vehicle control; <sup>1</sup>*p*<0.05, <sup>2</sup>*p*<0.01, <sup>3</sup>*p*<0.001, relative to MEHP treatment [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

DEGs	HepG2		MEHP	-HepG2	MEHP-HepG2-Co-M2	
	R-1	R-2	R-1	R-2	R-1	R-2
MT-ATP6	-0.34	-0.06	1.68	0.53	-0.63	-1.17
MT-ND2	-0.13	-0.03	1.43	0.84	-1.16	-0.95
MT-CYB	0.09	0.15	1.27	0.78	-0.94	-1.36
MT-ND4L	-0.20	0.88	1.05	0.52	-0.73	-1.51
ATP6V0E2	0.44	0.34	1.03	0.57	-0.72	-1.67
MT-ND1	-0.80	-0.73	1.69	0.72	-0.30	-0.59
MT-CO1	-1.00	-0.77	1.65	0.44	0.30	-0.62
MT-ND5	-0.68	-1.05	1.38	0.64	0.59	-0.88
MT-ND6	-1.02	-1.23	1.36	0.42	0.63	-0.15
MT-ATP8	0.03	-0.70	0.30	0.55	1.34	-1.52
ATP6V0A2	0.34	-0.01	-0.58	1.74	-0.28	-1.20

**Table S41**Relative expression of DEGs associated with OXPHOS in HepG2 cells treated withMEHP with/without M2 macrophages [Z scores; n=2/treatment].

Note: Correspond to Figure 7D. DEGs, differentially expressed genes; MEHP, mono-2-ethylhexyl phthalate (200  $\mu$ M); R, replicate.

DEGs	MEHP-HepG2			MEHP-HepG2-Co-M2			
	R-1	R-2	R-3		R-1	R-2	R-3
ALDOC	-0.97	-0.84	-0.90		1.09	0.92	0.71
LDHA	-1.01	-0.71	-1.01		0.94	0.96	0.81
HK1	-1.40	-0.53	-0.67		0.91	0.72	0.98
ENO2	-1.02	-0.49	-1.08		0.76	1.29	0.55
ENO1	-0.93	-0.66	-1.03		0.98	1.23	0.42
РКМ	-0.87	-0.94	-0.87		1.08	1.08	0.52
PGM1	-0.62	-1.12	-0.84		1.31	0.40	0.87
HK2	-0.39	-1.12	-0.84		1.30	-0.05	1.09
PGK1	-0.59	-0.99	-0.69		1.37	-0.22	1.12
TPI1	-0.84	-0.68	-1.11		1.12	1.08	0.43
GAPDH	-1.01	-0.38	-1.01		0.63	1.54	0.22
PFKL	-1.09	-0.38	-0.88		0.69	1.55	0.11
PFKP	-0.56	-1.16	-0.61		1.62	0.55	0.16
PFKFB4	-0.92	-0.80	-1.01		0.95	0.90	0.88
PFKFB3	-0.70	-0.95	-1.04		1.05	0.61	1.03
MPI	-1.14	-0.64	-0.71		0.69	1.49	0.32
ENOSF1	-0.98	-0.35	-1.09		0.44	1.52	0.47
PFKFB1	-1.00	-0.51	-1.00		0.19	1.27	1.04

**Table S42**Relative expression of DEGs associated with glycolysis in HepG2 cells treated withMEHP with/without M2 macrophages [Z scores; n=3/treatment].

Note: Correspond to Figure 7E. DEGs, differentially expressed genes; MEHP, mono-2-ethylhexyl phthalate (200  $\mu$ M); R, replicate.

Detherous	DECa	MEHP-HepG2			MEHP-HepG2-Co-M2		
rauiways	DEGS	R-1	R-2	R-3	R-1	R-2	R-3
Glycerolipid metabolism	ALDH1B1	1.16	0.43	0.88	-0.23	-1.38	-0.86
	GPD1L	0.83	0.17	0.98	-0.22	-1.81	0.04
	PCYT1A	0.97	0.33	0.98	-0.29	-1.68	-0.30
	GPD1	0.94	0.79	0.86	-1.27	-0.31	-1.02
	ETNK2	0.21	0.66	1.50	-0.94	-1.13	-0.31
	PCYT1B	0.68	0.42	1.41	-0.51	-0.73	-1.26
	ETNPPL	0.83	0.90	0.75	-0.56	-0.33	-1.58
	PLA2G12B	1.02	0.43	0.89	-0.15	-1.63	-0.56
	PLA2G12A	1.22	0.09	0.92	-1.33	-0.93	0.03
	PLPP3	1.06	0.30	0.78	-0.22	-1.76	-0.17
	PLPP1	0.71	0.98	0.98	-0.70	-1.24	-0.74
	GPAM	1.03	0.35	0.87	-0.46	-1.68	-0.11
	AGPAT3	1.27	0.69	0.67	-0.60	-1.07	-0.97
	DGKK	0.96	0.12	0.87	-0.28	-1.80	0.13
Fatty acid metabolism	ADH6	0.92	0.99	0.48	-0.40	-1.63	-0.36
	ADH4	0.72	0.85	1.05	-0.38	-1.26	-0.97
	EHHADH	0.89	0.28	0.64	-0.08	-1.92	0.18
	ACSL1	1.04	0.15	0.77	-0.08	-1.82	-0.06
	HACD2	0.87	0.13	0.85	-0.28	-1.83	0.26
	FADS1	0.82	-0.19	0.99	0.14	-1.81	0.05
	ICMT	0.88	0.78	1.00	-0.51	-1.17	-0.99
	IDI1	0.90	0.37	0.67	-0.27	-1.87	0.20
	HMGCR	0.93	0.15	0.72	0.04	-1.90	0.06
	ACAT2	0.86	0.74	1.09	-0.63	-0.96	-1.10
	HMGCS1	1.02	0.10	0.73	-0.03	-1.85	0.03
	MCAT	0.75	0.92	0.89	-1.34	-0.27	-0.95
	FASN	1.11	0.57	0.99	-1.09	-0.73	-0.85

**Table S43**Relative expression of DEGs associated with lipid metabolism in HepG2 cellstreated with MEHP with/without M2 macrophages [Z scores; n=3/treatment].

Note: Correspond to Figure 7F. DEGs, differentially expressed genes; MEHP, mono-2-ethylhexyl phthalate (200  $\mu$ M); R, replicate.

	5 8		18)	
Dose group	0 mg/kg BW	625 mg/kg BW	1250 mg/kg BW	2500 mg/kg BW
Week 0	22.4±1.2	23.1±0.8	22.8±1.2	22.9±0.8
Week 1	22.4±1.1	23.4±1.2	22.9±0.8	23.1±1.1
Week 2	23.1±1.3	22.7±1.4	23.5±1.2	22.8±1.2
Week 3	22.9±1.2	23.4±1.2	22.8±1.1	22.5±0.9
Week 4	22.9±1.1	24.0±0.0	23.0±1.0	22.0±0.9

**Table S44**Body weight fluctuation in DEHP treated mice [g; mean  $\pm$  SD; n=10/group].

Note: Correspond to Figure S2A. BW, body weight; DEHP, di(2-Ethylhexyl) phthalate; SD, standard deviation. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control [analysis of variance (one-way ANOVA) with post hoc multiple comparison correction using Dunnett's multiple comparisons test].

**Table S45**Data for Figure S2B: Enriched KEGG pathways (level 2) in DEHP treated mice[number of genes; provided as Excel Table S45].

Dethwaya	DECa	HepG2-CT			HepG2-MEHP		
r auiways	DEGS	R-1	R-2	R-3	<b>R-1</b>	<b>R-2</b>	R-3
Glycerolipid metabolism	PLA2G3	-1.03	-0.18	-1.25	0.51	0.68	1.27
	AGPAT2	-0.98	-0.51	-0.83	-0.15	1.38	1.08
	LPCAT3	-1.07	-0.31	-0.85	1.14	-0.21	1.30
	AKR1B10	-0.71	-0.79	-1.16	1.13	0.57	0.97
	PLA2G12A	-1.13	0.52	-0.51	1.35	-0.94	0.71
	PLPP3	-0.82	0.54	0.15	1.34	-1.46	0.25
	PLPP5	-0.69	-1.26	-0.36	-0.02	1.39	0.93
	LPIN1	-0.59	-1.50	-0.05	1.16	-0.05	1.02
Fatty acid metabolism	FADS1	-0.85	-0.96	-0.78	1.08	0.29	1.22
	TECR	-0.44	-0.62	-0.76	-0.58	1.75	0.66
	FASN	-1.16	-0.49	-0.94	1.16	0.43	1.00
	ACADVL	-0.81	-0.88	-0.86	0.53	1.50	0.53
	ACAT2	-0.81	-1.06	-0.84	0.87	0.75	1.09
	ACAA2	-1.11	-1.21	-0.13	1.10	0.96	0.38
	FDPS	-0.75	-0.88	-1.04	1.20	0.90	0.57
	MVD	-0.97	-0.83	-0.86	0.69	1.32	0.65
	IDI1	-0.72	-0.99	-0.87	1.34	0.34	0.91
	HMGCR	-0.42	-0.65	-0.17	1.57	-1.13	0.79
	MVK	-0.80	-0.79	-1.06	0.61	1.31	0.73
Steroid biosynthesis	EBP	-0.52	-0.96	-0.70	0.03	1.78	0.38
	DHCR7	-0.89	-0.81	-1.03	0.89	0.90	0.94
	SQLE	-0.78	-0.89	-0.83	1.33	0.10	1.07
	MSMO1	-0.71	-0.98	-0.91	1.40	0.68	0.51
	NSDHL	-0.32	-1.01	-1.02	0.73	1.51	0.10
	TM7SF2	-1.09	-0.64	-0.62	0.39	1.67	0.28
	SC5D	-0.77	-1.26	-0.19	1.41	-0.09	0.89
	HSD17B7	-0.29	-1.05	-1.20	0.47	1.10	0.97
	AKR1C2	-0.89	-0.84	-0.77	0.20	1.48	0.83
	AKR1C1	-1.02	-1.09	-0.43	0.31	1.23	1.00
	CYP1A1	-0.89	-0.85	-0.91	0.56	0.75	1.33

**Table S46**Relative expression of DEGs associated with lipid metabolism in HepG2 cellstreated with MEHP [Z scores; n=3/treatment].

Note: Correspond to Figure S2C. DEGs, differentially expressed genes; MEHP, mono-2-ethylhexyl phthalate (200  $\mu$ M); R, replicate.

**Table S47**Relative expression of DEGs associated with PPAR pathway in DEHP treated mice[Z scores; n=6/group; provided as <a href="mailto:Excel Table S47"><u>Excel Table S47</u></a>].

Pathway	Counts	Gene Ratio	Padj
Fatty acid metabolism	12	12/155	2.30E-07
PPAR signaling pathway	11	11/155	5.25E-05
Ferroptosis	8	8/155	0.00029333
Biosynthesis of unsaturated fatty acids	6	6/155	0.000666895
Terpenoid backbone biosynthesis	6	6/155	0.000746958
Fatty acid degradation	7	7/155	0.002394125
Peroxisome	9	9/155	0.006369618

**Table S48**Pathway enrichment analysis of DEGs in HepG2 cells treated with MEHP[n=3/treatment].

Note: Correspond to Figure S2E. DEGs, differentially expressed genes; MEHP, mono-2-ethylhexyl phthalate (200 µM).

Measurements	Treatment	СМС	DEHP
TG(mM)	WT	0.68±0.07 (5)	0.38±0.10 (5)**
	Mac-KO	0.60±0.11 (5)	$0.66 \pm 0.05 \ (5)^{a}$
CHO(mM)	WT	1.14±0.07 (5)	2.08±0.30 (5)***
	Mac-KO	1.37±0.07 (3)	1.77±0.14 (4) <sup>a</sup>
CHE(mM)	WT	2.97±0.38 (5)	4.07±0.30(5)***
	Mac-KO	3.34±0.19 (3)	3.82±0.15 (5)
ALP(U/L)	WT	102.83±7.57 (5)	150.86±23.18 (5)***
	Mac-KO	113.00±2.00 (3)	118.33±16.99 (5) <sup>a</sup>
HDLC(mM)	WT	0.71±0.07 (5)	1.23±0.19 (5)***
	Mac-KO	0.88±0.05 (3)	1.12±0.09 (5)
LDLC(mM)	WT	1.47±0.03 (5)	1.76±0.24 (5)**
	Mac-KO	1.53±0.02 (3)	1.66±0.10 (5)

Table S49Clinical chemistry panel in WT and Mac-KO mice  $[mean \pm SD (n)].$ 

Note: Correspond to Figure S4A. ALP, alkaline phosphatase; CHE, cholinesterase; CHO, cholesterol; CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; HDLC, high-density lipoprotein cholesterol; LDLC, low-density lipoprotein cholesterol; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; TG, triglycerides; U/L, units per liter; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control;  $^{a}p$ <0.05,  $^{b}p$ <0.01,  $^{c}p$ <0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

	-		
	Treatment	СМС	DEHP
CD11b <sup>high</sup> F4/80 <sup>low</sup>	WT	8.89±1.00 (5)	12.80±0.94 (5)***
Macrophages	Mac-KO	4.83±0.43 (5)	9.47±1.35 (5)***,b
CD11blowF4/80high	WT	24.94±3.93 (5)	21.74±3.50 (5)*
Macrophages	Mac-KO	27.30±2.48 (5)	31.22±2.78 (5) <sup>b</sup>
CD11bhighF4/80-Ly6g+	WT	4.89±1.89 (5)	4.19±0.25 (5)
Neutrophils	Mac-KO	3.66±1.01 (4)	4.10±2.03 (5)

Table S50Macrophages and neutrophils analysis in liver of WT and Mac-KO mice using flowcytometry [% of CD45+ cells; mean  $\pm$  SD (n)].

Note: Correspond to Figure S4B. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control; ap<0.05, bp<0.01, cp<0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

Subtype	Cytokine	Treatment	СМС	DEHP
M1 macrophages	H 12-70	WT	8.61±2.90	23.73±4.30***
	IL-12p/0	Mac-KO	11.45±3.22	$12.43 \pm 2.08^{b}$
	IL-18	WT	39.53±10.10	63.02±7.30***
		Mac-KO	54.88±7.96	$36.13 \pm 9.30^{b}$
	IL-10	WT	77.52±7.39	52.44±6.31***
		Mac-KO	67.11±4.54	62.22±5.66
M2 macrophages	ц ээ	WT	33.07±5.48	24.05±4.04**
	1L-23	Mac-KO	30.91±1.15	$32.41 \pm 3.52^{\circ}$

Table S51Macrophages featured cytokines analysis in liver of WT and Mac-KO mice using<br/>flow cytometry [pg/mg protein; mean  $\pm$  SD; n=5].

Note: Correspond to Figure S4C. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control; ap<0.05, bp<0.01, cp<0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

Table S52 $CD69^+$  M1 macrophages evaluation in BMDM from WT and Mac-KO mice [mean<br/>stain area ( $\mu$ m2); mean $\pm$ SD (n)].

Treatment	CT(DMSO)	DEHP	МЕНР
WT	131.62±10.59 (11)	126.57±13.17 (14)	121.86±12.28 (11)
Mac-KO	129.17±17.18 (12)	117.85±13.47 (12)	133.54±15.80 (9)

Note: Correspond to Figure S5B. n, technical replicates. CT, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; DMSO, dimethyl sulfoxide; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; MEHP, mono-2-ethylhexyl phthalate; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control;  $^ap$ <0.05,  $^bp$ <0.01,  $^cp$ <0.001, relative to WT DEHP or WT MEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Bonferroni's multiple comparisons test].

Table S53Relative expression of DEGs associated with M1 macrophages in MEHP treatedmacrophages [Z scores; n=3/treatment; provided as Excel Table S53].

	Treatment	СМС	DEHP	
CD36 <sup>high</sup>	WT	6.77±2.86	8.67±2.50	
Macrophages	Mac-KO	7.55±1.24	14.30±2.80 <sup>a</sup>	
LD <sup>high</sup>	WT	13.26±2.23	12.09±2.06	
Macrophages	Mac-KO	13.90±0.87	19.24±2.66 <sup>c</sup>	

Table S54Macrophages lipid metabolism analysis in liver of WT and Mac-KO mice using flowcytometry [% of CD45+ cells; mean  $\pm$  SD; n=5/treatment].

Note: Correspond to Figure S5D. CMC, sodium carboxymethylcellulose, vehicle control; DEHP, di(2-Ethylhexyl) phthalate; Mac-KO, macrophage-specific PPAR $\gamma$  knockout; SD, standard deviation; WT, wild type. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, relative to vehicle control;  $a_p$ <0.05,  $b_p$ <0.01,  $c_p$ <0.001, relative to WT DEHP [analysis of variance (two-way ANOVA) with post hoc multiple comparison correction using Tukey's multiple comparisons test].

# **Supplemental Figures**



Figure S1. Gating strategy for flow cytometry of mouse liver.



Figure S2. The effect of DEHP on hepatic lipid metabolism.

(A) Body weight of mice in the dose-response study of DEHP. C57BL/6J male mice were treated with 0.5% (wt/vol) sodium carboxymethylcellulose (CMC, vehicle control) or different doses of DEHP (625, 1250, and 2500 mg/kg bw) by daily gavage for 28 days. Data were expressed as mean  $\pm$  SD (n=10/group).The data for Figure S2A are located in Table S44.

(B) Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment (level
2) of differentially expressed genes (DEGs) from livers of mice treated with CMC (vehicle control, CT) or DEHP (625 mg/kg bw) for 28 days (n=6/group). The data for

Figure S2B are located in Table S45.

(C) Heatmap of gene expression associated with PPAR signaling pathway in HepG2 cells. The cells were treated with MEHP (200  $\mu$ M) for 24 h in triplicate wells (n=3/treatment). After treatment, cells were collected for RNA sequencing. The data for Figure S2C are located in Table S46.

(D) Heatmap of gene expression associated with PPAR signaling pathway in mice model. C57BL/6J male mice treated with CMC (vehicle control, CT) or DEHP (625 mg/kg bw) for 28 days (n=6/group).The data for Figure S2D are located in Table S47. (E) KEGG pathway enrichment for the HepG2 model. The cells were treated with MEHP (200  $\mu$ M) for 24 h in triplicate wells (n=3/treatment). After treatment, cells were collected for RNA sequencing. The data for Figure S2E are located in Table S48.



С

Comparison of the binding free energy of different complexes

Complex	ΔEvdw (kcal/mol)	ΔEele energy (kcal/mol)	ΔEpb (kcal/mol)	ΔEcavity (kcal/mol)	∆Ggas (kcal/mol)	∆Gsol (kcal/mol)	∆Gbinding (kcal/mol)
PPARα	-0.69	-9.29	2.50	-0.12	-9.98	2.38	-7.60
PPARγ	-1.07	-9.33	2.24	-0.17	-10.40	2.07	-8.33

VDWAALS = van der Waals contribution from molecular minimization (MM).

EEL = electrostatic energy as calculated by the MM force field.

EPB = the electrostatic contribution to the solvation free energy calculated by PB.

ECAVITY = nonpolar contribution to the solvation free energy calculated by an empirical model. DELTA G binding = final estimated binding free energy calculated from the terms above. (kCal/mol)

 $\Delta Ggas = \Delta Evdw + \Delta Eele, \quad \Delta Gsol = \Delta Epb + \Delta Ecavity, \quad \Delta Gbinding = \Delta Ggas + \Delta Gsol$ 

#### Figure S3. Activation of PPAR isoforms by DEHP and MEHP at the molecular

#### level.

(A) Sensorgram showing the binding response of PPAR $\delta$  with DEHP or MEHP as

determined by surface plasmon resonance (SPR) analysis.

(B) Root mean square deviation (RMSD) plots showing the stability of optimal docking

complex.

(C) The binding free energy of MEHP with PPAR $\alpha$  and PPAR $\gamma$  in molecular docking

and simulation.



Figure S4. The function of PPARy in hepatic macrophages.

Wild-type (WT, n=5/group) and macrophage-specific PPAR $\gamma$  knockout (Mac-KO, n=5/group) C57BL/6J male mice were treated with 0.5% (wt/vol) sodium carboxymethylcellulose (CMC, vehicle control) or DEHP (625 mg/kg bw) by daily gavage for 28 days.

(A) Plasma level of triglyceride (TG), cholesterol (CHO), high-density lipoprotein cholesterol (HDLC), low-density lipoprotein cholesterol (LDLC), alkaline phosphatase

(ALP), and cholinesterase (CHE) of WT and Mac-KO mice at the end of the treatment. The data for Figure S4A are located in Table S49.

(B) Representative flow cytometry gating graphs and quantification of macrophages and neutrophils in the liver of WT or Mac-KO mice at the end of the treatment. An unusual observation (outlier) was excluded in the Mac-KO-CMC group. The data for Figure S4B are located in Table S50.

(C) Quantification of cytokines IL-10 and IL-23, IL-12p70 and IL-18 in the liver of WT or Mac-KO mice at the end of the treatment (n=5/group). The data for Figure S4C are located in Table S51.

Data are represented as mean  $\pm$  SD; \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001, n.s, no significance, relative to vehicle control or WT DEHP; Two-way ANOVA with Turkey's post-test.



Figure S5. The effects of DEHP and MEHP on Macrophages polarization.

(A) Experimental scheme of polarization model with the bone marrow derived macrophages (BMDMs).

(B) Quantification of CD69 (M1 marker) expression in BMDMs from WT or Mac-KO mice. BMDMs were isolated from the Wild-type (WT, n=5/group) and macrophage-specific PPAR $\gamma$  knockout (Mac-KO, n=5/group) C57BL/6J male mice. BMDMs were

treated with 0.1% DMSO control (CT), DEHP (200  $\mu$ M) or MEHP (200  $\mu$ M) in combination with lipopolysaccharide (LPS, 100 ng/ml) and recombinant murine IFN- $\gamma$ (50 ng/ml) for 24 h in in triplicate wells (the plots contains all technical replicates from 3-5 imaging sites of each well). Cells with positive stain of CD69 were acquired with ImageXpress system and normalized to the total cells in each of the replicate wells. The data for Figure S5B are located in Table S52.

(C) Heatmap of M1 marker genes expression in THP-1 derived macrophages. THP-1 derived M0 macrophages were treated with MEHP (200  $\mu$ M) during the induction of M1 and M2 polarization. Cells from triplicate wells (n=3) were analyzed with RNA sequencing at the end of the treatment. The data for Figure S5C are located in Table S53.

(D) Representative flow cytometry gating graphs and quantification of CD36 expression and lipid droplets (LD) level in hepatic macrophages of WT and Mac-KO mice model. Wild-type (WT, n=5/group) and macrophage-specific PPAR $\gamma$  knockout (Mac-KO, n=5/group) C57BL/6J male mice were treated with 0.5% (wt/vol) sodium carboxymethylcellulose (CMC, vehicle control) or DEHP (625 mg/kg bw) by daily gavage for 28 days. The data for Figure S5D are located in Table S54.

Data are represented as mean  $\pm$  SD; \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001, n.s, no significance, relative to vehicle control or WT DEHP; Two-way ANOVA with Bonferroni's post-test.