

Supplementary Data

**Development of Targeted Nanoparticles Loaded with Antiviral Drugs
for SARS-CoV-2 Inhibition**

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Figure S1A. Structures of “alkylated ligands” as model/surrogate of the *tri*-block-PEGylated copolymers used for docking calculations: PEG-ACL (orange), PEG-DCL (green), PEG-CPL (blue), and PEG-TAPI-2 (magenta). The site and mode of conjugation, with the short chain to simulate the polymer (grey) are indicated.

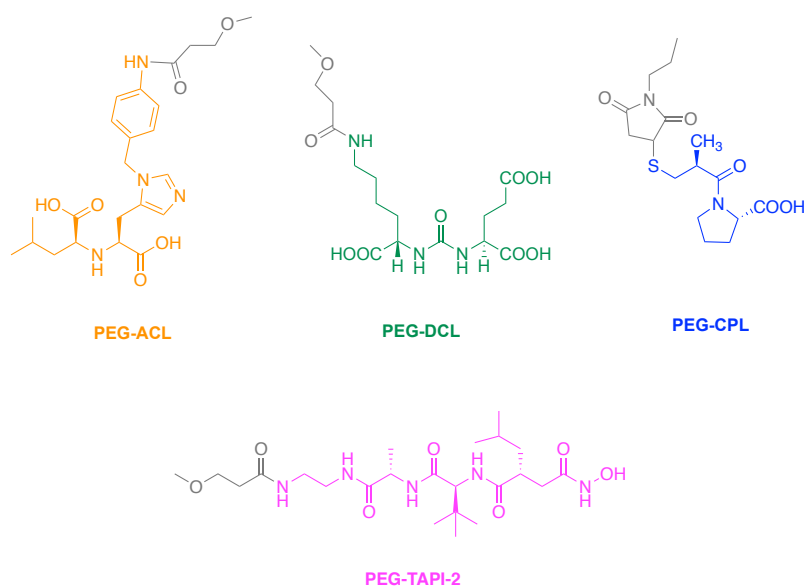
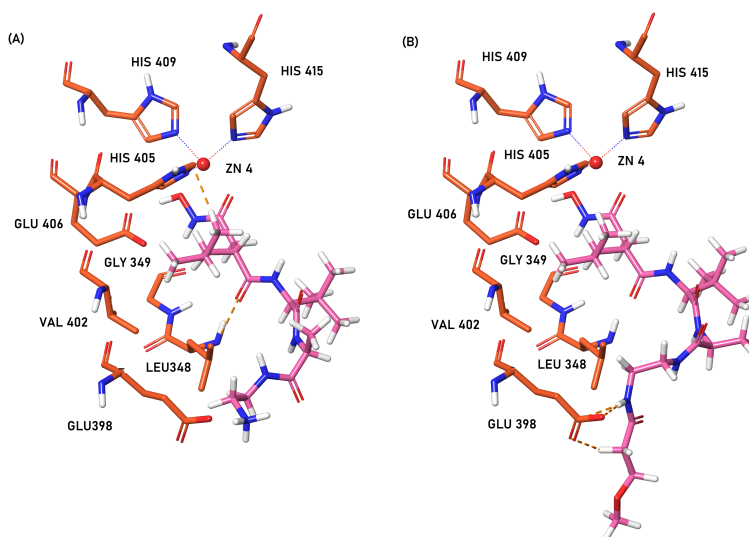
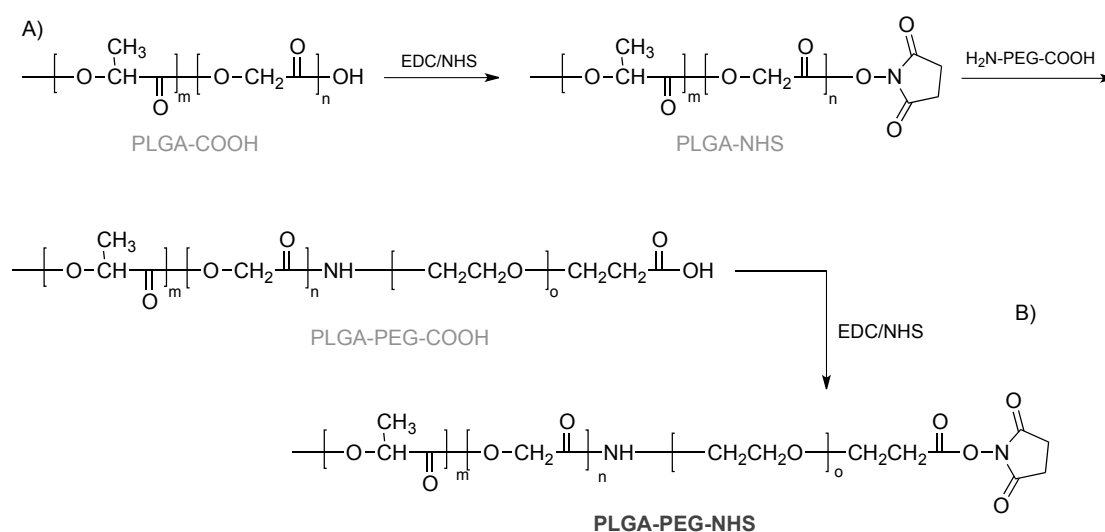


Figure S1B. 3D interaction images of the ligand **4** (A, magenta) and its PEGylated model (B), in the TACE active site. Zinc ion cofactor is represented as red sphere.



Scheme S1. Synthesis of copolymer PLGA-PEG-NHS.^a

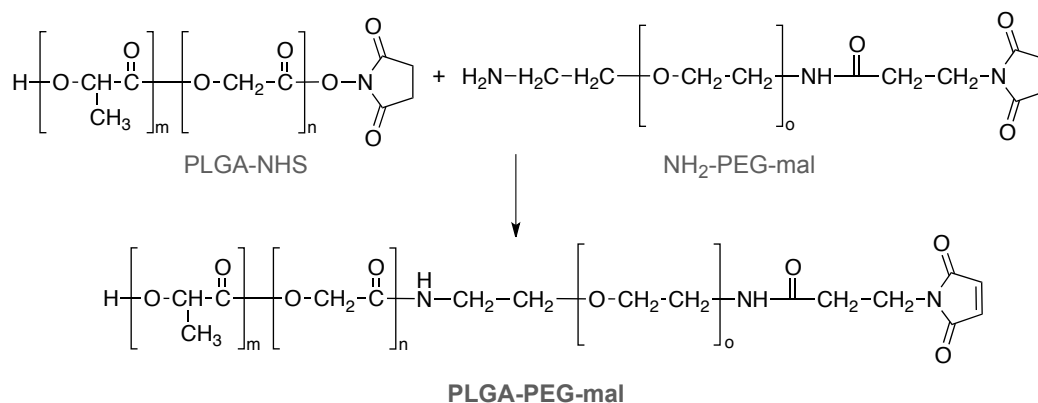


Reagents and conditions: A) *i*) NHS, CH_2Cl_2 , EDC•HCl, N_2 , room temperature for 12 h, N_2 ; *ii*) CHCl_3 , DIPEA, r.t. for 24 h, N_2 . B) NHS, CH_2Cl_2 , EDC•HCl, N_2 , r.t. for 12 h, N_2 .

A) Synthesis of PLGA-PEG-COOH di-Block-Copolymer [1-3]. Briefly, to a solution of PLGA-COOH (4.0 g, 0.221 mmol) in anhydrous CH_2Cl_2 (15 mL), *N*-hydroxysuccinimide (NHS, 0.102 g, 0.884 mmol., ~4 equiv) and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDC•HCl, 0.182 g, 0.950 mmol, ~4.3 equiv.) were added, and the reaction mixture was magnetically stirred at room temperature for 12 h under nitrogen atmosphere. PLGA-NHS was obtained by precipitation with cold diethyl ether (Et_2O , 15 mL) as a white solid, which was filtered, and repeatedly washed in a cold mixture of Et_2O and (few drops of) MeOH, then treated with nitrogen and dried under vacuum to remove solvent (~95% yield). The intermediate PLGA-NHS (3.5 g, 0.200 mmol) was dissolved in anhydrous CHCl_3 (12 mL), $\text{NH}_2\text{-PEG-COOH}$ (0.91 g, 0.26 mmol; 1.3 equiv) and *N,N*-diisopropylethylamine (DIPEA) (0.14 mL, 0.80 mmol; 4 equiv) were added, and the reaction reaction was magnetically stirred at room temperature for 24 h under nitrogen atmosphere. The mixture was then repeatedly treated with cold Et_2O to remove unreacted PEG, to afford the PLGA-PEG block copolymer (~95% yield), which was dried under vacuum, characterized, and used for NP preparation without any further purification. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 5.23 (m, OC-CH(CH_3)O, PLGA), 4.78 (m, OC- CH_2 O, PLGA), 3.65 (s, CH_2CH_2 O, PEG), 1.56 (brs, OC-CH CH_3 O, PLGA) [1-3].

B) Synthesis of PLGA-PEG-NHS di-Block-Copolymer [1-3]. To a solution of PLGA-PEG-COOH (2.5 g, 0.1225 mmol) in anhydrous CH₂Cl₂ (9 mL), NHS (56 mg, 0.490 mmol, ~4 equiv) and EDC•HCl (101 mg, 0.5267 mmol, ~4.3 equiv) were added, and the solution was magnetically stirred at room temperature for 12 h under nitrogen atmosphere. The activated PLGA-PEG-NHS copolymer was obtained by precipitation with cold diethyl ether (Et₂O, 10 mL) as a white solid (~92% yield), following the procedure previously used to obtain the PLGA- NHS intermediate.

Scheme S2. Synthesis of copolymer PLGA-PEG-mal.^a



Reagents and conditions: (i) CHCl₃, DIPEA, N₂, r.t. for 24 h.

Synthesis of PLGA-PEG-mal [4]. To a solution of PLGA-NHS (1.0 g, 0.057 mmol) in anhydrous CHCl₃ (7 mL), NH₂-PEG-maleimide (0.80 g, 0.228 mmol, 4 equiv.) in anhydrous CHCl₃ (5 mL) and was added DIPEA (0.1 mL, 0.57 mmol, 10 equiv.), and the solution was magnetically stirred at room temperature for 24 h under nitrogen atmosphere. The PLGA-PEG-mal copolymer was obtained by precipitation with cold Et₂O, then dried under vacuum, and used for NP preparation without further treatment (yield, 95%). ¹H-NMR (400 MHz, CDCl₃): δ 5.32-5.10 (m, 1H, -OC-CH(CH₃)O-, PLGA), 4.93-4.63 (m, 2H, -OC-CH₂O- PLGA), 3.64 (brs, 2H, -CH₂CH₂O-, PEG), 1.58 (brs, 3H, -OC-CH(CH₃)O-, PLGA) [4].

Figure S2. Characterization of PLGA-PEG-1: $^1\text{H-NMR}$ with pattern signals of PLGA-PEG fragment, and magnification of representative ACL (1) peaks (dotted window).

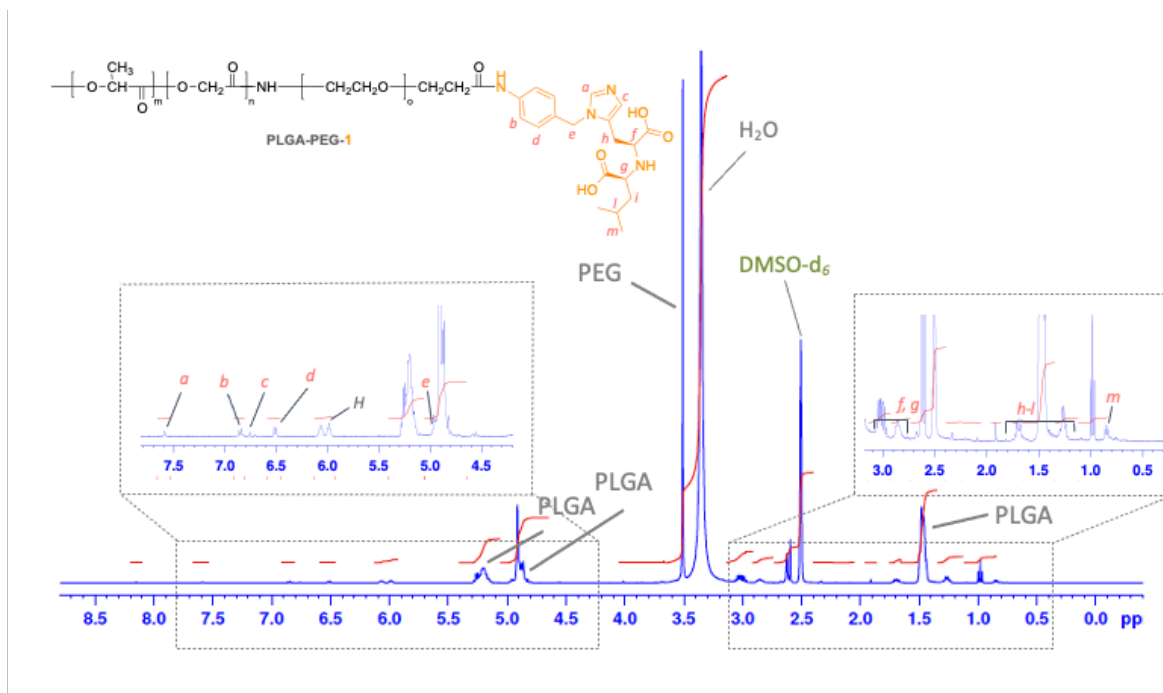


Figure S3. Characterization of PLGA-PEG-2: $^1\text{H-NMR}$ with pattern signals of PLGA-PEG fragment, and magnification of representative DCL (2) peaks (dotted window).

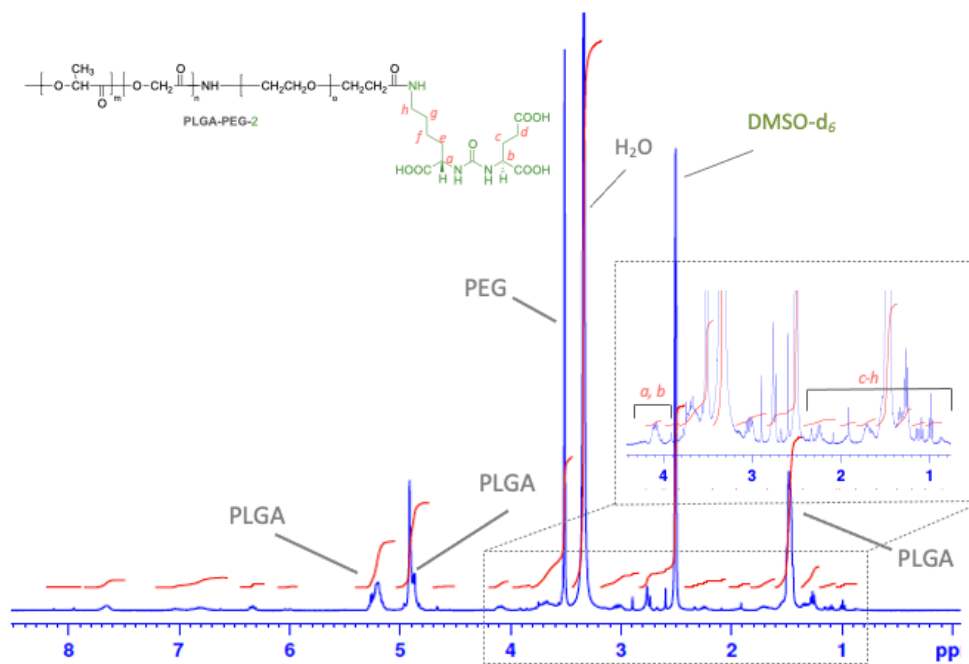


Figure S4. Characterization of PLGA-PEG-3: $^1\text{H-NMR}$ with pattern signals of PLGA-PEG fragment, and magnification of representative CPL (3) peaks (dotted window).

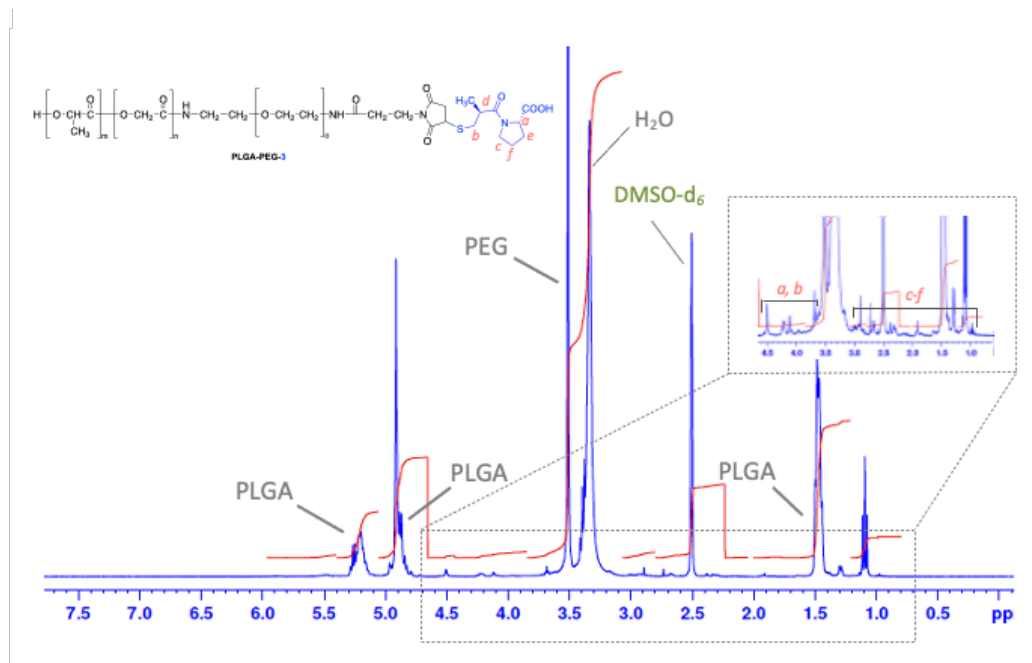


Figure S5. Characterization of PLGA-PEG-4: $^1\text{H-NMR}$ with pattern signals of PLGA-PEG fragment, and magnification of representative TAPI-2 (4) peaks (dotted window).

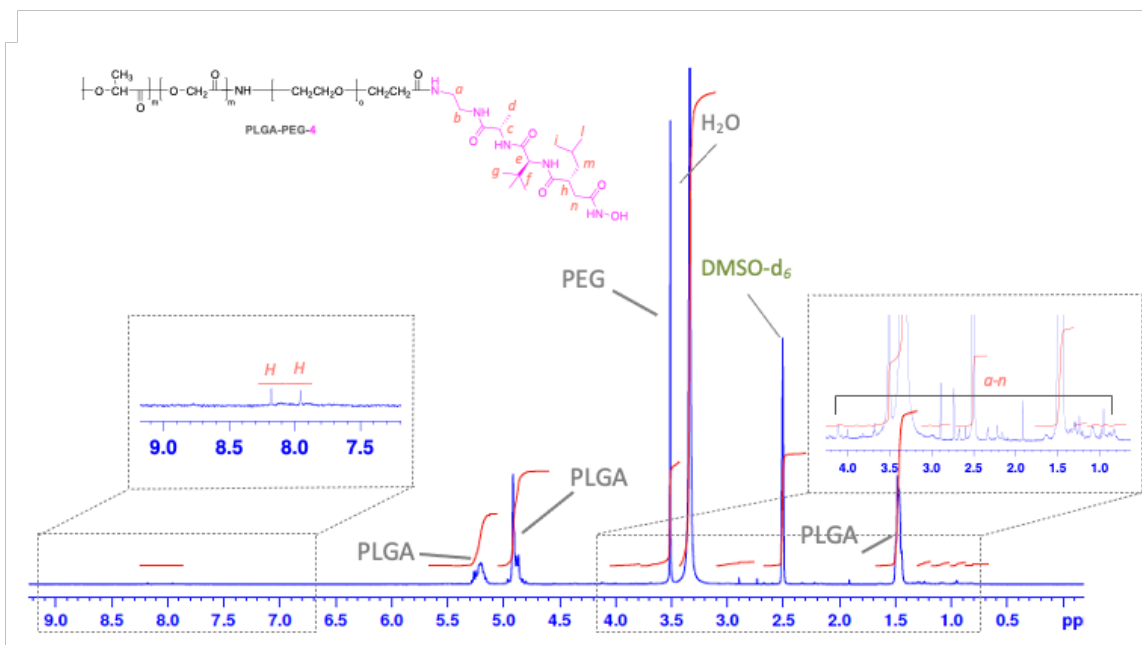


Figure S6. $^1\text{H-NMR}$ spectra of **1** recorded in DMSO-d_6 (a), $\text{DMSO-d}_6 + \text{D}_2\text{O}$ (b), and D_2O (c).

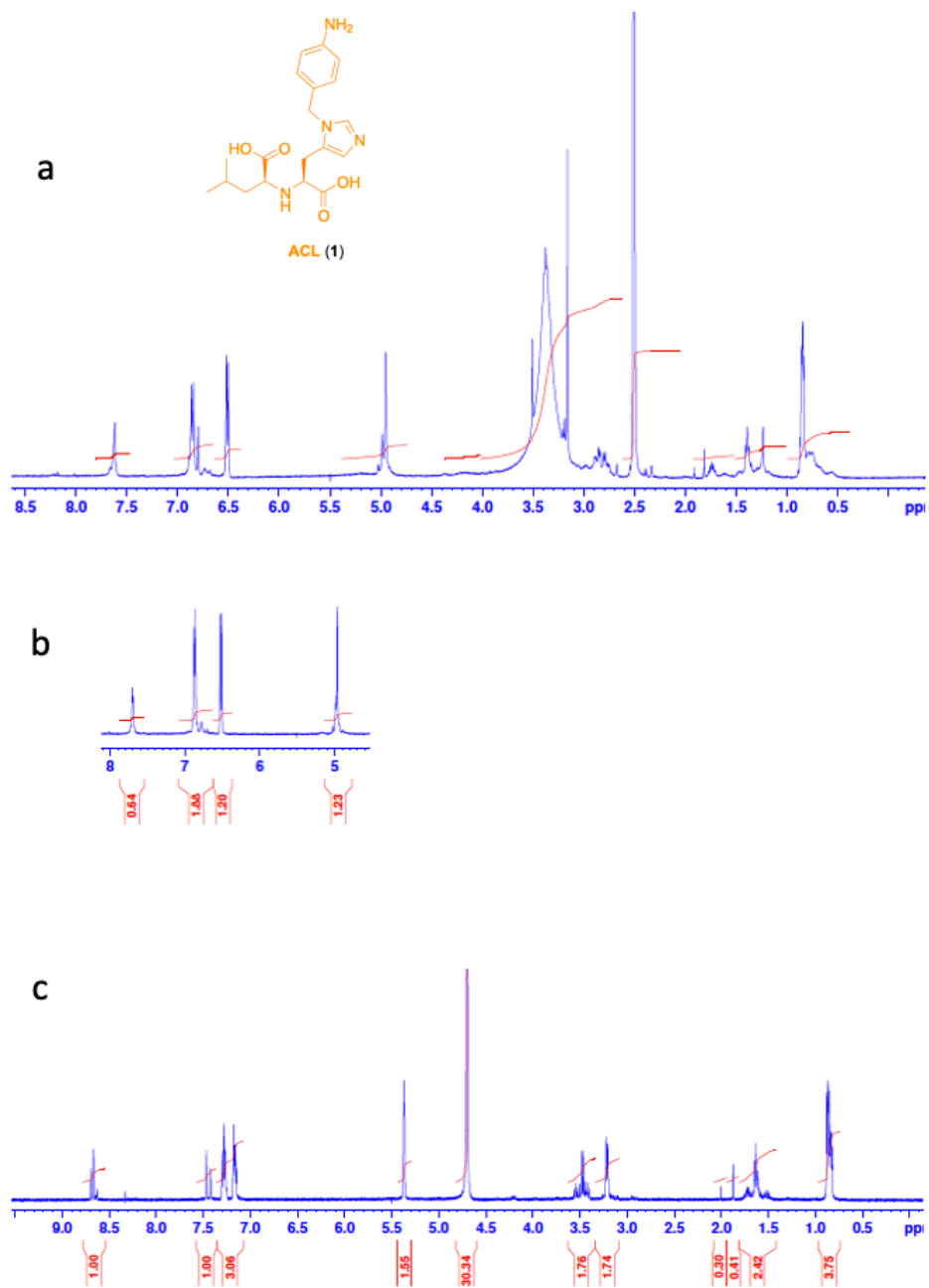


Figure S7. Top: 2D-NOESY spectrum of **1** recorded in D₂O (**a**), with significant crosspeak indicating a NOE correlation for **1** conformer in *S,S* configuration (**b**).

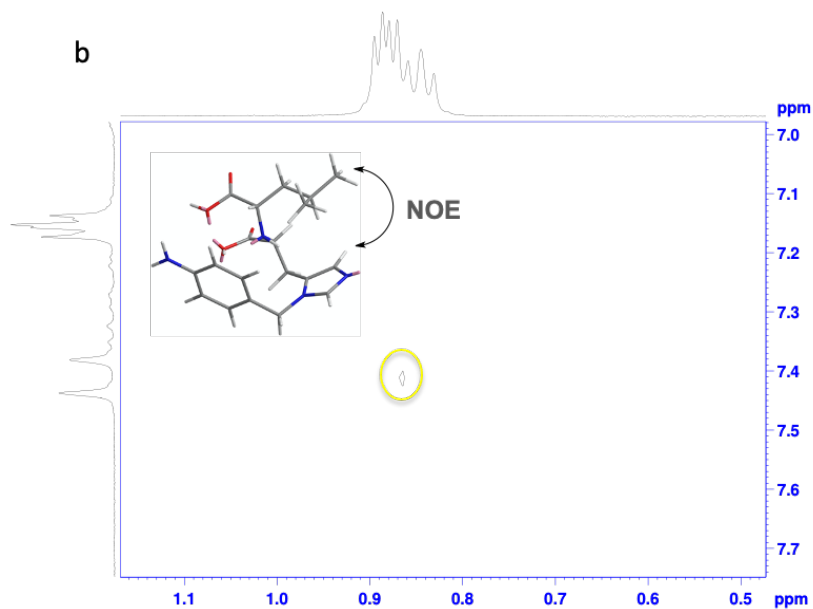
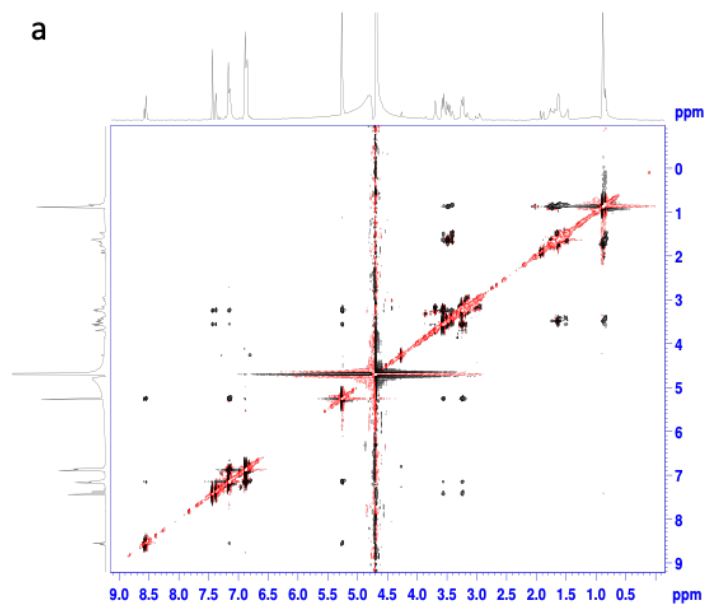


Figure S8. ^{13}C -NMR (a), COSY (b) and HSQC (c) spectra of **1** recorded in DMSO.

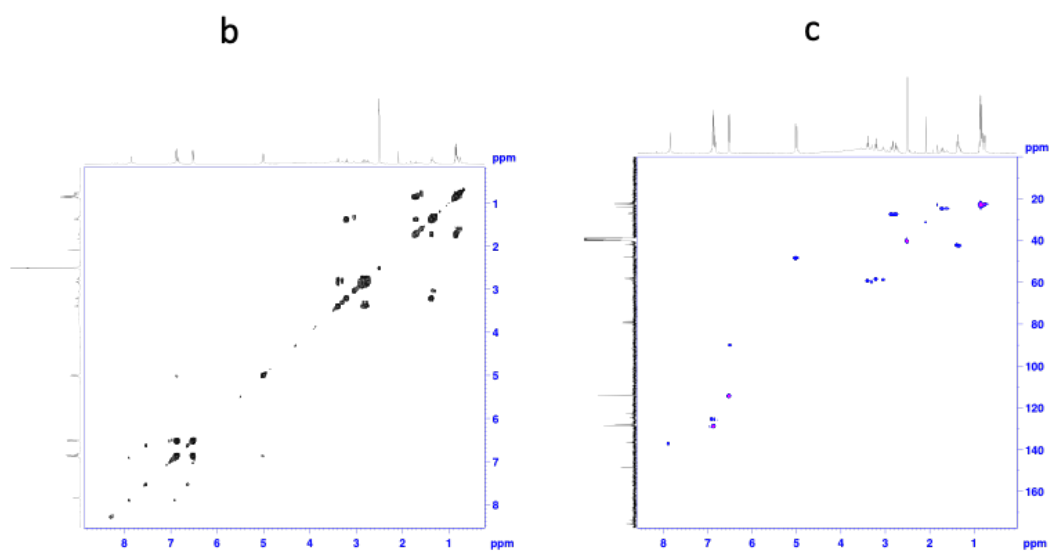
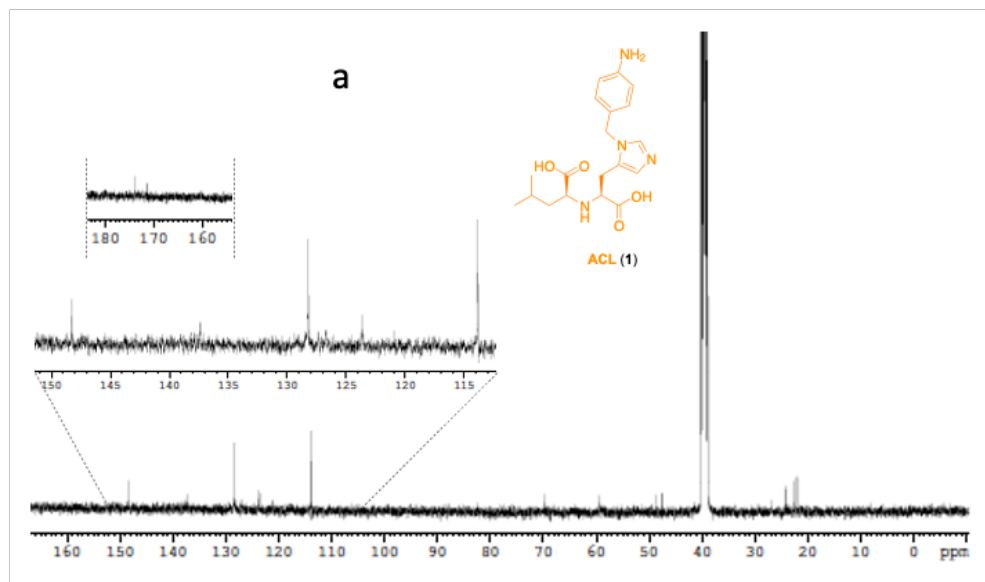


Figure S9. HRMS (ESI) spectra (**a**, **b**) of **1** calculated for $[C_{19}H_{26}N_4O_4+H]^+$ and $[C_{19}H_{26}N_4O_4+Na]^+$.

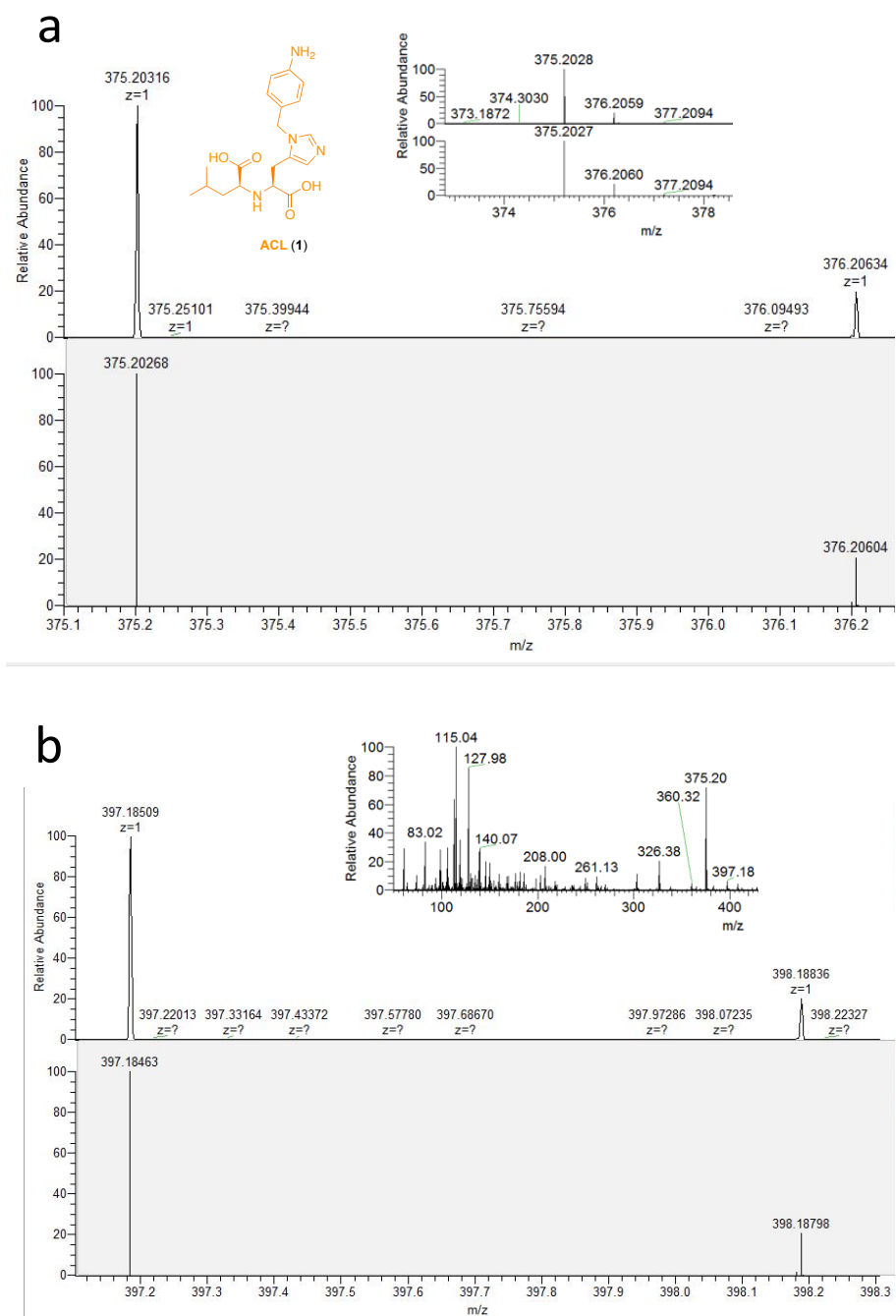
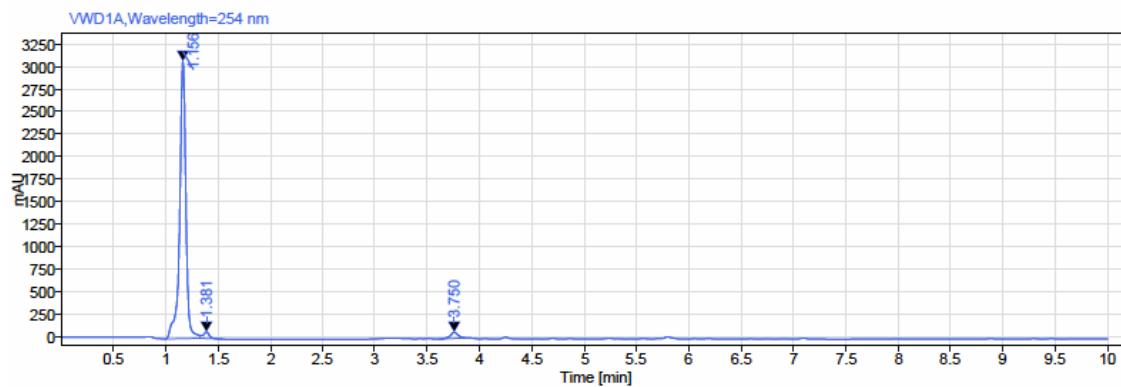


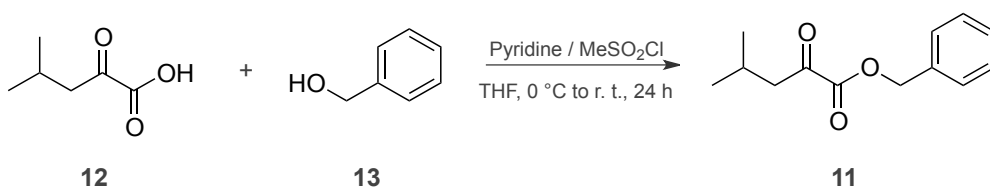
Figure S10. HPLC chromatogram of 1.



Signal: VWD1A, Wavelength=254 nm

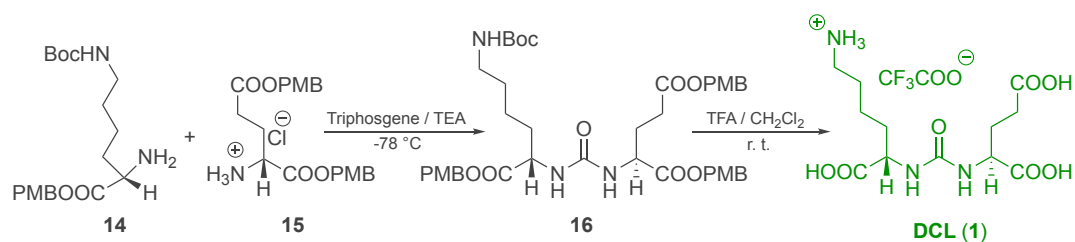
RT [min]	Type	Width [min]	Area	Height	Area%	Name
1.156	BM m	0.06	12718.58	3084.24	95.11	
1.381	MM m	0.06	304.63	71.92	2.28	
3.750	BM m	0.07	349.58	68.76	2.61	
		Sum	13372.78			

Scheme S3. Synthesis of the synthon **11**.^a



Synthesis of benzyl 4-methyl-2-oxopentanoate (11) [6]. Benzyl α -ketoester **11** was synthesized by dissolving the commercially available benzyl 4-methyl-2-oxopentanoate (**12**) (5.0 g., 4.74 mL, 38.42 mmol, 1 equiv), benzyl alcohol (**13**) (8.31 g., 7.95 mL, 76.84 mmol, 2 equiv) and pyridine (7.60 g., 7.67 mL, 96.05 mmol, 2.5 equiv), in THF (35 mL), cooled to 0 °C, under magnetic stirring, according to a previously reported method [5,6]. To this solution, mesyl chloride (5.28 g., 3.58 mL, 46.10 mmol, 1.2 equiv) was added dropwise, and the reaction mixture was then stirred at room temperature for 24 h. Next, the reaction was quenched with water (80 mL) and extracted with diethyl ether (3 x 50 mL). The organic layers were dried over Na₂SO₄, filtered and concentrated in vacuo. The resulting crude product was purified by flash column chromatography on silica gel (petroleum ether / ethyl acetate = 95:5) to afford **11** as a colorless oil (97% yield). ¹H-NMR (400 MHz, CDCl₃) δ 7.39-7.36 (5H, m), 5.27 (2H, s), 2.71 (2H, d), 2.18 (1H, m), 0.95 (6H, d). MS (ESI), calculated for C₁₃H₁₆O₃ ([M]⁺) 220 [6].

Scheme S4. Synthesis of **2**.



Preparation of 2-[[[(5-amino-1-carboxypentyl)carbamoyl]amino]pentanedioic acid (**2**) [1-3].

The intermediate ((10*S*,14*S*)-tris(4-methoxybenzyl)2,2-dimethyl-4,12-dioxo-3-oxa-5,11,13-triazahexadecane-10,14,16-tricarboxylate (**16**, 200 mg, 0.26 mmol) was dissolved in 5 mL 1:1 trifluoroacetic acid (TFA):CH₂Cl₂ solution and stirred at room temperature. After 3 h, the solution was evaporated, and the solid residue was triturated with dry Et₂O, filtered, and washed several times with dry Et₂O to yield **2** as a beige powder, which became amorphous after air exposure (yield: ~90%). ¹H-NMR (400 MHz, D₂O) δ 4.19– 4.12 (m, 2H), 2.93– 2.90 (m, 2H), 2.45– 2.41 (m, 2H), 2.14– 2.05 (m, 1H), 1.93– 1.76 (m, 2H), 1.67– 1.59 (m, 3H), 1.45–1.30 (m, 2H). ¹³C-NMR (D₂O) δ 177.3, 177.1, 176.9, 176.4, 159.3, 66.0, 52.9, 52.6, 39.2, 30.4, 30.1, 26.2, 26.1, 22.0, 14.1. HRMS (ESI), calculated for C₁₂H₂₂N₃O₇ ([M+H]⁺) 320.1450. Anal. (C₁₂H₂₁N₃O₇ • 1.15CF₃COOH • 0.65Et₂O) C, H, N [1-3,7].

Preparation of ((10*S*,14*S*)-tris(4-methoxybenzyl)2,2-dimethyl-4,12-dioxo-3-oxa-5,11,13-triazahexadecane-10,14,16-tricarboxylate (**16**).

The urea-based key intermediate **16** was synthesized by coupling of 2-amino-6-tert-butoxycarbonylamino-4-methoxybenzyl ester (**14**) with bis-4-methoxybenzyl-L-glutamate hydrochloride (**15**) following the previously described procedures [1-3]. Briefly, triphosgene (0.50 g, 1.71 mmol) was placed onto a flame dried flask cooled to 0 °C, under nitrogen atmosphere, and a solution of **15** (1.89 g, 5.17 mmol, 3 equiv) and TEA (1.50 mL, 10.32 mmol, 6 equiv) dissolved in CH₂Cl₂ (10 mL) was added. The reaction mixture was stirred at 0 °C for 20 min. A mixture of **14** (2.90 g, 6.82 mmol, 3 equiv) and TEA (1.94 mL, 13.63 mmol, 6 equiv) in CH₂Cl₂ (7 mL) was then added, and the resulting mixture was allowed to warm to room temperature and stirred for 2 h. Product was extracted by CH₂Cl₂, washed with water and brine, and dried over Na₂SO₄. Purification by flash chromatography (80/20 CH₂Cl₂/EtOAc) afforded a yellow oil that solidified upon standing (yield: ~75%). ¹H-NMR (400 MHz, CDCl₃) δ: 7.26 (d, 6H), 6.87 (d, 6H), 5.52 (d, 2H), 5.13-4.98 (m, 6H), 4.76 (brs, 1H), 4.53-

4.48 (m, 1H), 4.50-4.43 (m, 1H), 3.79 (s, 9H), 3.04-2.96 (m, 2H), 2.40-2.34 (m, 2H), 2.14-2.11 (m, 2H), 1.94-1.70 (m, 2H), 1.60-1.56 (m, 2H), 1.42 (s, 9H), 1.25-1.22 (m, 2H). HRMS (ESI), calculated for C₄₁H₅₃N₃O₁₂ ([M+H]⁺) 780 [1-3].

Figure S11. ^1H -NMR (a) and ^{13}C -NMR (b) spectra of **2** recorded in D_2O .

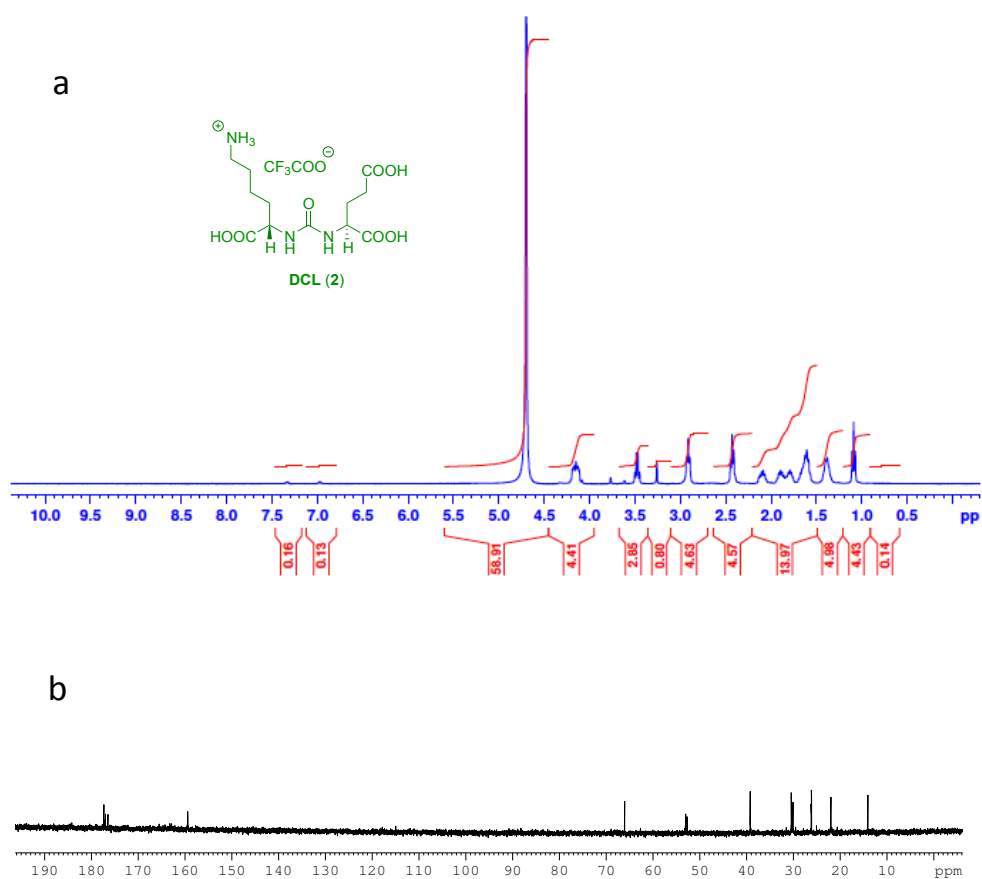


Figure S12. a, b) HRMS (ESI) of **2**, calculated for $C_{12}H_{22}N_3O_7$ ($[M+H]^+$).

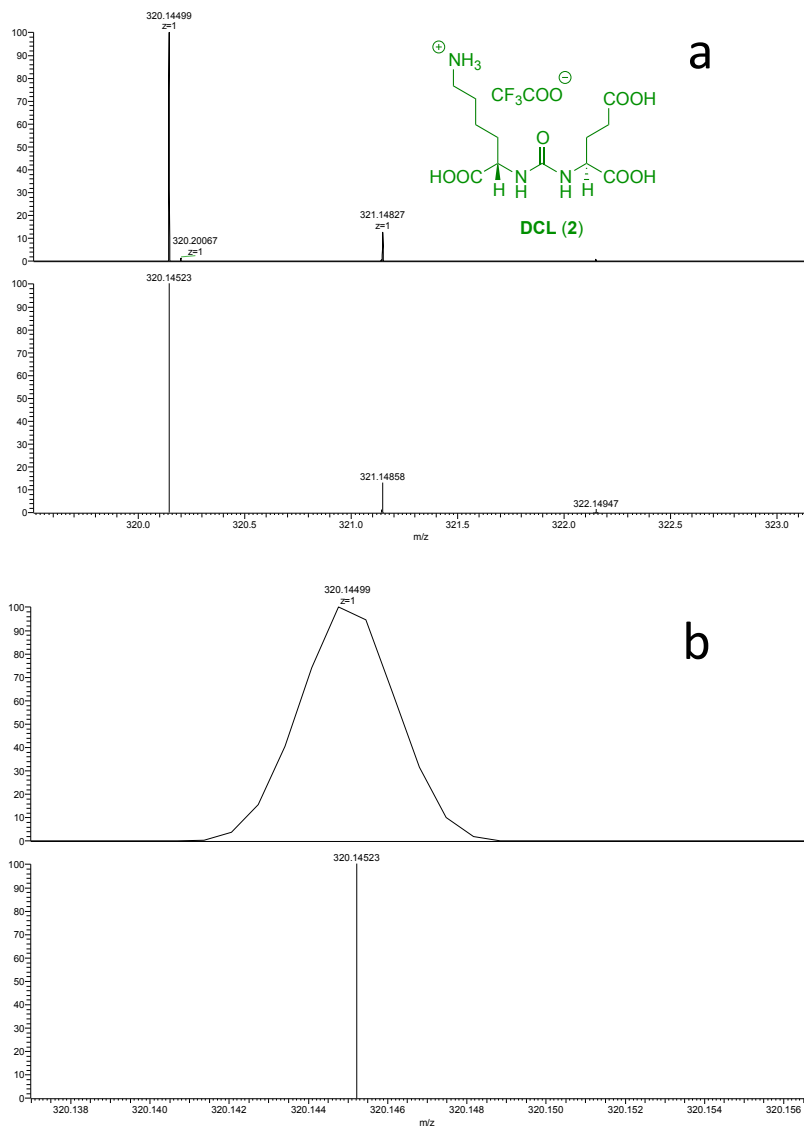


Figure S13. Microscopic images showing effect of TNP-1E and RDV on SARS-CoV-2 CPE induction. **(A)** Not infected VERO E6 cell monolayer as control (CTRL), and SARS-CoV-2 infected cells at 0.05_{MOI}, after 72 hours post infection. **(B)** SARS-CoV-2 infected VERO E6 after TNP-1E exposure at different concentrations. Unloaded NP were tested at the same NP w/v concentration ($\mu\text{g}/\text{mL}$) of the parent RDV-loaded nanosample (300, 60, 6 $\mu\text{g}/\text{mL}$, for TNP-1E, which correspond to 5, 1, 0.1 μM , for RDV-TNP-1, respectively). **(C)** SARS-CoV-2 infected VERO E6 after RDV exposure at different concentrations (5, 1, 0.1 μM) (scale bar 50 μm).

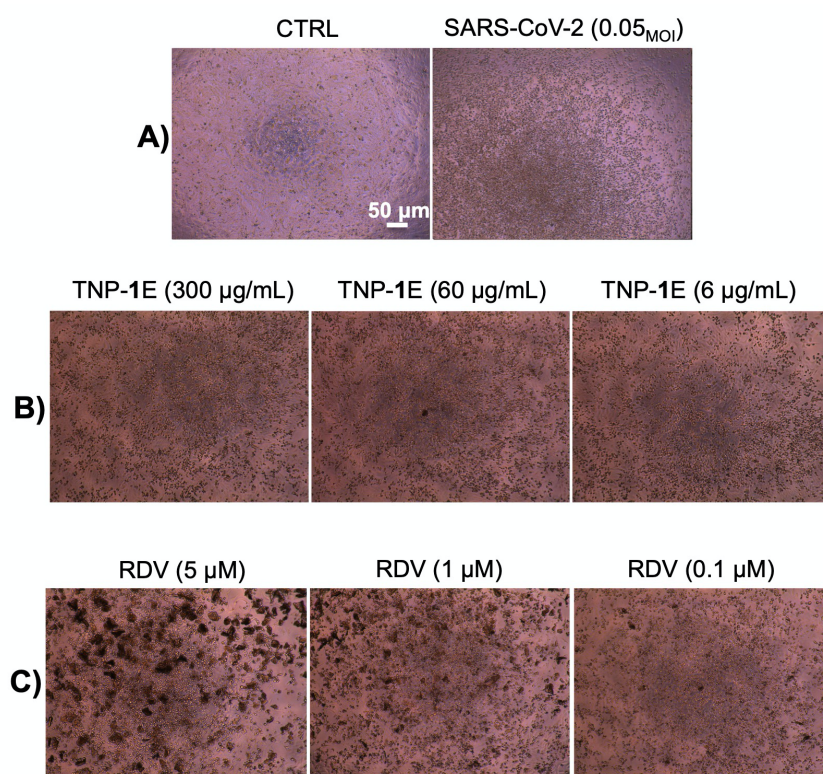
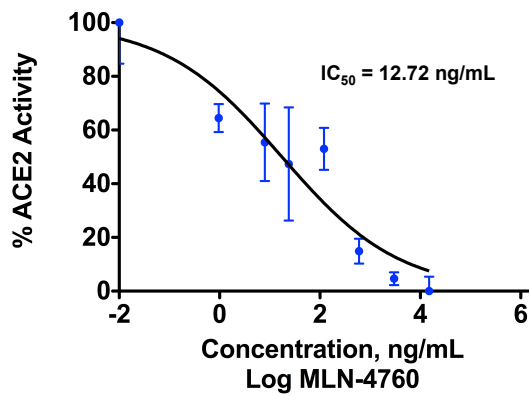


Figure S14. ACE2 exopeptidase activity by MLN-4760.



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