

Supplementary Materials for

Secondary loss of *miR-3607* reduced cortical progenitor amplification during rodent evolution

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Other Supplementary Material for this manuscript includes the following:

Tables S1 to S3
Movies S1 and S2

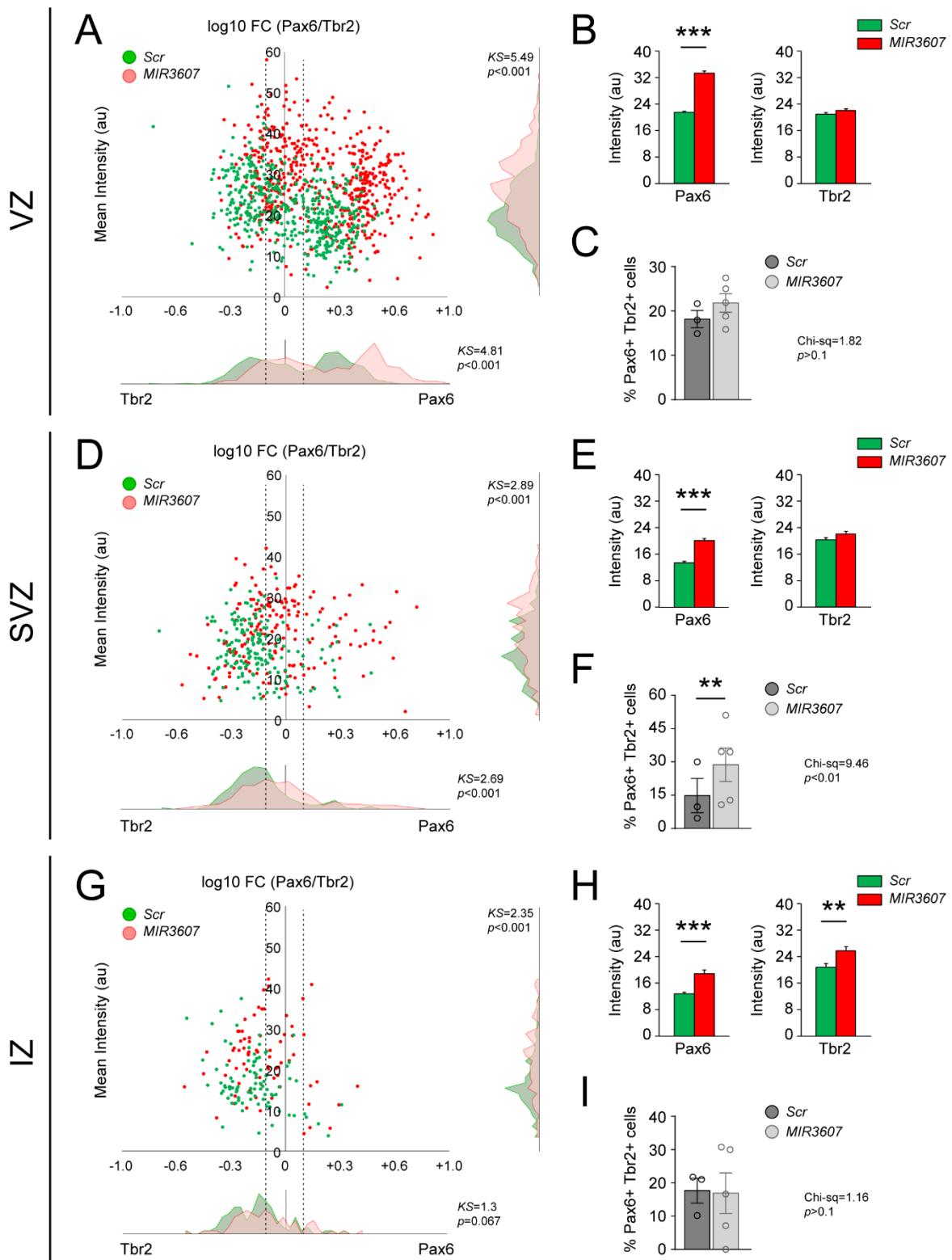


Fig. S1. *MIR3607* drives overexpression of Pax6 in cortical cells

(A,D,G) Scatter plots of ratio Pax6/Tbr2 expression level (log10 Fold Change) relative to mean intensity of both markers (arbitrary units) in individual GFP+ cells, and frequency distribution plots for each of the individual parameters, at the indicated layers of the developing cortex. Dashed vertical lines delimit Pax6/Tbr2 co-expression (-0.1<log10FC<+0.1).

(B,E,H) Average expression intensity of Pax6 and Tbr2 in individual cells at the indicated layers.

(C,F,I) Proportion of cells co-expressing Pax6 and Tbr2 (-0.1<log10FC<+0.1). Differences in Pax6 expression intensity were largest in VZ, but differences in Pax6/Tbr2 co-expression were largest in SVZ.

N = 471 cells VZ, 176 cells SVZ, 91 cells IZ, 3 embryos for *Scr*; 538 cells VZ, 176 cells SVZ, 73 cells IZ, 5 embryos for *MIR3607*. Kolmogorov-Smirnov test (A,D,G), t-test (B,E,H), X² test (C,F,I); ***p*<0.01, ****p*<0.001.

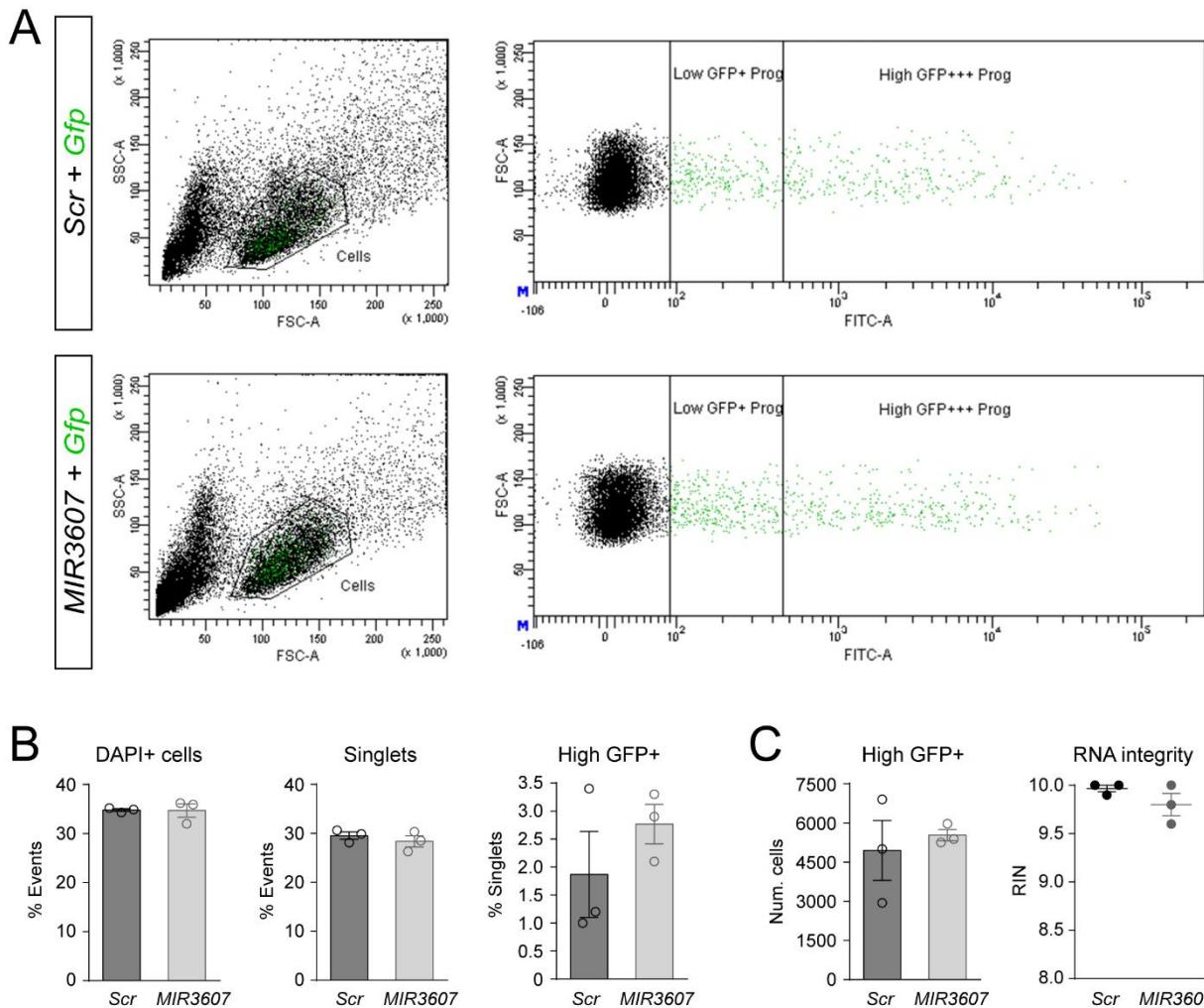


Fig. S2. FACS sorting of cortical cells for transcriptomic profiling

(A) Examples of sort profiles of cells from E15.5 cortex electroporated at E14.5 with the indicated plasmids. Left column shows the dot plots of cells in forward scatter (FSC) and side scatter (SSC) with the polygon indicating the gate selecting the healthy cells. Right column shows the dot plots of cells in forward scatter (FSC) with fluorescence intensity (FITC) indicated. Only High GFP+++ cells were selected for RNA-seq analysis.

(B) Quality control of FACS sorting, showing similar values between *Scr*- and *MIR3607*-electroporated embryos for frequency of DAPI+ elements, of singlets among total events, and frequency of high GFP+ cells among singlets.

(C) Quality control of RNA extraction experiments, showing a similar number of high GFP+ single cells used for RNA extraction, and similar integrity of extracted RNA between experimental groups, above 9.5 in all cases.

Histograms indicate mean \pm SEM, and circles within indicate values for individual embryos; n = 3 embryos per group; χ^2 tests (B) and t-tests (C) revealed non-significance ($p>0.05$).

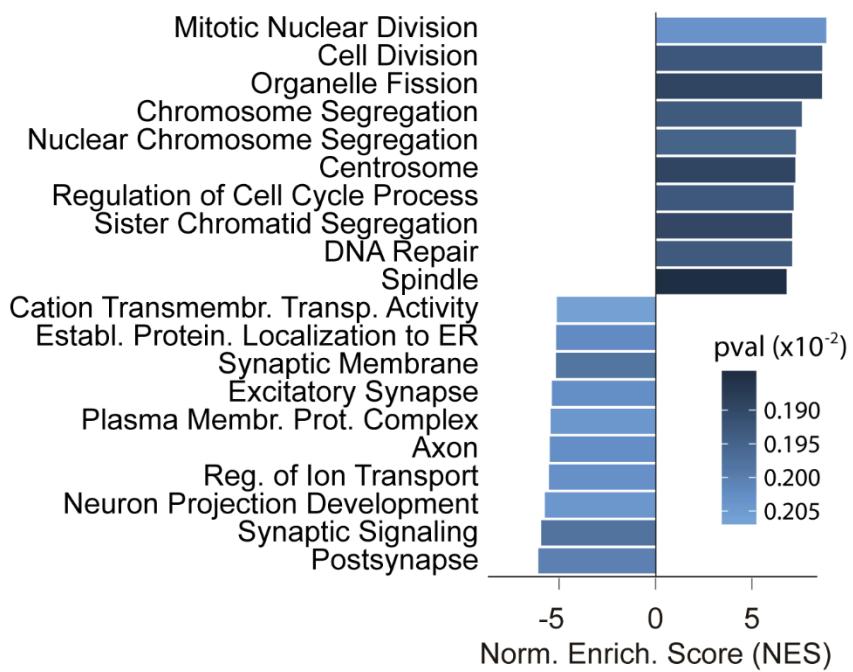


Fig. S3. Gene Ontology analysis of DEGs from GSEA

Bar chart of top ranked GO gene sets according to normalized enrichment score (NES), from gene set enrichment analysis (GSEA). Bars are color-coded according to statistical significance (nominal p value) as indicated.

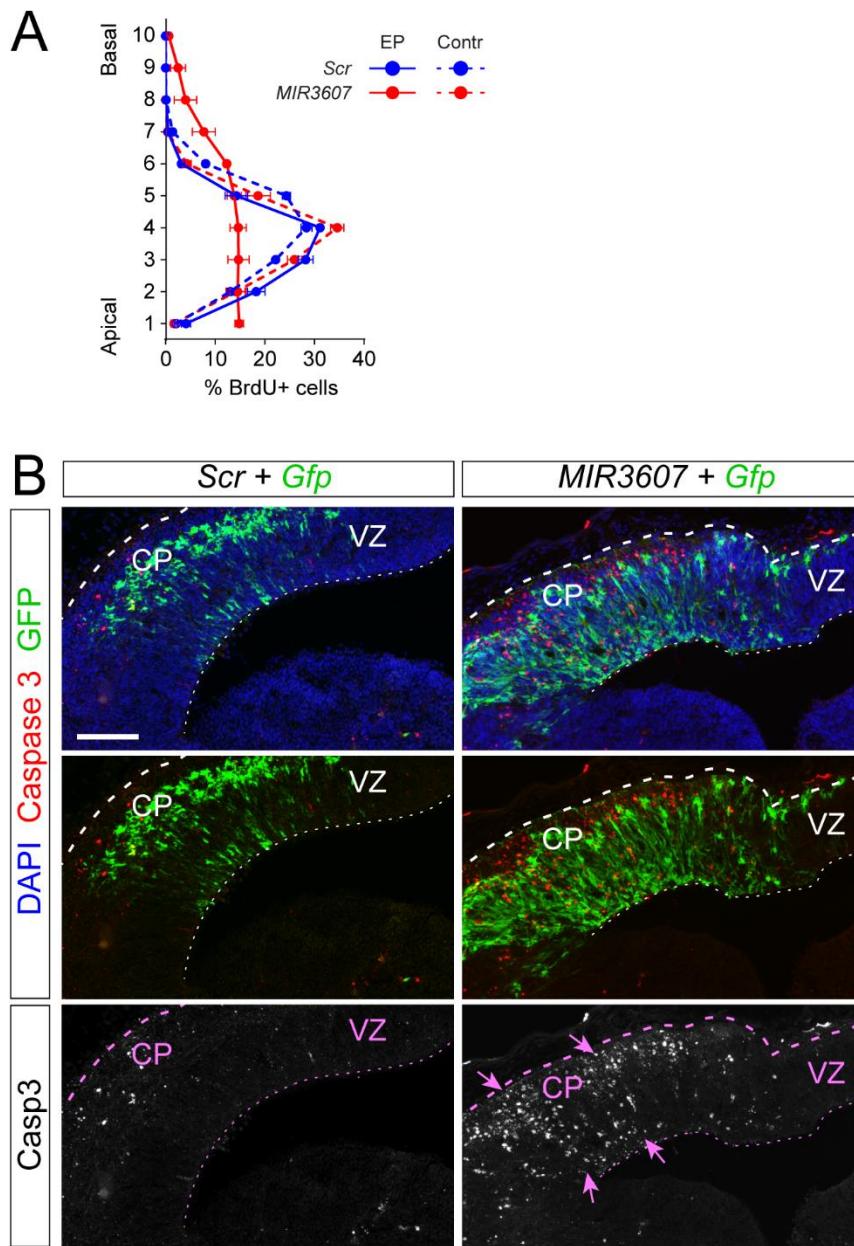


Fig. S4. *MIR3607* overexpression induces progenitor cell disorganization and apoptosis in the early embryonic mouse cortex

(A) Quantification of binned distribution of BrdU+ cells across the thickness of the parietal cortex of mouse embryos electroporated at E12.5 with the indicated plasmids, and analyzed at E13.5. Data is from electroporated hemispheres (solid lines) and from non-electroporated, contralateral hemispheres (dashed lines). The typical accumulation of BrdU-incorporating cells in the basal side of the VZ (bins 3-5) was observed in *Scr*-electroporated (solid blue line) and contralateral hemispheres (dashed lines), but severely disturbed in *MIR3607*-electroporated cortices (solid red line). Plots show mean \pm SEM; n = 2,673 cells ipsi, 3,583 cells contra, 2 embryos, *Scr*; 5,150 cells ipsi, 5,485 cells contra, 4 embryos, *MIR3607*.

(B) Sections through the parietal cortex of mouse embryos electroporated at E12.5 with the indicated plasmids, analyzed at E13.5 and stained as indicated. Expression of *MIR3607* caused a dramatic increase in apoptosis (Casp3+ cells), both in VZ and CP (arrows). Dotted lines indicate apical surface, dashed lines indicate pial surface.

Scale bar: 100 μ m.

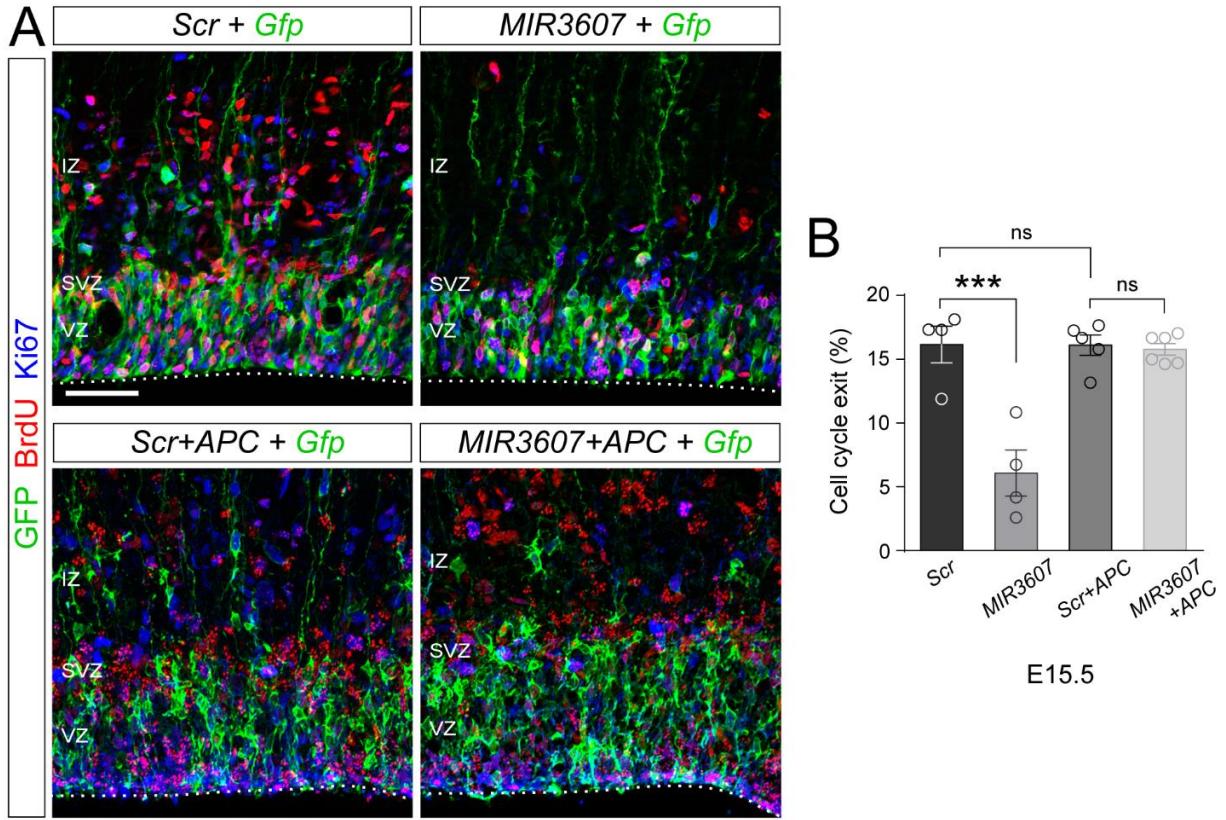


Fig. S5. Expression of APC rescues the changes in cell cycle exit caused by *MIR3607*

(A,B) Sections through the parietal cortex of E15.5 mouse embryos injected with a single pulse of BrdU at E14.5 followed 4hr later by electroporation with the indicated plasmids, and stained as indicated (A), and quantifications of cell cycle exit of GFP+ cells in any layer (B). Histograms indicate mean \pm SEM, and circles within indicate values for individual embryos; $n = 4-6$ embryos per group. X^2 test; *** $p < 0.001$; ns, not significant.

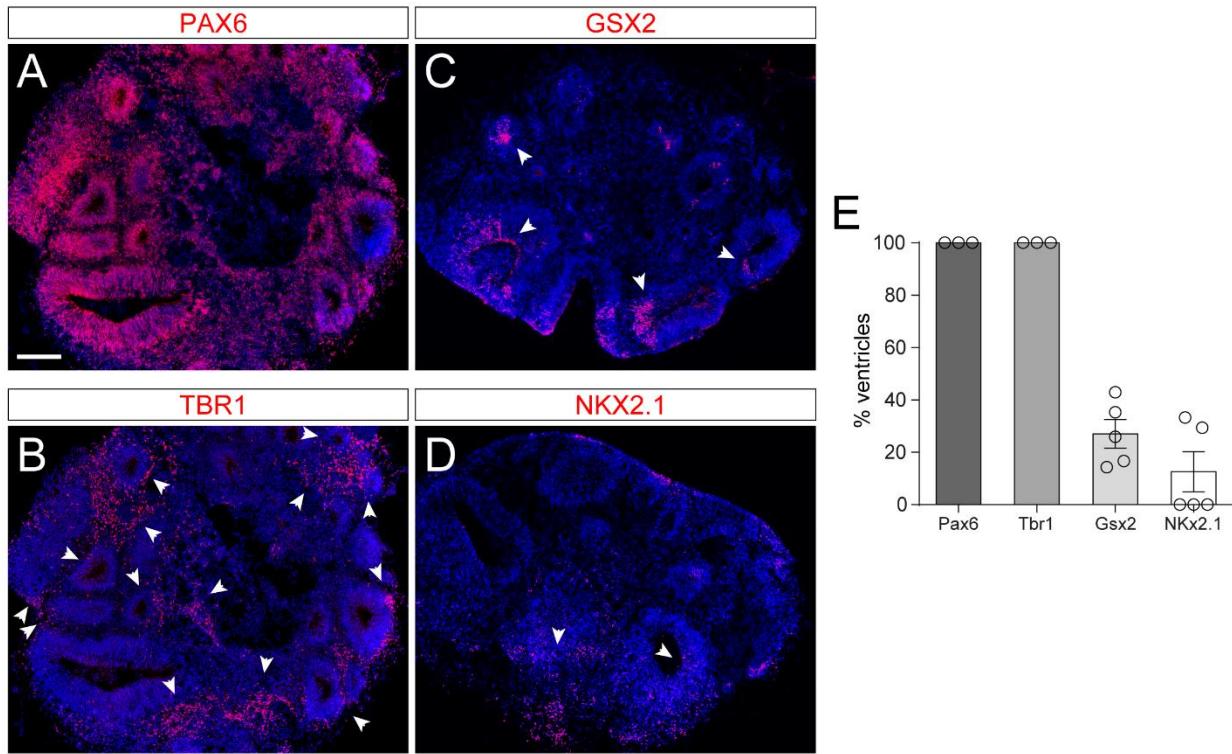


Fig. S6. Validation of cortical identity of human cerebral organoids

(A-D) Sections through human cerebral organoids immunostained for the indicated cortical (A,B) and subcortical (C,D) markers. Arrowheads indicate ventricles containing cells positive for the relevant marker. All ventricles are positive for the cortical markers PAX6 and TBR1, whereas only a minority have a few cells positive for the subcortical markers GSX2 and NKX2.1.

(E) Proportion of ventricles positive for the indicated markers within individual organoids. Histograms show mean \pm SEM; circles indicate values for individual organoids; $n = 3-5$ organoids per group, 5-21 ventricles per organoid.

Scale bar: 150 μ m.

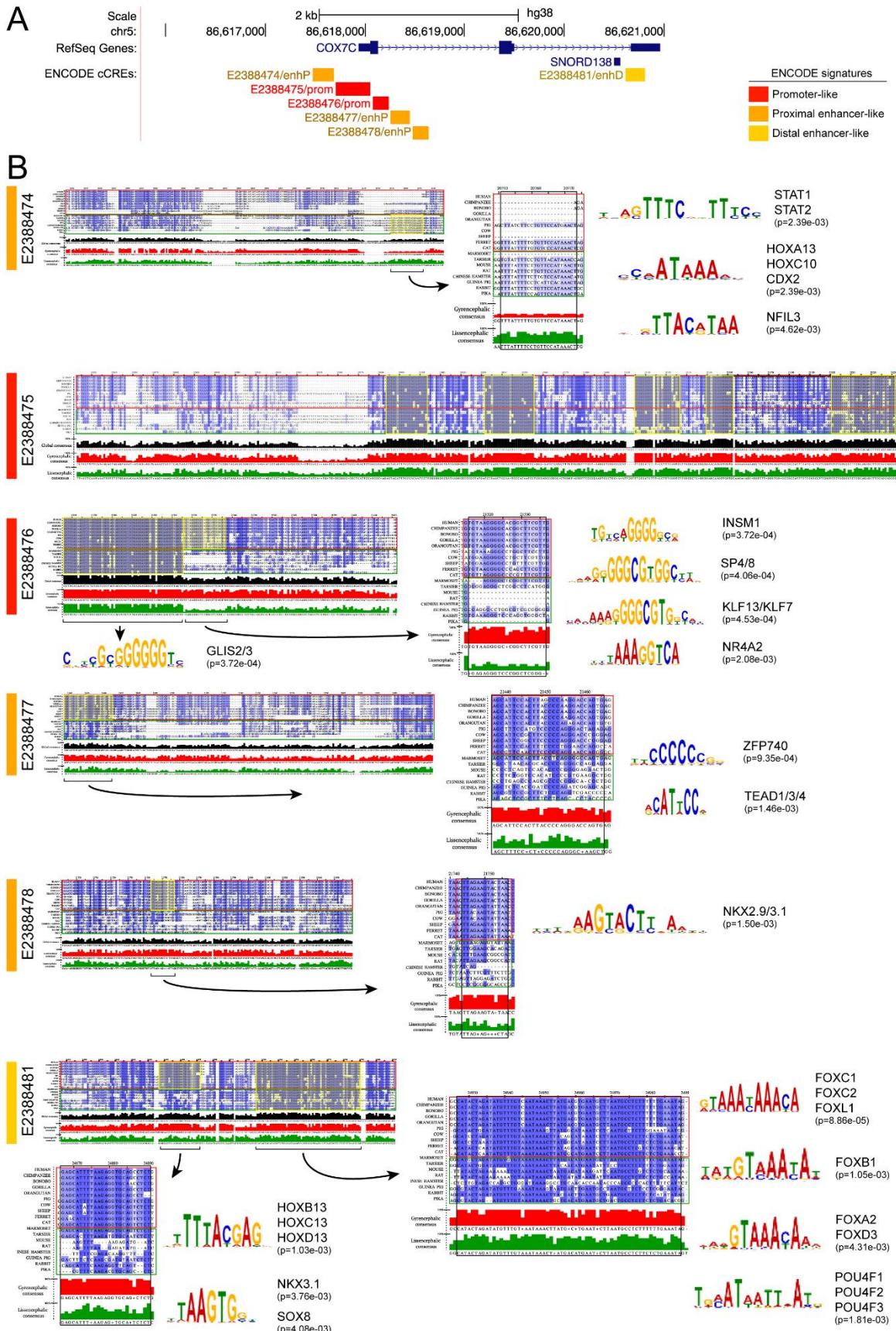


Fig. S7. Conservation and divergence of MIR3607 expression regulation

(A) Genomic region of MIR3607 (SNORD13B) in the human genome (GRCh38/hg38). Colored boxes indicate candidate cis-regulatory elements (cCREs) showing promoter-like, proximal enhancer-like and distal enhancer-like signatures (67).

(B) DNA sequences from -5kb to +1Kb downstream of MIR3607 TSS for 18 species, from UCSD Table browser (*xenoRefGene*). Gyrencephalic species are on top (red boxes): human (*Homo sapiens*), chimpanzee (*Pan troglodytes*), bonobo (*Pan paniscus*), gorilla (*Gorilla gorilla*), orangutan (*Pongo pygmaeus*), pig (*Sus domesticus*), cow (*Bos taurus*), sheep (*Ovis aries*), ferret (*Mustela furo*) and cat (*Felis silvestris*). Lissencephalis species are on bottom (green boxes): marmoset (*Callithrix jacchus*), tarsier (*Tarsius tarsier*), mouse (*Mus musculus*), rat (*Rattus norvegicus*), chinese hamster (*Cricetulus barabensis*), guinea pig (*Cavia porcellus*), rabbit (*Oryctolagus cuniculus*) and pika (*Ochotona princeps*). Sequences were aligned with Clustal Omega, conserved cCRE subregions were selected (yellow boxes) and putative transcription factor binding sites were identified using MEME (71). Sequence logos for motifs conserved across all species analyzed are shown (e.g. E2388481/enhD), along with those identified exclusively in gyrencephalic (e.g. E2388478/enhP) or lysencephalic species (e.g. E2388474/enhP). Top candidate transcription factors binding each motif, and their *p* value, are indicated. For each cCRE, the percent sequence identity histogram for gyrencephalic (red) and lysencephalic (green) species is plotted below the alignment overview.

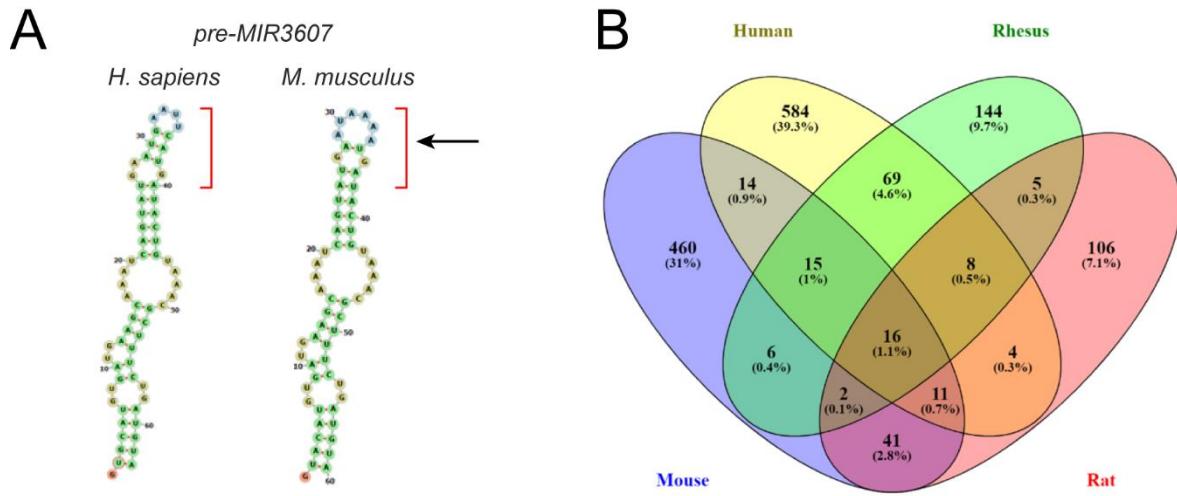


Fig. S8. Conservation and divergence of miRNAs targeting *APC*

(A) Schema of the predicted secondary structure of *pre-MIR3607* in human (*H. sapiens*) and mouse (*M. musculus*), according to (61). Sequence differences are predicted to affect the structure of the loop region (red brackets), shorter in mouse (arrow).

(B) Venn diagram of miRNAs computationally predicted to target *APC* in the indicated species. Intersections indicate the number and percentage of miRNAs common between the intersected species. Most targeting miRNAs are unique to each species, and the majority of common miRNAs are shared between primates (with a large and folded cortex), or between rodents (with a small and smooth cortex).

LEGENDS TO SUPPLEMENTARY TABLES

Supplementary Table S1. Sequence similarity of pre-*MIR3607* across mammals

Alignment of the sequence of pre-MIR3607 across mammalian species of main clades, spanning the 5p and 3p regions (green), and the loop region. Yellow shade indicates the seed sequence; purple shade indicates nucleotide mismatches compared to human (top); dashes indicate missing nucleotides.

Supplementary Table S2. Differential gene expression and functional enrichment analyses for RNAseq data between *MIR3607* and *Scrambled* conditions

Table parts contain: 1) Differential gene expression analysis (complete table without any filter: “FullTable”; table of significantly differentially expressed genes, padj. < 0.01, “FDR_0.01”). 2) Functional enrichment analysis with DAVID functional annotation clustering (DAVID defined default annotation categories for significant genes: “FDR_0.01_Full”; Gene Ontology for Biological Processes: “FDR_0.01_GOTERM_BP_ALL”; KEGG Pathway Ontology: “FDR_0.01_KEGG”; KEGG Pathway and Reactome Ontologies: “FDR_0.01_KEGG+Reactome”). 3) Cytoscape-ClueGo results for functionally grouped gene ontologies and pathway annotation networks (“ClueGO Results”). 4) MIR3607 human ortholog target genes (“HumanTargets”). 5) Differential gene expression results for predicted mouse *MIR3607* targets (“HumanOrthologTarget”). 6) GSEA enrichment analysis results (results for MSigDB Hallmark: “gsea_MSigDB_HALLMARK_H”; results for Gene Ontology, Biological Processes: “gsea_MsigDB_GO_BP_C5”).

Supplementary Table S3. Target sites for *MIR3607* in the 3’UTR of *APC* across selected vertebrates

Identification and sequence of putative target sites for *MIR3607* in two locations of the *APC* mRNA 3’UTR. Green shade indicates identical sequence compared to human (top); dashes indicate missing nucleotides. Mouse and ferret are highlighted with red and yellow shades.

LEGENDS TO SUPPLEMENTARY MOVIES

Supplementary Movie S1. Radial migration of control mouse cortical neurons

Videomicroscopy of migrating cortical neurons in an organotypic slice culture from the cerebral cortex of an E17.5 mouse embryo electroporated *in utero* at E14.5 with DNA plasmids encoding for *Scrambled* miRNA, *mGFP-flox* and *Cre*. Ventricular surface is down, pial surface is up. The slice was prepared 20hr after *in utero* electroporation, and imaging started 4hrs after slice preparation. Time between frames is 30 min. Total imaging time is 24.5hrs. Red dots track three migrating neurons through the movie.

Supplementary Movie S2. Radial migration of *MIR3607*-overexpressing mouse cortical neurons

Videomicroscopy of migrating cortical neurons in an organotypic slice culture from the cerebral cortex of an E17.5 mouse embryo electroporated *in utero* at E14.5 with DNA plasmids encoding for *MIR3607*, *mGFP-flox* and *Cre*. Ventricular surface is down, pial surface is up. The slice was prepared 20hr after *in utero* electroporation, and imaging started 4hrs after slice preparation. Time between frames is 30 min. Total imaging time is 31.5hrs. Red dots track two migrating neurons through the movie.

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