

Electrochemical Rearrangement of 3-Hydroxyoxindoles into Benzoxazinones

Supporting Information

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1 General Remarks

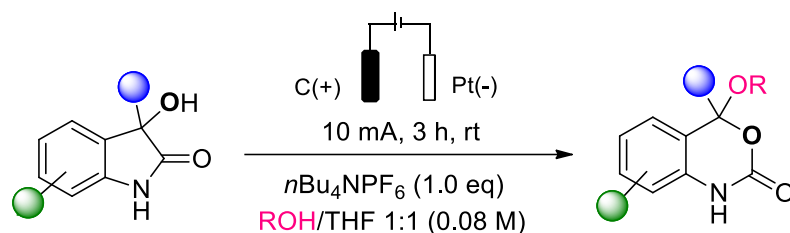
All solvents were distilled from appropriate drying agents prior to use or directly taken from commercial sealed bottles under an atmosphere of argon. All reagents were used as received from commercial suppliers (*Alfa Aesar*, *Sigma Aldrich* or *TCI*) unless otherwise stated. Reaction progress was monitored by thin layer chromatography (TLC) performed on aluminum plates coated with silica gel F₂₅₄ with 0.2 mm thickness. Chromatograms were visualized by fluorescence quenching with UV light at 254 nm and/or by staining using vanilin. Flash column chromatography was performed using silica gel 60 (230-400 mesh, Merck and co.). Yields refer to chromatographically and spectroscopically pure compounds. ¹H NMR, ¹³C NMR and ¹⁸F NMR spectra were recorded using a Bruker AV-400, AV-600 and AV-700 spectrometer at 300K. ¹H NMR chemical shifts are reported in ppm using residual solvent peak as reference (CDCl₃: δ = 7.26 ppm or DMSO-d₆: δ = 2.50 ppm). Data for ¹H NMR are presented as follows: chemical shift δ (ppm), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet, br = broad), coupling constant *J* (Hz) and integration; ¹³C NMR spectra were recorded at 100, 150 or 175 MHz using broadband proton decoupling and chemical shifts are reported in ppm using residual solvent peaks as reference (CDCl₃: δ = 77.16 ppm or DMSO-d₆: δ = 39.52 ppm). Multiplicity was defined by recorded a ¹³C NMR spectra using the attached proton test (APT). Neat infra-red spectra were recorded using a Bruker Vertex 70 FT-IR spectrometer. Wavenumbers are reported in cm⁻¹. Mass spectra were obtained using a Finnigan MAT 8200 or (70 eV) or an Agilent 5973 (70 eV) spectrometer, using electrospray ionization (ESI) and a maXis UHR-TOF analyzer.

Electrolysis general information

Electrochemical reactions were performed with ElectraSyn 2.0 package (IKA) using the constant current mode. The reactions were conducted in a 10 mL vial with a magnetic stir bar and a graphite-SK-50 (5.0 x 0.8 x 0.2 cm) working electrode and a platinum-plated (5.0 x 0.8 x 0.2 cm) counter-electrode with a distance of 0.6 cm between the two electrodes.

2 General Procedure for Electrochemical Formation of 3,1-Benzoxazin-2-ones

General procedure (A) to access 3,1-Benzoxazin-2-ones

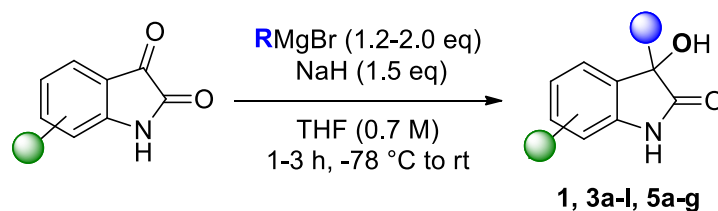


The ElectraSyn vial (10 mL) equipped with a stir bar was charged with 3-hydroxyoxindole derivatives **1**, **3a-l**, **5a-f** (0.40 mmol, 1.0 equiv.), $n\text{Bu}_4\text{NPF}_6$ (155 mg, 0.40 mmol, 1.0 equiv.), ROH (2.5 mL) and THF (2.5 mL). The ElectraSyn vial cap equipped with anode (graphite) and cathode (platinum) was inserted into the mixture. The reaction mixture was electrolyzed at a constant current of 10 mA for 3 h. The ElectraSyn vial cap was removed, and electrodes were rinsed with CH_2Cl_2 (2.0 mL), which was combined with the crude mixture. Then, the crude mixture was concentrated under reduced pressure and purified by FC over silica gel (20 g SiO_2 , heptane/EtOAc, 100/0 to 50/50, gradient) to furnish the desired **2a-g**, **4a-l**, **6a-f**.

Due to limitation of this specific electrochemical device, the maximum reaction limit is 0.4 mmol.

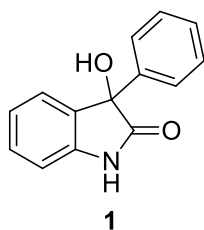
3 Experimental Procedures and Characterization Data of the Starting Materials **1**, **3a-l** and **5a-g**

General procedure (B) to access 3-hydroxysubstituted oxindoles (**1**, **3a-l**, **5a-g**)



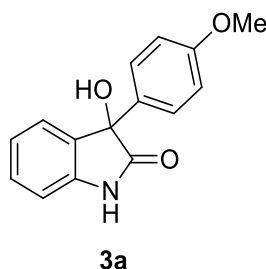
To a solution of the corresponding isatin derivative (1.0 equiv.) in THF (0.7 M), NaH (1.5 equiv.) was added at $-78\text{ }^\circ\text{C}$. The mixture was stirred for 30 min. Then, the Grignard reagent (1.2-2.0 equiv.) was added dropwise at this temperature. The reaction was warmed up to rt and stirred until completion by TLC (1 to 3 hours generally). The mixture was then quenched with a saturated aq. solution NH_4Cl , extracted with EtOAc, washed with H_2O , dried over MgSO_4 and concentrated under reduced pressure. CH_2Cl_2 was added to the solids obtained. The insoluble part was filtered and dried in vacuo to afford the desired product. If necessary, crude products were purified by column chromatography over silica gel (40 g SiO_2 , gradient, heptane/EtOAc, 100/0 to 50/50) to furnish the desired products.

3-Hydroxy-3-phenylindolin-2-one **1**



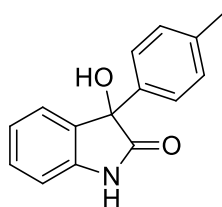
General procedure **B** was followed with isatin (2.0 g, 13.6 mmol) and phenylmagnesium bromide (3.0 M in Et₂O, 9.1 mL, 27.2 mmol, 2.0 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2a** as a yellow solid (1.2 g, 5.33 mmol, 39%). **¹H NMR (400 MHz, DMSO-*d*₆):** δ 10.40 (s, 1H), 7.33-7.23 (m, 6H), 7.10 (d, *J* = 6.9 Hz, 1H), 6.96 (td, *J* = 7.7, 1.1 Hz, 1H), 6.91 (d, *J* = 7.7 Hz, 1H), 6.63 (s, 1H). **¹³C NMR (100 MHz, DMSO-*d*₆):** δ 178.5 (C=O), 142.0 (C), 141.6 (C), 133.8 (C), 129.2 (CH), 128.1 (2CH), 127.4 (CH), 125.4 (2CH), 124.8 (CH), 122.1 (CH), 109.9 (CH), 77.3 (C). **FT-IR (neat, cm⁻¹):** 3408, 1703, 1615, 1467, 1360, 1338, 1180, 1156, 1119, 1067. **HRMS (ESI⁺):** *m/z* calcd. for C₁₄H₁₁NO₂Na [M+Na]⁺ 248.0682, found 248.0670. The data are in agreement with those previously reported in the literature.¹

3-Hydroxy-3-(4-methoxyphenyl)indolin-2-one **3a**



General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and 4-methoxyphenylmagnesium bromide (0.50 M in THF, 9.5 mL, 4.76 mmol, 1.4 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3a** as a white solid (0.65 mg, 2.55 mmol, 75%). **¹H NMR (400 MHz, DMSO-*d*₆):** δ 10.32 (s, 1H), 7.24 (td, *J* = 7.6, 1.1 Hz, 1H), 7.18 (d, *J* = 8.8 Hz, 2H), 7.10 (d, *J* = 7.2 Hz, 1H), 6.96 (t, *J* = 7.6 Hz, 1H), 6.89-6.85 (m, 3H), 6.51 (s, 1H), 3.71 (s, 3H). **¹³C NMR (100 MHz, DMSO-*d*₆):** δ 178.6 (C=O), 158.7 (C), 141.9 (C), 133.7 (C), 133.5 (C), 129.1 (CH), 126.8 (2CH), 124.8 (CH), 121.9 (CH), 113.4 (2CH), 109.8 (CH), 76.9 (C), 55.1 (CH₃). **FT-IR (neat, cm⁻¹):** 3270, 1718, 1684, 1610, 1509, 1466, 1347, 1297, 1250, 1177, 1123, 1106, 1076, 1033. **HRMS (ESI⁺):** *m/z* calcd. for C₁₅H₁₃NO₃Na [M+Na]⁺ 278.0788, found 278.0775. The data are in agreement with those previously reported in the literature.²

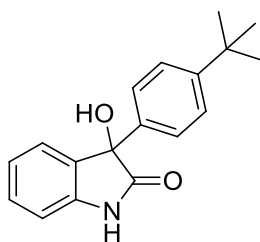
3-Hydroxy-3-(*p*-tolyl)indolin-2-one **3b**



3b

General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and *p*-tolylmagnesium bromide prepared from 4-bromotoluene (0.75 mL, 6.12 mmol, 1.8 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3b** as an off white solid (0.20 g, 0.83 mmol, 24%). **¹H NMR (600 MHz, CDCl₃):** δ 8.87 (brs, 1H), 7.24 (t, *J* = 7.8 Hz, 1H), 7.16 (d, *J* = 7.9 Hz, 2H), 7.13-7.10 (m, 3H), 7.02 (t, *J* = 7.5 Hz, 1H), 6.93 (d, *J* = 7.8 Hz, 1H), 4.60 (s, 1H), 2.34 (s, 3H). **¹³C NMR (150 MHz, CDCl₃):** δ 179.1 (C=O), 141.8 (C), 137.5 (C), 133.6 (C), 130.0 (C), 129.8 (2CH), 128.5 (2CH), 128.4 (CH), 125.4 (CH), 122.8 (CH), 110.1 (CH), 52.5 (CH), 21.3 (CH₃). **HRMS (ESI⁺):** *m/z* calcd. for C₁₅H₁₃NO₂Na [M+Na]⁺ 262.0838, found 262.0839. The data are in agreement with those previously reported in the literature.²

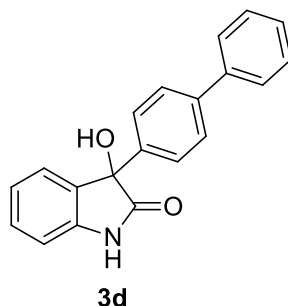
3-(4-(*Tert*-butyl)phenyl)-3-hydroxyindolin-2-one **3c**



3c

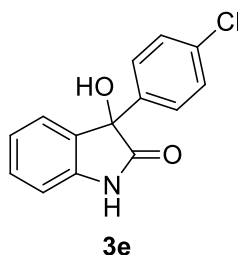
General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and (4-(*tert*-butyl)phenyl)magnesium bromide prepared from 1-bromo-4-*tert*-butylbenzene (1.1 mL, 6.12 mmol, 1.8 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3c** as a yellow solid (0.59 g, 2.08 mmol, 61%). **¹H NMR (600 MHz, DMSO-*d*₆):** δ 10.36 (s, 1H), 7.32 (d, *J* = 8.5 Hz, 2H), 7.24 (td, *J* = 7.7, 0.8 Hz, 1H), 7.19 (d, *J* = 8.5 Hz, 2H), 7.10 (d, *J* = 7.4 Hz, 1H), 6.96 (t, *J* = 7.4 Hz, 1H), 6.88 (d, *J* = 7.7 Hz, 1H), 6.53 (s, 1H), 1.24 (s, 9H). **¹³C NMR (150 MHz, DMSO-*d*₆):** δ 178.6 (C=O), 149.8 (C), 141.9 (C), 138.6 (C), 133.8 (C), 129.2 (CH), 125.2 (2CH), 124.9 (2CH), 124.8 (CH), 122.0 (CH), 109.8 (CH), 77.2 (C), 34.2 (C), 31.1 (3CH₃). **HRMS (ESI⁺):** *m/z* calcd. for C₁₈H₁₉NO₂Na [M+Na]⁺ 304.1308, found 304.1306. The data are in agreement with those previously reported in the literature.³

3-([1,1'-Biphenyl]-4-yl)-3-hydroxyindolin-2-one **3d**



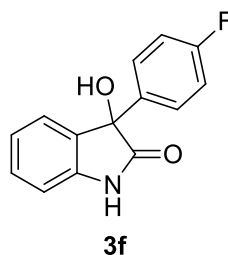
General procedure **B** was followed with isatin (1.0 g, 6.80 mmol) and [1,1'-biphenyl]-4-ylmagnesium bromide prepared from 4-bromobiphenyl (3.4 mL, 13.6 mmol, 2.0 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3d** as a white solid (1.1 g, 3.54 mmol, 52%). ¹H NMR (600 MHz, DMSO-d₆): δ 10.43 (s, 1H), 7.67-7.57 (m, 4H), 7.50-7.40 (m, 2H), 7.40-7.31 (m, 3H), 7.31-7.22 (m, 1H), 7.14 (d, *J* = 6.7 Hz, 1H), 7.03-6.87 (m, 2H), 6.68 (s, 1H). ¹³C NMR (150 MHz, DMSO-d₆): δ 178.4 (C=O), 142.0 (C), 140.7 (C), 139.9 (C), 139.4 (C), 133.7 (C), 129.3 (CH), 128.9 (2CH), 127.5 (CH), 126.7 (2CH), 126.5 (2CH), 126.1 (2CH), 124.8 (CH), 122.1 (CH), 109.9 (CH), 77.2 (C). HRMS (ESI⁺): *m/z* calcd. for C₂₀H₁₅NO₂Na [M+Na]⁺ 324.0995, found 324.0997. The data are in agreement with those previously reported in the literature.³

3-(4-Chlorophenyl)-3-hydroxyindolin-2-one **3e**



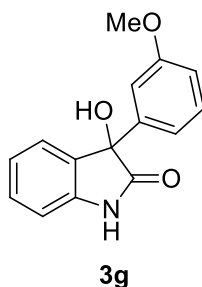
General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and chlorophenylmagnesium bromide (1.0 M in THF, 6.8 mL, 6.80 mmol, 2.0 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3e** as a beige solid (0.20 g, 0.77 mmol, 23%). ¹H NMR (600 MHz, DMSO-d₆): δ 10.45 (s, 1H), 7.32 (d, *J* = 8.5 Hz, 2H), 7.28-7.24 (m, 3H), 7.10 (d, *J* = 7.3 Hz, 1H), 6.98 (t, *J* = 7.5 Hz, 1H), 6.91 (d, *J* = 7.8 Hz, 1H), 6.74 (s, 1H). ¹³C NMR (150 MHz, DMSO-d₆): δ 178.1 (C=O), 142.0 (C), 140.5 (C), 133.2 (C), 132.2 (C), 129.5 (CH), 128.2 (2CH), 127.4 (2CH), 124.8 (CH), 122.2 (CH), 110.0 (CH), 76.9 (C). FT-IR (neat, cm⁻¹): 3189, 1714, 1620, 1489, 1471, 1396, 1181, 1102. HRMS (ESI⁺): *m/z* calcd. for C₁₄H₁₀ClNO₂Na [M+Na]⁺ 282.0292, found 282.0293.

3-(4-Fluorophenyl)-3-hydroxyindolin-2-one **3f**



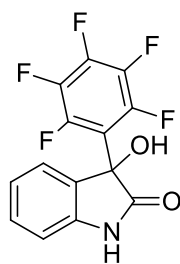
General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and fluorophenylmagnesium bromide (1.0 M in THF, 6.1 mL, 6.12 mmol, 1.8 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3f** as a light yellow solid (0.45 g, 1.86 mmol, 55%). **¹H NMR (600 MHz, DMSO-*d*₆):** δ 10.42 (s, 1H), 7.32-7.27 (m, 2H), 7.26 (td, $J = 7.7, 1.2$ Hz, 1H), 7.17-7.09 (m, 3H), 6.98 (td, $J = 7.5, 0.7$ Hz, 1H), 6.90 (d, $J = 7.7$ Hz, 1H), 6.69 (s, 1H). **¹³C NMR (150 MHz, DMSO-*d*₆):** δ 178.3 (C=O), 161.6 (d, $J^{\text{CF}} = 243.5$ Hz, C), 141.9 (C), 137.7 (d, $J^{\text{CF}} = 2.8$ Hz, C), 133.4 (C), 129.4 (CH), 127.6 (d, $J^{\text{CF}} = 8.3$ Hz, 2CH), 124.8 (CH), 122.2 (CH), 114.9 (d, $J^{\text{CF}} = 21.4$ Hz, 2CH), 110.0 (CH), 76.8 (C). **¹⁹F NMR (565 MHz, DMSO-*d*₆):** δ -115.2. **FT-IR (neat, cm⁻¹):** 3189, 1714, 1620, 1489, 1471, 1396, 1181, 1102. **HRMS (ESI⁺):** m/z calcd. for C₁₄H₁₀FN₂O₂Na [M+Na]⁺ 266.0588, found 266.0591. All analytical data were in accordance with literature.³

3-Hydroxy-3-(3-methoxyphenyl)indolin-2-one **3g**



General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and 3-methoxyphenylmagnesium bromide (1.0 M in 2-MeTHF, 4.8 mL, 4.76 mmol, 1.4 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3g** as a white solid (0.2 g, 0.78 mmol, 23%). **¹H NMR (400 MHz, DMSO-*d*₆):** δ 10.39 (s, 1H), 7.24 (td, $J = 7.7, 1.1$ Hz, 1H), 7.19 (t, $J = 8.0$ Hz, 1H), 7.09 (d, $J = 7.2$ Hz, 1H), 6.96 (m, 2H), 6.89 (d, $J = 7.7$ Hz, 1H), 6.83 (dd, $J = 8.0, 2.2$ Hz, 1H), 6.69 (d, $J = 7.8$ Hz, 1H), 6.62 (s, 1H), 3.72 (s, 3H). **¹³C NMR (100 MHz, DMSO-*d*₆):** δ 178.3 (C=O), 159.1 (C), 143.1 (C), 141.9 (C), 133.7 (C), 129.2 (CH), 129.2 (CH), 124.7 (CH), 122.0 (CH), 117.5 (CH), 112.5 (CH), 111.5 (CH), 109.8 (CH), 77.2 (C), 55.0 (CH₃). **FT-IR (neat, cm⁻¹):** 3405, 1703, 1615, 1607, 1585, 1485, 1467, 1433, 1338, 1285, 1245, 1179, 1149, 1119, 1098, 1076, 1044. **HRMS (ESI⁺):** m/z calcd. for C₁₅H₁₃NO₃Na [M+Na]⁺ 278.0788, found 278.0804. The data are in agreement with those previously reported in the literature.⁴

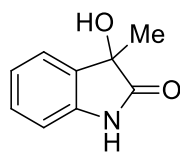
3-Hydroxy-3-(perfluorophenyl)indolin-2-one **3h**



3h

General procedure **B** was followed with isatin (0.50 g, 2.04 mmol) and (perfluorophenyl)magnesium bromide prepared from bromopentafluorobenzene (1.2 M in Et₂O, 6.0 mL, 6.12 mmol, 1.8 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3h** as a yellow solid (0.95 g, 3.01 mmol, 89%). **¹H NMR (700 MHz, DMSO-*d*₆):** δ 10.67 (brs, 1H), 7.27 (td, *J* = 7.7, 1.2 Hz, 1H), 7.24 (d, *J* = 7.4 Hz, 1H), 7.17 (s, 1H), 6.97 (td, *J* = 7.6, 0.9 Hz, 1H), 6.90 (d, *J* = 7.7 Hz, 1H). **¹³C NMR (176 MHz, DMSO-*d*₆):** δ 175.6 (C=O), 144.5 (dm, *J* = 251.6 Hz, 2C), 141.8 (C), 139.9 (dm, *J* = 252.0 Hz, C), 137.3 (dm, *J* = 248.3 Hz, 2C), 130.4 (CH), 130.4 (C), 124.8 (CH), 122.2 (CH), 115.6 (m, C), 110.2 (CH), 75.2 (C). **¹⁹F NMR (659 MHz, DMSO-*d*₆):** δ 139.5 (d, *J* = 19.9 Hz), -155.2 (t, *J* = 22.4 Hz), -162.5 (dd, *J* = 22.4, 19.9 Hz). **HRMS (ESI⁺):** *m/z* calcd. for C₁₄H₆F₅NO₂Na [M+Na]⁺ 338.0211, found 338.0208.

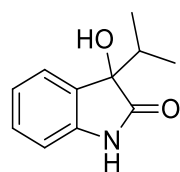
3-Hydroxy-3-methylindolin-2-one **3i**



3i

General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and methylmagnesium bromide (3.0 M in Et₂O, 2.3 mL, 6.80 mmol, 2.0 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3i** as a light yellow solid (0.39 g, 2.39 mmol, 70%). **¹H NMR (600 MHz, DMSO-*d*₆):** 10.19 (s, 1H), 7.28 (d, *J* = 7.3 Hz, 1H), 7.18 (t, *J* = 7.6 Hz, 1H), 6.96 (t, *J* = 7.4 Hz, 1H), 6.80 (d, *J* = 7.7 Hz, 1H), 5.84 (s, 1H), 1.35 (s, 3H). **¹³C NMR (150 MHz, DMSO-*d*₆):** δ 179.7 (C=O), 141.1 (C), 133.6 (C), 128.8 (CH), 123.4 (CH), 121.6 (CH), 109.6 (CH), 72.6 (C), 24.5 (CH₃). **HRMS (ESI⁺):** *m/z* calcd. for C₉H₉NO₂Na [M+Na]⁺ 186.0525, found 186.0524. The data are in agreement with those previously reported in the literature.⁵

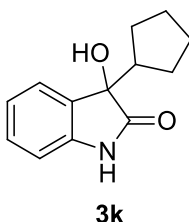
3-Hydroxy-3-isopropylindolin-2-one **3j**



3j

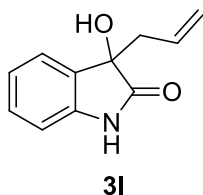
General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and isopropylmagnesium chloride (2.0 M in THF, 2.4 mL, 4.76 mmol, 1.4 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3j** as a light yellow solid (70 mg, 0.37 mmol, 11%). **¹H NMR (400 MHz, DMSO-*d*₆):** δ 10.19 (s, 1H), 7.23-7.17 (m, 2H), 6.94 (td, *J* = 7.6, 0.8 Hz, 1H), 6.79 (d, *J* = 7.6 Hz, 1H), 5.76 (s, 1H), 2.11-2.01 (m, 1H), 0.96 (d, *J* = 6.9 Hz, 3H), 0.63 (d, *J* = 6.9 Hz, 3H). **¹³C NMR (100 MHz, DMSO-*d*₆):** δ 179.5 (C=O), 142.2 (C), 130.6 (C), 128.7 (CH), 124.6 (CH), 121.3 (CH), 109.3 (CH), 78.3 (C), 34.8 (CH), 16.2 (CH₃), 15.8 (CH₃). **FT-IR (neat, cm⁻¹):** 3346, 1699, 1621, 1469, 1357, 1194, 1179, 1123, 1095, 1076. **HRMS (ESI⁺):** *m/z* calcd. for C₁₁H₁₃NO₂Na [M+Na]⁺ 214.0838, found 214.0853. The data are in agreement with those previously reported in the literature.⁶

3-Cyclopentyl-3-hydroxyindolin-2-one **3k**



General procedure **B** was followed with isatin (0.30 g, 2.04 mmol) and cyclopentylmagnesium bromide (2.0 M in THF, 1.4 mL, 2.85 mmol, 1.4 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3k** as a light orange solid (0.15 g, 0.69 mmol, 34%). **¹H NMR (500 MHz, CDCl₃):** δ 8.01 (brs, 1H), 7.40 (d, *J* = 7.3 Hz, 1H), 7.25 (td, *J* = 7.7, 1.2 Hz, 1H), 7.05 (t, *J* = 7.3 Hz, 1H), 6.87 (d, *J* = 7.7 Hz, 1H), 2.51-2.41 (m, 1H), 1.82-1.72 (m, 1H), 1.72-1.60 (m, 2H), 1.58-1.45 (m, 4H), 1.32-1.21 (m, 1H). **¹³C NMR (126 MHz, CDCl₃):** δ 180.7 (C=O), 140.7 (C), 130.2 (C), 129.6 (CH), 125.1 (CH), 123.0 (CH), 110.2 (CH), 78.6 (C), 47.7 (CH), 26.6 (CH₂), 26.5 (CH₂), 25.7 (CH₂), 25.5 (CH₂). **HRMS (ESI⁺):** *m/z* calcd. for C₁₃H₁₅NO₂Na [M+Na]⁺ 240.0995, found 240.0996.

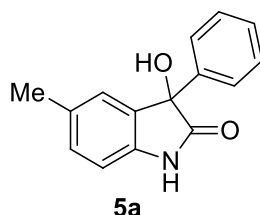
3-Allyl-3-hydroxyindolin-2-one **3l**



General procedure **B** was followed with isatin (0.50 g, 3.40 mmol) and allylmagnesium bromide (1.0 M in Et₂O, 6.8 mL, 6.80 mmol, 2.0 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **3k** as a light yellow solid (0.37 g, 1.94 mmol, 57%). **¹H NMR (400 MHz, CDCl₃):** δ 8.33 (brs, 1H), 7.37 (d, *J* = 7.5 Hz, 1H), 7.26 (dt, *J* = 8.8, 6.5 Hz, 1H), 7.07 (t, *J* = 7.5 Hz, 1H), 6.88 (d, *J* = 7.7 Hz, 1H), 5.73-5.61 (m, 1H), 5.15-5.08 (m, 2H), 3.36 (s, 1H), 2.75 (dd, *J* = 13.4, 6.4 Hz, 1H), 2.62 (dd, *J* = 13.4, 8.2 Hz, 1H). **¹³C NMR (100 MHz, CDCl₃):** δ 180.3 (C=O), 140.4 (C), 130.4 (CH), 130.3 (C), 129.8 (CH), 124.6 (CH), 123.2 (CH), 120.7 (CH₂), 110.5 (CH), 76.4 (C), 43.0 (CH₂). **FT-IR (neat, cm⁻¹):** 3322, 3221, 3064,

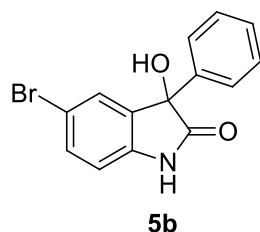
2714, 1681, 1624, 1472, 1355, 1274, 1188, 1110, 1091. **HRMS (ESI⁺):** m/z calcd. for C₁₁H₁₁NO₂Na [M+Na]⁺ 212.0682, found 212.0675.

3-Hydroxy-5-methyl-3-phenylindolin-2-one **5a**



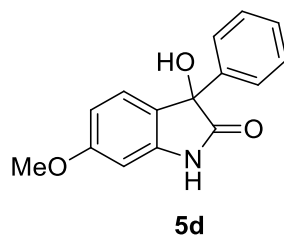
General procedure **B** was followed with 5-methylisatin (0.50 g, 3.10 mmol) and phenylmagnesium bromide (3.0 M in Et₂O, 2.1 mL, 6.21 mmol, 2.0 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **5a** as a white solid (0.46 g, 1.91 mmol, 61%). **¹H NMR (400 MHz, DMSO-*d*₆):** δ 10.30 (s, 1H), 7.33-7.23 (m, 5H), 7.04 (d, $J = 7.8$ Hz, 1H), 6.90 (s, 1H), 6.79 (d, $J = 7.8$ Hz, 1H), 6.57 (s, 1H), 2.21 (s, 3H). **¹³C NMR (100 MHz, DMSO-*d*₆):** δ 178.5 (C=O), 141.7 (C), 139.4 (C), 133.9 (C), 130.9 (C), 129.4 (CH), 128.1 (2CH), 127.3 (CH), 125.4 (2CH), 125.3 (CH), 109.6 (CH), 77.4 (C), 20.60 (CH₃). **FT-IR (neat, cm⁻¹):** 3209, 1697, 1624, 1491, 1202, 1190, 1149, 1139, 1128. **HRMS (ESI⁺):** m/z calcd. for C₁₅H₁₃NO₂Na [M+Na]⁺ 262.0838, found 262.0853. The data are in agreement with those previously reported in the literature.²

5-Bromo-3-hydroxy-3-phenylindolin-2-one **5b**



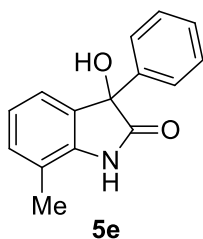
General procedure **B** was followed with 5-bromoisatin (0.32 g, 1.39 mmol) and phenylmagnesium bromide (3.0 M in Et₂O, 0.93 mL, 2.79 mmol, 2.0 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **5b** as a light yellow solid (0.30 mg, 0.98 mmol, 71%). **¹H NMR (600 MHz, DMSO-*d*₆):** δ 10.57 (brs, 1H), 7.44 (dd, $J = 8.3, 2.1$ Hz, 1H), 7.36-7.31 (m, 2H), 7.30-7.26 (m, 3H), 7.20 (d, $J = 2.1$ Hz, 1H), 6.88 (d, $J = 8.3$ Hz, 1H), 6.79 (s, 1H). **¹³C NMR (150 MHz, DMSO-*d*₆):** δ 178.0 (C=O), 141.3 (C), 140.8 (C), 136.2 (C), 132.0 (CH), 128.3 (2CH), 127.7 (CH), 127.4 (CH), 125.3 (2CH), 113.7 (C), 112.1 (CH), 77.4 (C). **HRMS (ESI⁺):** m/z calcd. for C₁₄H₁₀BrNO₂Na [M+Na]⁺ 325.9787, found 325.9786. The data are in agreement with those previously reported in the literature.³

3-Hydroxy-6-methoxy-3-phenylindolin-2-one **5d**



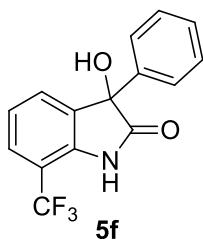
General procedure **B** was followed with 6-methoxyisatin (0.30 g, 1.69 mmol) and phenylmagnesium bromide (3.0 M in Et₂O, 0.84 mL, 1.49 mmol, 1.5 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **5d** as a light yellow solid (0.13 g, 0.51 mmol, 30%). **¹H NMR (600 MHz, DMSO)** δ 10.34 (brs, 1H), 7.32-7.28 (m, 2H), 7.28-7.22 (m, 3H), 6.99 (d, *J* = 8.2 Hz, 1H), 6.51 (dd, *J* = 8.2, 2.3 Hz, 1H), 6.49 (brs, 1H), 6.45 (d, *J* = 2.3 Hz, 1H), 3.75 (s, 3H). **¹³C NMR (150 MHz, DMSO)** δ 178.9 (C=O), 160.3 (C), 143.3 (C), 141.9 (C), 128.0 (2CH), 127.3 (CH), 125.7 (CH), 125.7 (C), 125.5 (2CH), 106.8 (CH), 96.6 (CH), 77.0 (C), 55.3 (CH₃). **HRMS (ESI⁺):** *m/z* calcd. for C₁₅H₁₃NO₃Na [M+Na]⁺ 278.0788, found 278.0790. The data are in agreement with those previously reported in the literature.⁷

3-Hydroxy-7-methyl-3-phenylindolin-2-one **5e**



General procedure **B** was followed with 7-methylisatin (0.20 g, 1.24 mmol) and phenylmagnesium bromide (3.0 M in Et₂O, 0.50 mL, 1.49 mmol, 1.2 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **5e** as a light yellow solid (0.12 g, 0.54 mmol, 44%). **¹H NMR (400 MHz, DMSO-*d*₆):** δ 10.43 (s, 1H), 7.33-7.22 (m 5H), 7.07 (d, *J* = 7.0 Hz, 1H), 6.93-6.84 (m, 2H), 6.57 (s, 1H), 2.25 (s, 3H). **¹³C NMR (100 MHz, DMSO-*d*₆):** δ 178.9 (C=O), 141.7 (C), 140.4 (C), 133.4 (C), 130.4 (CH), 128.0 (2CH), 127.3 (CH), 125.4 (2CH), 122.1 (CH), 122.0 (CH), 119.2 (C), 77.5 (C), 16.4 (CH₃). **FT-IR (neat, cm⁻¹):** 3165, 1720, 1626, 1606, 1486, 1448, 1381, 1300, 1209, 1191, 1163, 1118, 1102, 1070. **HRMS (ESI⁺):** *m/z* calcd. for C₁₅H₁₃NO₂Na [M+Na]⁺ 262.0838, found 262.0852. The data are in agreement with those previously reported in the literature.⁷

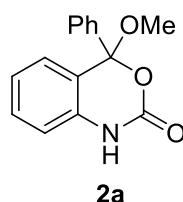
3-Hydroxy-3-phenyl-7-(trifluoromethyl)indolin-2-one **5f**



General procedure **B** was followed with 7-(trifluoromethyl)isatin (0.30 g, 1.39 mmol) and phenylmagnesium bromide (3.0 M in Et₂O, 0.93 mL, 2.79 mmol, 2.0 equiv.). Purification was performed with flash column chromatography over silica gel (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **5f** as a light yellow solid (0.20 g, 0.70 mmol, 50%). **¹H NMR (600 MHz, DMSO-*d*₆):** δ 10.92 (brs, 1H), 7.56 (d, *J* = 8.0 Hz, 1H), 7.38 (d, *J* = 7.3 Hz, 1H), 7.36-7.31 (m, 2H), 7.31-7.24 (m, 3H), 7.16 (t, *J* = 7.7 Hz, 1H), 6.85 (s, 1H). **¹³C NMR (150 MHz, CDCl₃):** 178.8 (C=O), 140.6 (C), 139.4 (q, *J*^{CF} = 2.2 Hz, C), 135.6 (C), 128.9 (CH), 128.3 (2CH), 127.8 (CH), 125.7 (q, *J*^{CF} = 4.6 Hz, CH), 125.4 (2CH), 123.6 (q, *J*^{CF} = 271.8 Hz, C), 122.4 (CH), 111.0 (q, *J*^{CF} = 32.8 Hz, C), 76.0 (C). **¹⁹F NMR (471 MHz, CDCl₃):** δ -59.9. **FT-IR (neat, cm⁻¹):** 3219, 2921, 2851, 2360, 2342, 1720, 1614, 1443, 1317, 1198. **HRMS (ESI⁺):** *m/z* calcd. for C₁₅H₁₀F₃NO₂Na [M+Na]⁺ 316.0556, found 316.0556.

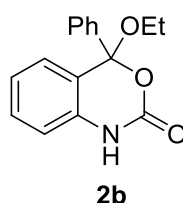
4 Characterization data of 3,1-Benzoxazin-2-ones 2a-g, 4a-l and 6a-f

4-Methoxy-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one 2a



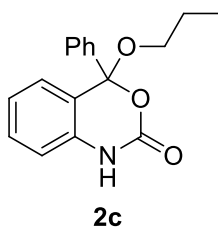
General procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2a** as a white solid (92.7 mg, 0.36 mmol, 91%). **¹H NMR (400 MHz, CDCl₃):** δ 9.01 (brs, 1H), 7.55-7.51 (m, 2H), 7.45-7.38 (m, 3H), 7.34 (ddd, $J = 8.4, 7.4, 1.8$ Hz, 1H), 7.08-7.01 (m, 2H), 6.95 (d, $J = 8.4$ Hz, 1H), 3.44 (s, 3H). **¹³C NMR (100 MHz, CDCl₃):** δ 151.6 (C=O), 138.6 (C), 135.1 (C), 130.6 (CH), 129.3 (CH), 128.6 (2CH), 126.9 (3CH), 123.7 (CH), 121.0 (C), 114.8 (CH), 107.5 (C), 52.0 (CH₃). **FT-IR (neat, cm⁻¹):** 1709, 1603, 1492, 1449, 1434, 1346, 1256, 1215, 1177, 1093, 1076, 1007. **HRMS (ESI⁺):** m/z calcd. for C₁₅H₁₃NO₃Na [M+Na]⁺ 278.0788, found 278.0789.

4-Ethoxy-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one 2b



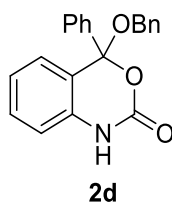
General procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and EtOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2b** as a white solid (83.3 mg, 0.31 mmol, 77%). **¹H NMR (400 MHz, CDCl₃):** δ 9.73 (brs, 1H), 7.59-7.52 (m, 2H), 7.44-7.37 (m, 3H), 7.32 (ddd, $J = 8.4, 7.4, 1.8$ Hz, 1H), 7.08-7.02 (m, 2H), 6.99 (d, $J = 8.4$ Hz, 1H), 3.71 (qd, $J = 7.1, 2.2$ Hz, 2H), 1.28 (t, $J = 7.1$ Hz, 3H). **¹³C NMR (100 MHz, CDCl₃):** δ 152.3 (C=O), 139.3 (C), 136.0 (C), 130.4 (CH), 129.2 (CH), 128.5 (2CH), 126.8 (2CH), 126.8 (CH), 123.6 (CH), 121.3 (C), 114.9 (CH), 107.5 (C), 60.5 (CH₂), 15.4 (CH₃). **FT-IR (neat, cm⁻¹):** 1710, 1601, 1490, 1449, 1342, 1255, 1215, 1135, 1095, 1076, 1026, 1002. **HRMS (ESI⁺):** m/z calcd. for C₁₆H₁₅NO₃Na [M+Na]⁺ 292.0944, found 292.0941.

4-Phenyl-4-propoxy-1H-benzo[d][1,3]oxazin-2(4H)-one 2c



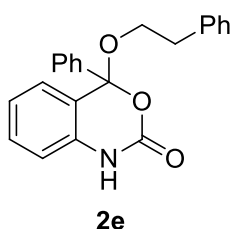
General procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and *n*PrOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2c** as a white solid (89.3 mg, 0.32 mmol, 79%). **¹H NMR (400 MHz, CDCl₃):** δ 9.72 (brs, 1H), 7.55-7.52 (m, 2H), 7.43-7.36 (m, 3H), 7.30 (ddd, *J* = 8.4, 7.4, 1.8 Hz, 1H), 7.08-7.00 (m, 2H), 6.99 (d, *J* = 8.4 Hz, 1H), 3.58 (qt, *J* = 8.8, 7.4 Hz, 2H), 1.73-1.63 (m, 2H), 0.95 (t, *J* = 7.4 Hz, 3H). **¹³C NMR (100 MHz, CDCl₃):** δ 152.2 (C=O), 139.6 (C), 136.1 (C), 130.4 (CH), 129.2 (CH), 128.5 (2CH), 126.9 (CH), 126.8 (2CH), 123.6 (CH), 121.2 (C), 114.9 (CH), 107.4 (C), 66.2 (CH₂), 23.1 (CH₂), 10.7 (CH₃). **FT-IR (neat, cm⁻¹):** 1708, 1601, 1491, 1449, 1434, 1341, 1254, 1216, 1180, 1133, 1094, 1077, 1036, 1002. **HRMS (ESI⁺):** *m/z* calcd. for C₁₇H₁₇NO₃Na [M+Na]⁺ 306.1101, found 306.1102.

4-(Benzyloxy)-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one 2d



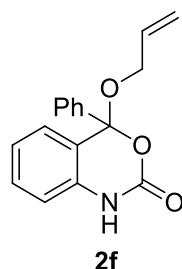
The general procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and BzOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2d** as a white solid (64.6 mg, 0.12 mmol, 49%). **¹H NMR (400 MHz, CDCl₃):** δ 9.77 (brs, 1H), 7.54-7.47 (m, 2H), 7.36-7.29 (m, 3H), 7.28-7.10 (m, 6H), 7.04 (d, *J* = 7.5 Hz, 1H), 6.98-6.87 (m, 2H), 4.62 (d, *J* = 2.7 Hz, 2H). **¹³C NMR (100 MHz, CDCl₃):** δ 152.1 (C=O), 139.1 (C), 137.1 (C), 135.1 (C), 130.6 (CH), 129.4 (CH), 128.6 (2CH), 128.5 (2CH), 127.9 (3CH), 126.9 (3CH), 123.7 (CH), 120.9 (C), 115.0 (CH), 107.5 (C), 66.6 (CH₂). **HRMS (ESI⁺):** *m/z* calcd. for C₂₁H₁₇NO₃Na [M+Na]⁺ 354.1101, found 354.1101.

4-Phenethoxy-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one 2e



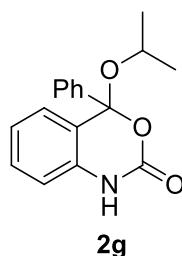
General procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and 2-phenylethanol (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2e** as a white solid (46.2 mg, 0.14 mmol, 33%). **¹H NMR (400 MHz, CDCl₃):** δ 9.24 (brs, 1H), 7.39-7.34 (m, 2H), 7.31-7.27 (m, 3H), 7.24-7.17 (m, 3H), 7.14-7.09 (m, 3H), 6.91 (ddd, *J* = 8.4, 7.4, 1.8 Hz, 1H), 6.88-6.84 (m, 2H), 3.82-3.72 (m, 2H), 2.90 (t, *J* = 7.1 Hz, 2H). **¹³C NMR (100 MHz, CDCl₃):** δ 151.8 (C=O), 139.3 (C), 138.5 (C), 136.0 (C), 130.4 (CH), 129.2 (2CH), 129.2 (CH), 128.6 (2CH), 128.5 (2CH), 126.8 (CH), 126.8 (2CH), 126.5 (CH), 123.6 (CH), 121.0 (C), 114.8 (CH), 107.4 (C), 66.5 (CH₂), 36.4 (CH₂). **HRMS (ESI⁺):** *m/z* calcd. for C₂₂H₁₉NO₃Na [M+Na]⁺ 368.1257, found 368.1256.

4-(Allyloxy)-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one **2f**



General procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and allyl alcohol (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2f** as a white solid (86.6 mg, 0.31 mmol, 77%). **¹H NMR (400 MHz, CDCl₃):** δ 9.31 (brs, 1H), 7.57-7.53 (m, 2H), 7.44-7.38 (m, 3H), 7.31 (ddd, *J* = 8.4, 7.4, 1.8 Hz, 1H), 7.08-7.00 (m, 2H), 6.96 (d, *J* = 8.4 Hz, 1H), 5.96 (ddt, *J* = 17.2, 10.5, 5.5 Hz, 1H), 5.35-5.29 (m, 1H), 5.21-5.16 (m, 1H), 4.18 (ddd, *J* = 5.5, 1.7, 1.6 Hz, 2H). **¹³C NMR (100 MHz, CDCl₃):** δ 151.8 (C=O), 138.9 (C), 136.0 (C), 133.7 (CH), 130.5 (CH), 129.3 (CH), 128.6 (2CH), 126.9 (3CH), 123.7 (CH), 121.2 (C), 117.5 (CH₂), 114.9 (CH), 107.4 (C), 66.9 (CH₂). **FT-IR (neat, cm⁻¹):** 1709, 1602, 1491, 1449, 1347, 1256, 1213, 1133, 1105, 1090, 1074, 1036, 1005. **HRMS (ESI⁺):** *m/z* calcd. for C₁₇H₁₅NO₃Na [M+Na]⁺ 304.0944, found 304.0942.

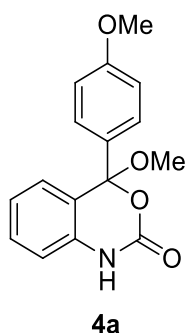
4-Isopropoxy-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one **2g**



General procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and *i*PrOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2g** as a brown solid (57.8 mg, 0.20 mmol, 51%). **¹H NMR (400 MHz, CDCl₃):** δ 9.79 (brs, 1H), 7.57-7.50 (m, 2H), 7.42-7.35 (m, 3H), 7.29 (ddd, *J* = 7.6, 7.6, 1.4 Hz, 1H), 7.15 (dd, *J* = 7.6, 1.4 Hz, 1H), 7.02 (ddd, *J* = 7.6, 7.6, 1.1

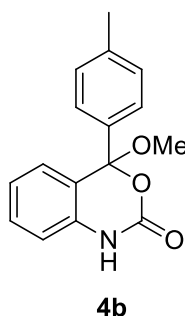
Hz, 1H), 6.98 (dd, $J = 7.6, 1.1$ Hz, 1H) 4.12-4.02 (m, 1H), 1.30 (d, $J = 6.1$ Hz, 3H), 1.23 (d, $J = 6.1$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3): δ 152.3 (C=O), 140.2 (C), 135.0 (C), 130.4 (CH), 129.1 (CH), 128.4 (2CH), 127.1 (CH), 126.9 (2CH), 123.4 (CH), 121.4 (C), 114.9 (CH), 108.3 (C), 69.2 (CH), 24.2 (2CH₃). FT-IR (neat, cm^{-1}): 1707, 1599, 1494, 1361, 1258, 1202, 1068, 1012. HRMS (ESI⁺): m/z calcd. for $\text{C}_{17}\text{H}_{17}\text{NO}_3\text{Na}$ $[\text{M}+\text{Na}]^+$ 306.1101, found 306.1105.

4-Methoxy-4-(4-methoxyphenyl)-1H-benzo[d][1,3]oxazin-2(4H)-one 4a



The general procedure **A** was followed with **3a** (102 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO_2 , heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4a** as a light brown solid (98.7 mg, 0.35 mmol, 87%). ^1H NMR (400 MHz, CDCl_3): δ 8.79 (brs, 1H), 7.43 (d, $J = 8.9$ Hz, 2H), 7.31 (ddd, $J = 8.0, 7.0, 1.9$ Hz, 1H), 7.10-7.00 (m, 2H), 6.94-6.90 (m, 3H), 3.82 (s, 3H), 3.43 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3): δ 160.3 (C=O), 151.6 (C), 136.1 (C), 130.9 (C), 130.5 (CH), 128.4 (2CH), 126.9 (CH), 123.6 (CH), 121.3 (C), 114.7 (CH), 113.9 (2CH), 107.6 (C), 55.5 (CH₃), 52.0 (CH₃). FT-IR (neat, cm^{-1}): 1700, 1604, 1513, 1505, 1494, 1432, 1353, 1319, 1305, 1251, 1209, 1185, 1169, 1153, 1095, 1032, 1009. HRMS (ESI⁺): m/z calcd. for $\text{C}_{16}\text{H}_{15}\text{NO}_4\text{Na}$ $[\text{M}+\text{Na}]^+$ 308.0893, found 308.0887.

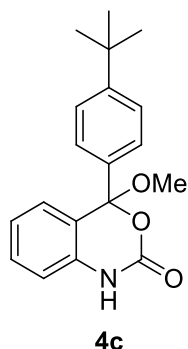
4-Methoxy-4-(p-tolyl)-1H-benzo[d][1,3]oxazin-2(4H)-one 4b



General procedure **A** was followed with **3b** (95.7 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO_2 , heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4b** as a light yellow solid (99.0 mg, 0.37 mmol, 92%). ^1H NMR (400 MHz, CDCl_3): δ 9.66 (brs, 1H), 7.40 (d, $J = 8.3$ Hz, 2H), 7.30 (td, $J = 8.0, 1.5$ Hz, 1H), 7.22 (d, $J = 8.0$ Hz, 2H), 7.06 (dd, $J = 7.7, 1.5$ Hz, 1H), 7.02 (td, $J = 7.7, 1.0$ Hz, 1H), 6.99 (dd, $J = 8.0, 1.0$ Hz, 1H), 3.43 (s, 3H), 2.37 (s, 3H). ^{13}C NMR

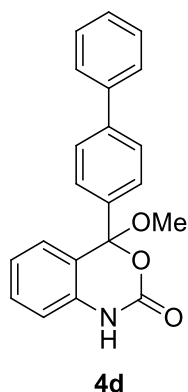
(150 MHz, CDCl₃): δ 152.3 (C=O), 139.3 (C), 135.9 (C), 135.2 (C), 130.5 (CH), 129.3 (2CH), 126.9 (2CH), 126.8 (CH), 123.7 (CH), 121.1 (C), 115.0 (CH), 107.7 (C), 52.0 (CH₃), 21.4 (CH₃). FT-IR (neat, cm⁻¹): 3096, 3004, 2924, 2360, 2342, 1714, 1603, 1493, 1345, 1255, 1182. HRMS (ESI⁺): m/z calcd. for C₁₆H₁₅NO₃Na [M+Na]⁺ 292.0944, found 292.0943.

4-(4-(Tert-butyl)phenyl)-4-methoxy-1H-benzo[d][1,3]oxazin-2(4H)-one 4c



The general procedure **A** was followed with **3c** (113 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4c** as a light yellow solid (89.9 mg, 0.29 mmol, 72%). ¹H NMR (600 MHz, DMSO-d₆): δ 10.62 (brs, 1H), 7.46 (d, J = 8.5 Hz, 2H), 7.36-7.29 (m, 3H), 7.02 (t, J = 7.5 Hz, 1H), 7.00-6.95 (m, 2H), 3.25 (s, 3H), 1.27 (s, 9H). ¹³C NMR (150 MHz, DMSO-d₆): δ 151.5 (C=O), 149.3 (C), 136.7 (C), 135.8 (C), 130.3 (CH), 126.3 (CH), 125.9 (2CH), 125.3 (2CH), 122.7 (CH), 120.1 (C), 114.4 (CH), 106.0 (C), 51.0 (CH₃), 34.4 (C), 31.0 (3CH₃). FT-IR (neat, cm⁻¹): 3100, 2950, 2359, 2342, 1720, 1602, 1494, 1347, 1275, 1263, 1258. HRMS (ESI⁺): m/z calcd. for C₁₉H₂₂NO₃ [M+H]⁺ 312.1594, found 312.1595.

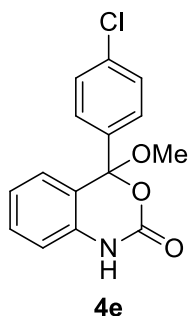
4-([1,1'-Biphenyl]-4-yl)-4-methoxy-1H-benzo[d][1,3]oxazin-2(4H)-one 4d



The general procedure **A** was followed with **3d** (121 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4d** as a light yellow solid (91.1 mg, 0.28 mmol, 69%). ¹H NMR (600 MHz, DMSO-d₆): δ 10.69 (brs, 1H), 7.74 (d, J = 8.4 Hz, 2H), 7.68 (d, J = 7.3 Hz, 2H), 7.50 (d, J = 8.4 Hz, 2H), 7.48 (t, J = 7.7 Hz, 2H), 7.39 (t, J = 7.3 Hz, 1H), 7.35 (ddd, J = 8.6, 6.1, 2.8 Hz, 1H), 7.08-7.02 (m, 2H), 7.00 (d, J = 8.1

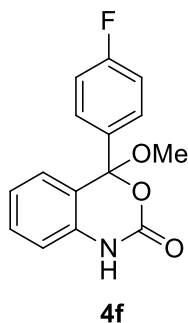
Hz, 1H), 3.29 (s, 3H). ^{13}C NMR (150 MHz, DMSO- d_6): δ 149.2 (C=O), 140.8 (C), 139.3 (C), 138.6 (C), 135.8 (C), 130.4 (CH), 129.0 (2CH), 127.8 (CH), 126.8 (2CH), 126.8 (2CH), 126.8 (2CH), 126.3 (CH), 122.8 (CH), 119.9 (C), 114.5 (CH), 105.9 (C), 51.0 (CH₃). FT-IR (neat, cm^{-1}): 3006, 2989, 2360, 2342, 1716, 1603, 1492, 1343, 1275, 1258, 1216, 1186. HRMS (ESI⁺): m/z calcd. for C₂₁H₁₇NO₃Na [M+Na]⁺ 354.1101, found 354.1097.

4-(4-Chlorophenyl)-4-methoxy-1H-benzo[d][1,3]oxazin-2(4H)-one **4e**



General procedure **A** was followed with **3e** (104 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4e** as a light yellow solid (60.0 mg, 0.21 mmol, 52%). ^1H NMR (600 MHz, CDCl₃): δ 9.66 (s, 1H), 7.46 (d, J = 8.6 Hz, 2H), 7.38 (d, J = 8.6 Hz, 2H), 7.33 (td, J = 7.6, 1.5 Hz, 1H), 7.07–7.03 (m, 1H), 7.03–6.97 (m, 2H), 3.42 (s, 3H). ^{13}C NMR (150 MHz, CDCl₃): δ 151.8 (C=O), 137.5 (C), 135.4 (C), 135.1 (C), 130.8 (CH), 128.8 (2CH), 128.4 (2CH), 126.8 (CH), 123.8 (CH), 120.1 (C), 115.0 (CH), 107.1 (C), 52.0 (CH₃). FT-IR (neat, cm^{-1}): 3093, 2989, 2925, 2359, 2342, 1703, 1601, 1491, 1343, 1267, 1253, 1219. HRMS (ESI⁺): m/z calcd. for C₁₅H₁₂ClNO₃Na [M+Na]⁺ 312.0398, found 312.0398.

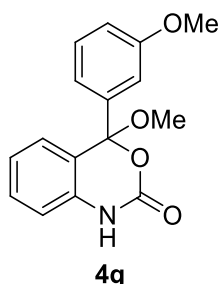
4-(4-Fluorophenyl)-4-methoxy-1,4-dihydro-benzo[d][1,3]oxazin-2-one **4f**



General procedure **A** was followed with **3f** (97.3 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4f** as a light yellow solid (88.5 mg, 0.32 mmol, 81%). ^1H NMR (700 MHz, DMSO- d_6): δ 10.70 (brs, 1H), 7.48–7.45 (m, 2H), 7.35 (dd, J = 7.8, 1.5 Hz, 1H), 7.29–7.25 (m, 2H), 7.02 (dd, J = 8.0, 1.1 Hz, 1H), 6.99 (d, J = 8.0 Hz, 1H), 6.94 (dd, J = 7.8, 1.1 Hz, 1H), 3.25 (s, 3H). ^{13}C NMR (176 MHz, DMSO): δ 162.7 (d, J^{CF} = 245.8 Hz, C), 149.1 (C=O), 135.8 (d, J^{CF} = 4.0 Hz, C), 135.8 (C), 130.5 (CH), 128.6 (d, J^{CF} = 8.6 Hz, 2CH), 126.3 (CH), 122.8 (CH), 119.7 (C), 115.4 (d, J^{CF} = 21.7 Hz, 2CH),

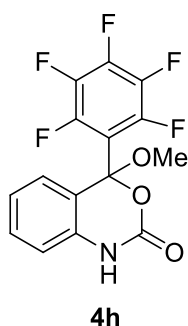
114.5 (CH), 105.5 (C), 51.0 (CH₃). ¹⁹F NMR (659 MHz, DMSO-d₆): δ -113.0. FT-IR (neat, cm⁻¹): 3094, 2990, 2930, 2360, 2332, 1716, 1708, 1602, 1507, 1494, 1345, 1276, 1228, 1217. HRMS (ESI⁺): *m/z* calcd. for C₁₅H₁₂FNO₃Na [M+Na]⁺ 296.0693, found 296.0692.

4-Methoxy-4-(3-methoxyphenyl)-1H-benzo[d][1,3]oxazin-2(4H)-one **4g**



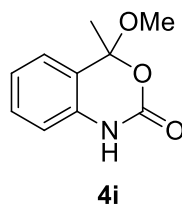
The general procedure **A** was followed with **3g** (102 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4g** as a light brown solid (61.3 mg, 0.22 mmol, 54%). ¹H NMR (600 MHz, CDCl₃): δ 9.38 (brs, 1H), 7.35-7.29 (m, 2H), 7.10-7.07 (m, 3H), 7.03 (ddd, *J* = 8.0, 7.0, 1.9 Hz, 1H), 6.97 (dd, *J* = 8.0, 0.9 Hz, 1H), 6.93 (ddd, *J* = 8.1, 2.5, 0.9, 1H), 3.82 (s, 3H), 3.45 (s, 3H). ¹³C NMR (150 MHz, CDCl₃): δ 159.9 (C=O), 151.8 (C), 140.0 (C), 135.0 (C), 130.6 (CH), 129.6 (CH), 126.8 (CH), 123.7 (CH), 120.8 (C), 119.3 (CH), 115.0 (CH), 114.9 (CH), 112.6 (CH), 107.3 (C), 55.5 (CH₃), 52.0 (CH₃). FT-IR (neat, cm⁻¹): 1702, 1604, 1589, 1486, 1433, 1355, 1313, 1293, 1273, 1263, 1200, 1171, 1086, 1052, 1041, 1015. HRMS (ESI⁺): *m/z* calcd. for C₁₆H₁₅NO₄Na [M+Na]⁺ 308.0893, found 308.0898.

4-Methoxy-4-(perfluorophenyl)-1H-benzo[d][1,3]oxazin-2(4H)-one **4h**



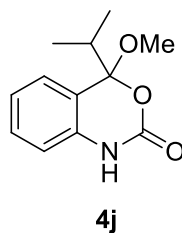
General procedure **A** was followed with **3h** (126 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4h** as a light yellow solid (27.0 mg, 0.078 mmol, 20%). ¹H NMR (700 MHz, CDCl₃): δ 9.07 (brs, 1H), 7.38-7.35 (m, 1H), 7.13-7.04 (m, 2H), 6.94 (d, *J* = 8.0 Hz, 1H), 3.45 (s, 3H). ¹³C NMR (176 MHz, CDCl₃): δ 149.9 (C=O), 144.9 (dm, *J* = 257.1 Hz, 2C), 141.8 (dm, *J* = 257.2 Hz, C), 138.2 (dm, *J* = 251.7 Hz, 2C), 134.5 (C), 131.5 (CH), 126.1 (CH), 124.3 (CH), 118.0 (C), 114.9 (CH), 114.8 (m, C), 105.7 (C), 51.8 (CH₃). ¹⁹F NMR (659 MHz, CDCl₃): δ -139.1 (m), -151.7 (m), -160.7 (m). FT-IR (neat, cm⁻¹): 3240, 3111, 2931, 1716, 1606, 1521, 1489, 1353, 1309, 1273, 1252, 1148. HRMS (ESI⁺): *m/z* calcd. for C₁₅H₈F₅NO₃Na [M+Na]⁺ 368.0317, found 368.0317.

4-Methoxy-4-methyl-1H-benzo[d][1,3]oxazin-2(4H)-one **4i**



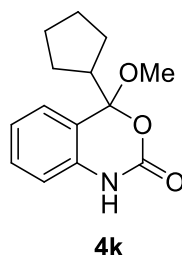
General procedure **A** was followed with **3i** (65.3 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4i** as a light yellow solid (37.8 mg, 0.20 mmol, 49%). **¹H NMR (400 MHz, CDCl₃):** δ 9.81 (s, 1H), 7.31 (td, $J = 7.7, 1.4$ Hz, 1H), 7.27 (dd, $J = 7.6, 1.6$ Hz, 1H), 7.12 (td, $J = 7.6, 1.0$ Hz, 1H), 6.93 (dt, $J = 7.9$ Hz, 1H), 3.31 (s, 3H), 1.87 (s, 3H). **¹³C NMR (100 MHz, CDCl₃):** δ 152.3 (C=O), 134.8 (C), 130.5 (CH), 125.2 (CH), 123.8 (CH), 120.4 (C), 114.9 (CH), 107.0 (C), 51.1 (CH₃), 27.1 (CH₃). **FT-IR (neat, cm⁻¹):** 3094, 2923, 2359, 2332, 1704, 1601, 1493, 1349, 1274, 1258. **HRMS (ESI⁺):** m/z calcd. for C₁₀H₁₁NO₃Na [M+Na]⁺ 216.0631, found 216.0629.

4-Isopropyl-4-methoxy-1H-benzo[d][1,3]oxazin-2(4H)-one **4j**



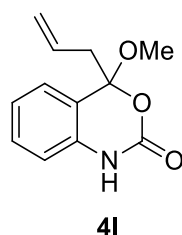
General procedure **A** was followed with **3j** (76.5 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4j** as a light orange solid (52.0 mg, 0.25 mmol, 59%). **¹H NMR (600 MHz, CDCl₃):** δ 9.32 (brs, 1H), 7.31 (td, $J = 7.8, 1.4$ Hz, 1H), 7.20 (dd, $J = 7.0, 0.7$ Hz, 1H), 7.10 (td, $J = 7.6, 1.0$ Hz, 1H), 6.91 (dd, $J = 8.0, 0.7$ Hz, 1H), 3.21 (s, 3H), 2.27 (m, 1H), 1.08 (d, $J = 6.8$ Hz, 3H), 0.87 (d, $J = 6.8$ Hz, 3H). **¹³C NMR (150 MHz, CDCl₃):** δ 151.3 (C=O), 138.1 (C), 129.4 (CH), 125.1 (CH), 122.5 (CH), 116.9 (C), 113.5 (CH), 111.0 (C), 50.6 (CH₃), 38.9 (CH), 15.7 (CH₃), 14.5 (CH₃). **FT-IR (neat, cm⁻¹):** 1703, 1601, 1494, 1461, 1434, 1373, 1332, 1271, 1252, 1248, 1143, 1132, 1096, 1042, 1012. **HRMS (ESI⁺):** m/z calcd. for C₁₂H₁₅NO₃Na [M+Na]⁺ 244.0944, found 244.0947.

4-Cyclopentyl-4-methoxy-1H-benzo[d][1,3]oxazin-2(4H)-one **4k**



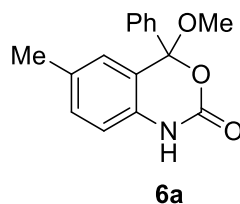
General procedure **A** was followed with **3k** (86.9 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4k** as a light orange solid (35.1 mg, 0.14 mmol, 35%). **¹H NMR (700 MHz, CDCl₃):** δ 9.57 (brs, 1H), 7.29 (td, *J* = 7.9, 1.4 Hz, 1H), 7.23 (d, *J* = 7.6 Hz, 1H), 7.09 (td, *J* = 7.6, 1.0 Hz, 1H), 6.93 (dd, *J* = 8.0, 0.6 Hz, 1H), 3.21 (s, 3H), 2.61-2.56 (m, 1H), 1.83 (dd, *J* = 12.9, 8.4 Hz, 1H), 1.74-1.70 (m, 1H), 1.66-1.63 (m, 1H), 1.62-1.57 (m, 1H), 1.55-1.44 (m, 3H), 1.42-1.36 (m, 1H). **¹³C NMR (176 MHz, CDCl₃)** δ 152.5 (C=O), 135.7 (C), 130.3 (CH), 126.0 (CH), 123.6 (CH), 118.8 (C), 114.7 (CH), 111.4 (C), 51.5 (CH₃), 51.1 (CH₃), 27.2 (CH₂), 26.1 (CH₂), 25.9 (CH₂), 25.6 (CH₂). **FT-IR (neat, cm⁻¹):** 2922, 2359, 2333, 1703, 1602, 1493, 1346, 1247. **HRMS (ESI⁺):** *m/z* calcd. for C₁₄H₁₇NO₃Na [M+Na]⁺ 270.1101, found 270.1100.

4-Allyl-4-methoxy-1H-benzo[d][1,3]oxazin-2(4H)-one **4l**



The general procedure **A** was followed with **3l** (75.7 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4l** as a light yellow solid (14.0 mg, 0.064 mmol, 16%). **¹H NMR (700 MHz, CDCl₃):** δ 9.47 (brs, 1H), 7.31 (ddd, *J* = 7.9, 6.3, 1.3 Hz, 1H), 7.24 (d, *J* = 7.7 Hz, 1H), 7.13-7.08 (m, 1H), 6.92 (d, *J* = 7.9 Hz, 1H), 5.66 (ddt, *J* = 17.3, 10.2, 7.2 Hz, 1H), 5.12-5.09 (m, 1H), 5.09-5.07 (m, 1H), 3.28 (s, 3H), 2.87 (dd, *J* = 14.3, 7.5 Hz, 1H), 2.81 (dd, *J* = 14.3, 6.9 Hz, 1H). **¹³C NMR (176 MHz, CDCl₃):** δ 151.7 (C=O), 135.4 (C), 130.4 (CH), 130.2 (CH), 125.7 (CH), 123.5 (CH), 120.4 (C), 118.1 (C), 114.5 (CH), 108.5 (CH₂), 51.2 (CH), 45.9 (CH₂). **FT-IR (neat, cm⁻¹):** 3246, 3103, 2982, 2938, 1704, 1602, 1494, 1358, 1256, 1148. **HRMS (ESI⁺):** *m/z* calcd. for C₁₂H₁₃NO₃Na [M+Na]⁺ 242.0788, found 242.0789.

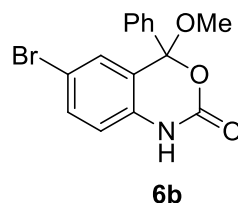
4-Methoxy-6-methyl-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one **6a**



The general procedure **A** was followed with **5a** (95.7 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **6a** as a white solid (35.7 mg, 0.13 mmol, 33%). **¹H NMR (400 MHz, CDCl₃):** δ 9.08 (brs, 1H), 7.52 (dd, *J* = 7.7, 1.6 Hz, 2H), 7.45-7.38 (m, 3H), 7.11 (d, *J* = 8.2 Hz, 1H), 6.84 (d, *J* = 7.8 Hz, 2H), 3.43 (s, 3H), 2.24 (s, 3H). **¹³C NMR (100 MHz, CDCl₃):** δ 151.7 (C=O), 138.8 (C), 133.4 (C), 132.7 (C), 131.2 (CH), 129.3

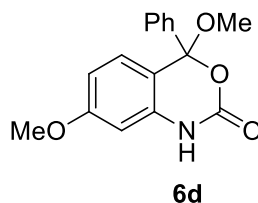
(CH), 128.6 (2CH), 127.0 (CH), 126.9 (2CH), 120.6 (C), 114.7 (CH), 107.5 (C), 51.9 (CH₃), 21.0 (CH₃). **FT-IR (neat, cm⁻¹):** 1708, 1607, 1512, 1450, 1347, 1251, 1226, 1195, 1164, 1095, 1075, 1008. **HRMS (ESI⁺):** *m/z* calcd. for C₁₆H₁₅NO₃Na [M+Na]⁺ 292.0944, found 292.0941.

6-Bromo-4-methoxy-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one **6b**



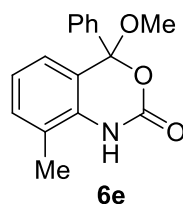
The general procedure **A** was followed with **5b** (122 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **6b** as a light yellow solid (60.9 mg, 0.18 mmol, 46%). **¹H NMR (500 MHz, DMSO-*d*₆):** δ 10.85 (brs, 1H), 7.54 (dd, *J* = 8.6, 2.2 Hz, 1H), 7.52-7.42 (m, 5H), 7.03 (d, *J* = 8.6 Hz, 1H), 6.95 (d, *J* = 2.2 Hz, 1H), 3.27 (s, *J* = 6.6 Hz, 3H). **¹³C NMR (126 MHz, DMSO-*d*₆):** δ 148.8 (C=O), 138.5 (C), 135.2 (C), 133.3 (CH), 129.3 (CH), 128.7 (2CH), 128.5 (CH), 126.2 (2CH), 122.4 (C), 116.8 (CH), 114.0 (C), 105.1 (C), 51.1 (CH₃). **HRMS (ESI⁺):** *m/z* calcd. for C₁₅H₁₂NO₃BrNa [M+Na]⁺ 355.9893, found 355.9886. The structure was analysed by X-Ray diffraction.

4,7-Dimethoxy-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one **6d**



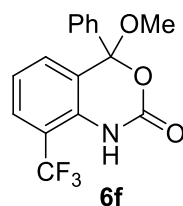
The general procedure **A** was followed with **5d** (102 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **6d** as a light yellow solid (59.0 mg, 0.21 mmol, 52%). **¹H NMR (400 MHz, CDCl₃):** δ 9.85 (brs, 1H), 7.51 (dd, *J* = 7.9, 1.6 Hz, 2H), 7.43-7.35 (m, 3H), 6.92 (d, *J* = 8.6 Hz, 1H), 6.57 (dd, *J* = 8.6, 2.4 Hz, 1H), 6.49 (d, *J* = 2.3 Hz, 1H), 3.78 (s, 3H), 3.42 (s, 3H). **¹³C NMR (100 MHz, CDCl₃):** δ 161.4 (C), 152.4 (C=O), 139.3 (C), 136.5 (C), 129.2 (CH), 128.5 (2CH), 128.2 (CH), 126.8 (2CH), 112.7 (C), 110.4 (CH), 107.8 (C), 99.5 (CH), 55.7 (CH₃), 51.8 (CH₃). **FT-IR (neat, cm⁻¹):** 3220, 3080, 2937, 2836, 1711, 1624, 1598, 1517, 1449, 1339, 1290, 1260. **HRMS (ESI⁺):** *m/z* calcd. for C₁₆H₁₅NO₄Na [M+Na]⁺ 308.0893, found 308.0891

4-Methoxy-8-methyl-4-phenyl-1H-benzo[d][1,3]oxazin-2(4H)-one **6e**



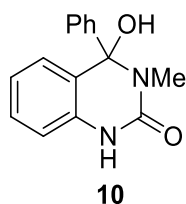
The general procedure **E** was followed with **5e** (95.7 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **6e** as a light yellow solid (32.3 mg, 0.12 mmol, 30%). **¹H NMR (600 MHz, CDCl₃):** δ 8.32 (brs, 1H), 7.52 (dd, $J = 8.0, 1.2$ Hz, 2H), 7.43-7.37 (m, 3H), 7.16 (d, $J = 7.3$ Hz, 1H), 6.94 (t, $J = 7.6$ Hz, 1H), 6.89 (d, $J = 7.6$ Hz, 1H), 3.44 (s, 3H), 2.34 (s, 3H). **¹³C NMR (150 MHz, CDCl₃):** δ 151.1 (C=O), 138.6 (C), 133.4 (C), 131.8 (CH), 129.2 (CH), 128.5 (2CH), 127.0 (2CH), 124.7 (CH), 123.1 (CH), 122.8 (C), 121.0 (C), 107.2 (C), 52.0 (CH₃), 16.8 (CH₃). **FT-IR (neat, cm⁻¹):** 1706, 1600, 1493, 1476, 1250, 1348, 1256, 1221, 1082, 1067, 1010. **HRMS (ESI⁺):** m/z calcd. for C₁₆H₁₅NO₃Na [M+Na]⁺ 292.0944, found 292.0941.

4-Methoxy-4-phenyl-8-(trifluoromethyl)-1H-benzo[d][1,3]oxazin-2(4H)-one **6f**



The general procedure **E** was followed with **5f** (117 mg, 0.40 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **6f** as a light yellow solid (74.7 mg, 0.23 mmol, 58%). **¹H NMR (700 MHz, CDCl₃):** δ 7.64 (brs, 1H), 7.60 (d, $J = 7.8$ Hz, 1H), 7.54-7.50 (m, 2H), 7.47-7.42 (m, 3H), 7.24 (d, $J = 7.8$ Hz, 1H), 7.11 (t, $J = 7.8$ Hz, 1H), 3.44 (s, 3H). **¹³C NMR (176 MHz, CDCl₃):** δ 149.1 (C=O), 137.0 (C), 132.9 (q, $J^{\text{CF}} = 1.3$ Hz, C), 131.0 (CH), 129.7 (CH), 128.9 (2CH), 127.7 (q, $J^{\text{CF}} = 4.8$ Hz, CH), 127.1 (2CH), 123.3 (q, $J^{\text{CF}} = 272.9$ Hz, C), 123.7 (C), 123.0 (CH), 115.1 (q, $J^{\text{CF}} = 32.8$ Hz, C), 106.3 (C), 52.1 (CH₃). **¹⁹F NMR (376 MHz, CDCl₃):** δ -60.3. **FT-IR (neat, cm⁻¹):** 3264, 3218, 3157, 2359, 2332, 1732, 1602, 1452, 1324, 1275, 1266, 1167, 1117. **HRMS (ESI⁺):** m/z calcd. for C₁₆H₁₂F₃NO₃Na [M+Na]⁺ 346.0661, found 346.0663.

4-Hydroxy-3-methyl-4-phenyl-3,4-dihydroquinazolin-2(1H)-one **10**

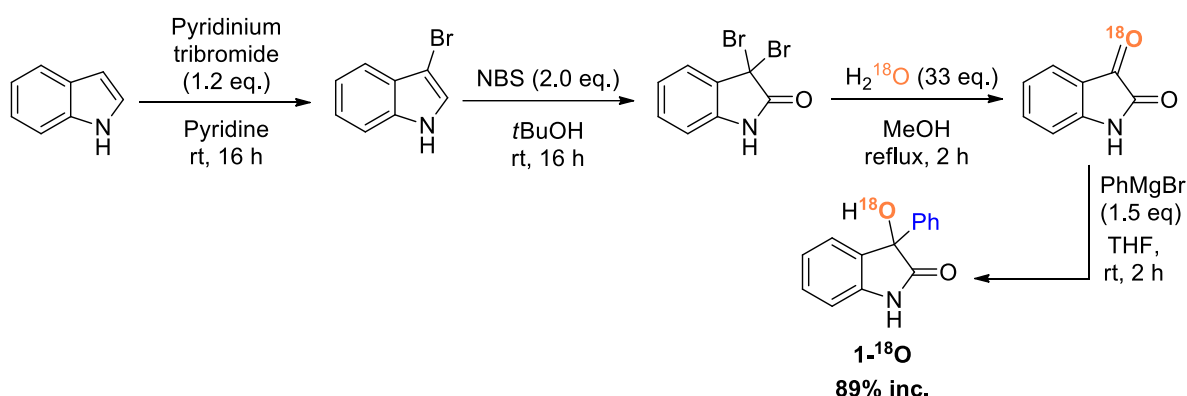


The general procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and MeNH₂ (1.0 M in THF, 4.0 mL, 20 equiv.) in 1.0 mL of THF. Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **10** as a yellow solid (64.7 mg, 0.25 mmol, 64%). ¹H NMR (400 MHz, DMSO-d₆): δ 9.73 (brs, 1H), 7.42-7.38 (m, 2H), 7.37-7.32 (m, 2H), 7.25 (dd, *J* = 7.3, 6.5 Hz, 1H), 7.11 (ddd, *J* = 7.7, 7.7, 1.4 Hz, 1H), 7.00 (s, 1H), 6.88 (dd, *J* = 7.7, 1.5 Hz, 1H), 6.82 (dd, *J* = 7.7, 1.4 Hz, 1H), 6.77 (ddd, *J* = 7.4, 7.3, 1.4 Hz, 1H), 2.60 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆): δ 151.6 (C=O), 145.3 (C), 134.8 (C), 128.5 (CH), 128.2 (2CH), 128.0 (CH), 127.4 (CH), 125.7 (2CH), 124.4 (C), 120.9 (CH), 113.3 (CH), 86.8 (C), 28.0 (CH₃). FT-IR (neat, cm⁻¹): 1651, 1605, 1513, 1499, 1469, 1445, 1431, 1398, 1347, 1327, 1278, 1186, 1168, 1023. HRMS (ESI⁺): *m/z* calcd. for C₁₅H₁₄N₂O₂ [M-OH]⁺ 237.1022, found 237.1024.

5 Mechanistic Experiments (Scheme 3)

5.1 ¹⁸O labelled experiments

Synthesis of 1-¹⁸O



3-Bromo-1H-indole

Following a reported procedure,⁸ a 100 mL round bottom flask was charged with indole (2.0 g, 17.1 mmol, 1.0 equiv.) in pyridine (20 mL). Then, pyridinium tribromide (6.6 g, 20.5 mmol, 1.2 equiv) in pyridine (20 mL) was added dropwise over 10 min after which the reaction was left to stir 16 h at rt. Next, ice water was added, and the reaction mixture was extracted with Et₂O (20 mL). The organic layers were washed with 6N HCl (20 mL) and saturated aq. NaHCO₃ (20 mL), dried with MgSO₄, filtered and concentrated in vacuo. The

desired product (1.99 g, 10.2 mmol, 60%) is used directly in the next step without further purification.

3,3-Dibromoindolin-2-one

Following a reported procedure,⁹ a 50 mL round bottom flask was charged with 3-bromo-1*H*-indole (1.0 g, 5.1 mmol, 1.0 equiv.) in *t*BuOH (10 mL). Then, *N*-bromosuccinimide (1.8 g, 10.2 mmol, 2.0 equiv) was added portion wise and reaction was stirred at rt for 16 h. Next, a saturated solution of Na₂S₂O₃ was added, and the reaction mixture was extracted with Et₂O (20 mL). The organic layers were washed with brine (20 mL), dried with MgSO₄, filtered and concentrated in vacuo. The desired product (1.6 g, 5.5 mmol, quant.) is used directly in the next step without further purification.

¹⁸O-Labelled Isatin

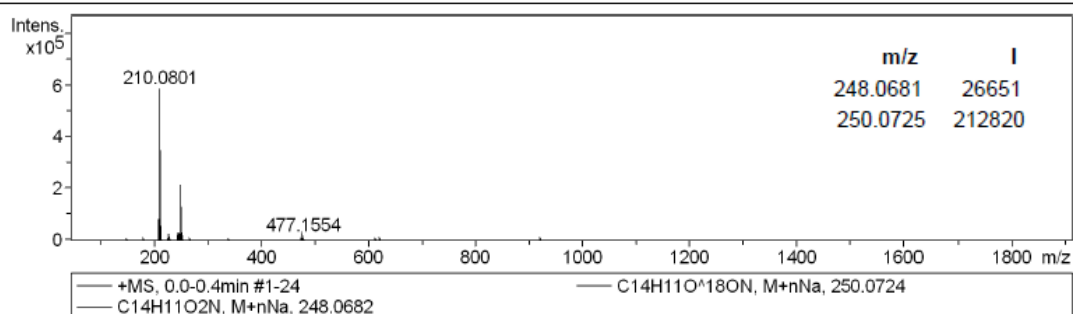
In a sealed tube, 3,3-dibromoindolin-2-one (1.6 g, 5.5 mmol, 1.0 equiv.) was introduced followed by MeOH (10 ml) and H₂¹⁸O (3.3 ml, 181 mmol, 33 equiv.). The mixture was heated at reflux for 2 h and then concentrated in vacuo. Purification by FC (20 g SiO₂, heptane/EA: 100/0 to 60/40) afforded the desired product as an orange solid (150 mg, 1.02 mmol, 19%, 89% incorporation). ¹H NMR (400 MHz, CDCl₃): δ 7.88 (brs, 1H), 7.63 (d, *J* = 7.6 Hz, 1H), 7.57 (t, *J* = 7.9 Hz, 1H), 7.13 (t, *J* = 7.6 Hz, 1H), 6.91 (d, *J* = 7.9 Hz, 1H). HRMS (ESI⁺): *m/z* calcd. for C₈H₅NO¹⁸ONa [M+Na]⁺ 172.0255, found 172.0254.

3-Hydroxy-3-phenylindolin-2-one 1-¹⁸O

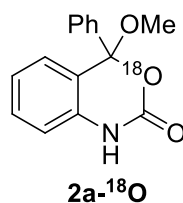
General procedure **B** was followed with ¹⁸O-Labelled Isatin (150 mg, 1.02 mmol) and phenylmagnesium bromide (3.0 M in Et₂O, 0.68 mL, 2.04 mmol, 2.0 equiv.) to afford **1-¹⁸O** as a yellow solid (90.4 mg, 0.40 mmol, 39%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.40 (s, 1H), 7.33-7.23 (m, 6H), 7.10 (d, *J* = 6.9 Hz, 1H), 6.96 (td, *J* = 7.7, 1.1 Hz, 1H), 6.91 (d, *J* = 7.7 Hz, 1H), 6.63 (s, 1H). HRMS (ESI⁺): *m/z* calcd. for C₁₄H₁₁NO¹⁸ONa [M+Na]⁺ 250.0724, found 250.0725.

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4200 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	1900 m/z	Set Charging Voltage	0 V	Set Divert Valve	Source
		Set Corona	0 nA	Set APCI Heater	0 °C



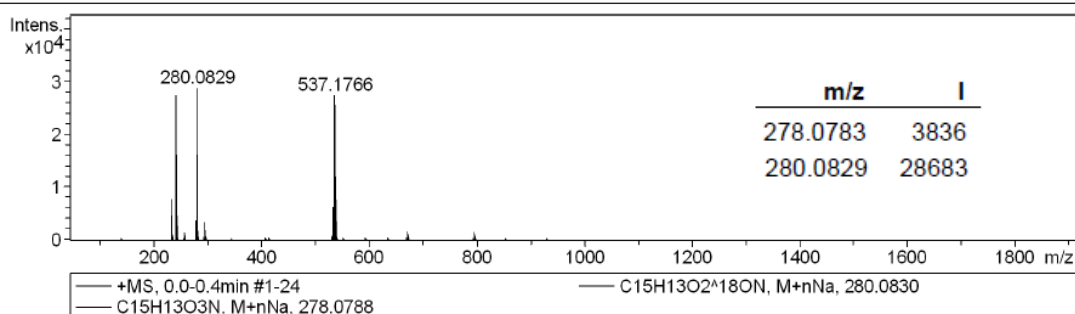
Characterization data of **2a-¹⁸O**



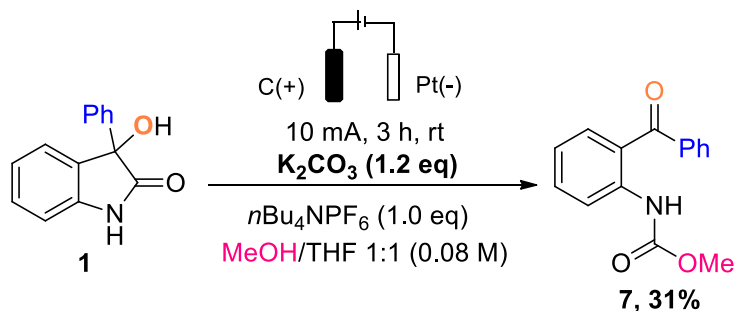
General procedure **A** was followed with **1-¹⁸O** (55.0 mg, 0.38 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **2a-¹⁸O** as a white solid (35.0 mg, 0.14 mmol, 37%, 88% incorporation). ¹H NMR (400 MHz, CDCl₃): δ 9.62 (brs, 1H), 7.53 (dd, *J* = 7.9, 1.7 Hz, 2H), 7.45-7.35 (m, 3H), 7.34-7.28 (m, 1H), 7.08-7.00 (m, 2H), 6.99 (d, *J* = 8.0 Hz, 1H), 3.44 (s, 3H). ¹³C NMR (100 MHz, CDCl₃): δ 153.0 (C=O), 138.7 (C), 135.1 (C), 130.6 (CH), 129.3 (CH), 128.6 (2CH), 126.9 (3CH), 123.7 (CH), 120.8 (C), 114.9 (CH), 107.5 (C), 52.0 (CH₃). HRMS (ESI⁺): *m/z* calcd. for C₁₅H₁₃NO¹⁸ONa [M+Na]⁺ 280.0830, found 280.0829.

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4200 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	1900 m/z	Set Charging Voltage	0 V	Set Divert Valve	Source
		Set Corona	0 nA	Set APCI Heater	0 °C



5.2 Formation of methyl (2-benzoylphenyl)carbamate **7**

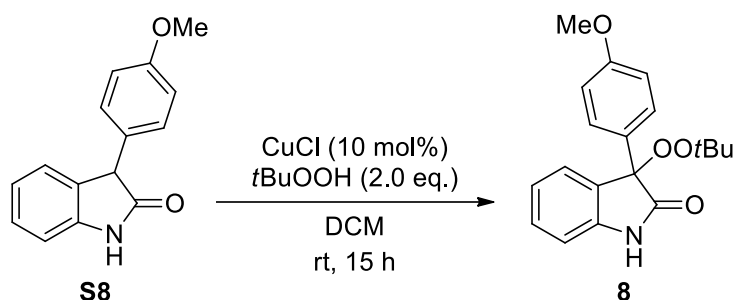


General procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and MeOH (2.5 mL) in presence of K₂CO₃ (1.2 equiv.). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford

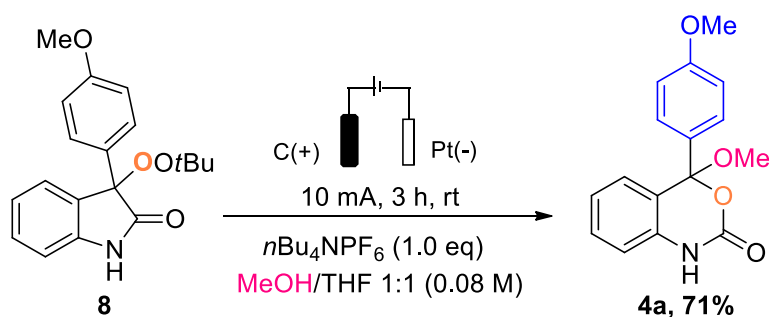
7 as a white solid (31.4 mg, 0.12 mmol, 31%). **¹H NMR (400 MHz, CDCl₃):** δ 10.29 (brs, 1H), 8.42 (d, *J* = 8.4 Hz, 1H), 7.69 (d, *J* = 7.1 Hz, 2H), 7.61-7.50 (m, 3H), 7.47 (t, *J* = 7.6 Hz, 2H), 7.03 (t, *J* = 7.6 Hz, 1H), 3.79 (s, 3H). **¹³C NMR (100 MHz, CDCl₃):** δ 199.4 (C=O), 154.4 (C=O), 141.1 (C), 138.9 (C), 134.4 (CH), 133.7 (CH), 132.4 (CH), 130.0 (2CH), 128.4 (2CH), 123.0 (C), 121.3 (CH), 120.0 (CH), 52.5 (CH₃). **HRMS (ESI⁺):** *m/z* calcd. for C₁₅H₁₃NO₃Na [M+Na]⁺ 278.0788, found 278.0789.

5.3 Reaction of peroxide **8** in the standard conditions

Synthesis of 3-(*tert*-butylperoxy)-3-(4-methoxyphenyl)indolin-2-one **8**



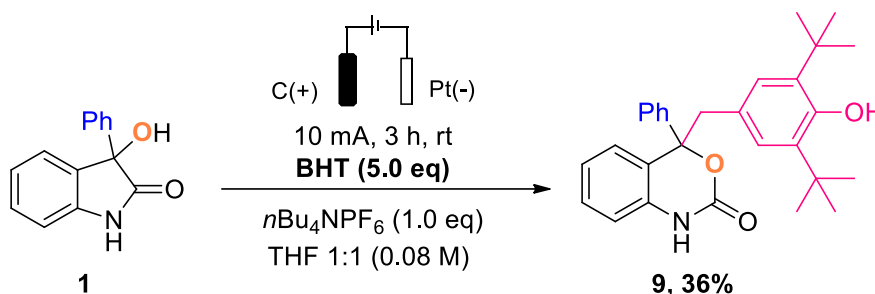
Following a reported procedure,¹⁰ a flamed-dried schlenk tube was charged with 3-(4-methoxyphenyl)indolin-2-one **S8** (300 mg, 1.25 mmol, 1 equiv) and copper(I) chloride (12.4 mg, 0.13 mmol, 0.1 equiv). The tube is sealed, then evacuated and backfilled with argon (3 times). Then, CH₂Cl₂ (12 mL) is added, followed by dropwise addition of *tert*-butyl hydroperoxide (5.5 M in decane, 2.5 mL, 2 equiv) and the reaction is stirred at rt for 15 h. The reaction mixture is dry-loaded onto silica gel and purified directly by FC (40 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **8** (210 mg, 0.64 mmol, 51%). **¹H NMR (700 MHz, CDCl₃):** δ 8.36 (brs, 1H), 7.40 (d, *J* = 9.0 Hz, 2H), 7.34 (d, *J* = 7.4 Hz, 1H), 7.30 (td, *J* = 7.7, 1.2 Hz, 1H), 7.09 (td, *J* = 7.6, 0.9 Hz, 1H), 6.89 (d, *J* = 7.8 Hz, 1H), 6.86 (d, *J* = 9.0 Hz, 2H), 3.78 (s, 3H), 1.18 (s, 9H). **¹³C NMR (176 MHz, CDCl₃):** δ 176.6 (C=O), 160.3 (C), 141.8 (C), 129.8 (CH), 128.9 (C), 128.9 (2CH), 127.9 (C), 126.6 (CH), 122.6 (CH), 114.0 (2CH), 110.3 (CH), 86.2 (C), 80.9 (C), 55.4 (CH₃), 26.7 (3CH₃). **HRMS (ESI⁺):** *m/z* calcd. for C₁₉H₂₁NO₄Na [M+Na]⁺ 350.1363, found 350.1364.



General procedure **A** was followed with **8** (100 mg, 0.31 mmol) and MeOH (2.5 mL). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4a** as a white solid (62.0 mg, 0.22 mmol,

71%). **¹H NMR (400 MHz, CDCl₃):** δ 8.79 (brs, 1H), 7.43 (d, J = 8.9 Hz, 2H), 7.31 (ddd, J = 8.0, 7.0, 1.9 Hz, 1H), 7.10-7.00 (m, 2H), 6.94-6.90 (m, 3H), 3.82 (s, 3H), 3.43 (s, 3H). **¹³C NMR (100 MHz, CDCl₃):** δ 160.3 (C=O), 151.6 (C), 136.1 (C), 130.9 (C), 130.5 (CH), 128.4 (2CH), 126.9 (CH), 123.6 (CH), 121.3 (C), 114.7 (CH), 113.9 (2CH), 107.6 (C), 55.5 (CH₃), 52.0 (CH₃). **FT-IR (neat, cm⁻¹):** 1700, 1604, 1513, 1505, 1494, 1432, 1353, 1319, 1305, 1251, 1209, 1185, 1169, 1153, 1095, 1032, 1009. **HRMS (ESI⁺):** m/z calcd. for C₁₆H₁₅NO₄Na [M+Na]⁺ 308.0893, found 308.0887.

5.4 BHT trapping experiment



General procedure **A** was followed with **1** (90.1 mg, 0.40 mmol) and MeOH (2.5 mL) in the presence of BHT (441 mg, 2.0 mmol, 5.0 equiv.). Purification was performed with flash column chromatography over silica gel (20 g SiO₂, heptane/EtOAc, 100/0 to 50/50, gradient) to afford **4a** as a white solid (66.2 mg, 0.15 mmol, 37%). **1a** was also recovered (52.8 mg, 0.23 mmol, 59%). **¹H NMR (400 MHz, CDCl₃):** δ 7.46-7.35 (m, 2H), 7.33-7.23 (m, 5H), 7.11 (s, 2H), 7.03 (t, J = 7.5 Hz, 1H), 6.86 (d, J = 7.8 Hz, 1H), 5.17 (s, 1H), 5.12 (d, J = 15.4 Hz, 1H), 4.55 (d, J = 15.4 Hz, 1H), 3.60 (s, 1H), 1.39 (s, 18H). **¹³C NMR (100 MHz, CDCl₃):** δ 177.8 (C=O), 153.5 (C), 143.1 (C), 140.6 (C), 136.4 (2C), 131.9 (C), 129.9 (CH), 128.8 (2CH), 128.4 (CH), 126.3 (C), 125.3 (2CH), 125.0 (CH), 124.2 (2CH), 123.5 (CH), 109.9 (CH), 78.1 (C), 44.3 (CH₂), 34.4 (2C), 30.4 (6CH₃). **HRMS (ESI⁺):** m/z calcd. for C₂₉H₃₃NO₃Na [M+Na]⁺ 466.2353, found 466.2355.

6 X-Ray Diffraction Analysis of compound 6b

The X-ray intensity data was measured on Bruker D8 Venture diffractometer equipped with multilayer monochromator, Mo K α INCOATEC micro focus sealed tube and Oxford cooling system. The structure was solved by *Charge Flipping*. Non-hydrogen atoms were refined with *anisotropic displacement parameters*. Hydrogen atoms were inserted at calculated positions and refined with riding model. The following software was used: *Bruker SAINT software package*¹¹ using a narrow-frame algorithm for frame integration, *SADABS*¹² for absorption correction, *OLEX2*¹³ for structure solution, refinement, molecular diagrams and graphical user-interface, *Shelxle*¹⁴ for refinement and graphical user-interface *SHELXS-2015*¹⁵ for structure solution, *SHELXL-2015*¹⁵ for refinement, *Platon*¹⁶ for symmetry check. Experimental data and CCDC-Codes Experimental data (Available online: <http://www.ccdc.cam.ac.uk/conts/retrieving.html>) can be found in **Table S1**. Crystal data, data collection parameters, and structure refinement details are given in **Table S2**. Asymmetric Units visualized in **Figure S1**.

The sample was prepared by slow liquid-liquid diffusion using a mixture of CH₂Cl₂/MeOH/AcOEt (1/1/1).

Table S1. Experimental parameter and CCDC-Code.

Sample	Machine	Source	Temp.	Detector Distance	Time/Frame	#Frames	Frame width	CCDC
	Bruker		[K]	[mm]	[s]		[°]	
6b	D8	Mo	100	30	3	2299	0.360	2102358

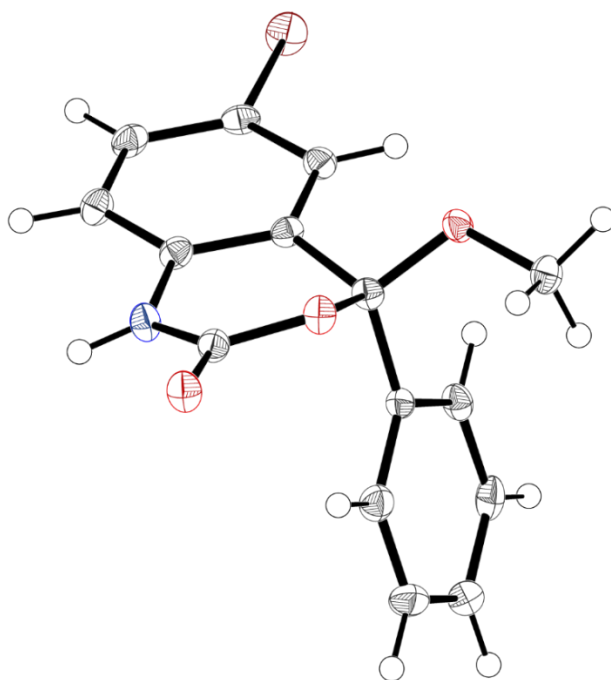


Figure S1. Crystal structure of **6b** drawn with 50% displacement ellipsoid. Only one part visualized. The bond precision for C-C single bonds is 0.0057Å.

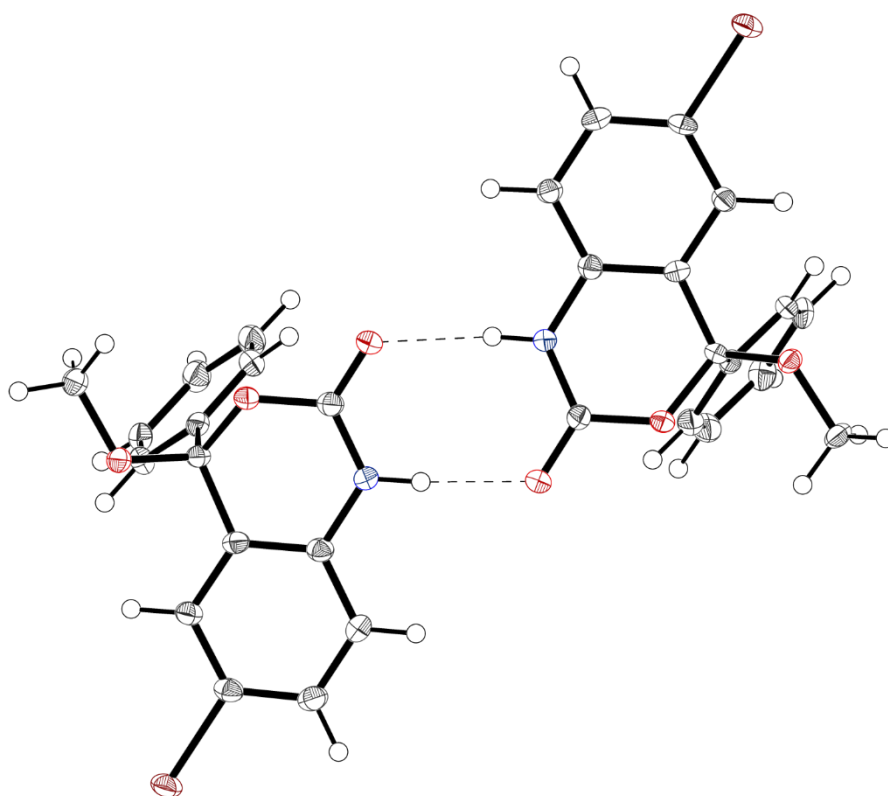


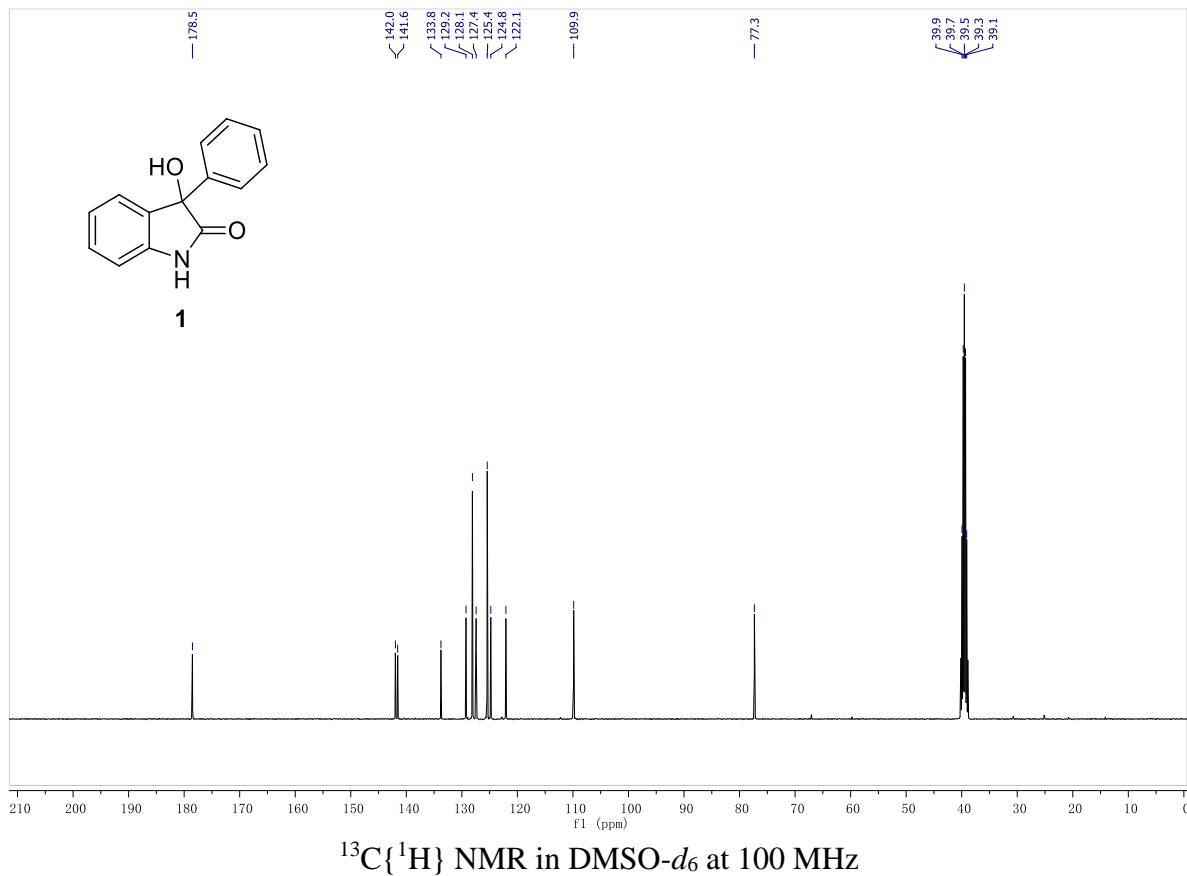
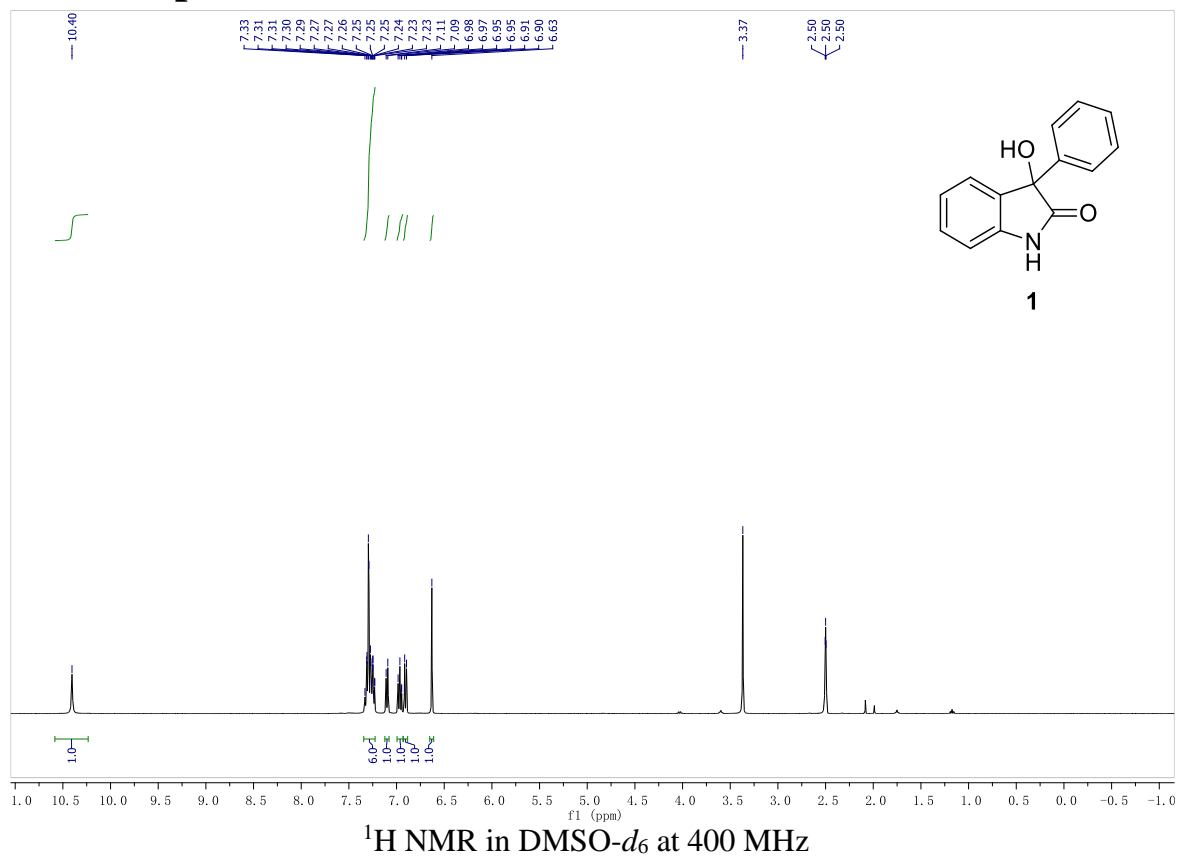
Figure S2. Crystal structure of **6b** drawn with 50% displacement ellipsoid. The bond precision for C-C single bonds is 0.0057Å.

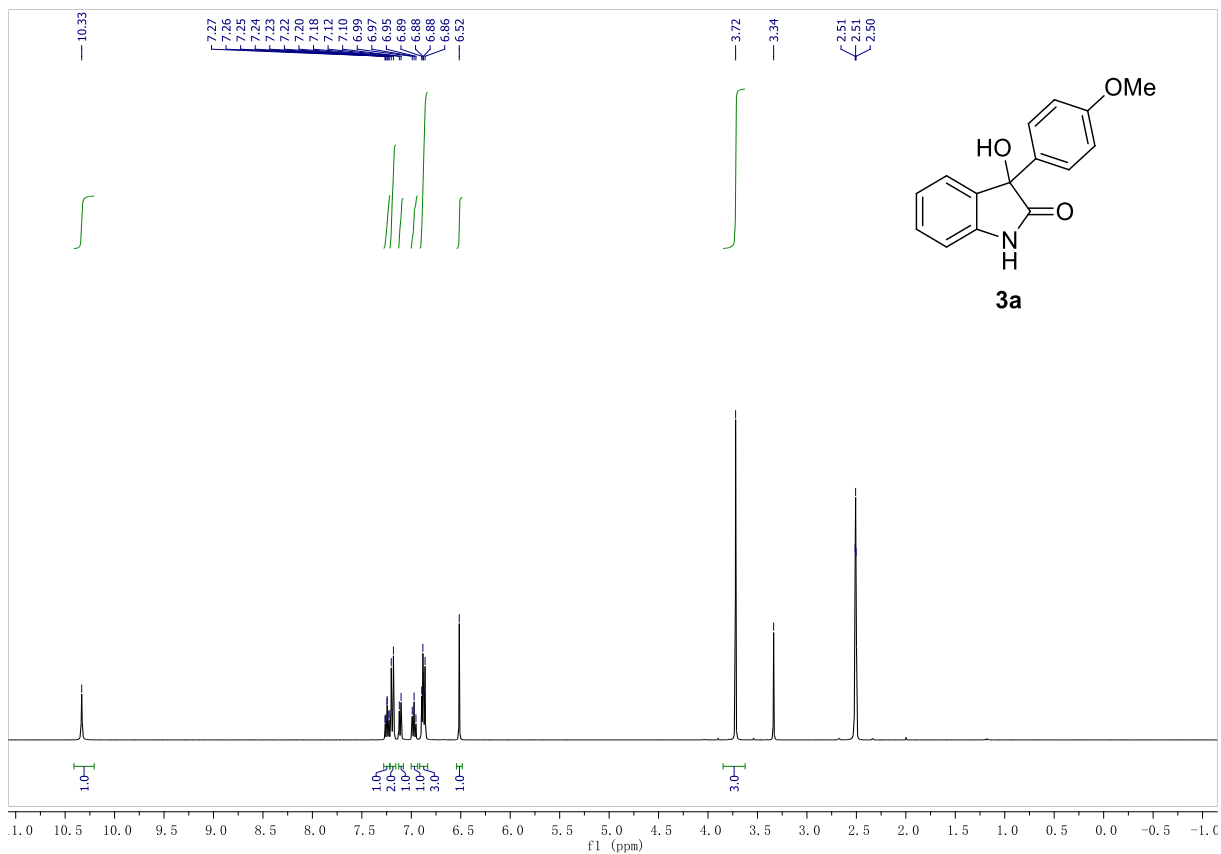
Table S2. Sample and crystal data, data collection and structure refinement. More detailed information can be found in the Cif Code of CCDC: **2102358**.

Identification code	mipa361_P-1
Empirical formula	C ₁₅ H ₁₂ BrNO ₃
Formula weight	334.17
Temperature/K	100.0
Crystal system	triclinic
Space group	P-1
a/Å	9.9912(6)
b/Å	11.1573(6)
c/Å	12.8256(7)
α /°	99.968(2)
β /°	105.709(2)
γ /°	99.578(2)

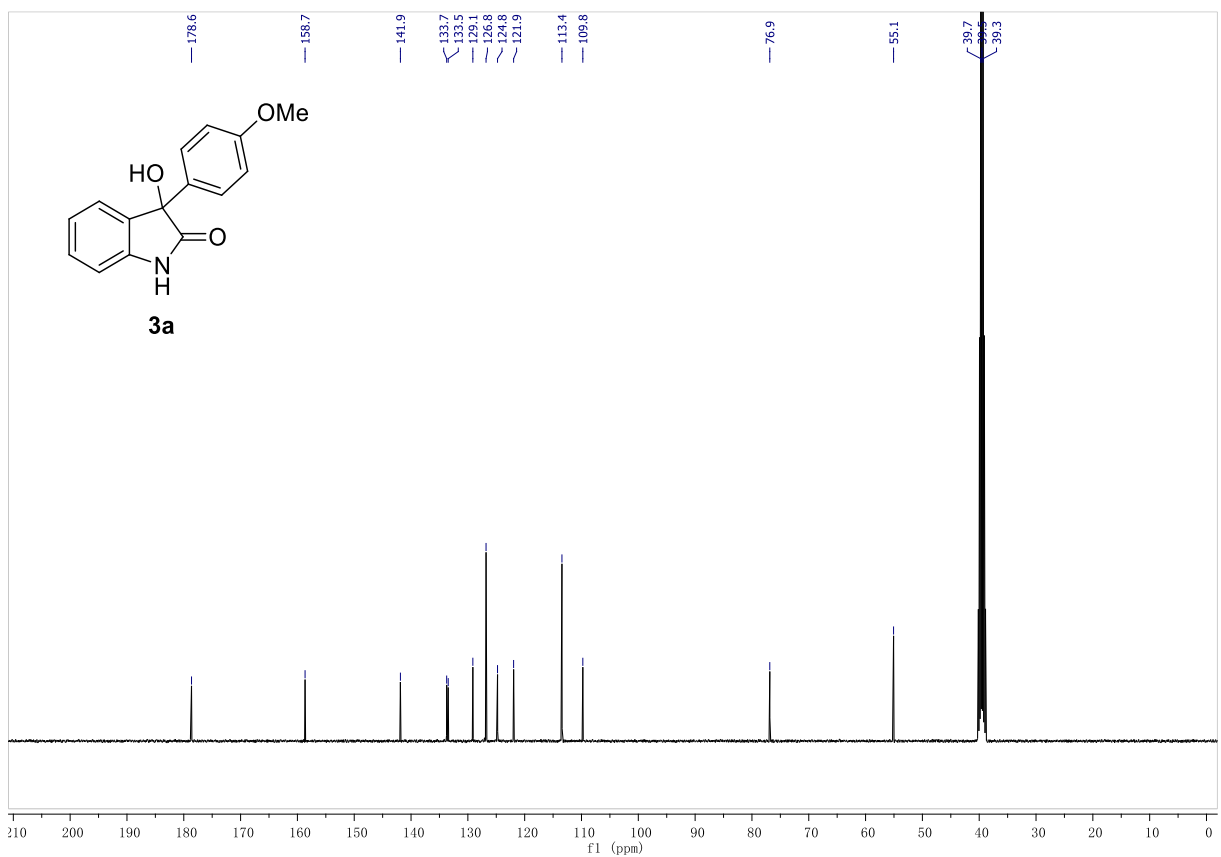
Volume/Å ³	1320.82(13)
Z	4
$\rho_{\text{calc}}/\text{cm}^3$	1.680
μ/mm^{-1}	3.118
F(000)	672.0
Crystal size/mm ³	0.358 × 0.313 × 0.124
Radiation	MoK α ($\lambda = 0.71073$)
2 Θ range for data collection/°	3.804 to 56.564
Index ranges	-13 ≤ h ≤ 12, -14 ≤ k ≤ 14, 0 ≤ l ≤ 17
Reflections collected	9148
Independent reflections	6510 [R _{int} = 0.1797, R _{sigma} = 0.0928]
Data/restraints/parameters	6510/0/363
Goodness-of-fit on F ²	1.067
Final R indexes [I ≥ 2 σ (I)]	R ₁ = 0.0763, wR ₂ = 0.1391
Final R indexes [all data]	R ₁ = 0.1015, wR ₂ = 0.1492
Largest diff. peak/hole / e Å ⁻³	2.29/-1.28

7 NMR Spectra

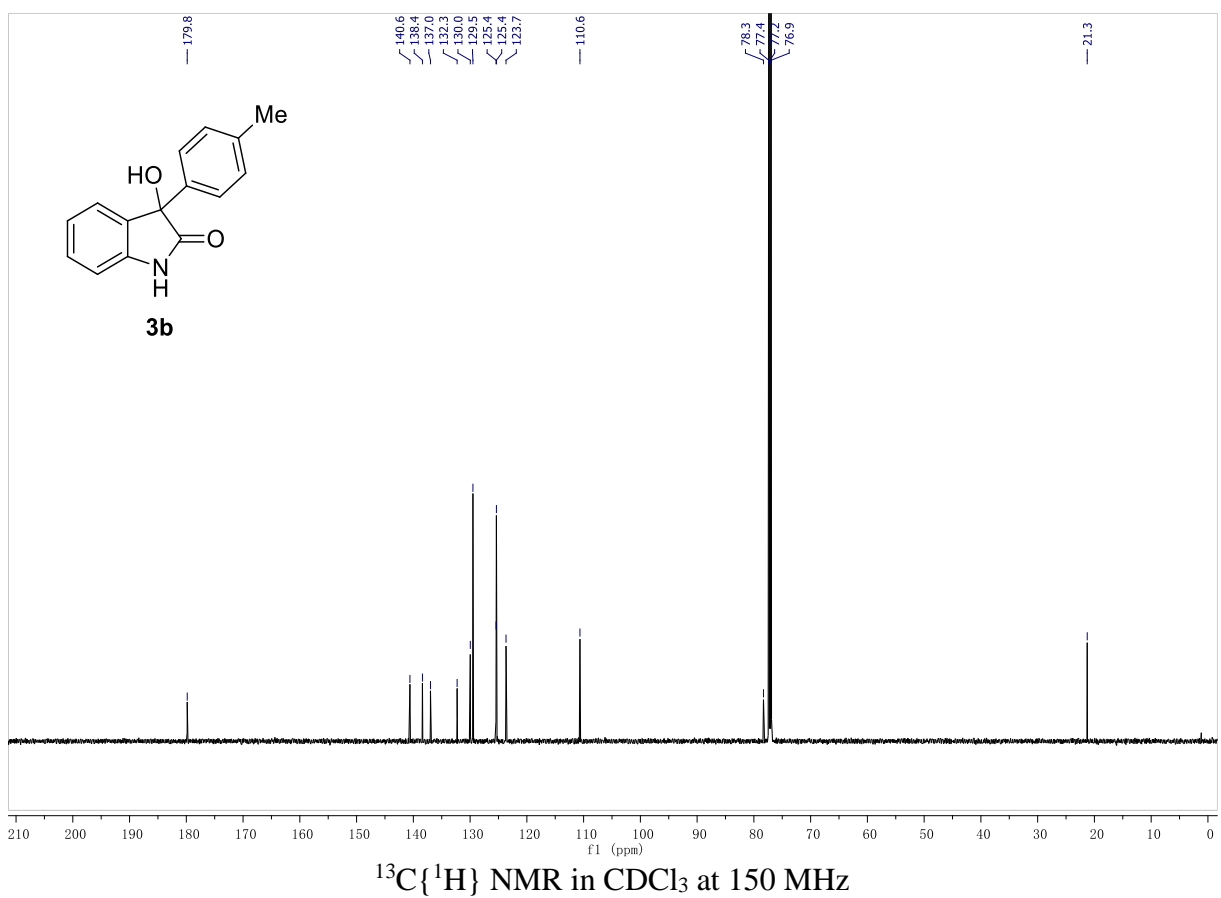
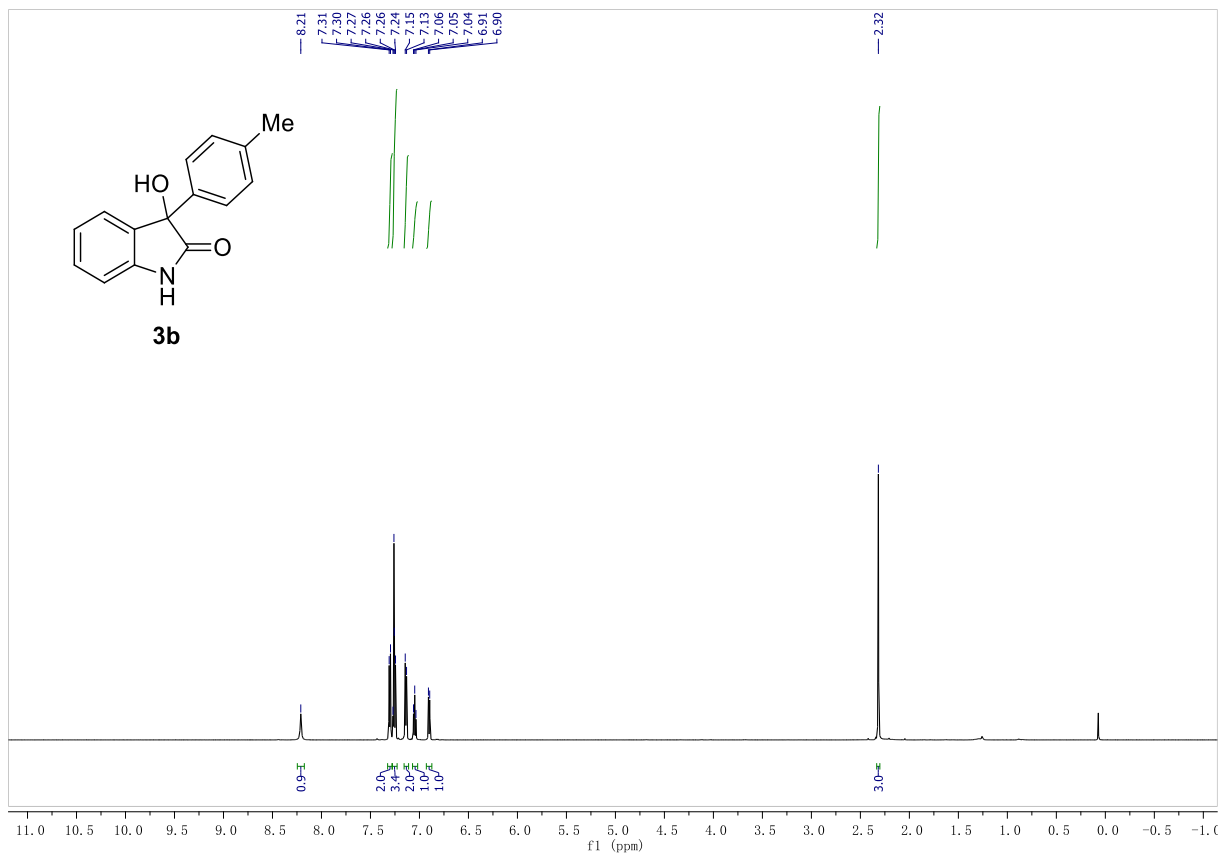


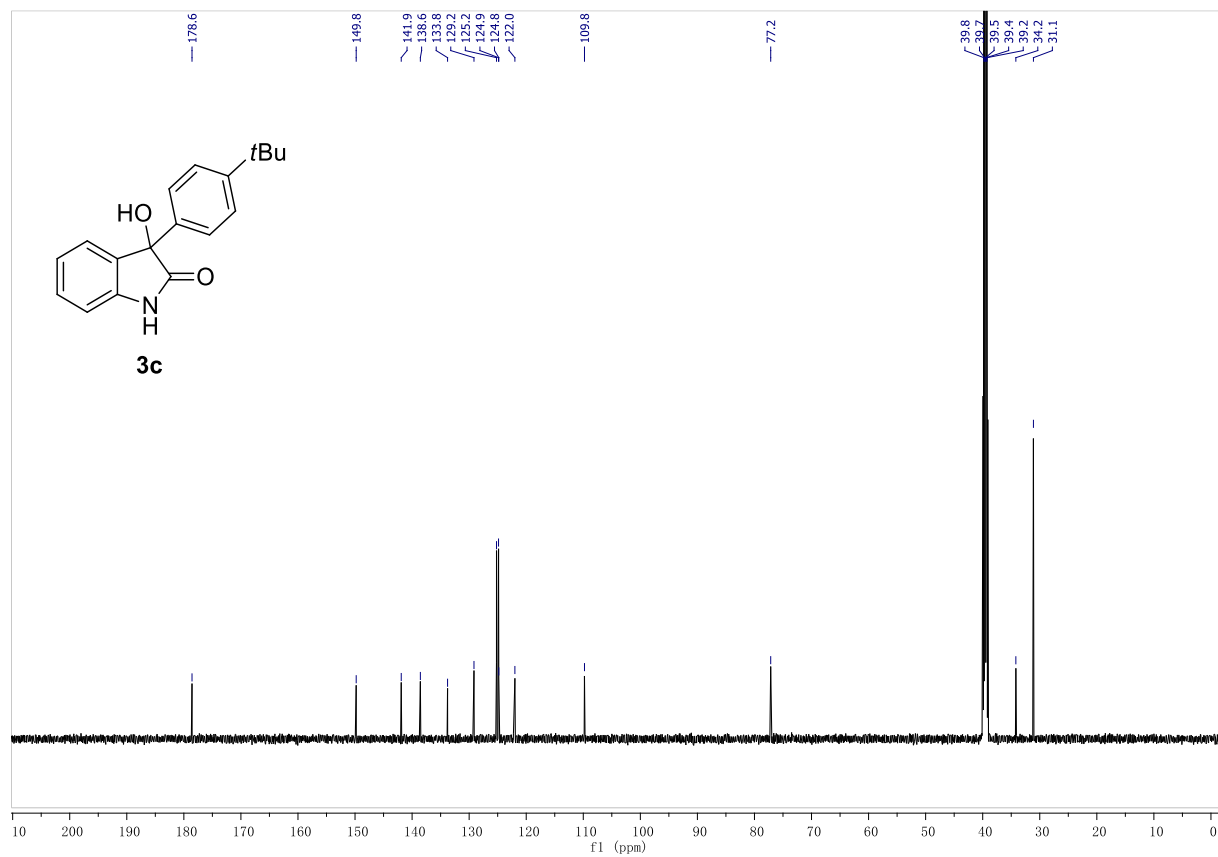
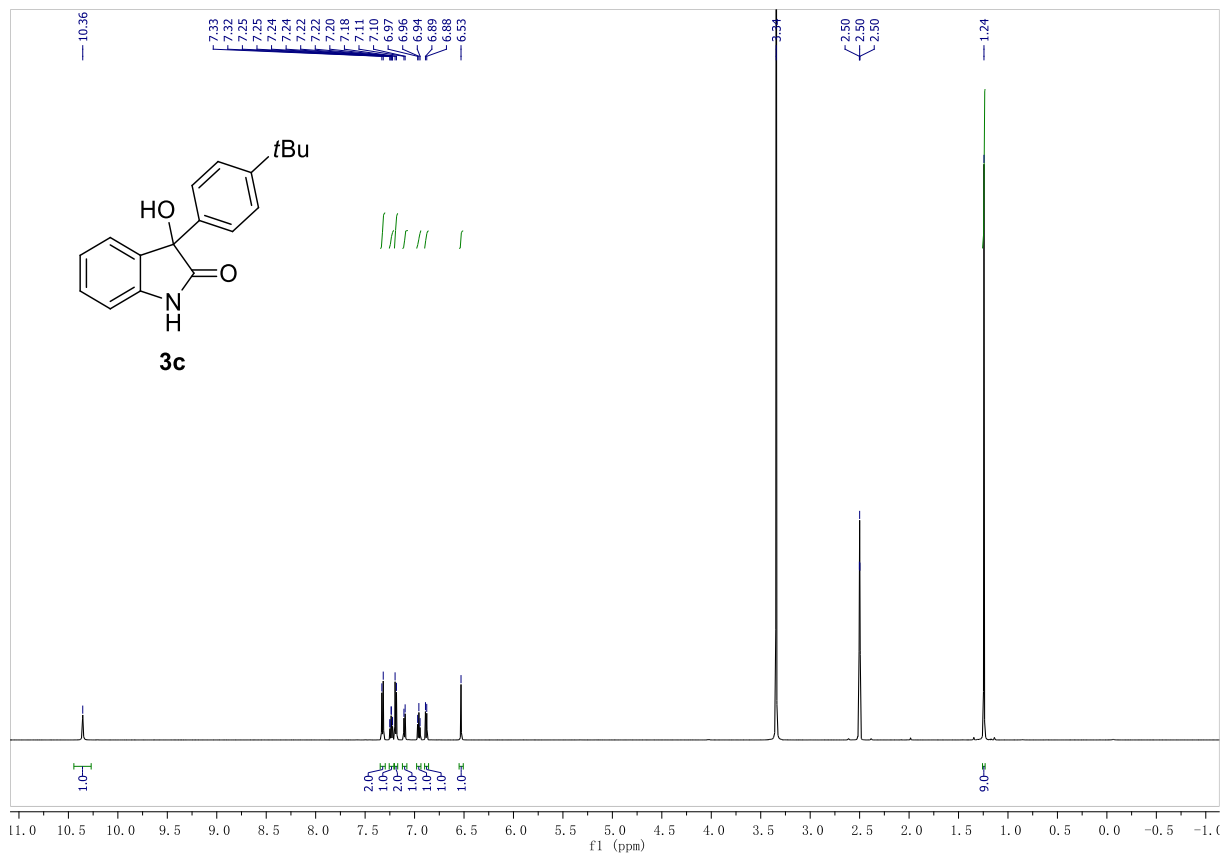


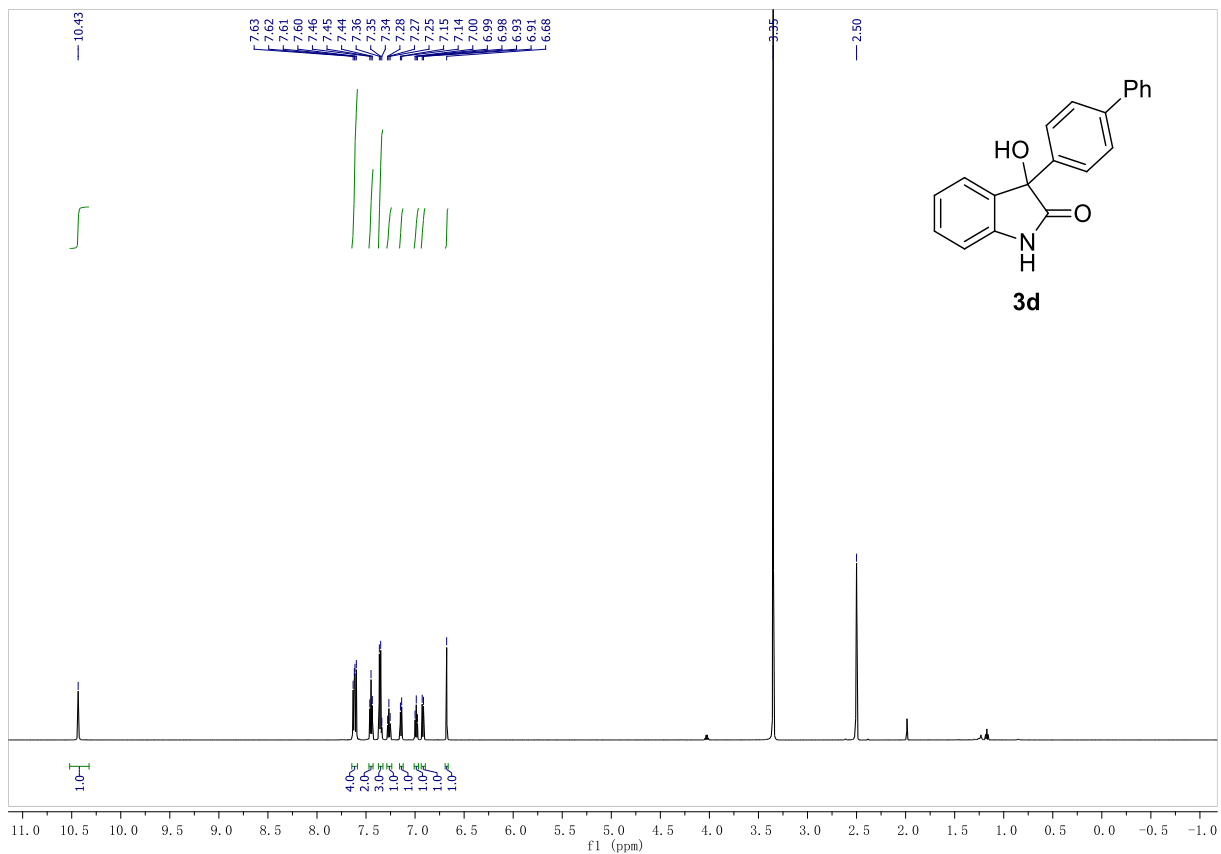
^1H NMR in $\text{DMSO-}d_6$ at 400 MHz



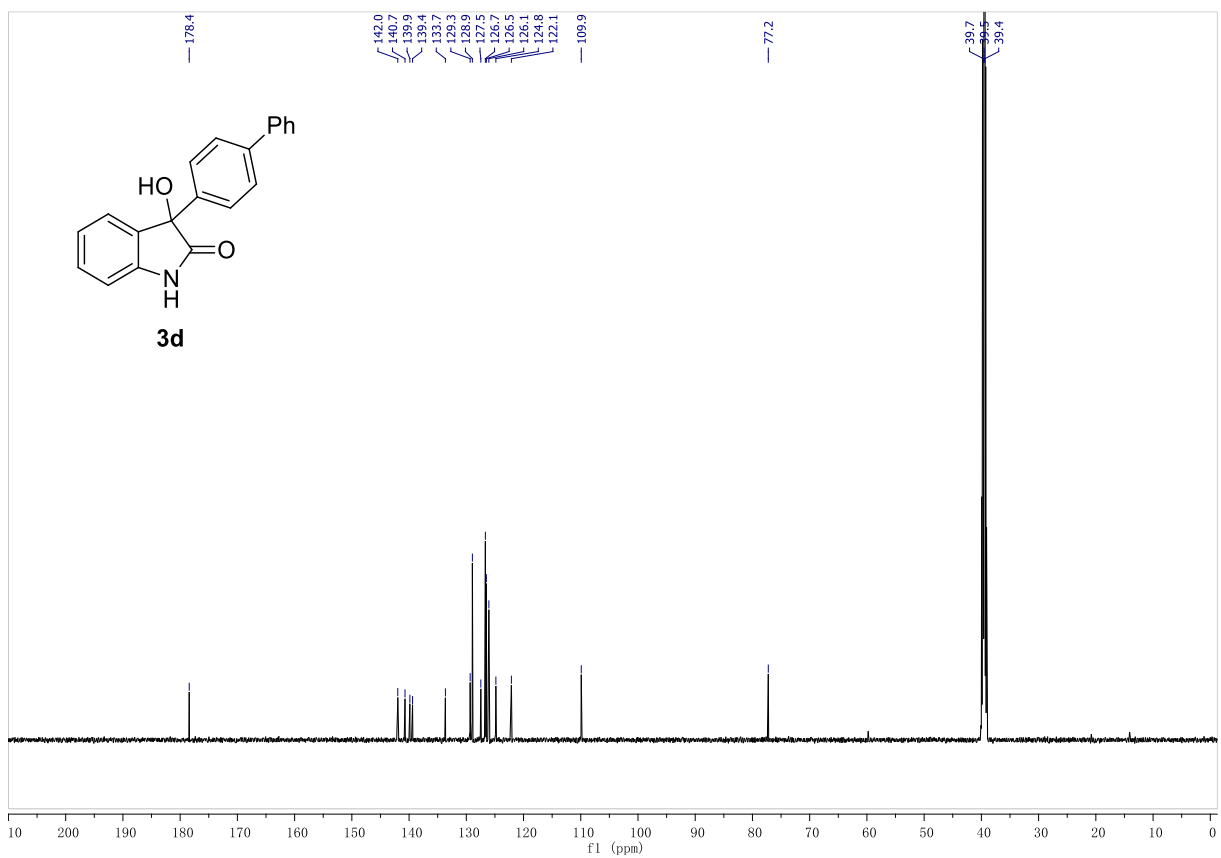
$^{13}\text{C}\{^1\text{H}\}$ NMR in $\text{DMSO-}d_6$ at 100 MHz



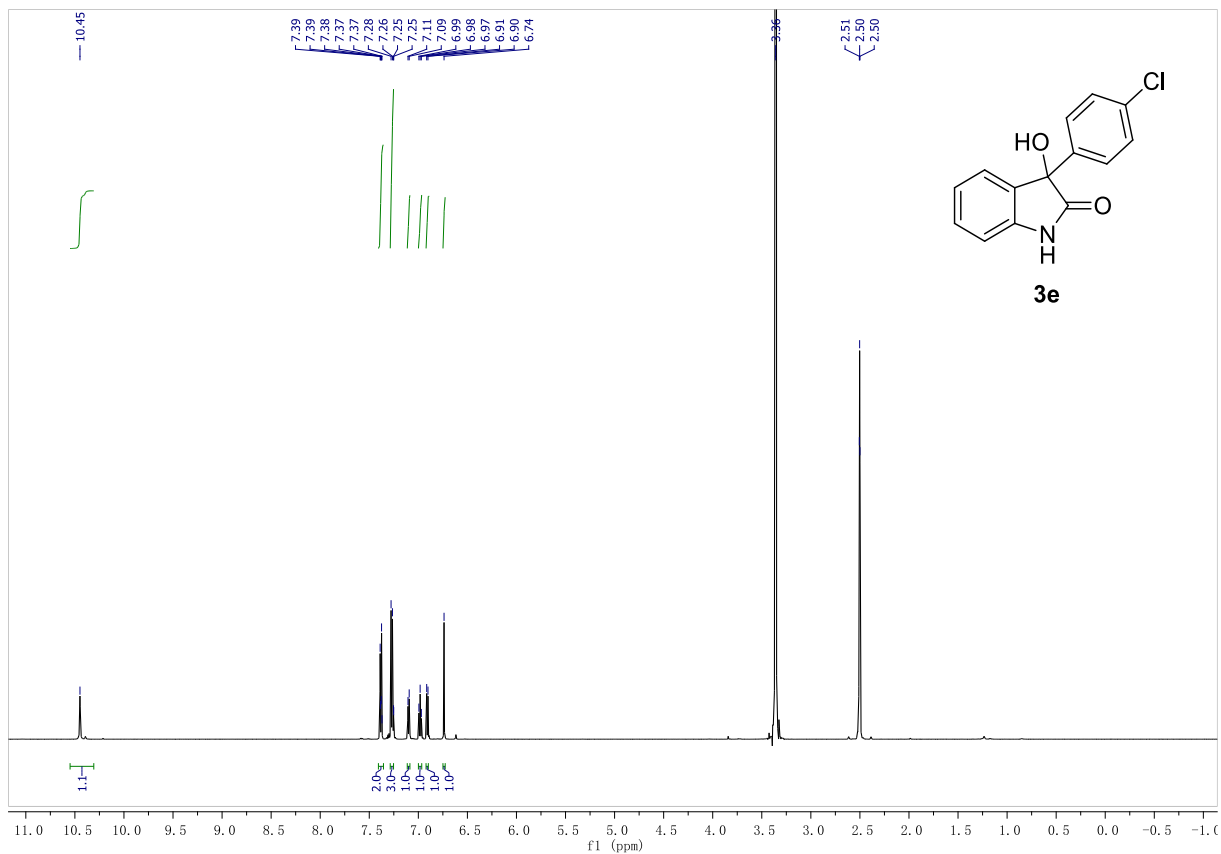




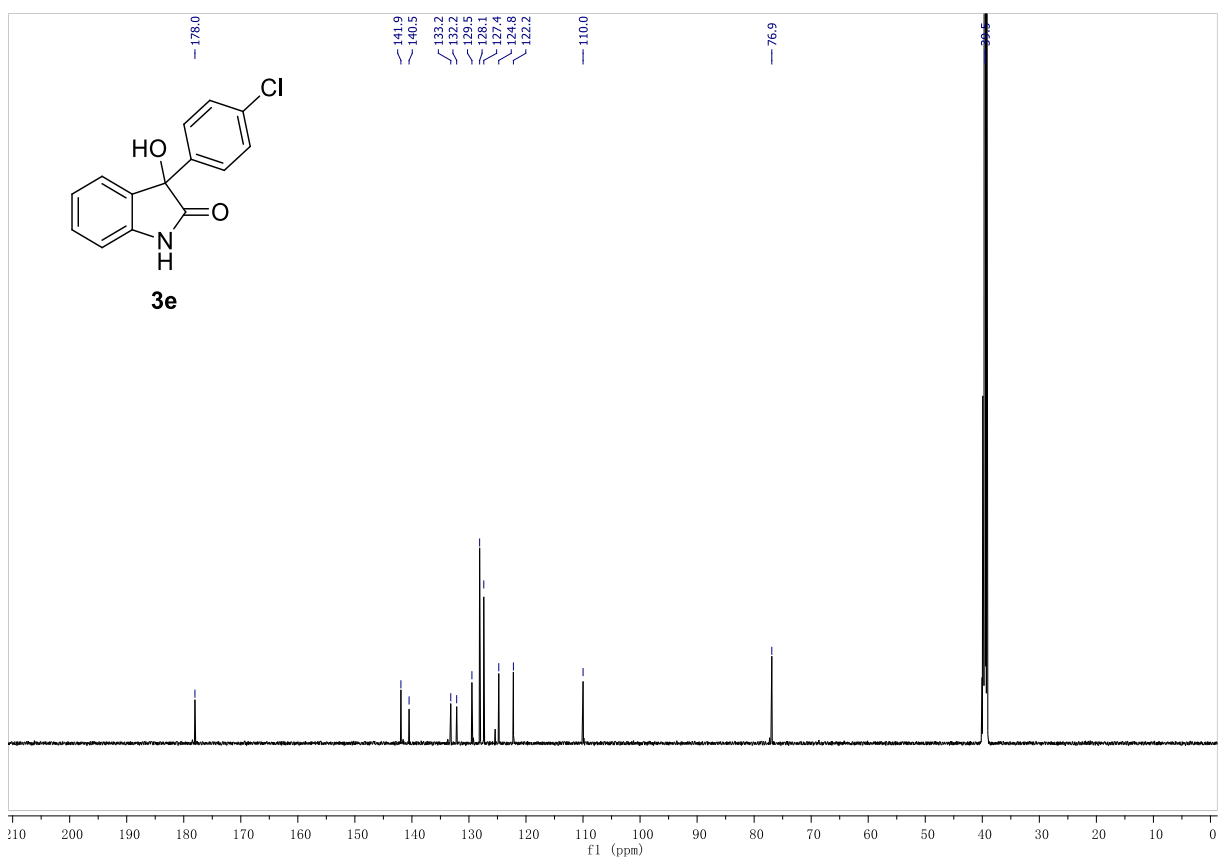
^1H NMR in $\text{DMSO-}d_6$ at 600 MHz



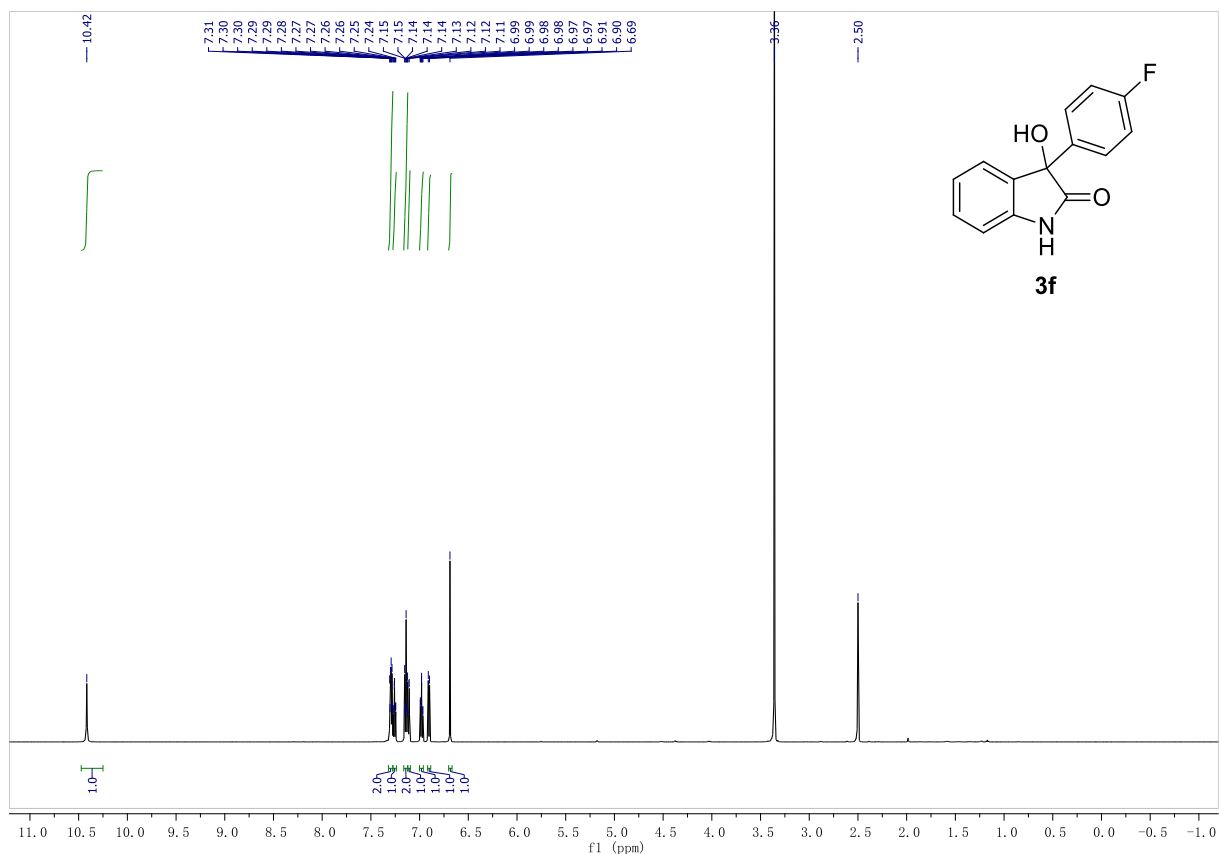
$^{13}\text{C}\{^1\text{H}\}$ NMR in $\text{DMSO-}d_6$ at 150 MHz



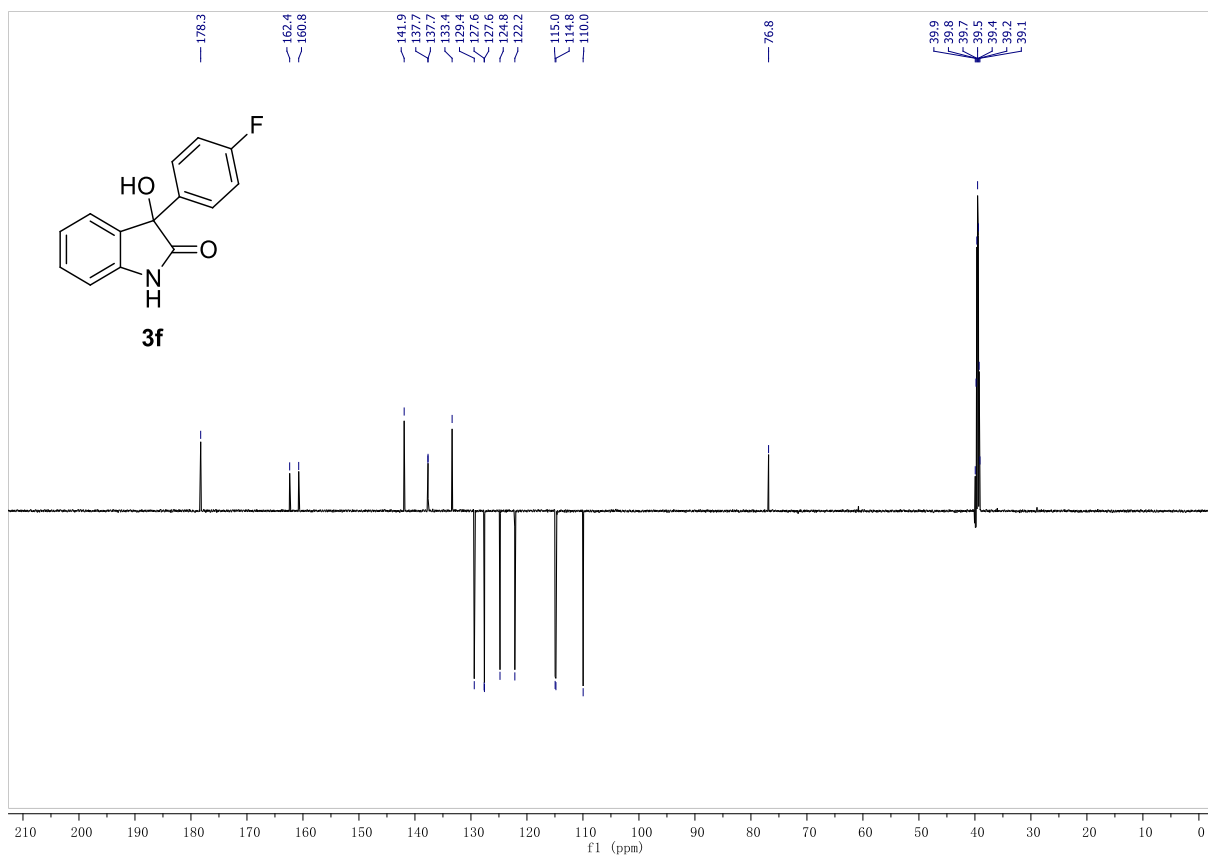
^1H NMR in $\text{DMSO-}d_6$ at 600 MHz



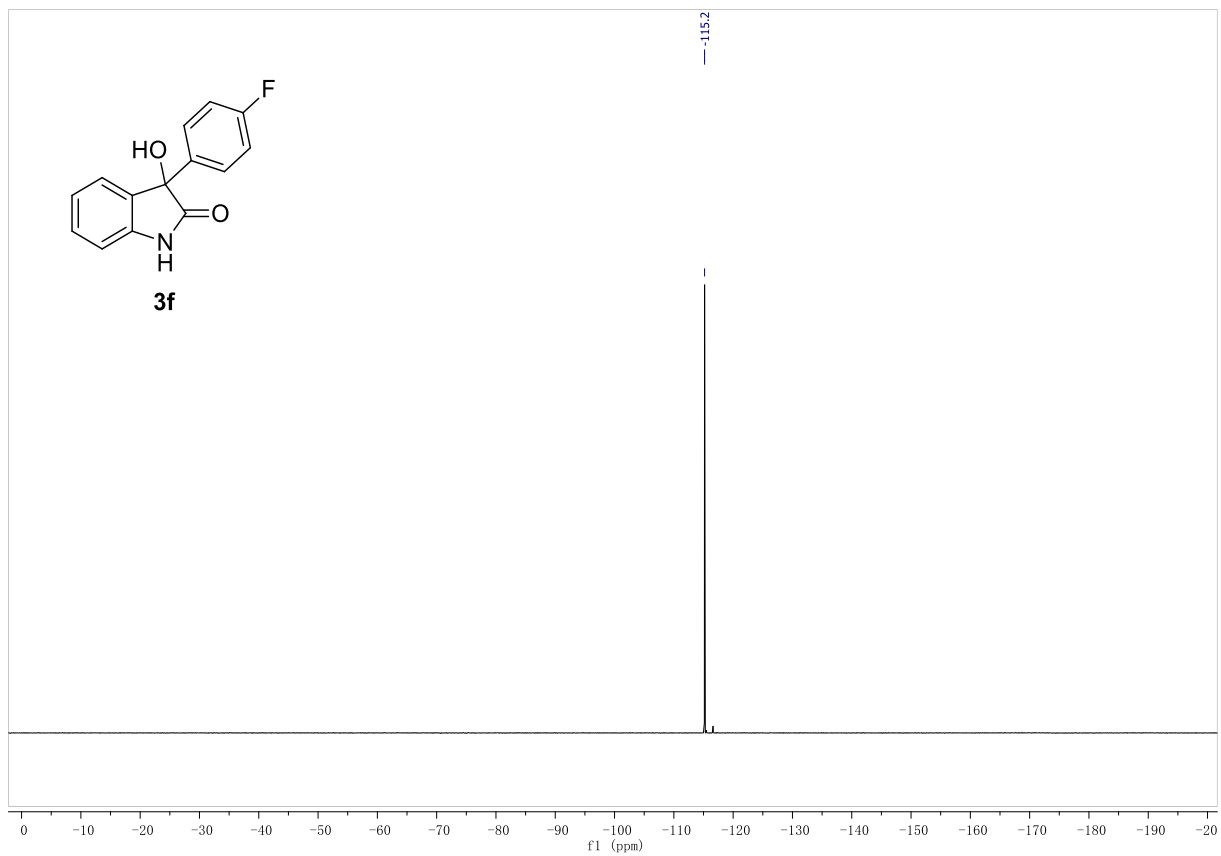
$^{13}\text{C}\{^1\text{H}\}$ NMR in $\text{DMSO-}d_6$ at 150 MHz



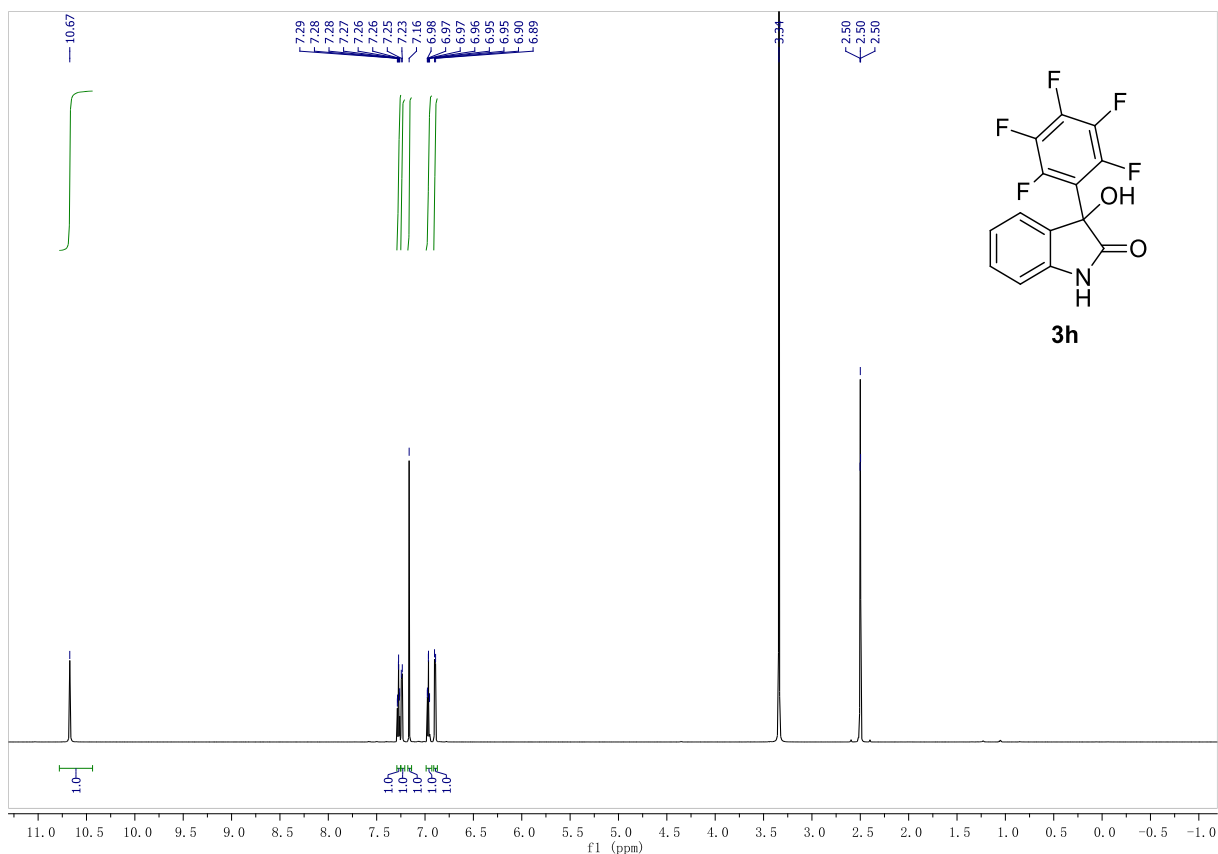
^1H NMR in $\text{DMSO-}d_6$ at 600 MHz



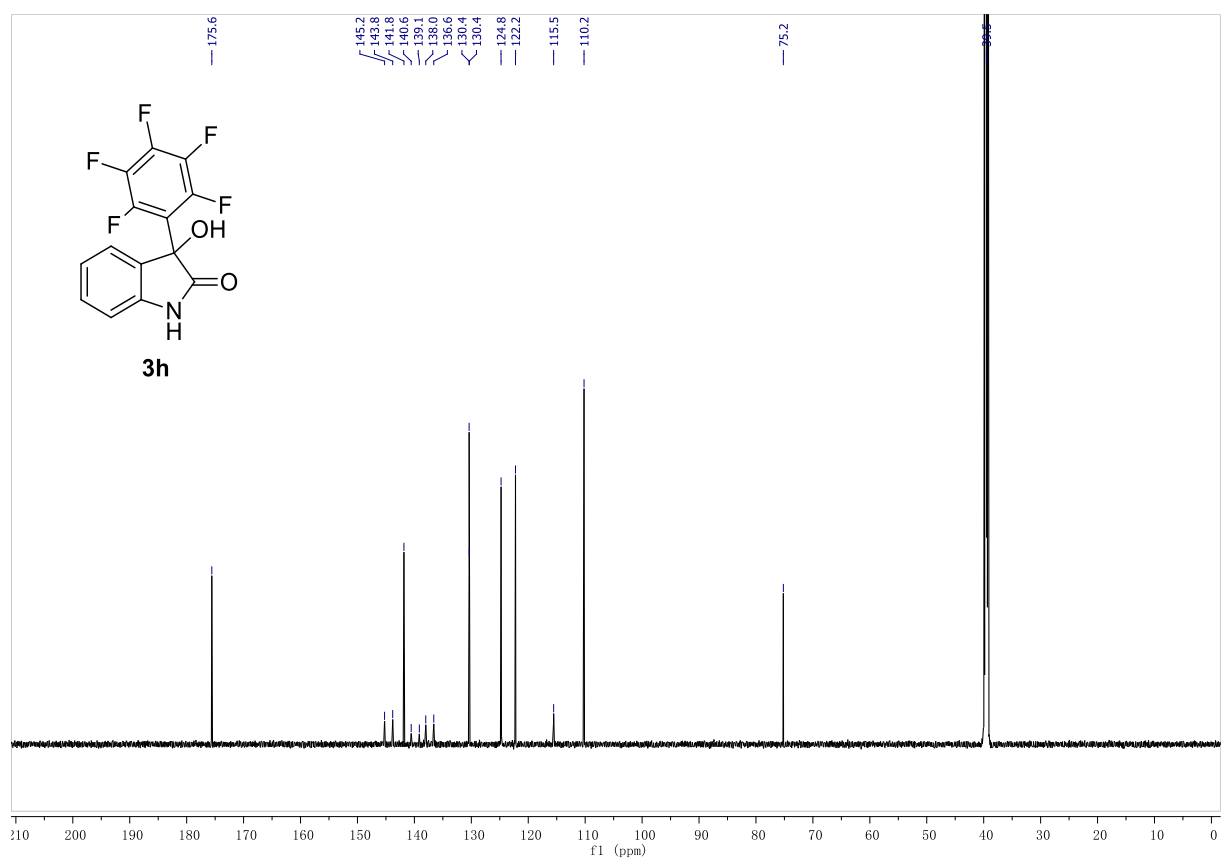
$^{13}\text{C}\{^1\text{H}\}$ NMR in $\text{DMSO-}d_6$ at 150 MHz



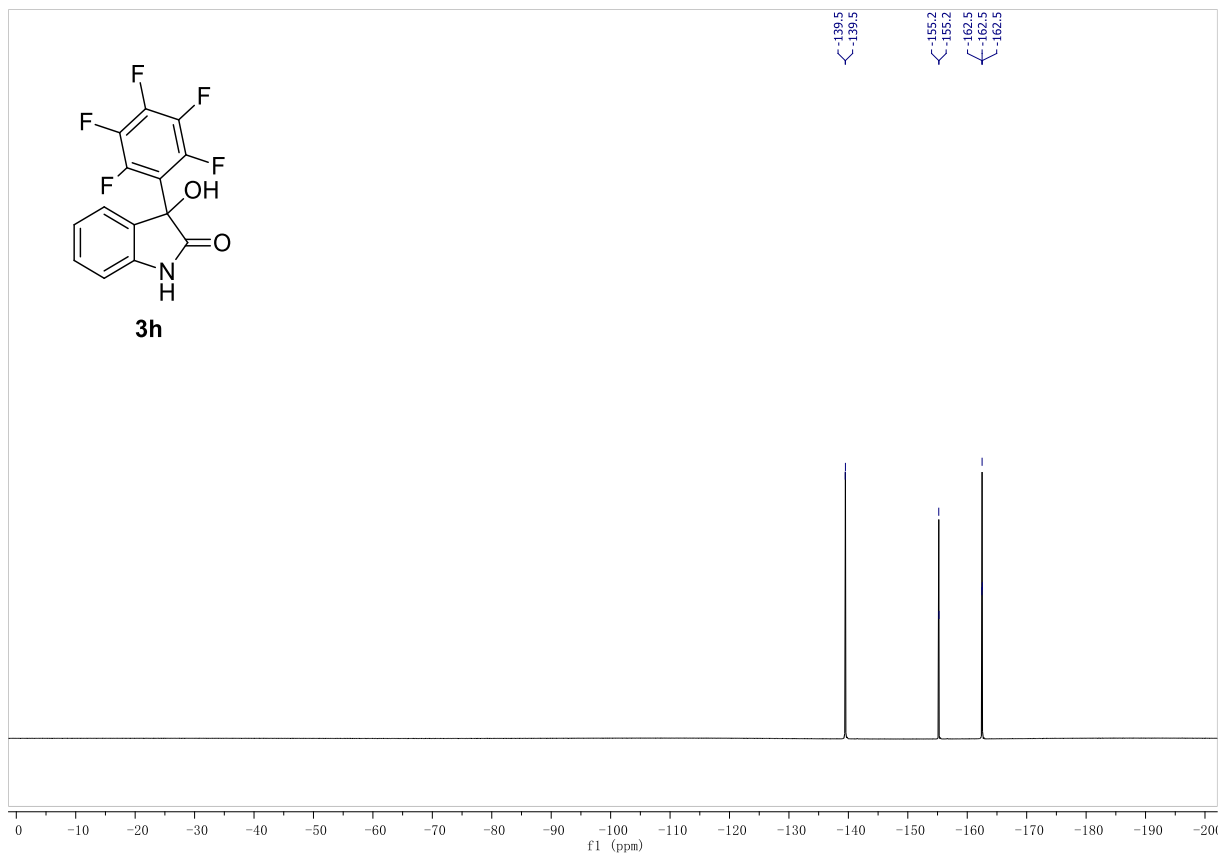
^{19}F NMR in $\text{DMSO-}d_6$ at 565 MHz



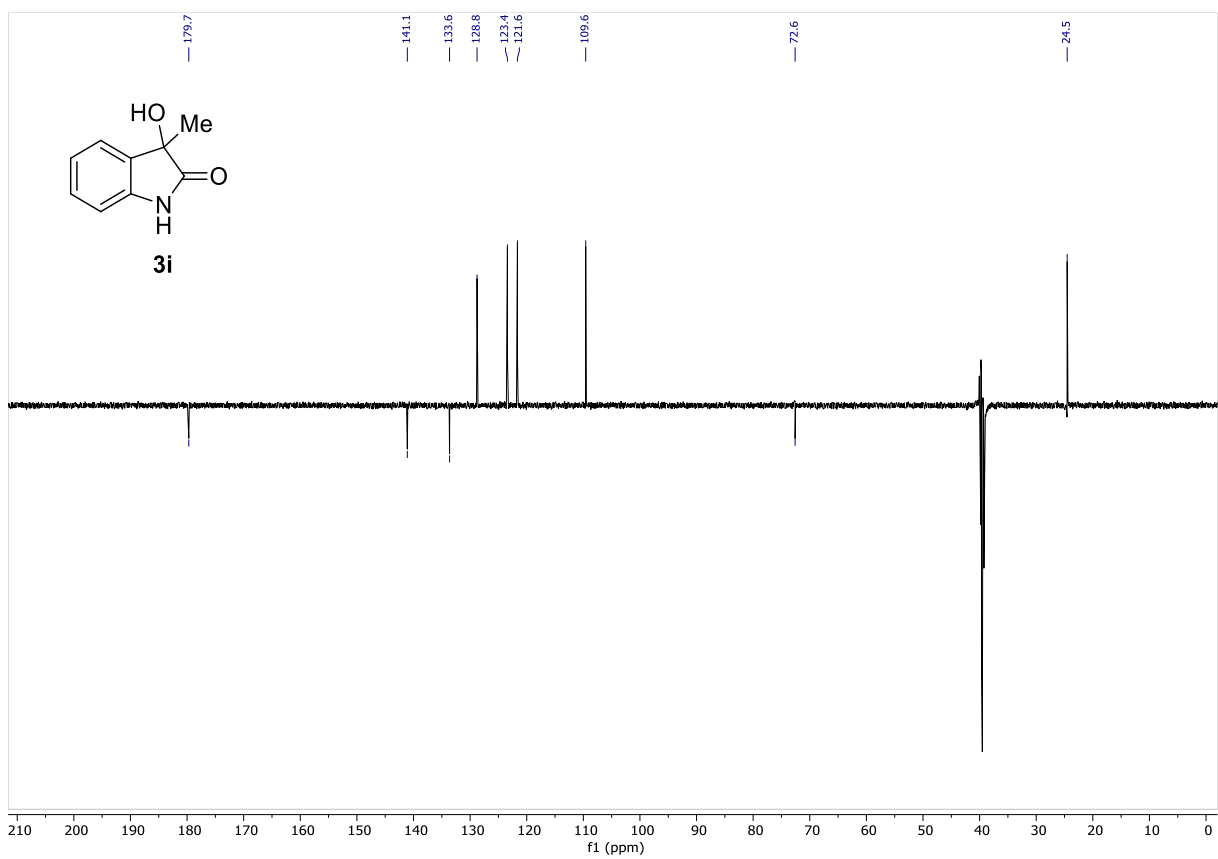
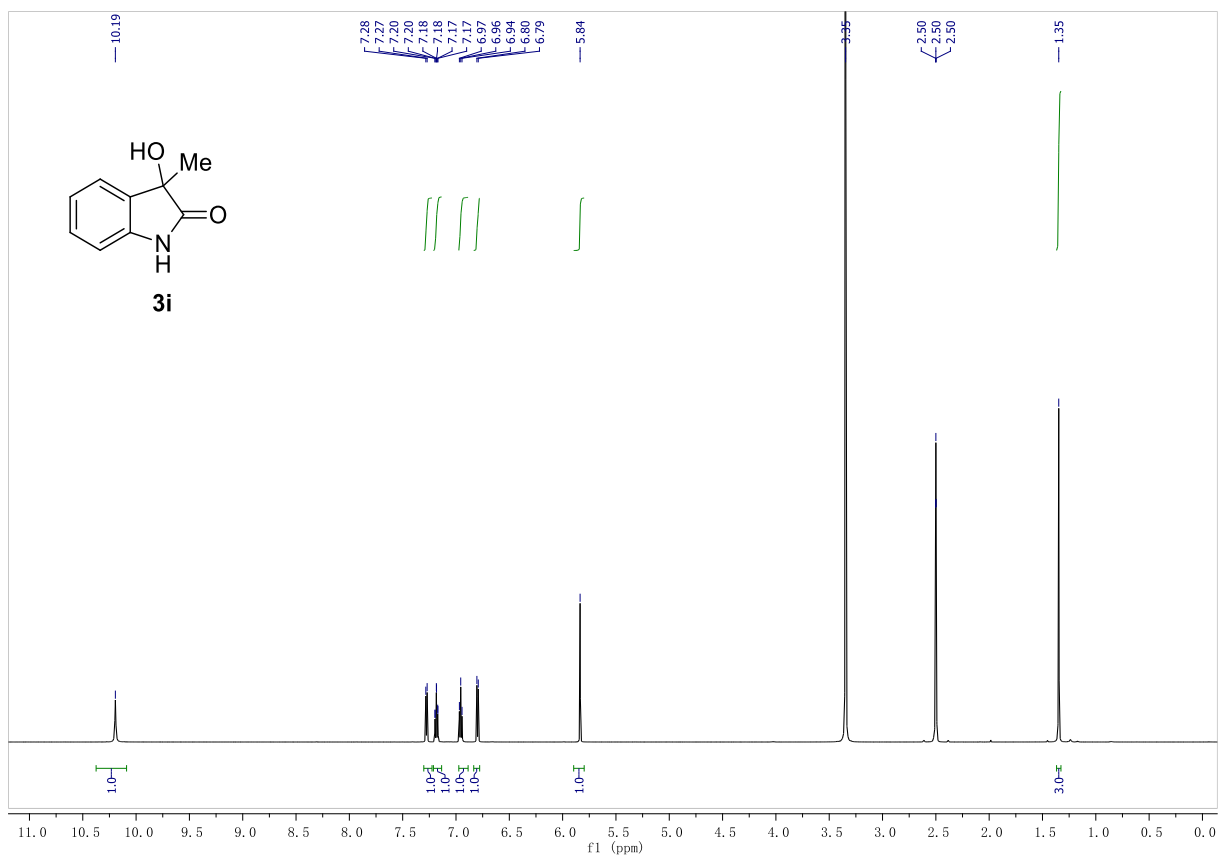
¹H NMR in DMSO-*d*₆ at 700 MHz

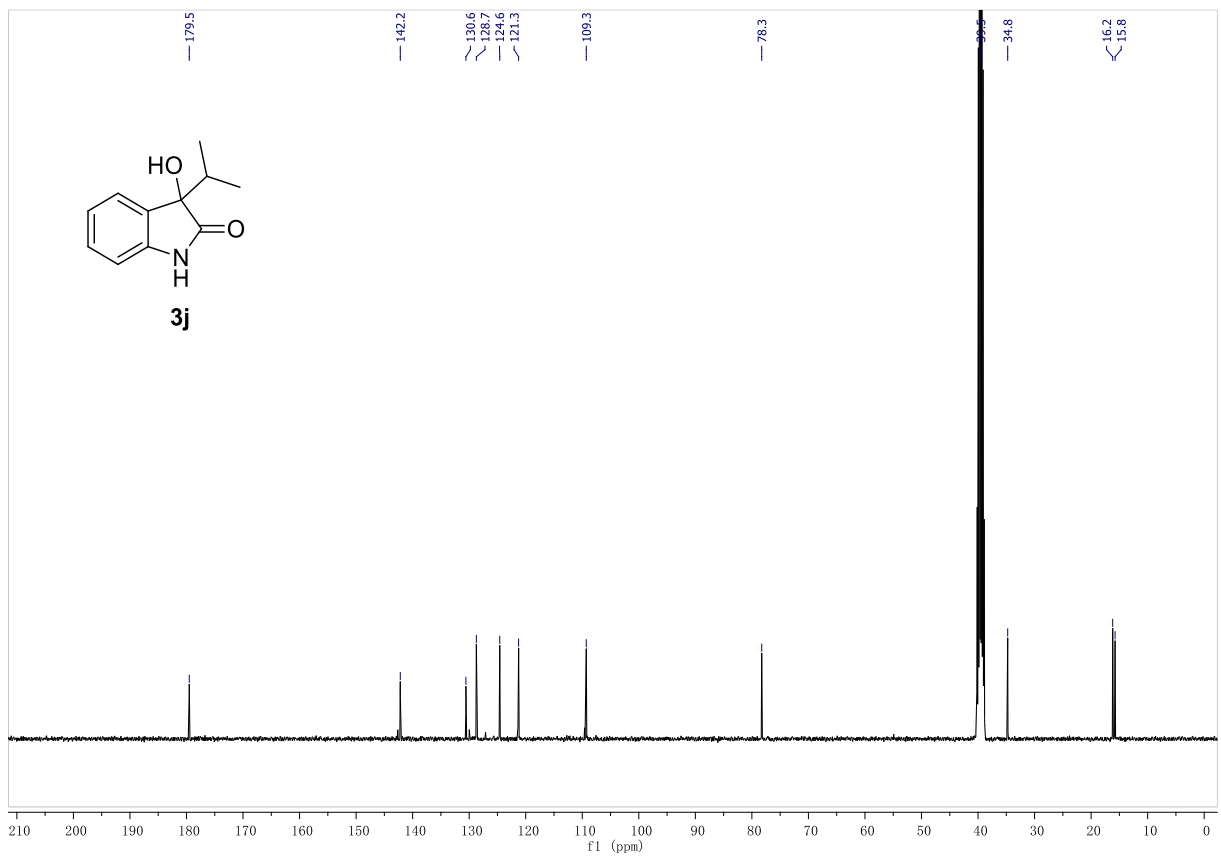
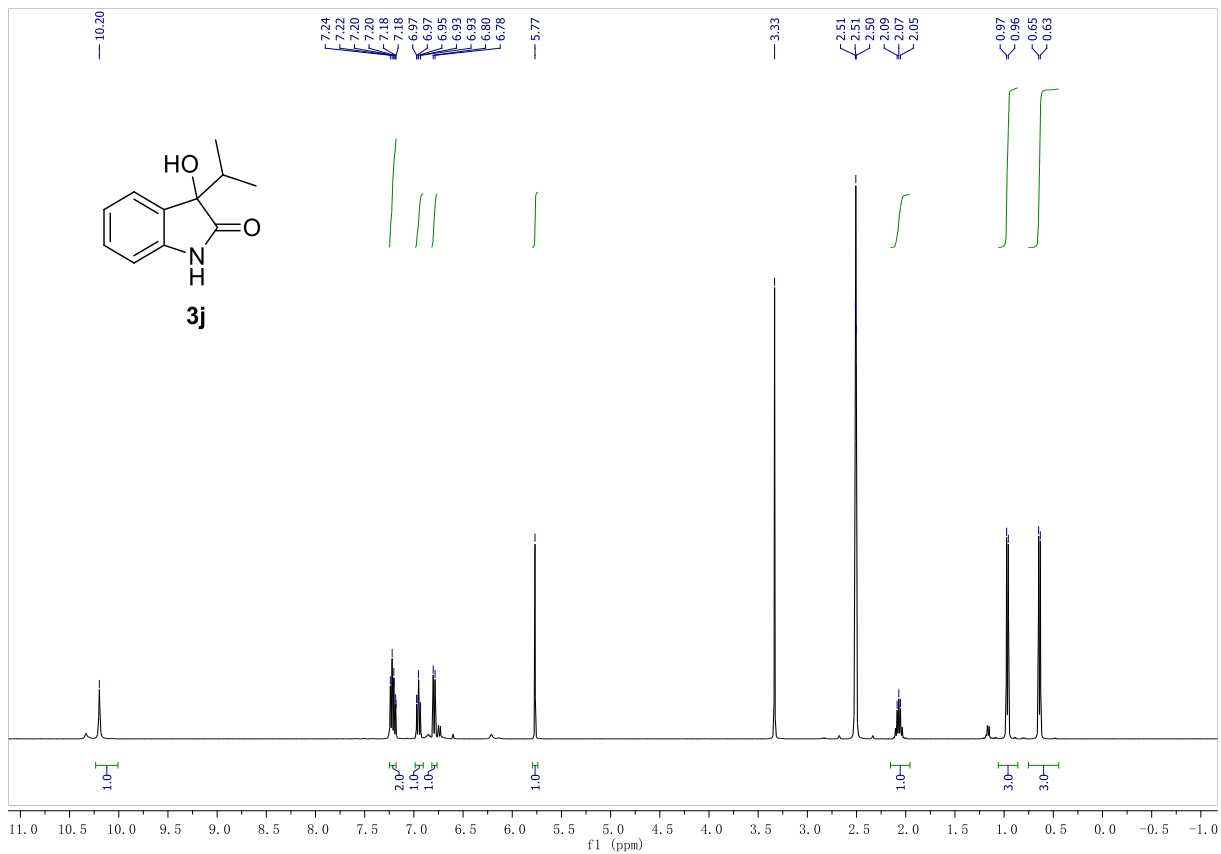


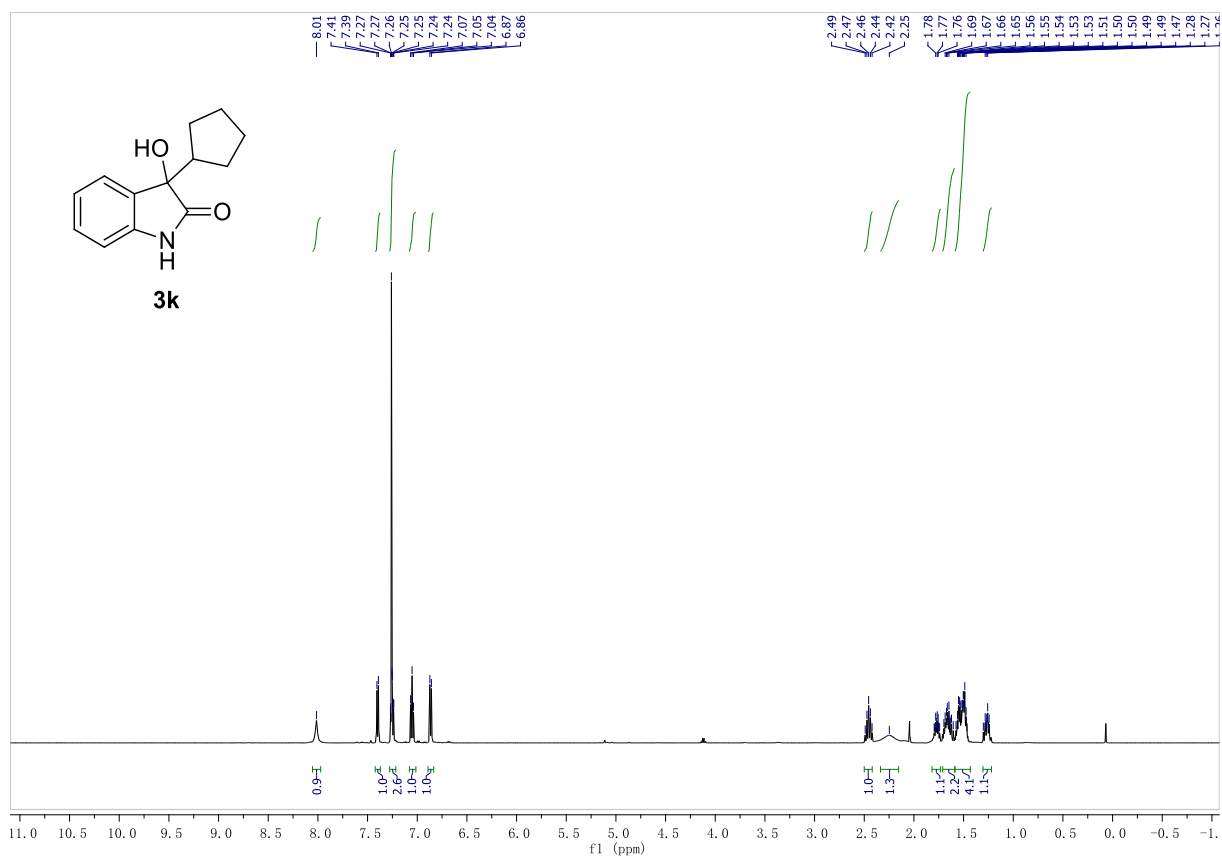
¹³C {¹H} NMR in DMSO-*d*₆ at 175 MHz



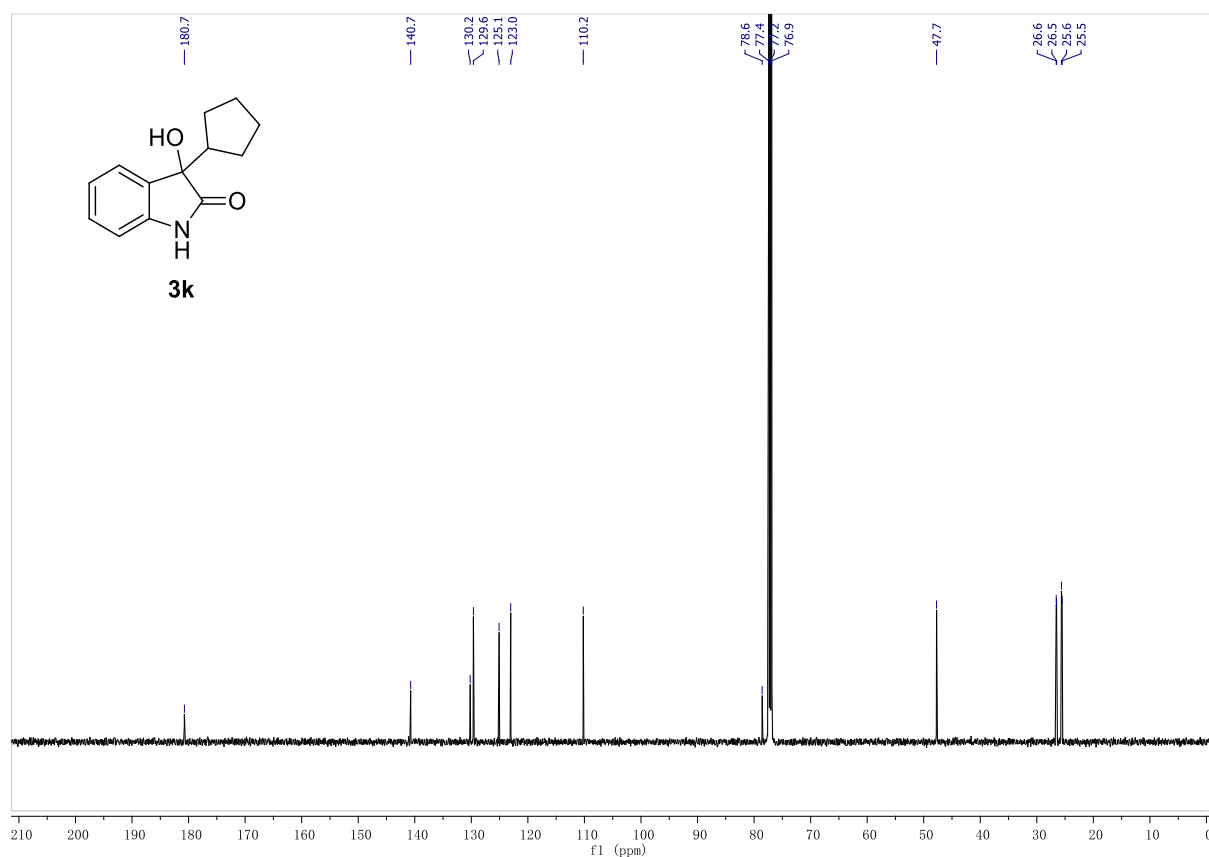
^{19}F NMR in $\text{DMSO-}d_6$ at 659 MHz



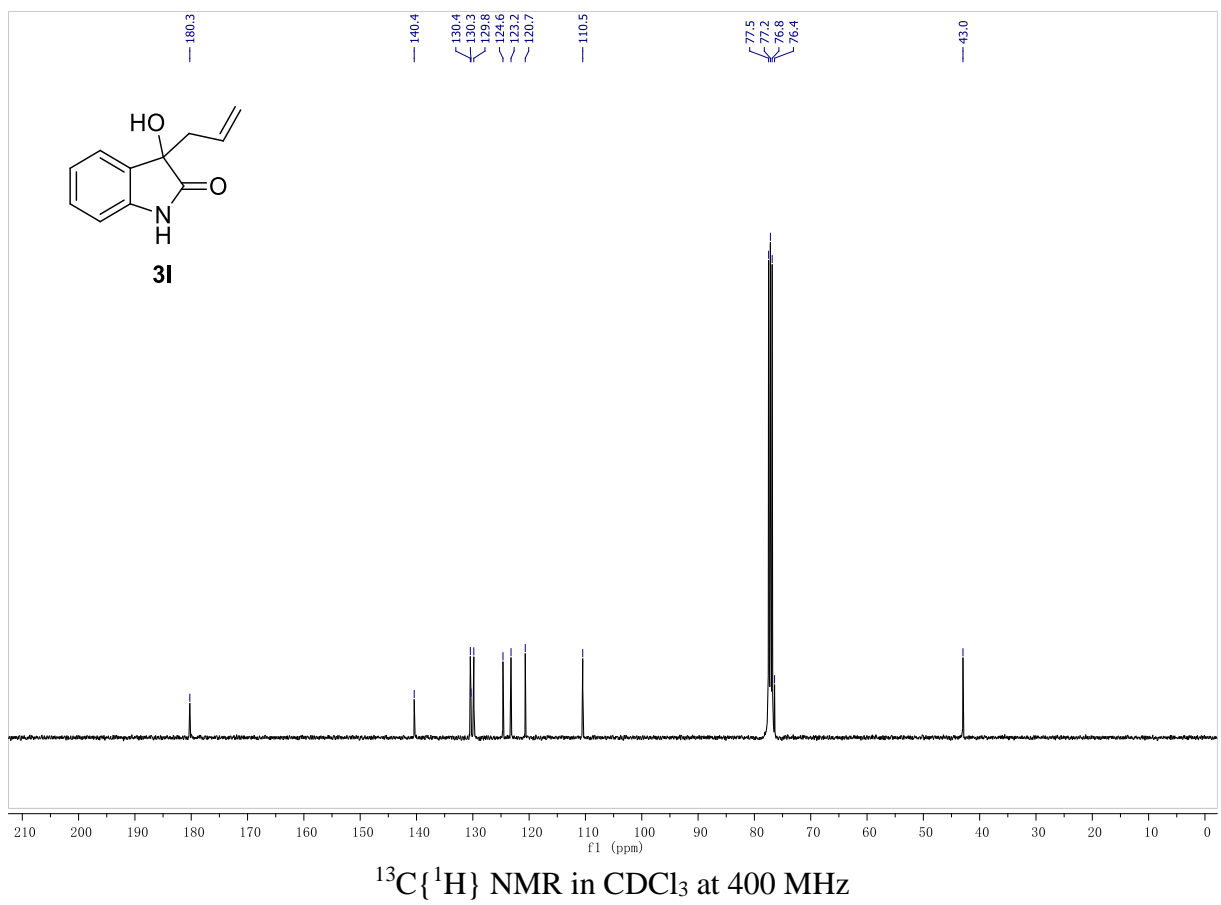
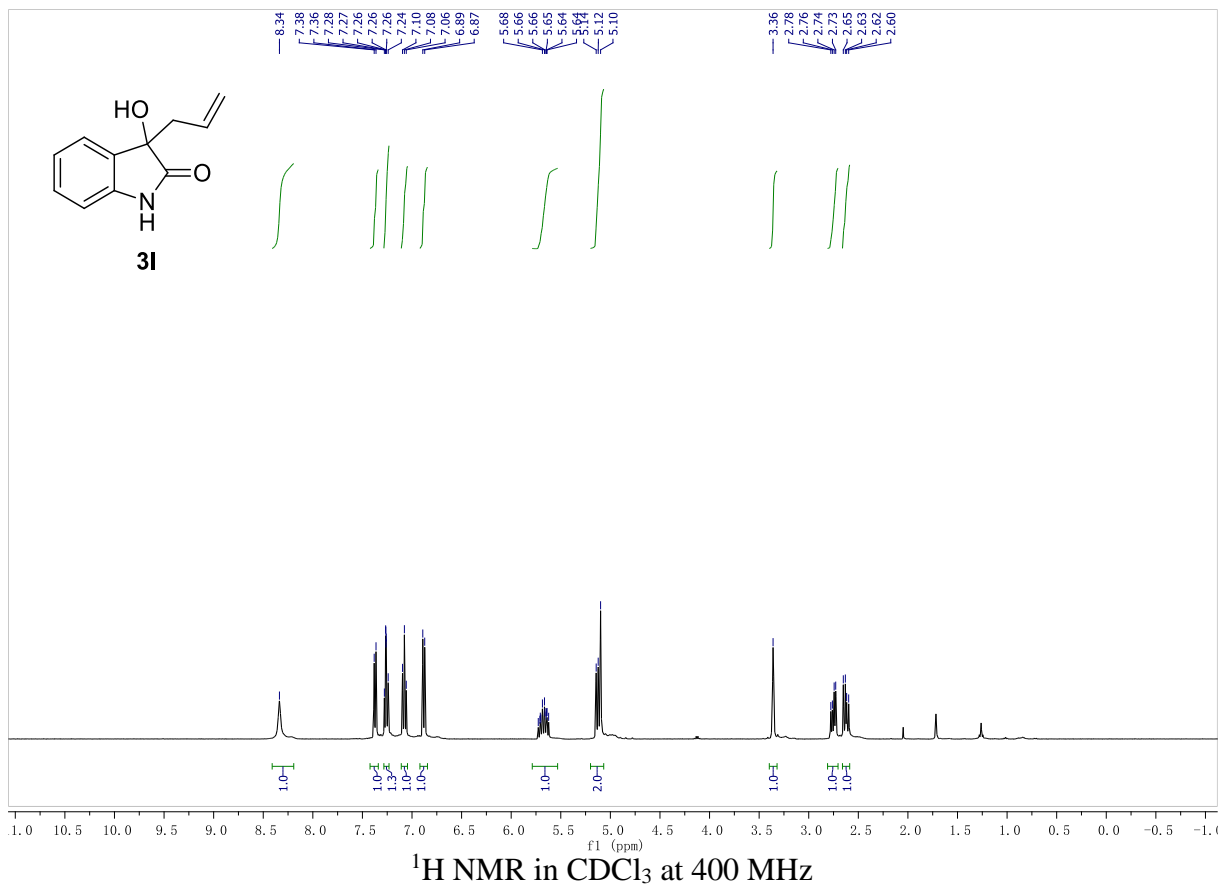


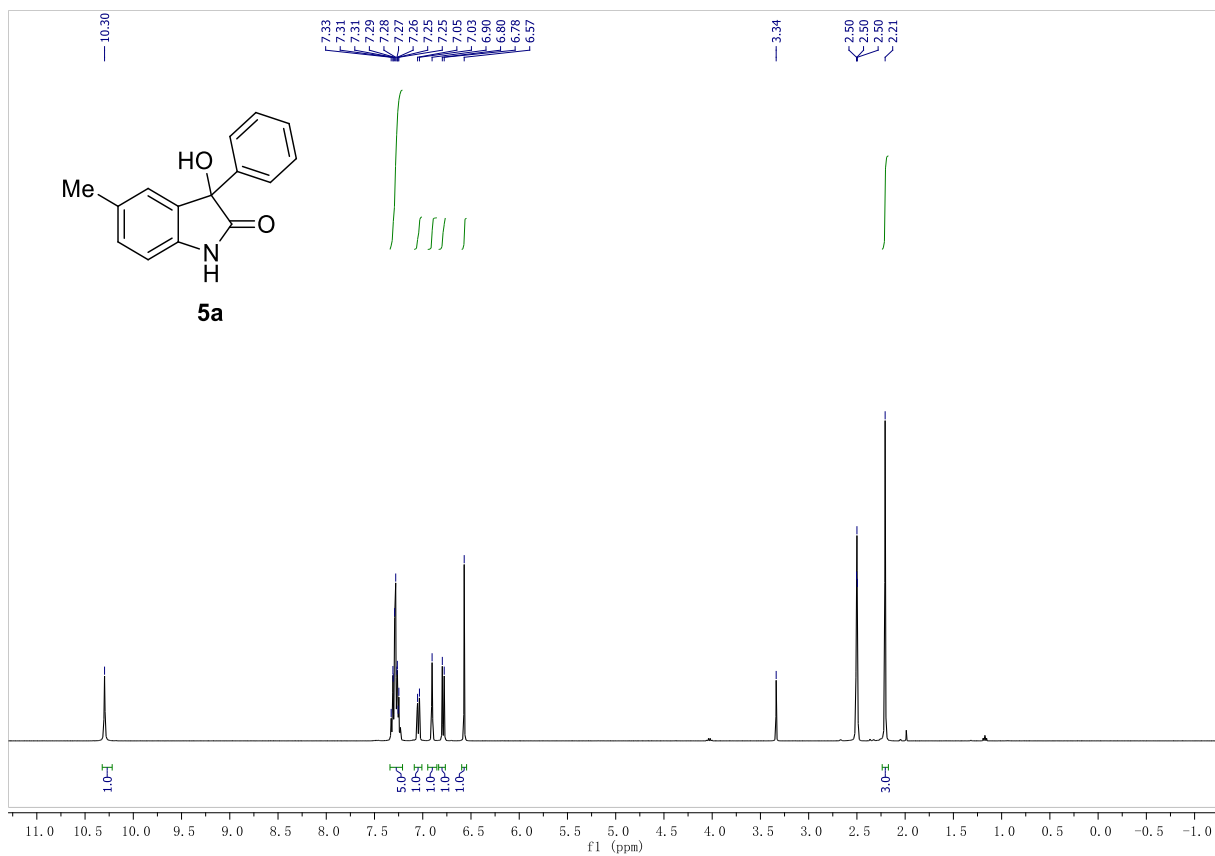


^1H NMR in CDCl_3 at 400 MHz

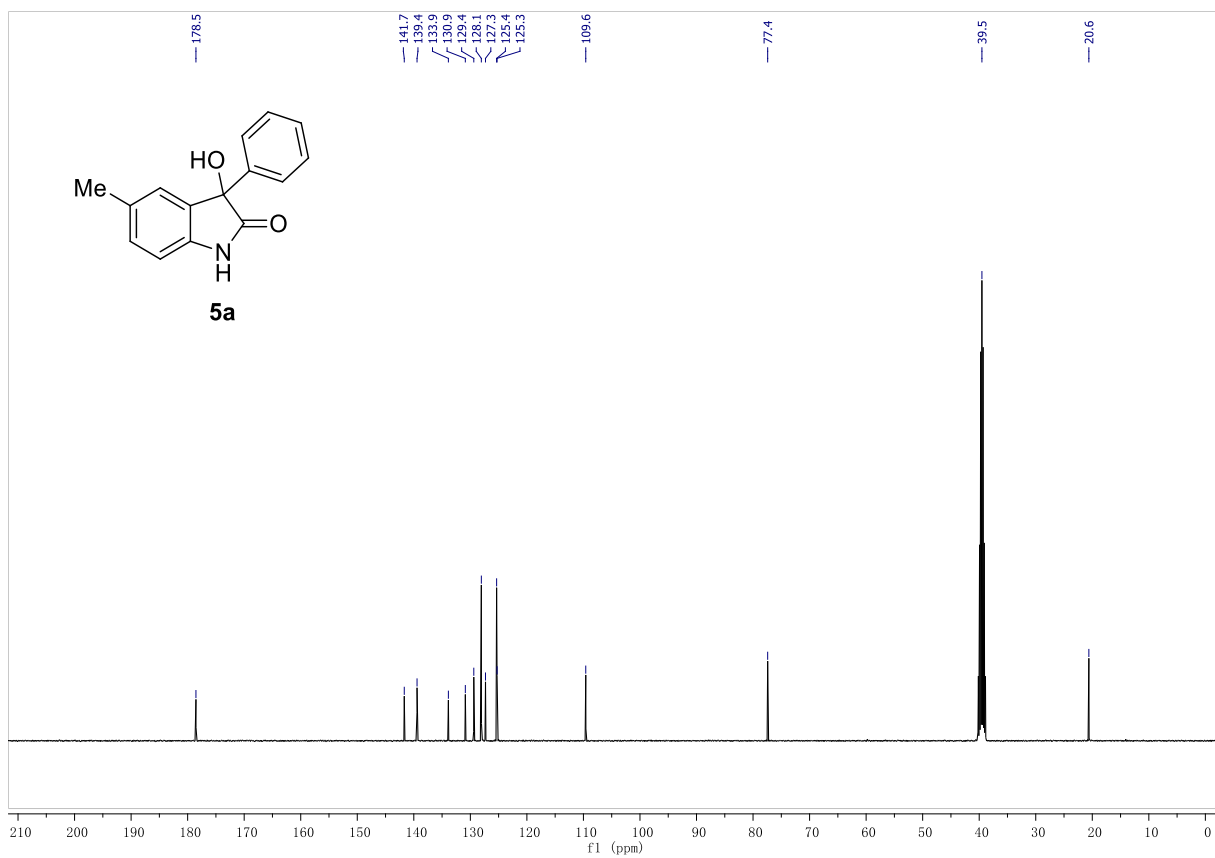


$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 100 MHz

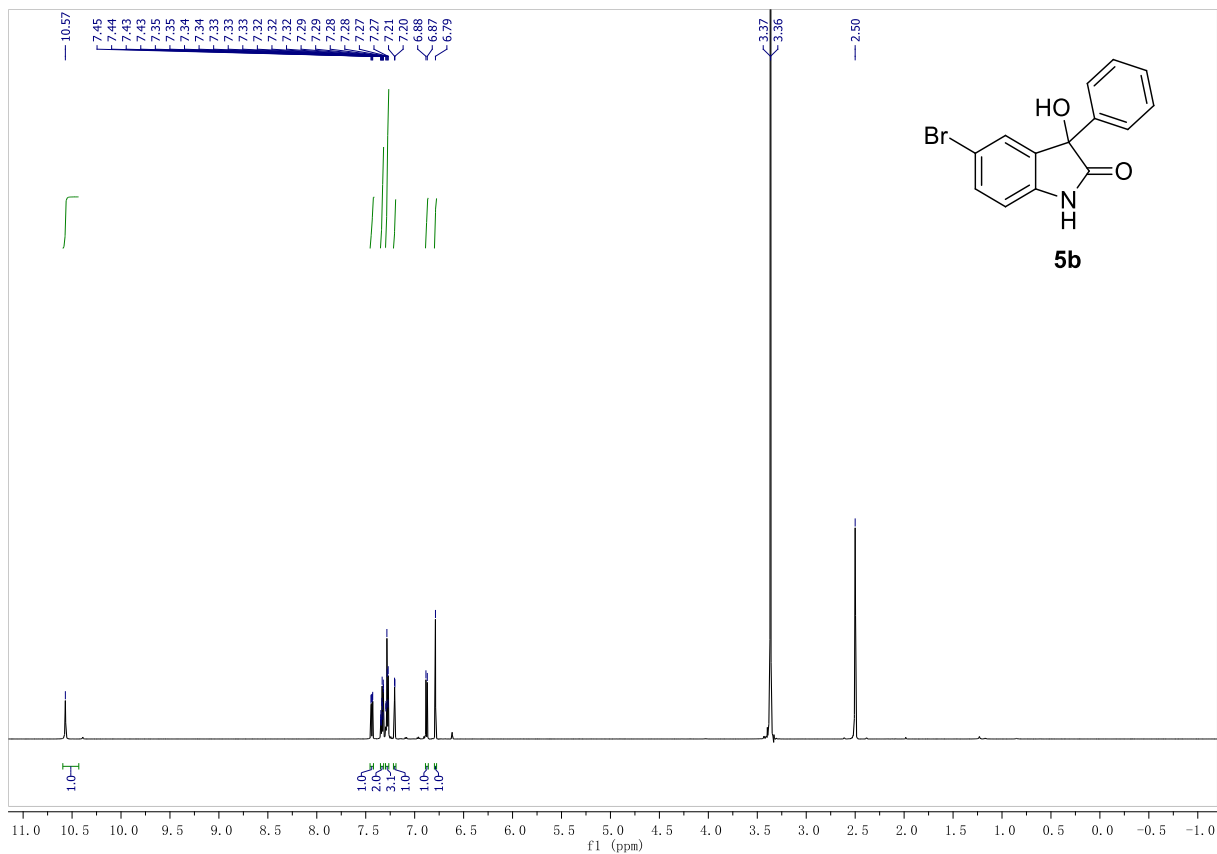




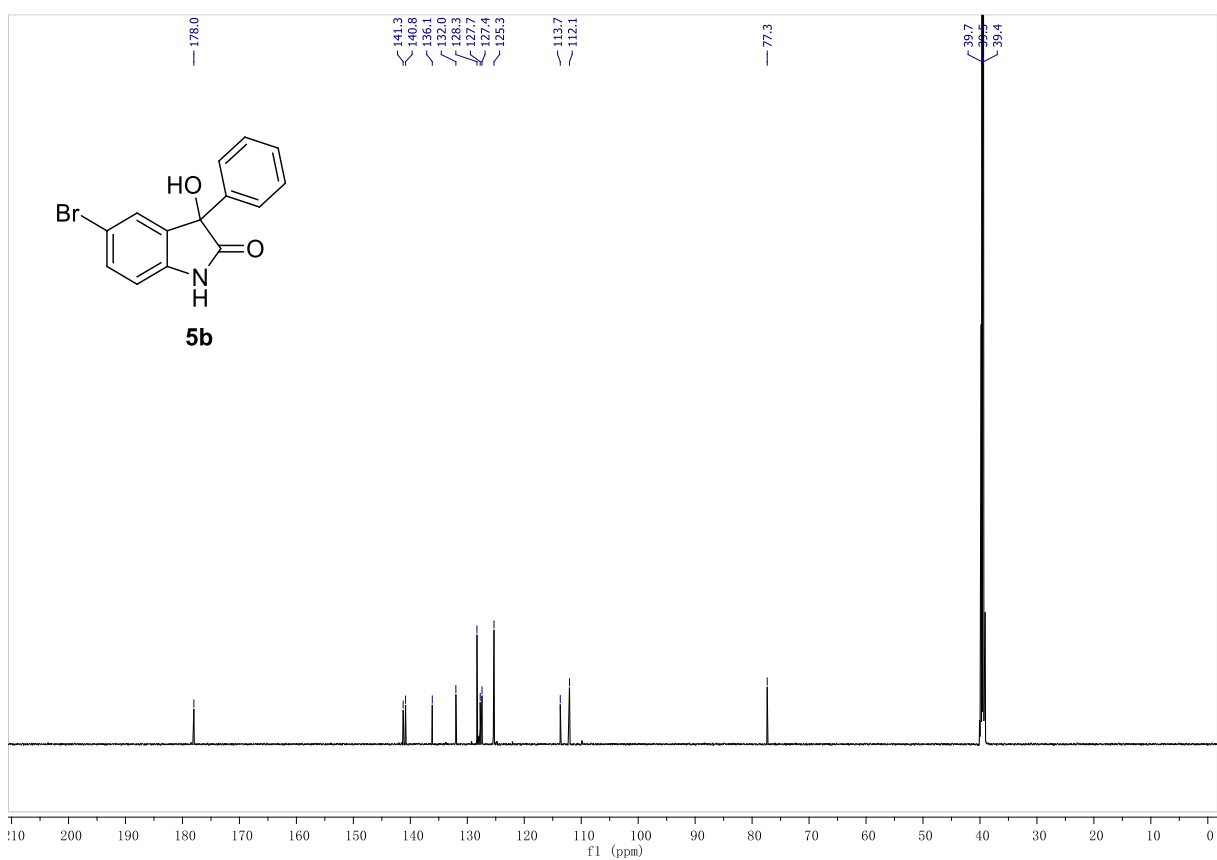
^1H NMR in DMSO- d_6 at 400 MHz



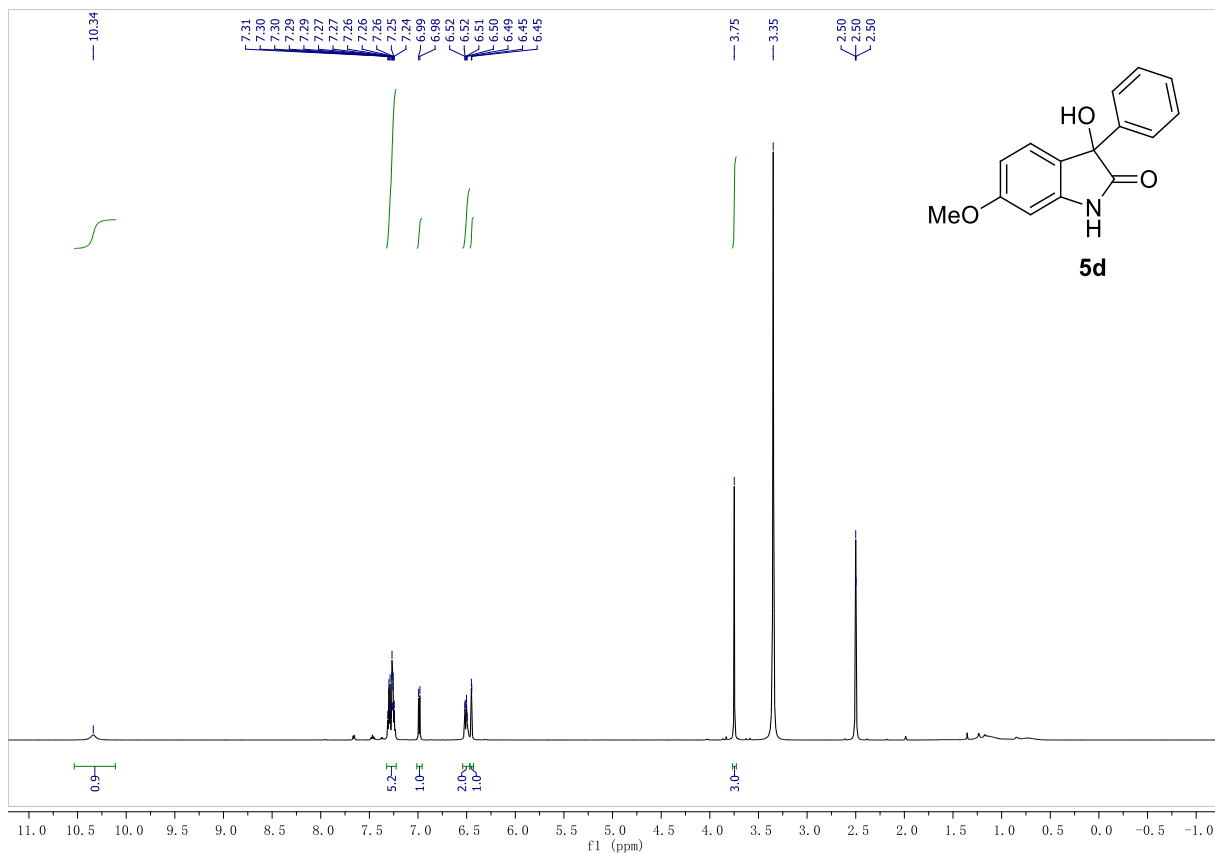
$^{13}\text{C}\{^1\text{H}\}$ NMR in DMSO- d_6 at 100 MHz



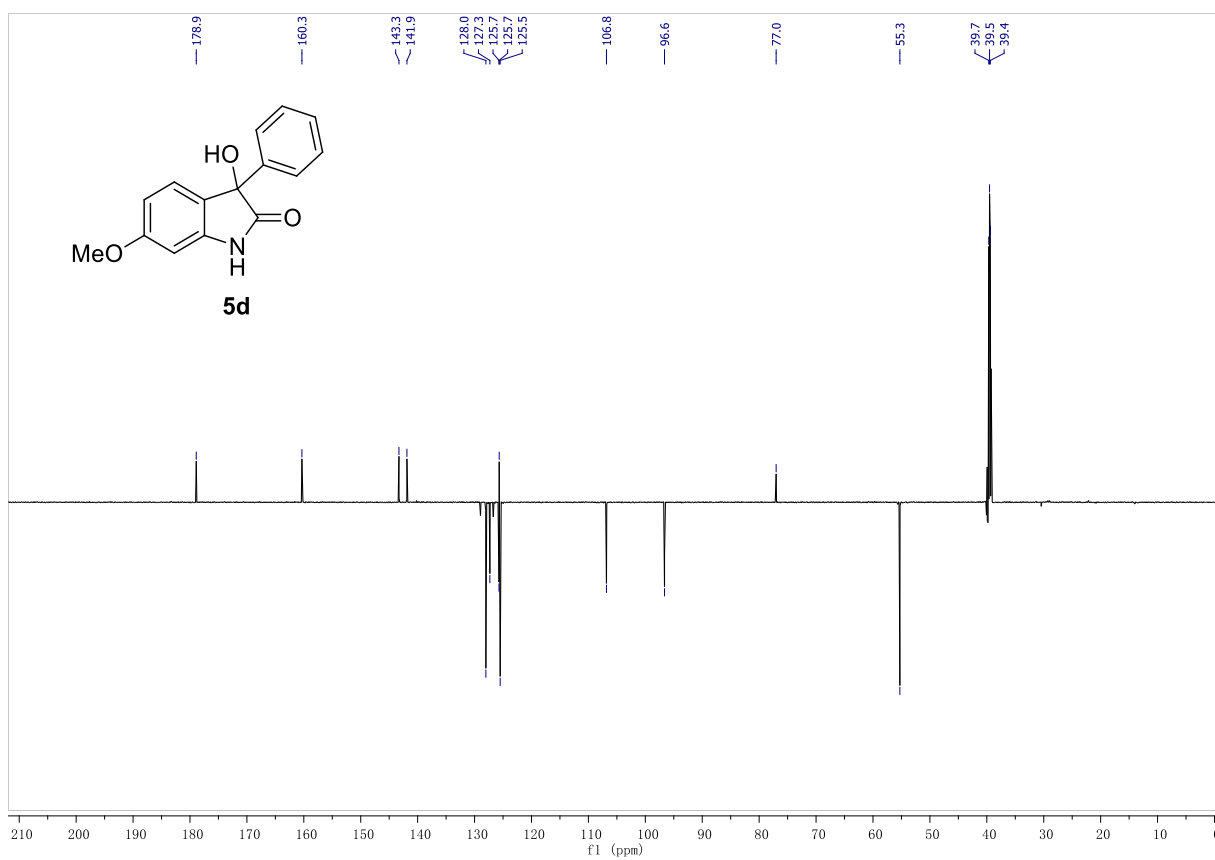
^1H NMR in $\text{DMSO-}d_6$ at 600 MHz



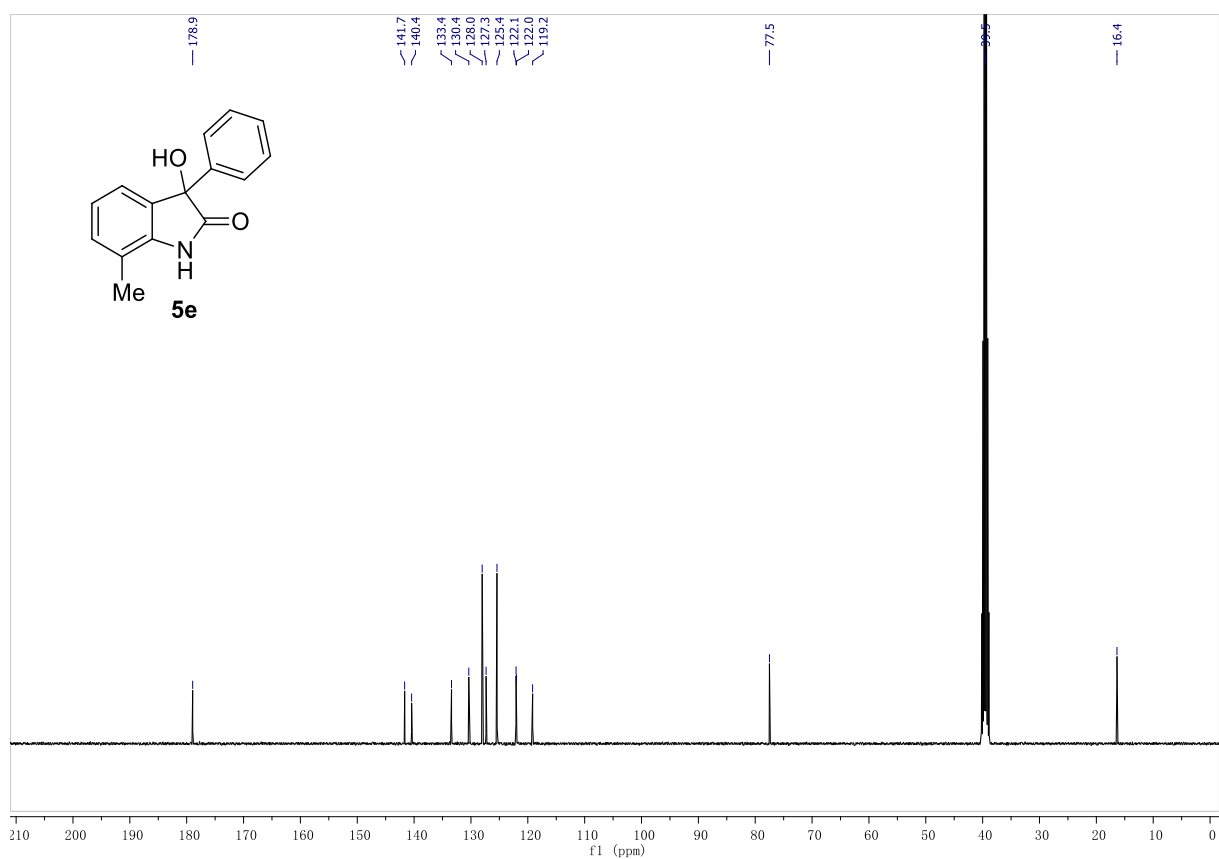
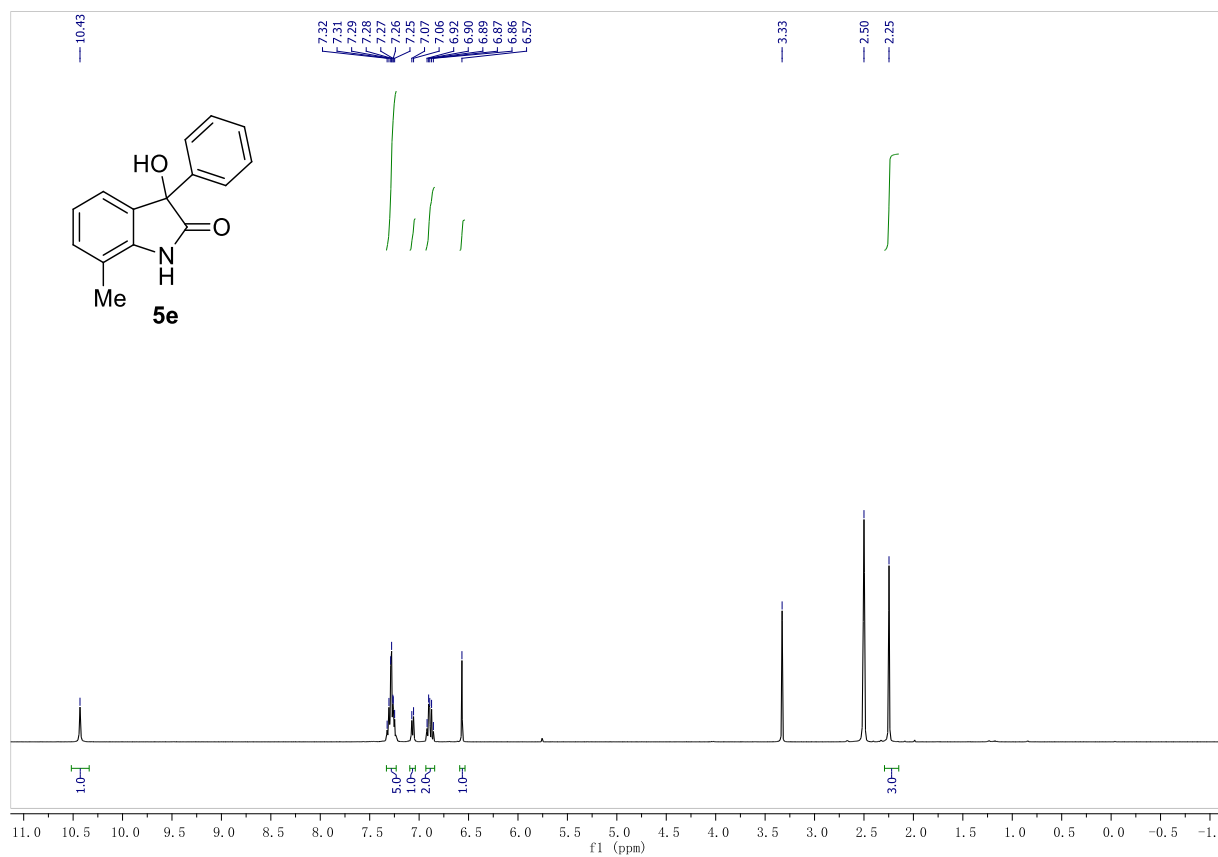
$^{13}\text{C}\{^1\text{H}\}$ NMR in $\text{DMSO-}d_6$ at 150 MHz

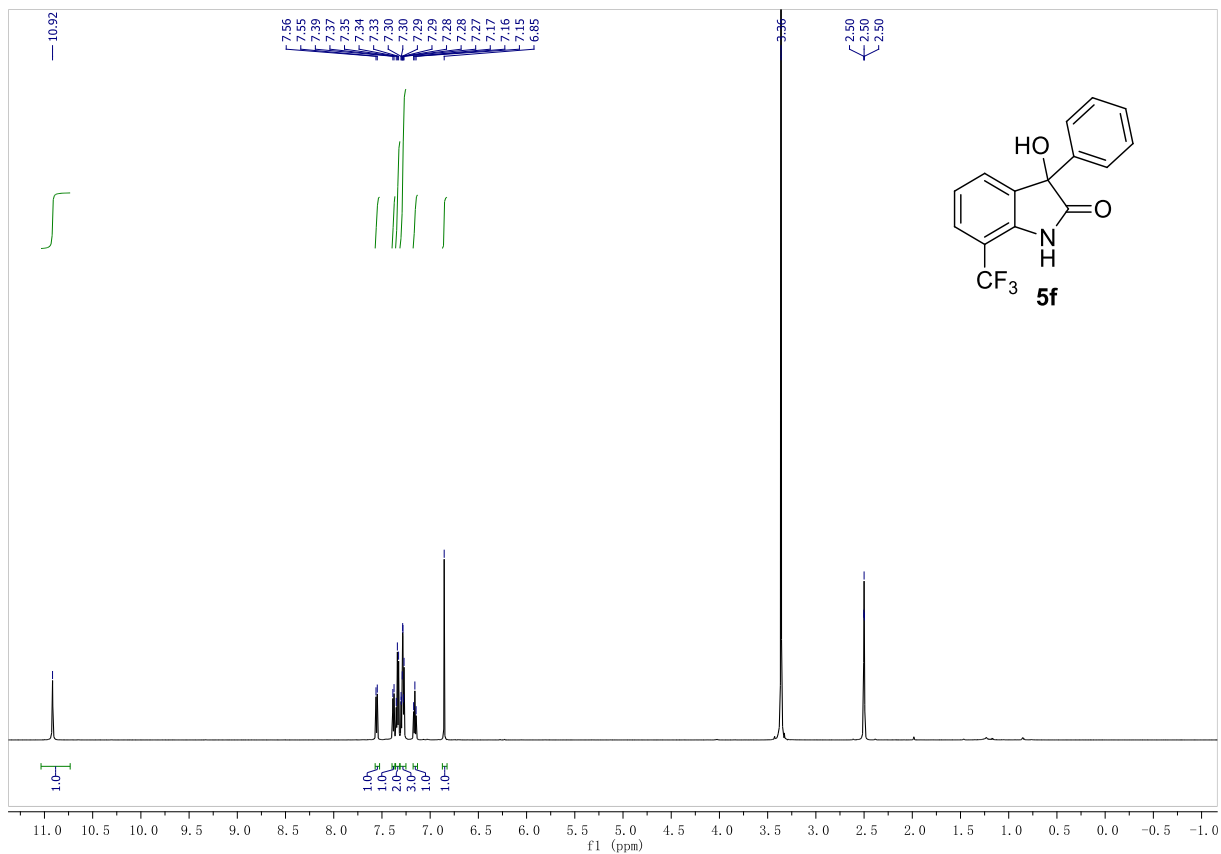


$^1\text{H NMR}$ in $\text{DMSO-}d_6$ at 600 MHz

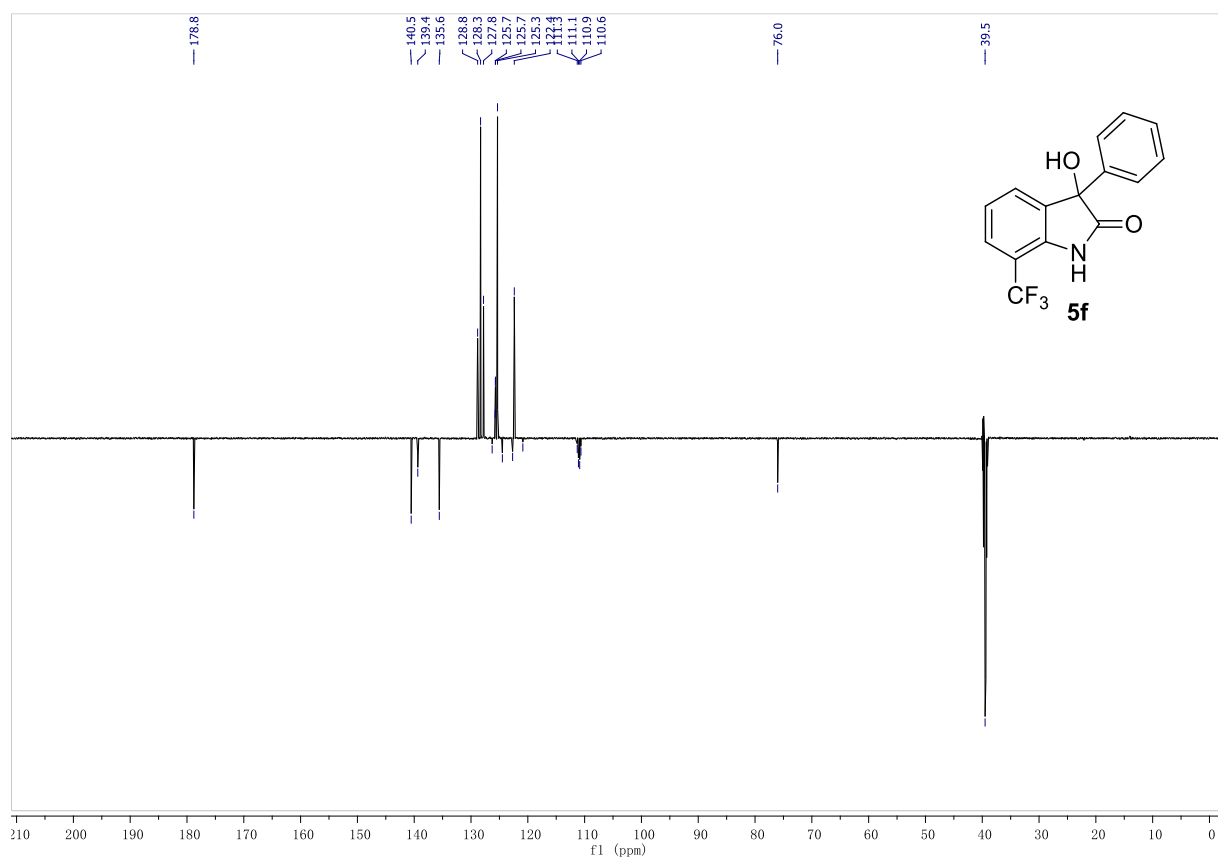


$^{13}\text{C}\{^1\text{H}\}$ NMR in $\text{DMSO-}d_6$ at 150 MHz

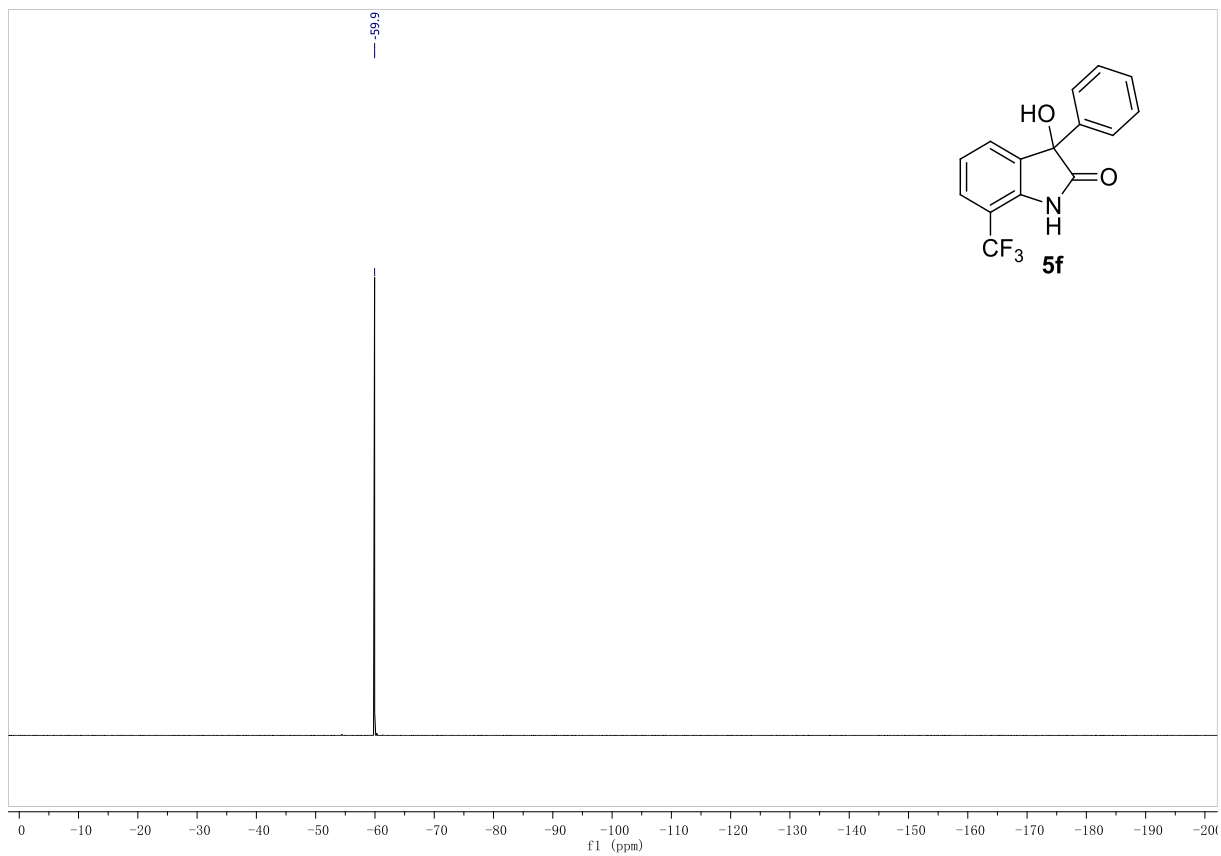


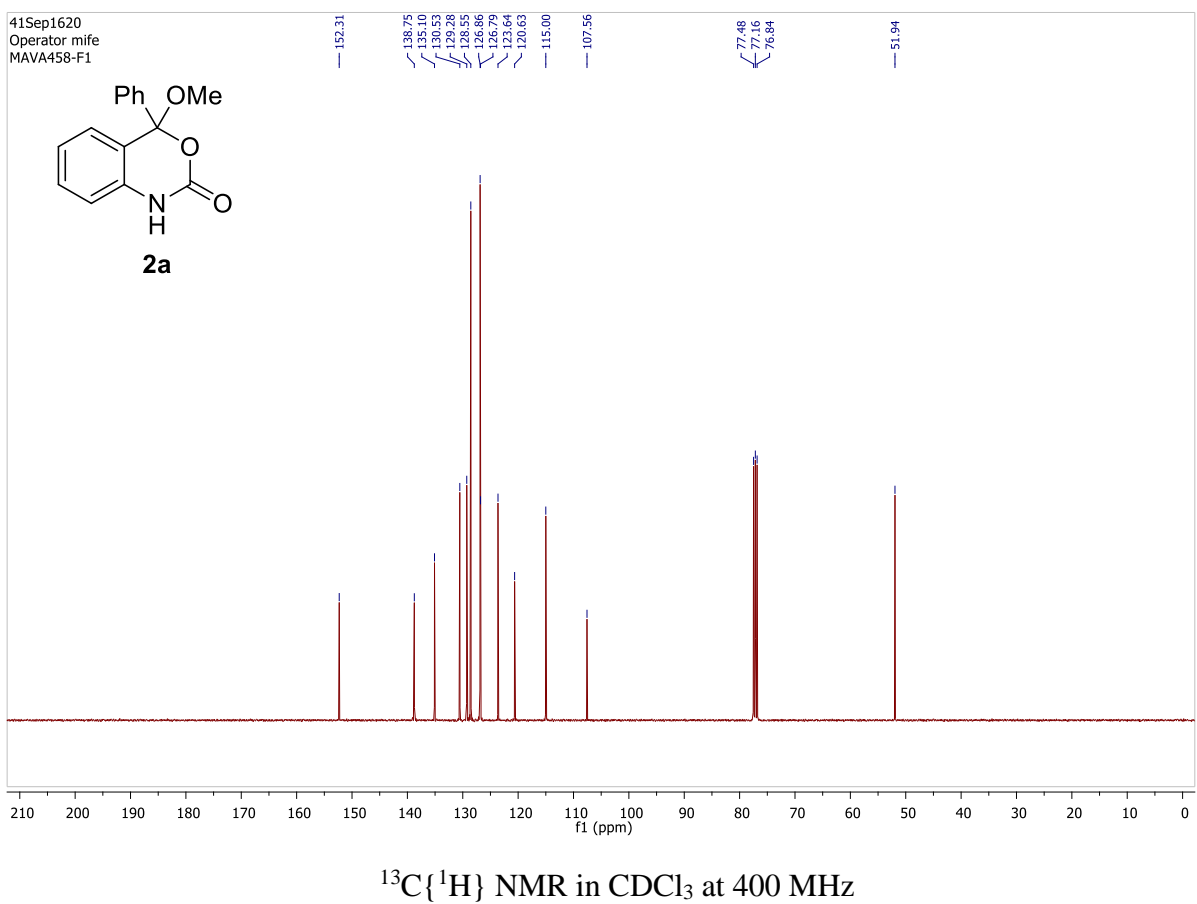
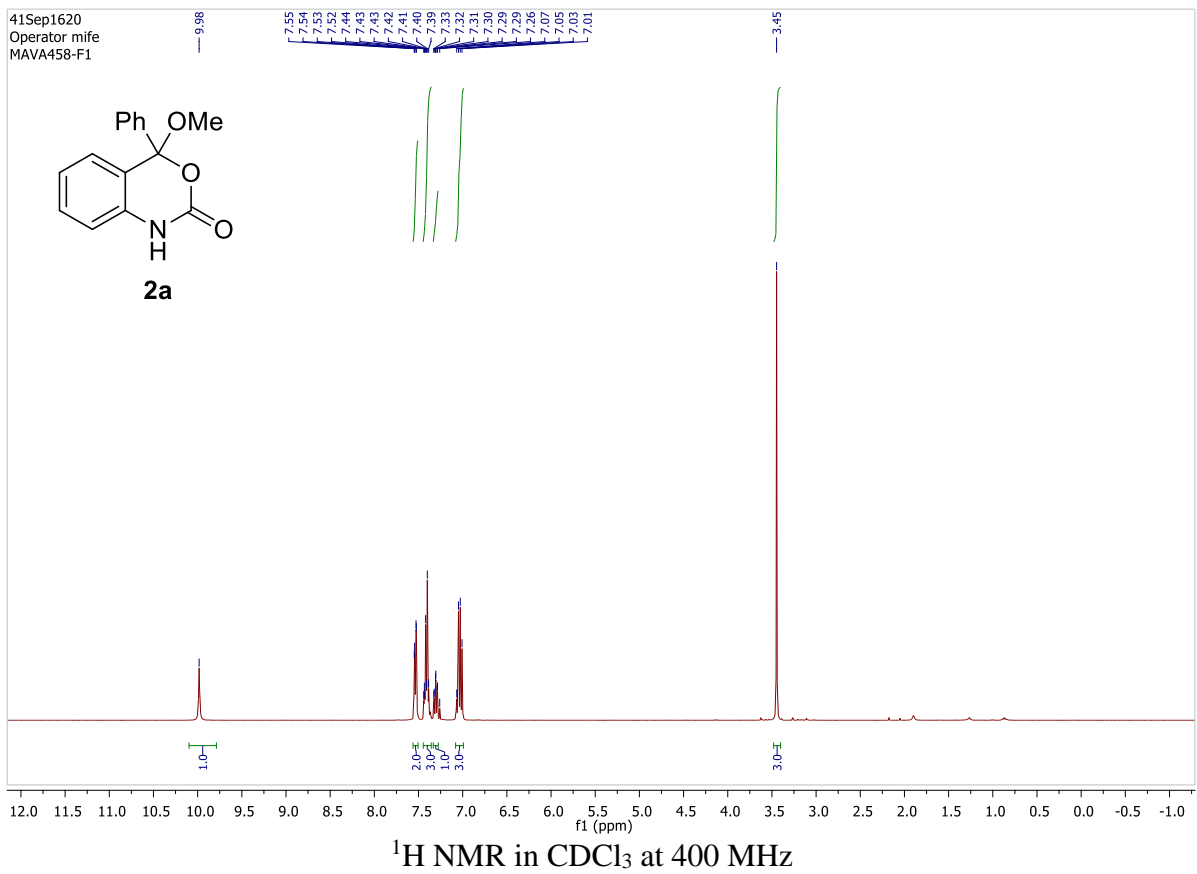


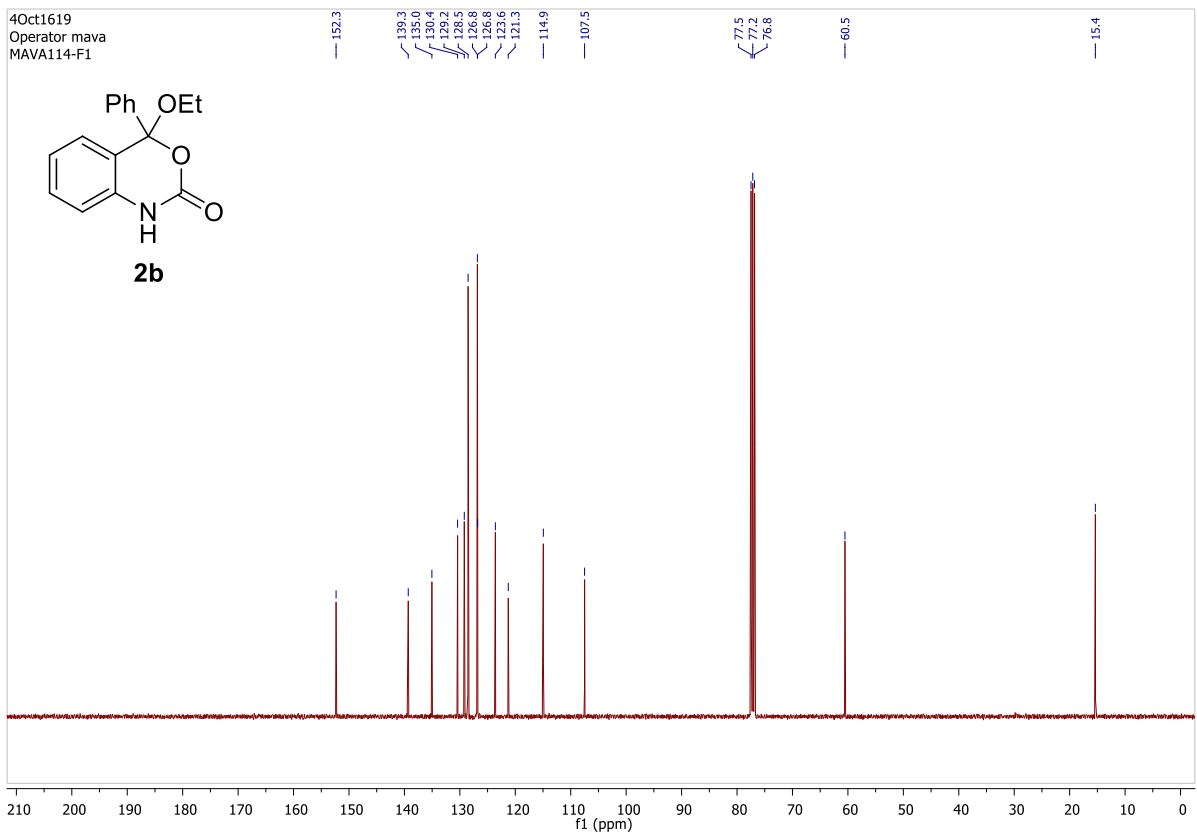
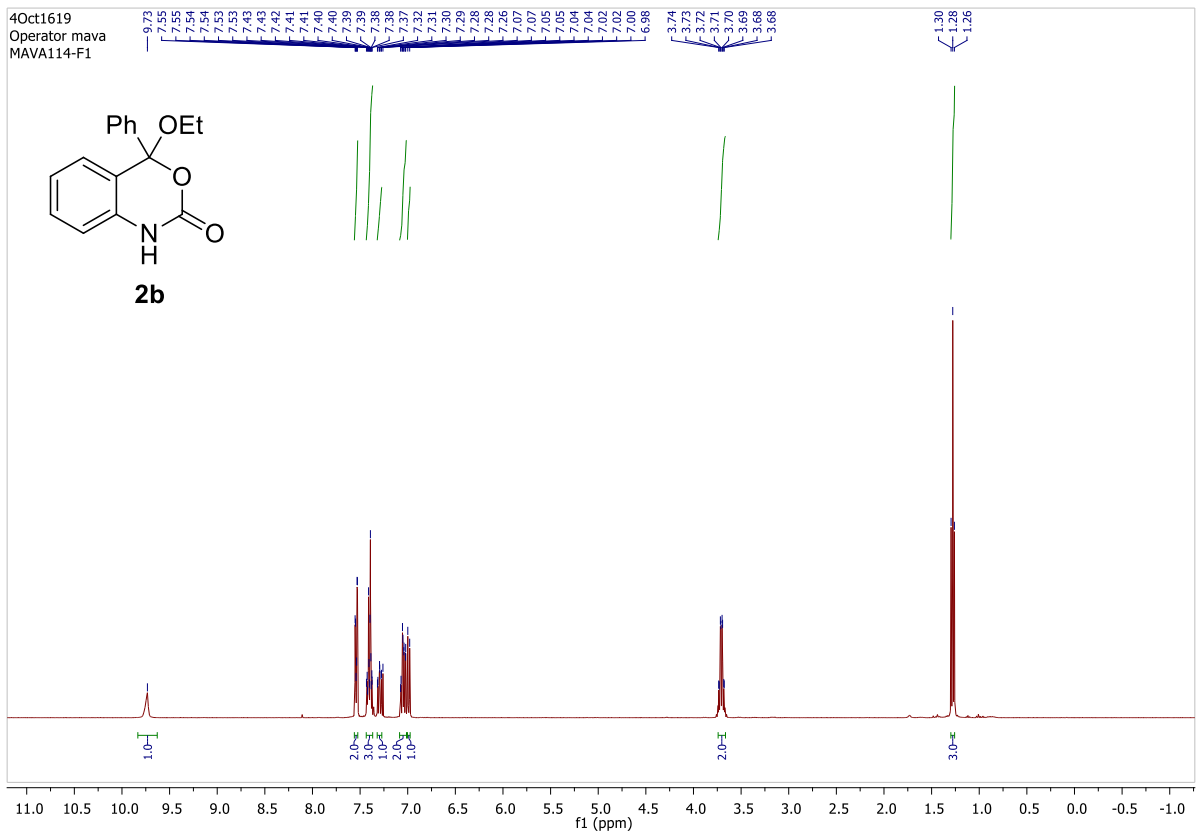
^1H NMR in $\text{DMSO-}d_6$ at 600 MHz

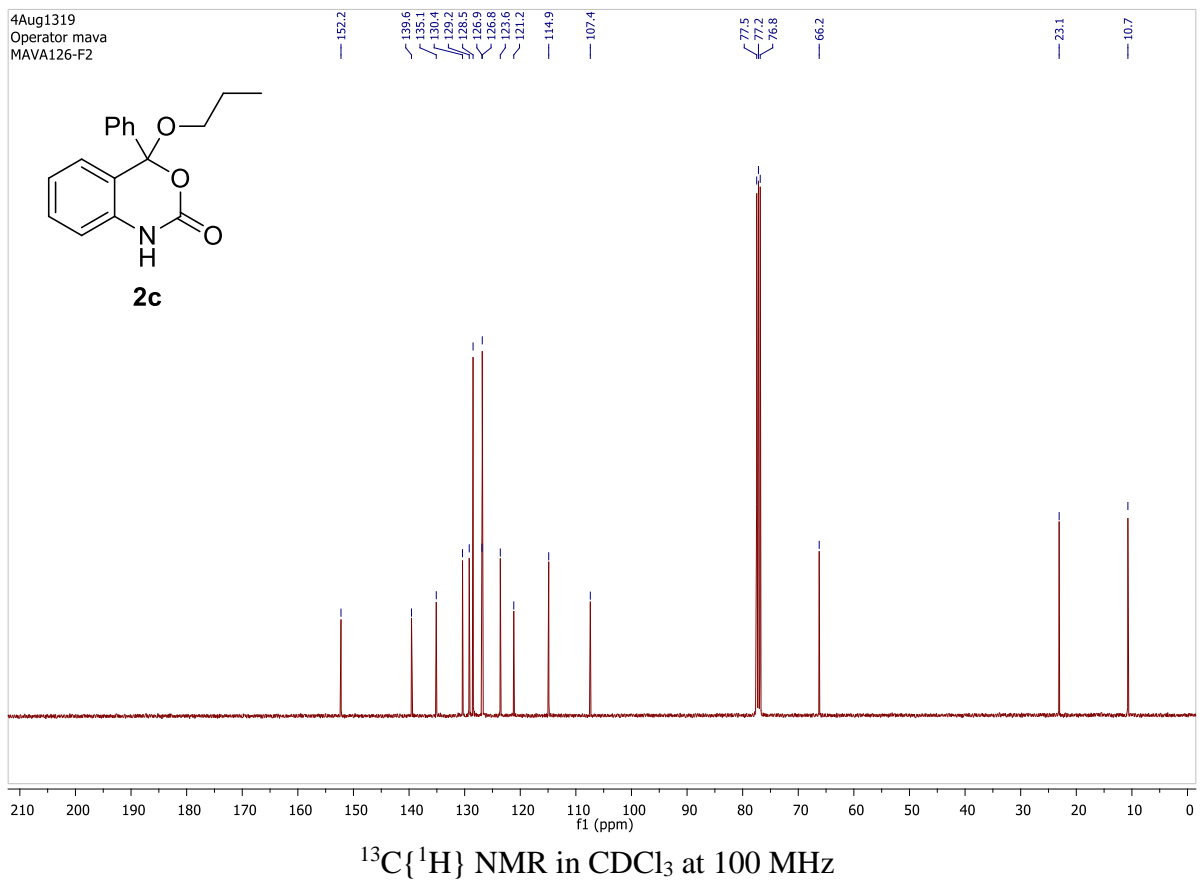
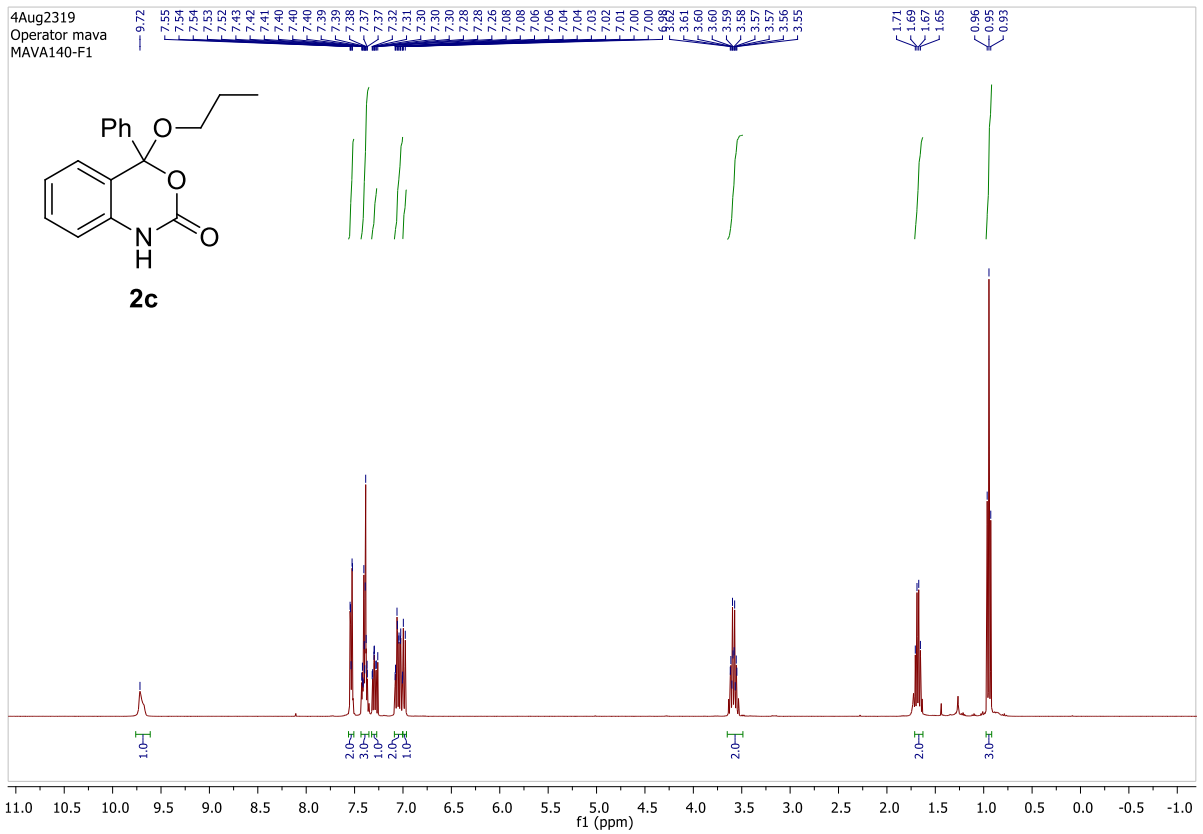


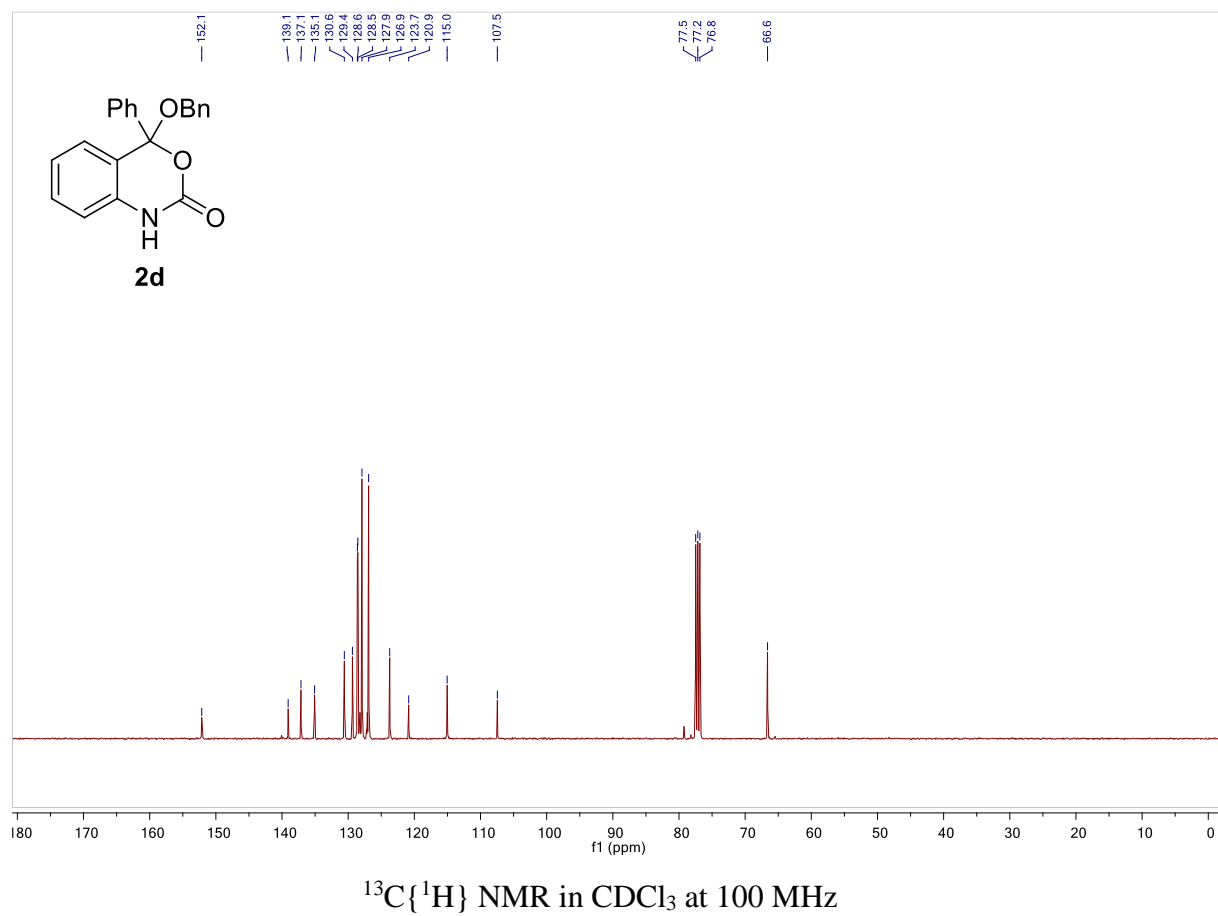
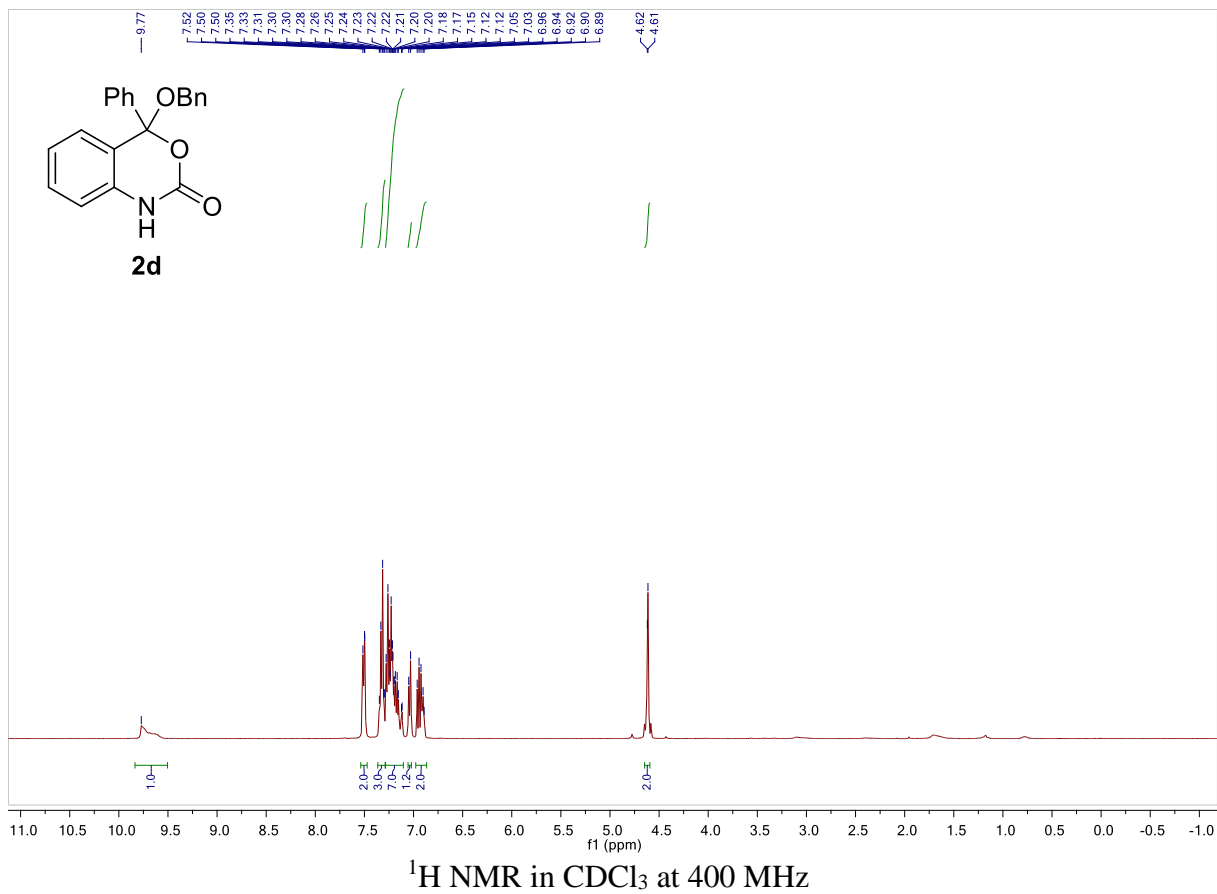
$^{13}\text{C}\{^1\text{H}\}$ NMR in $\text{DMSO-}d_6$ at 150 MHz

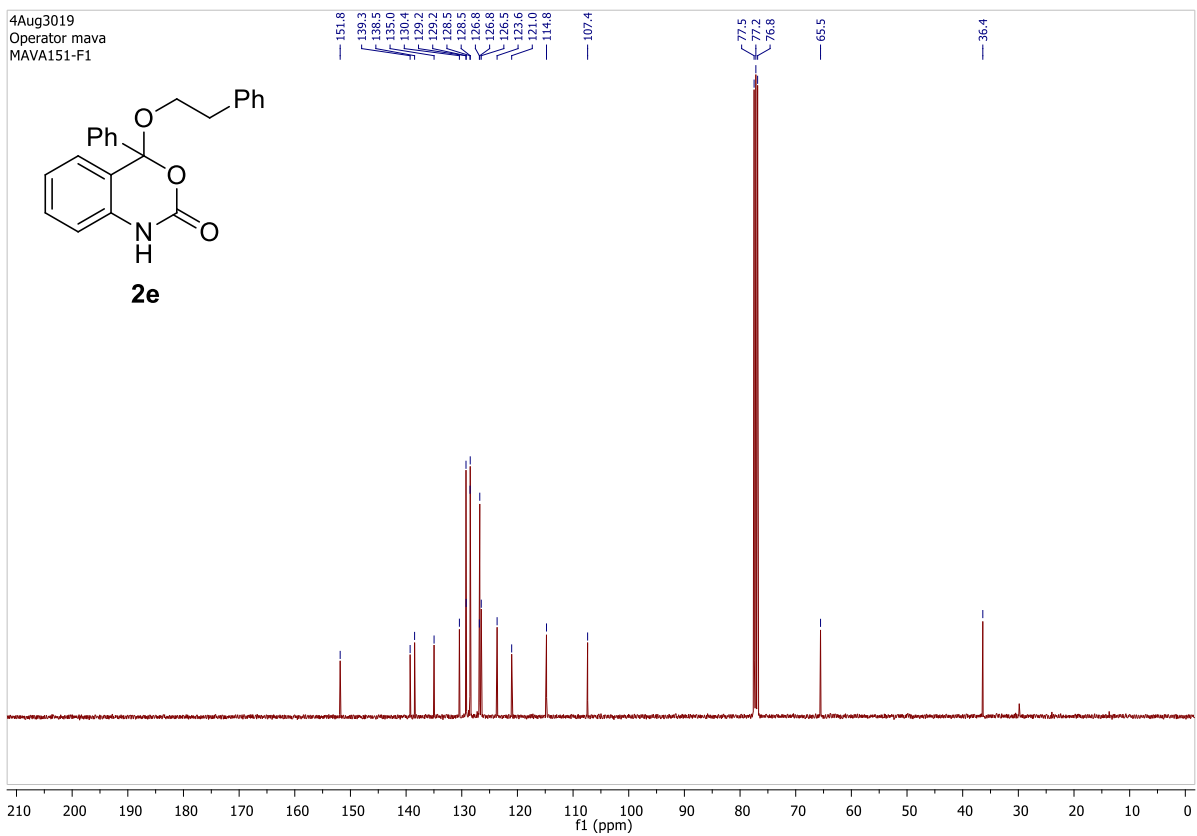
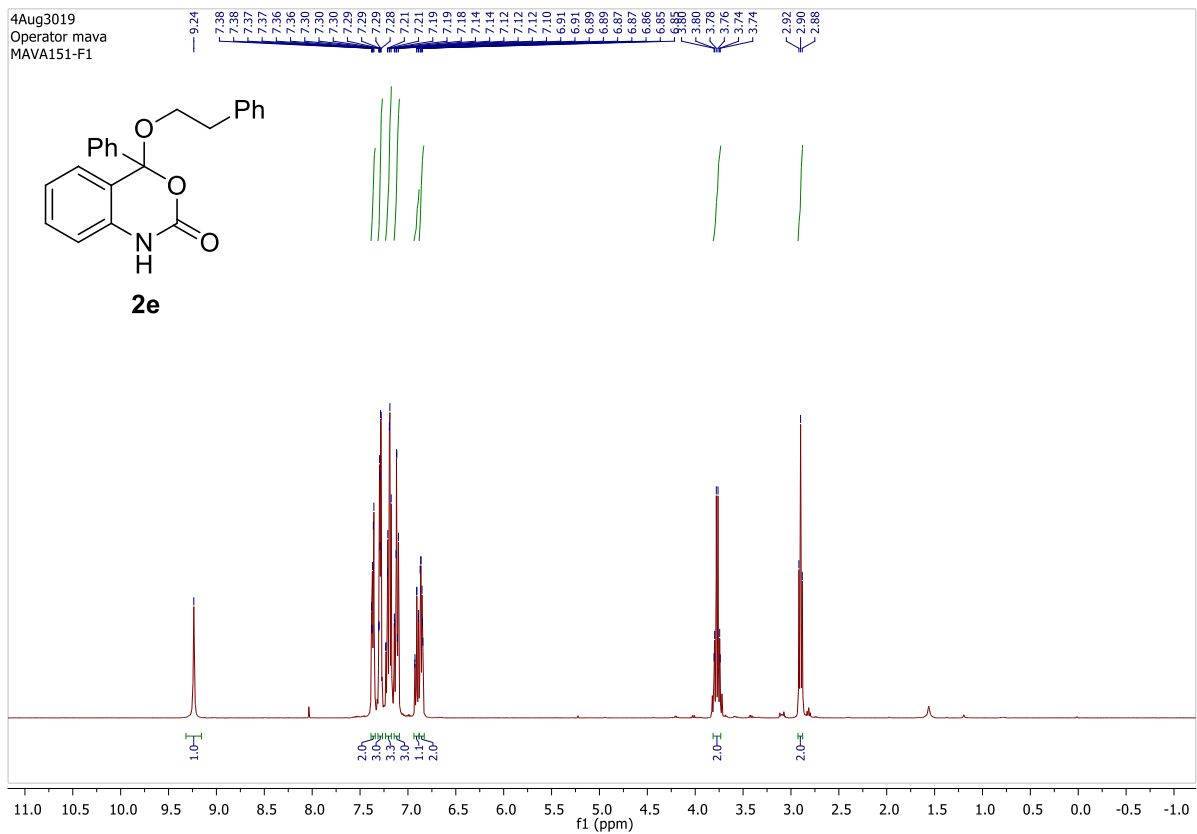


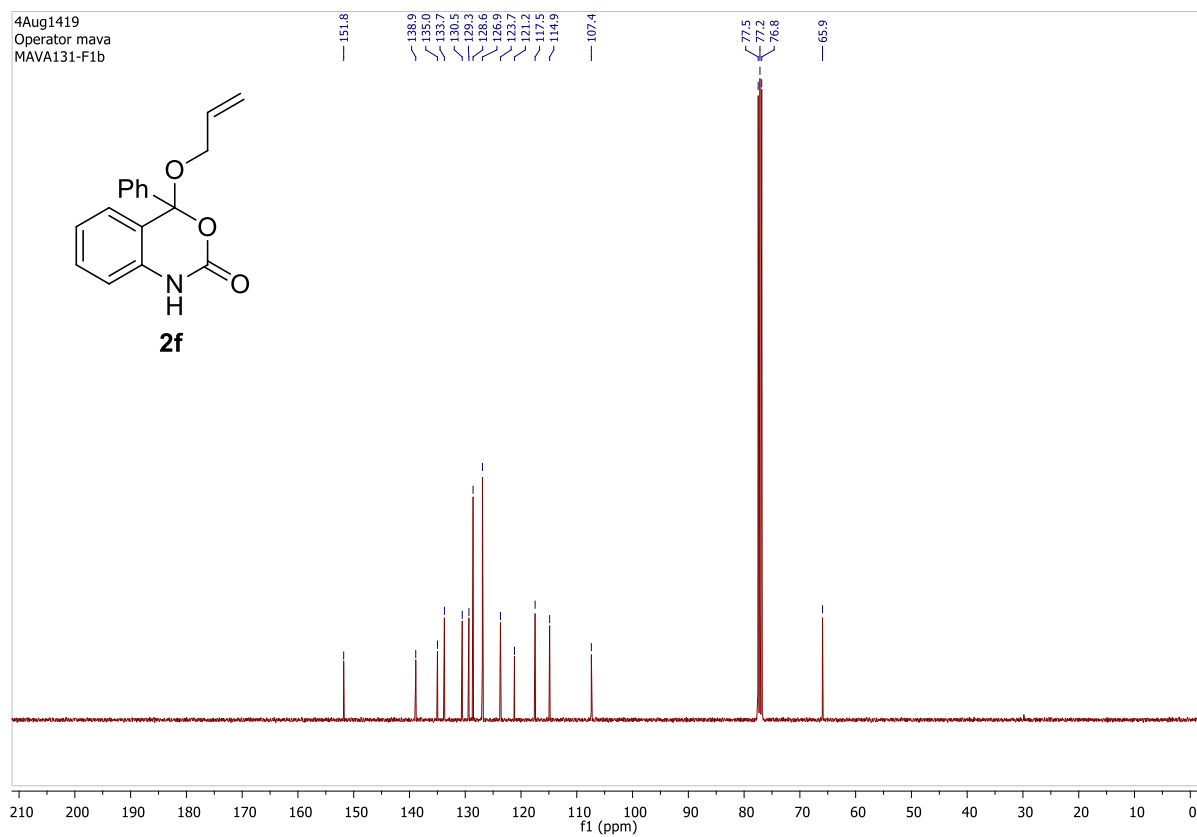
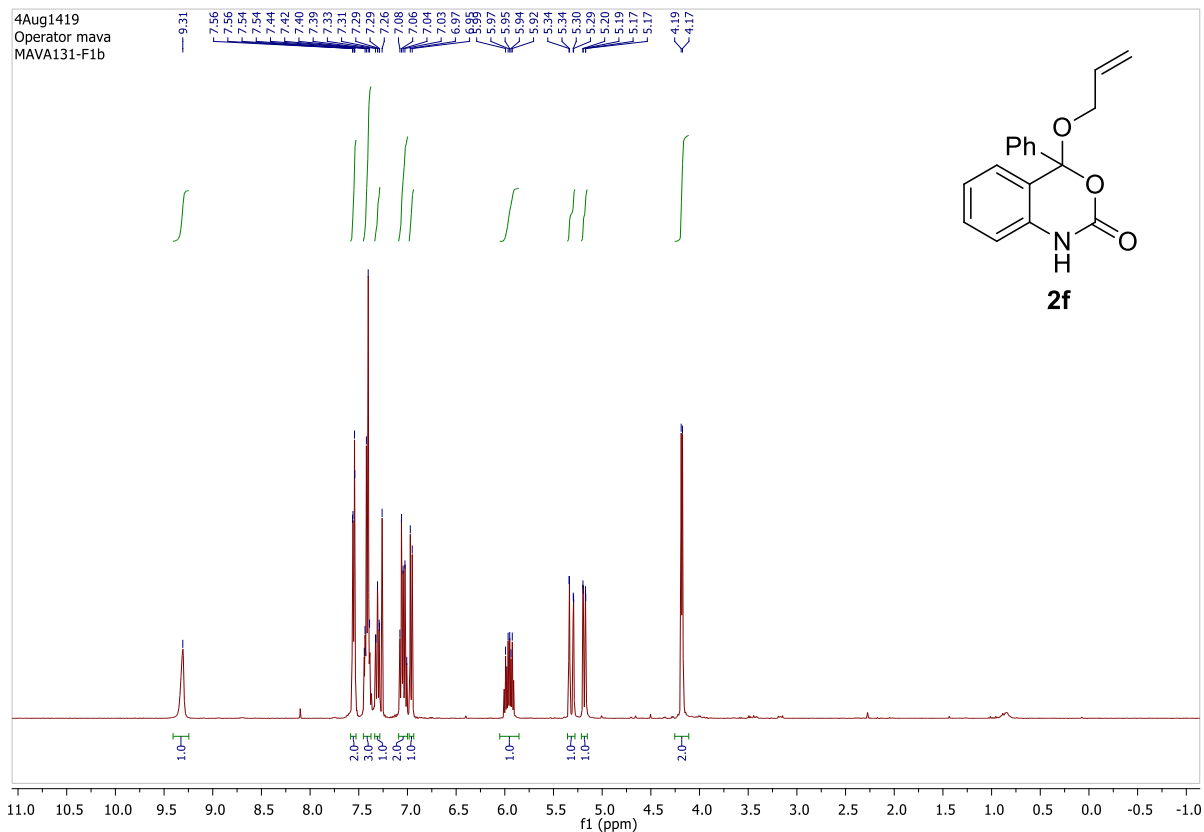


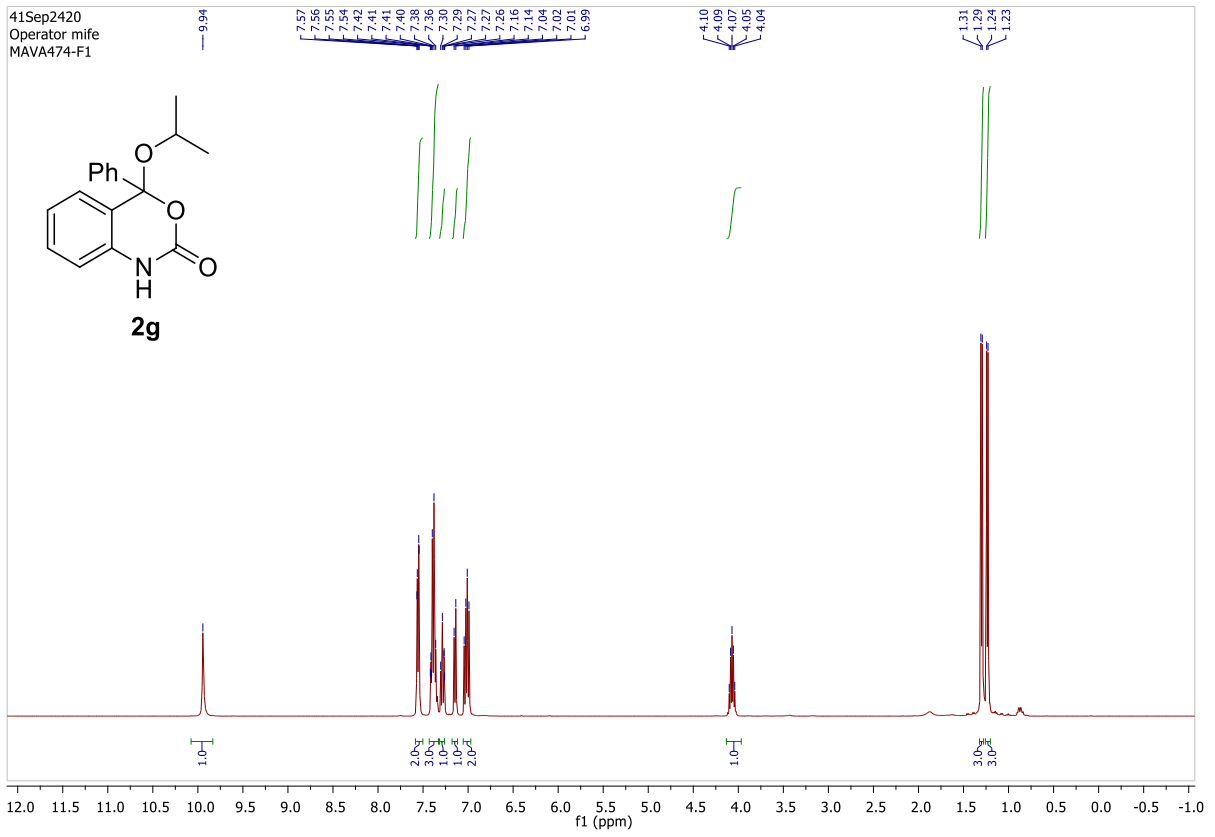




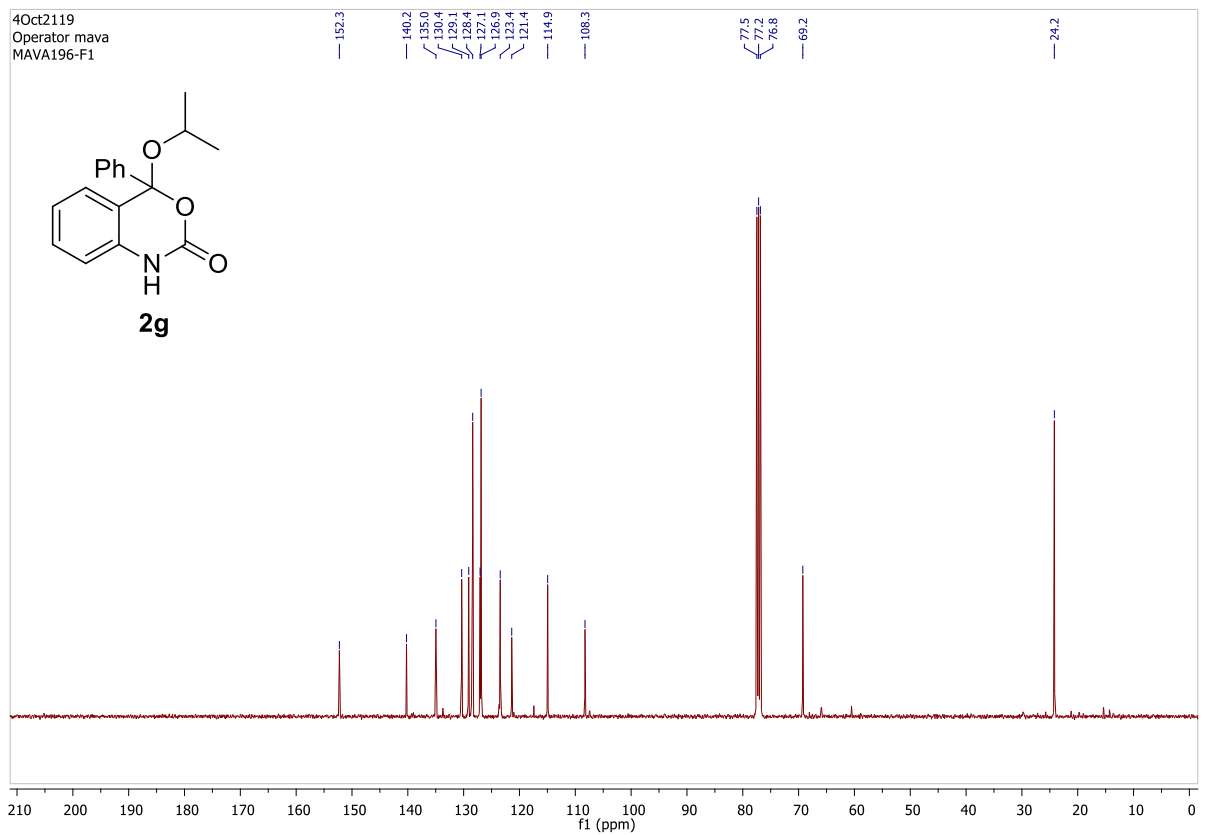




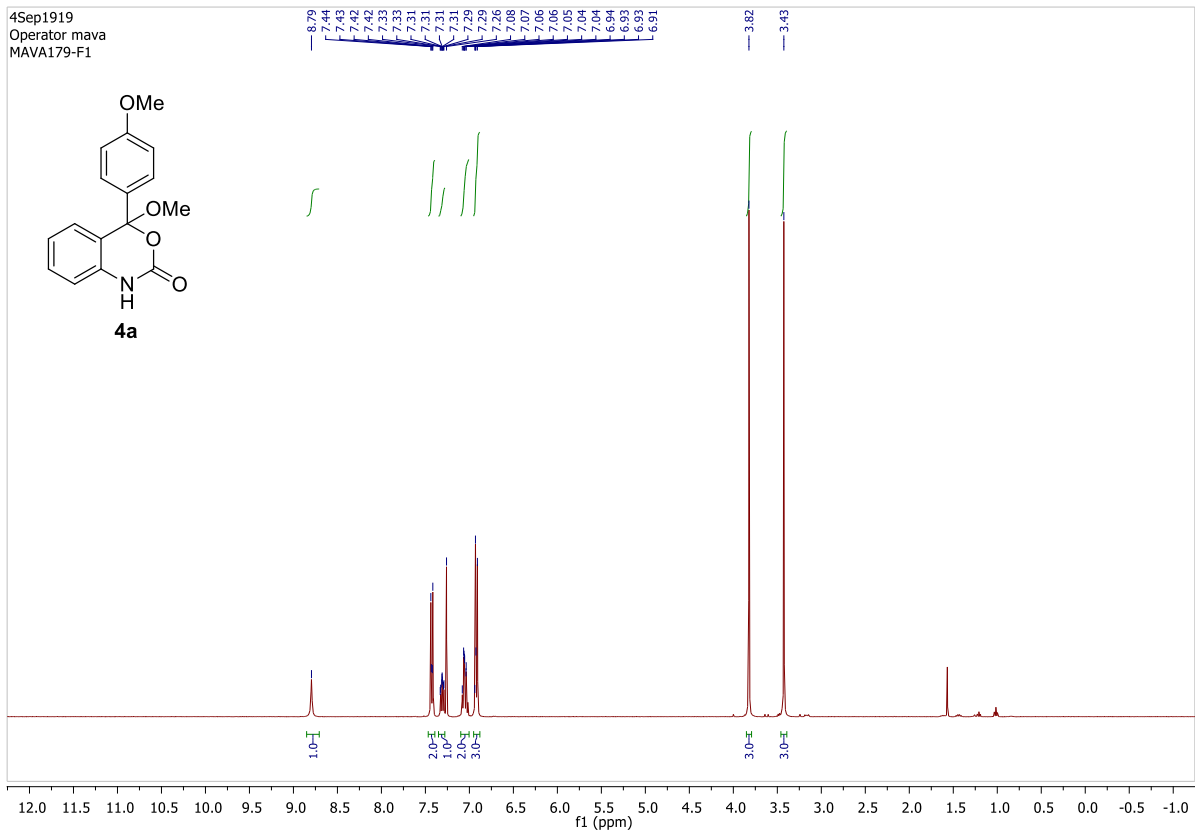




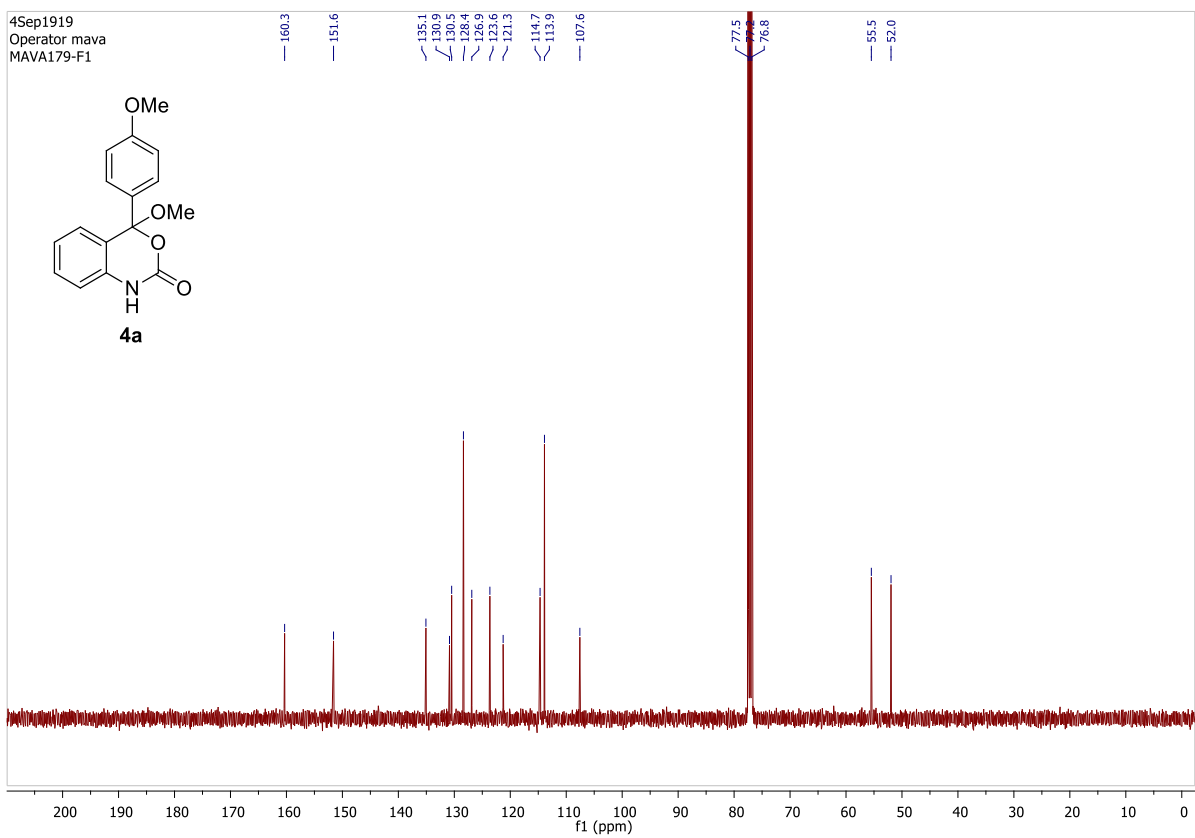
^1H NMR in CDCl_3 at 400 MHz



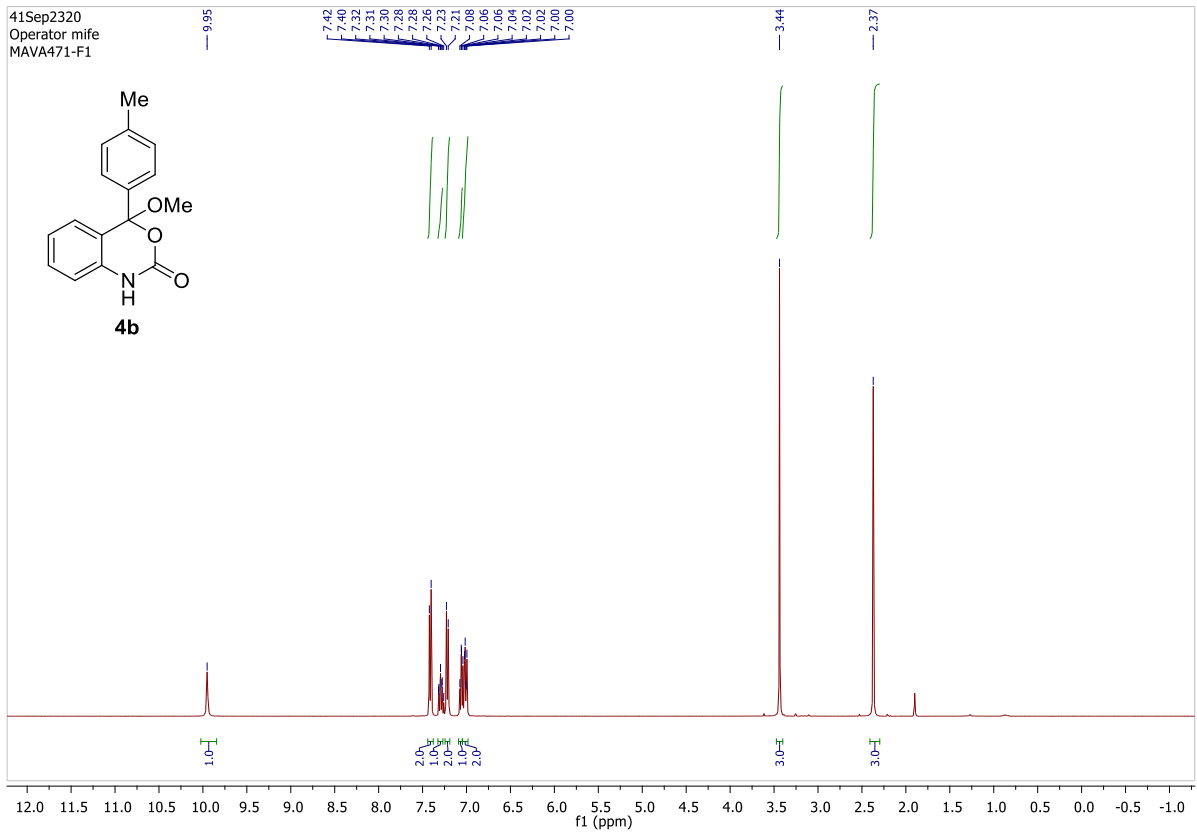
$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 100 MHz



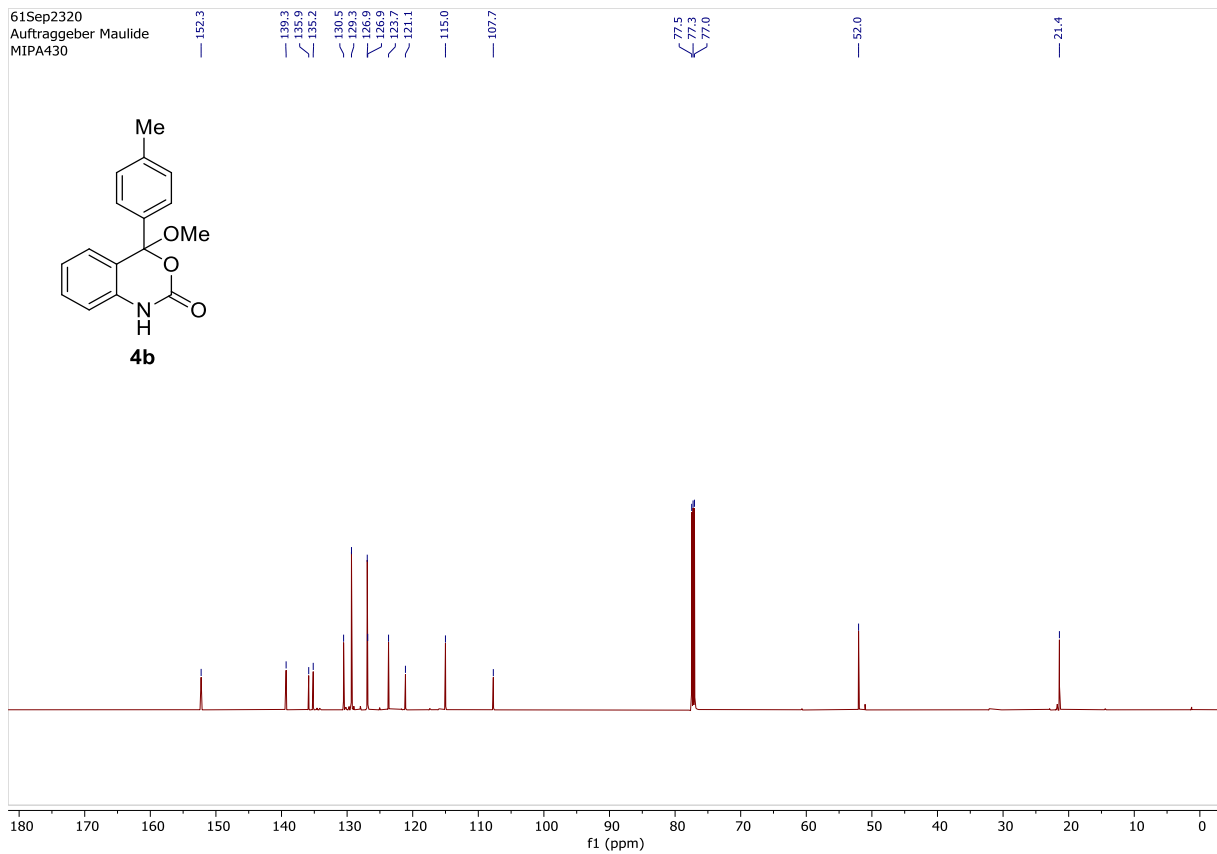
^1H NMR in CDCl_3 at 400 MHz



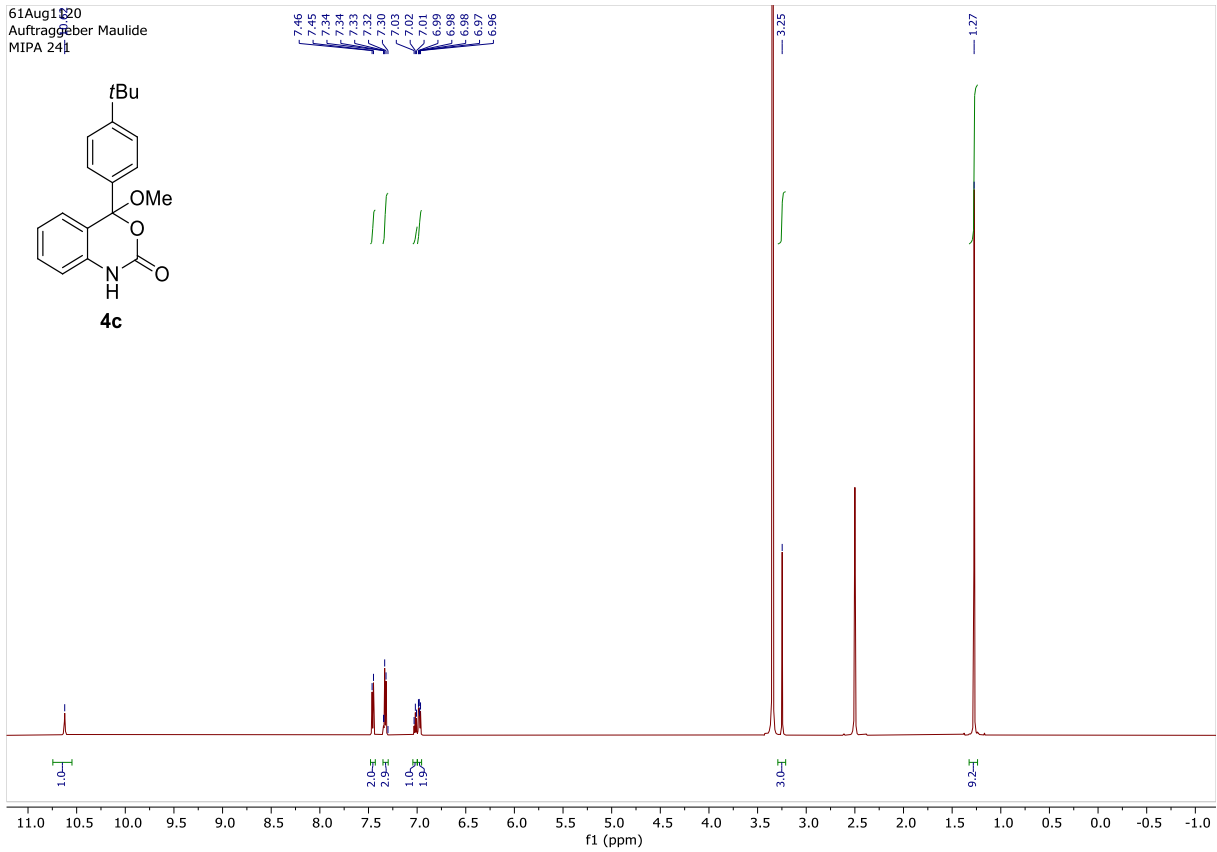
$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 100 MHz



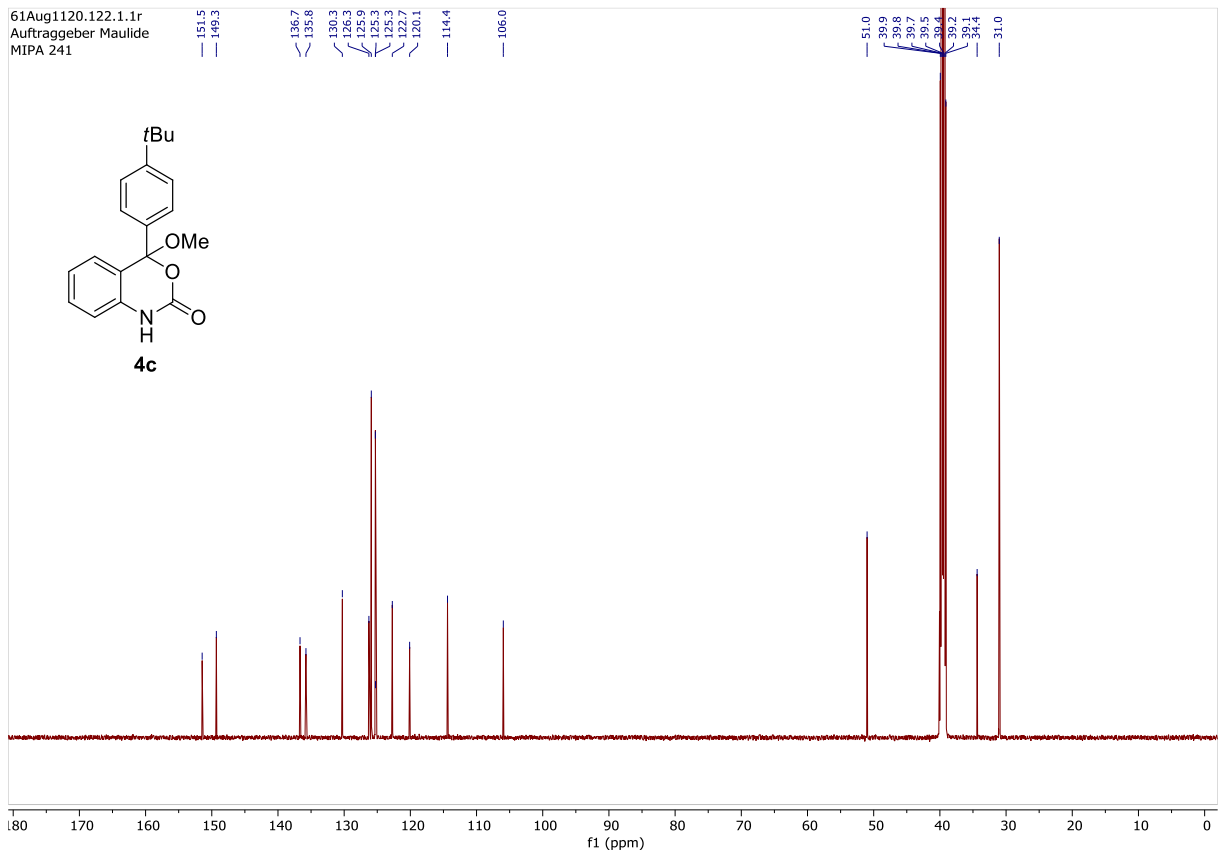
^1H NMR in CDCl_3 at 400 MHz



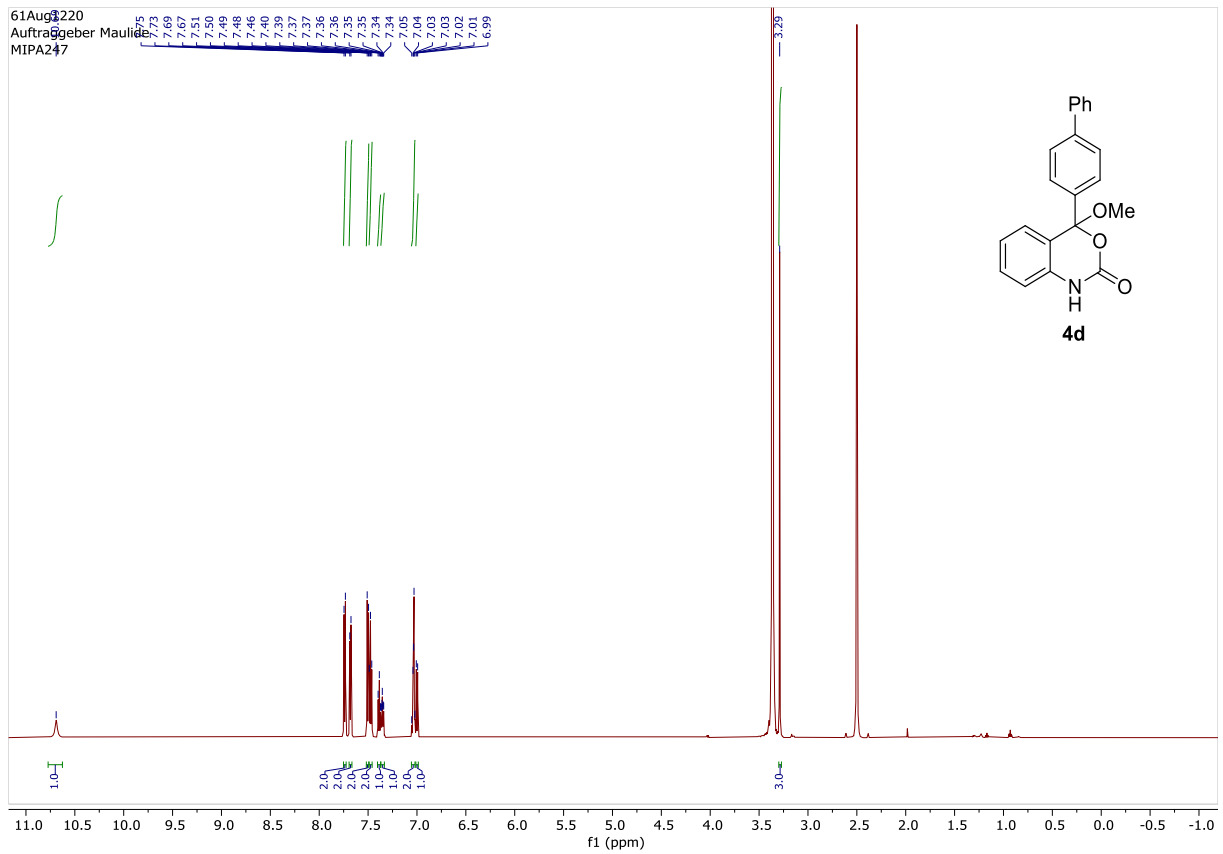
$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 150 MHz



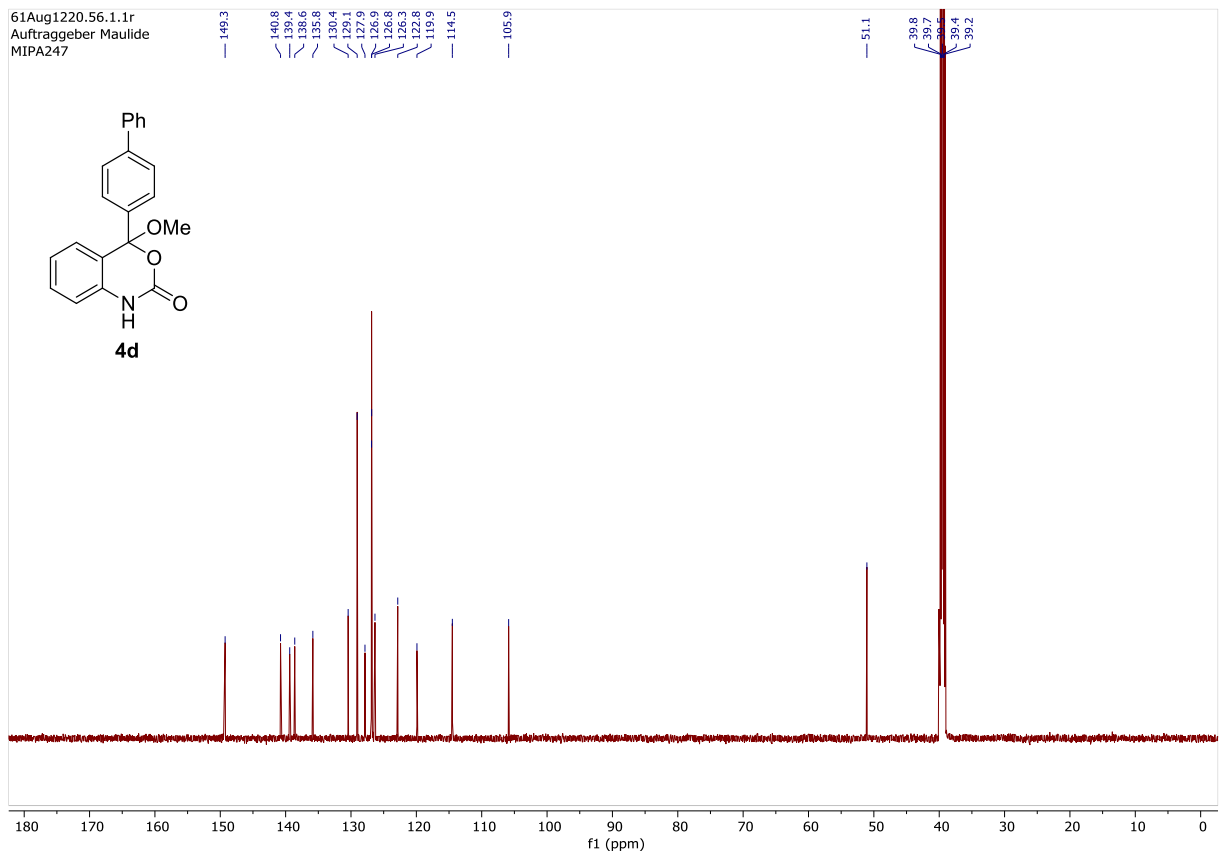
^1H NMR in CDCl_3 at 600 MHz



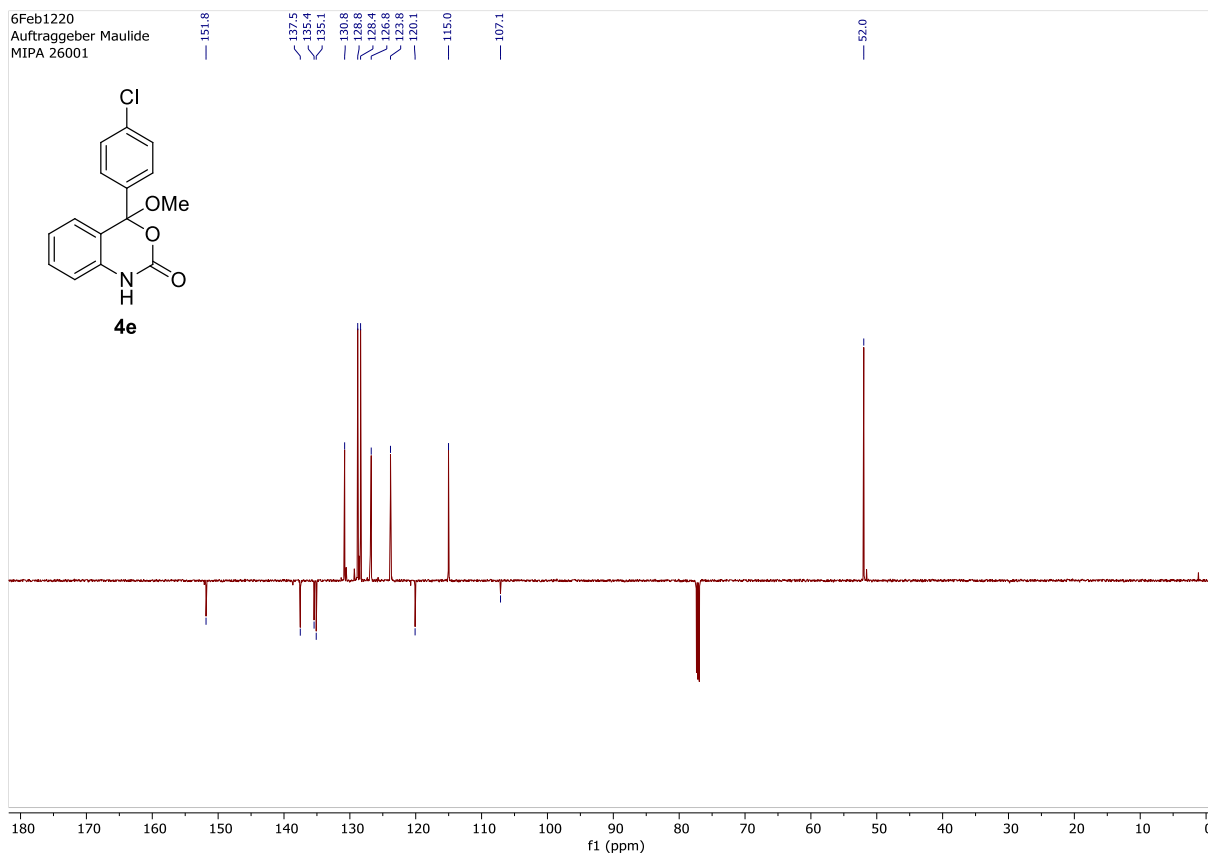
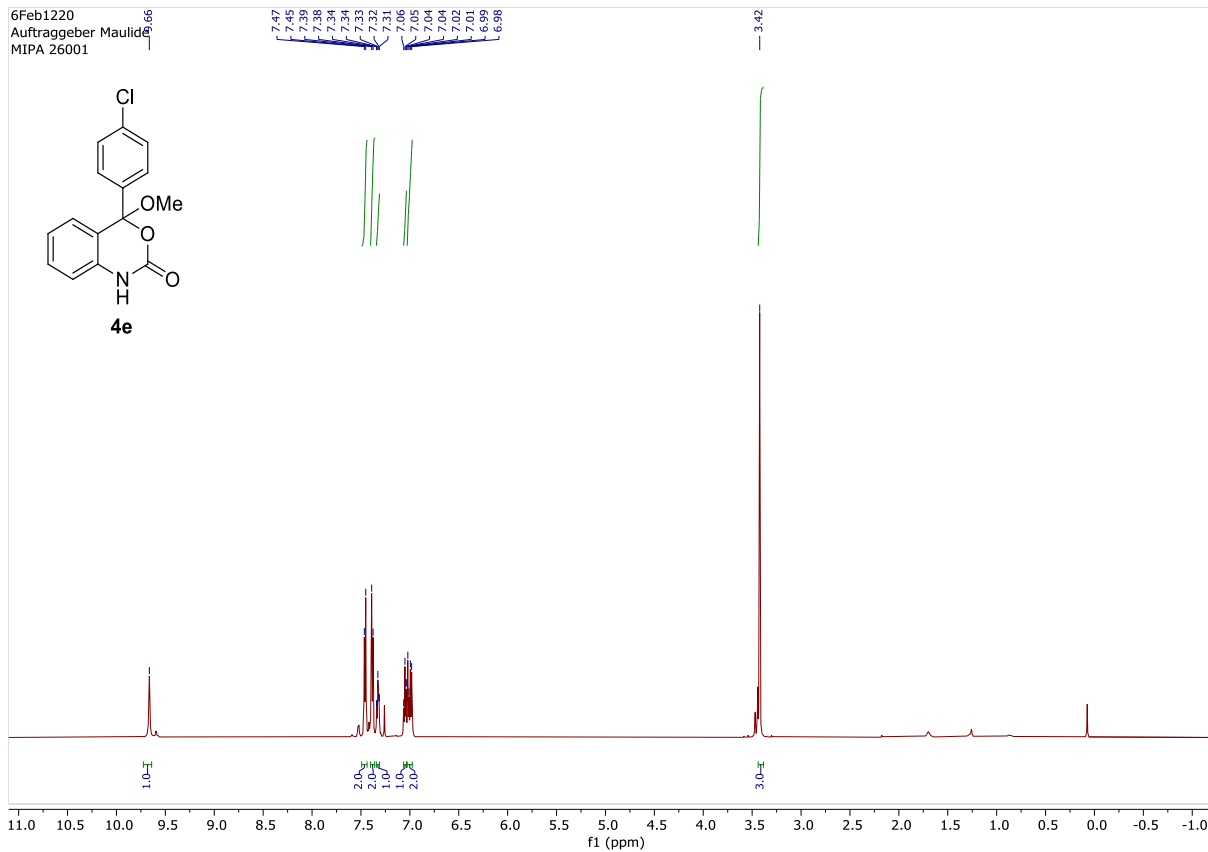
$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 150 MHz

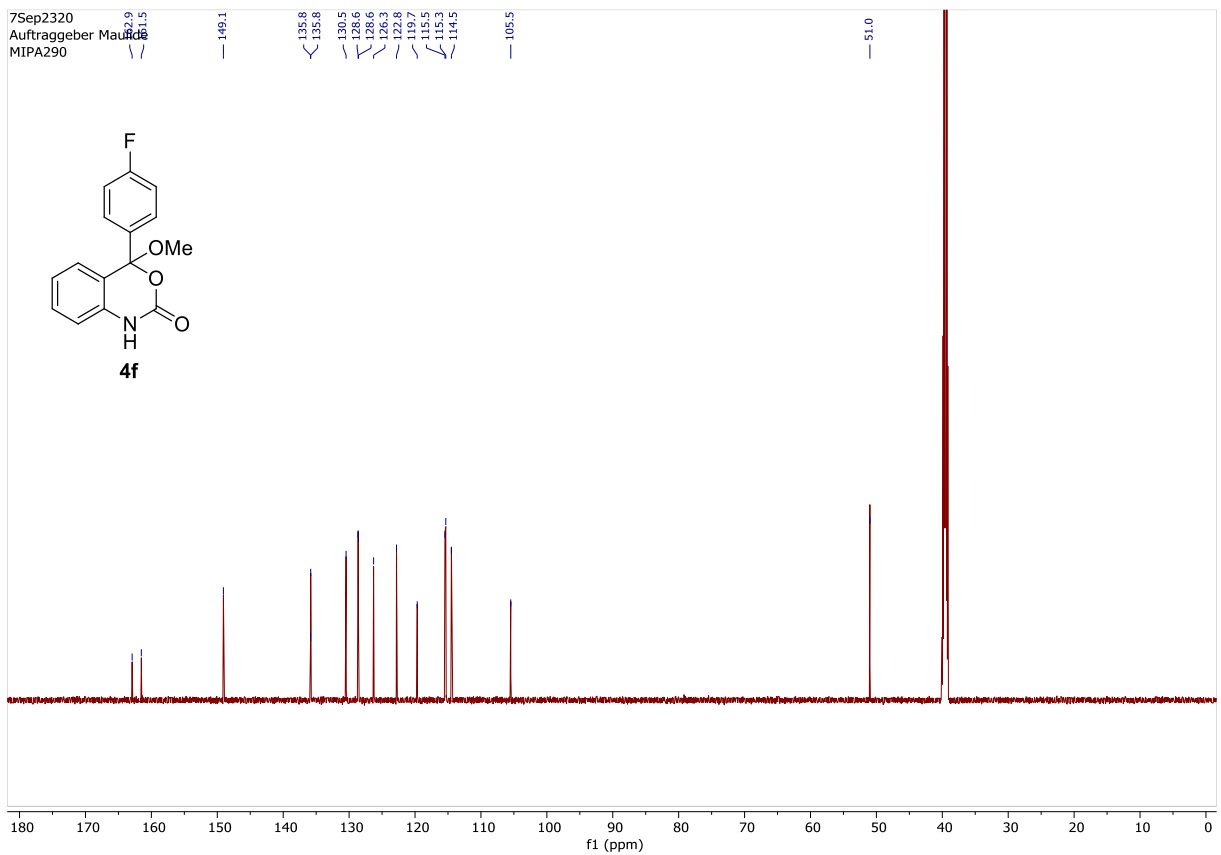
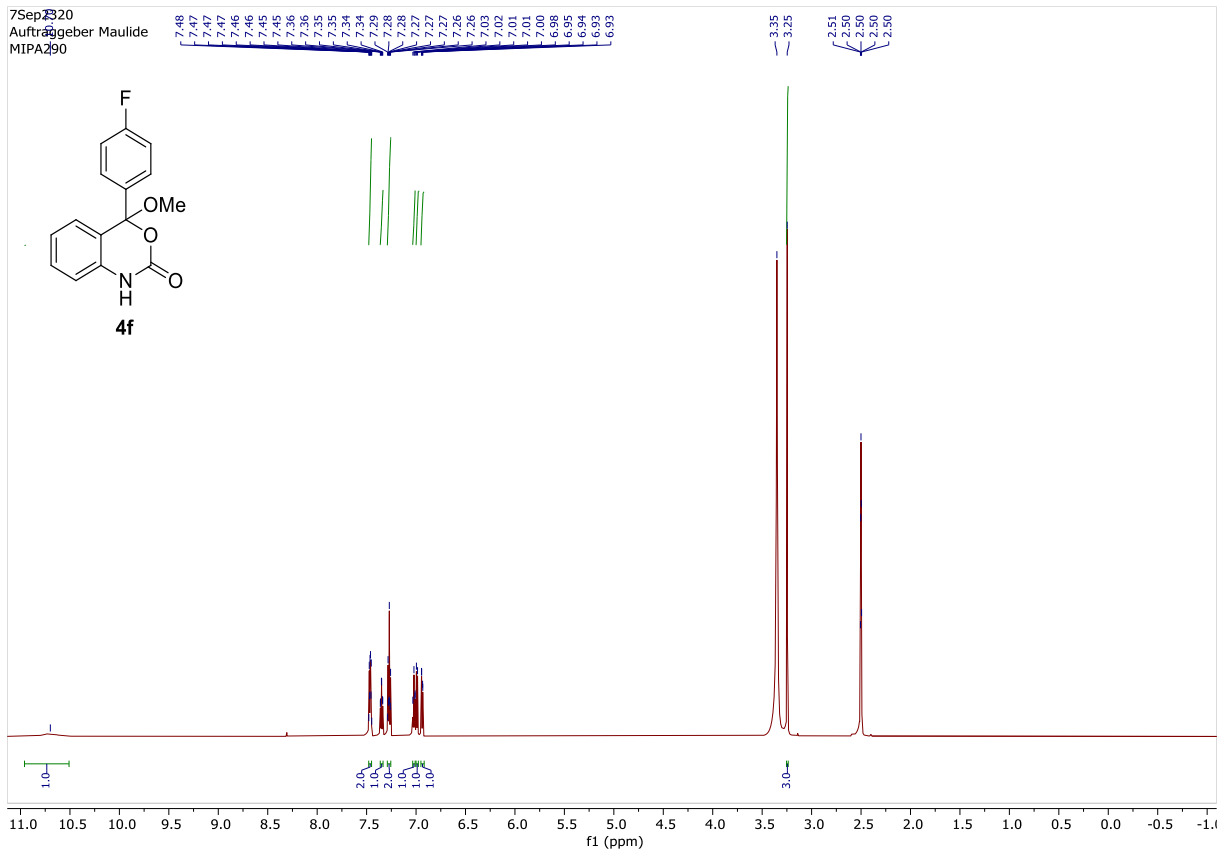


^1H NMR in CDCl_3 at 600 MHz

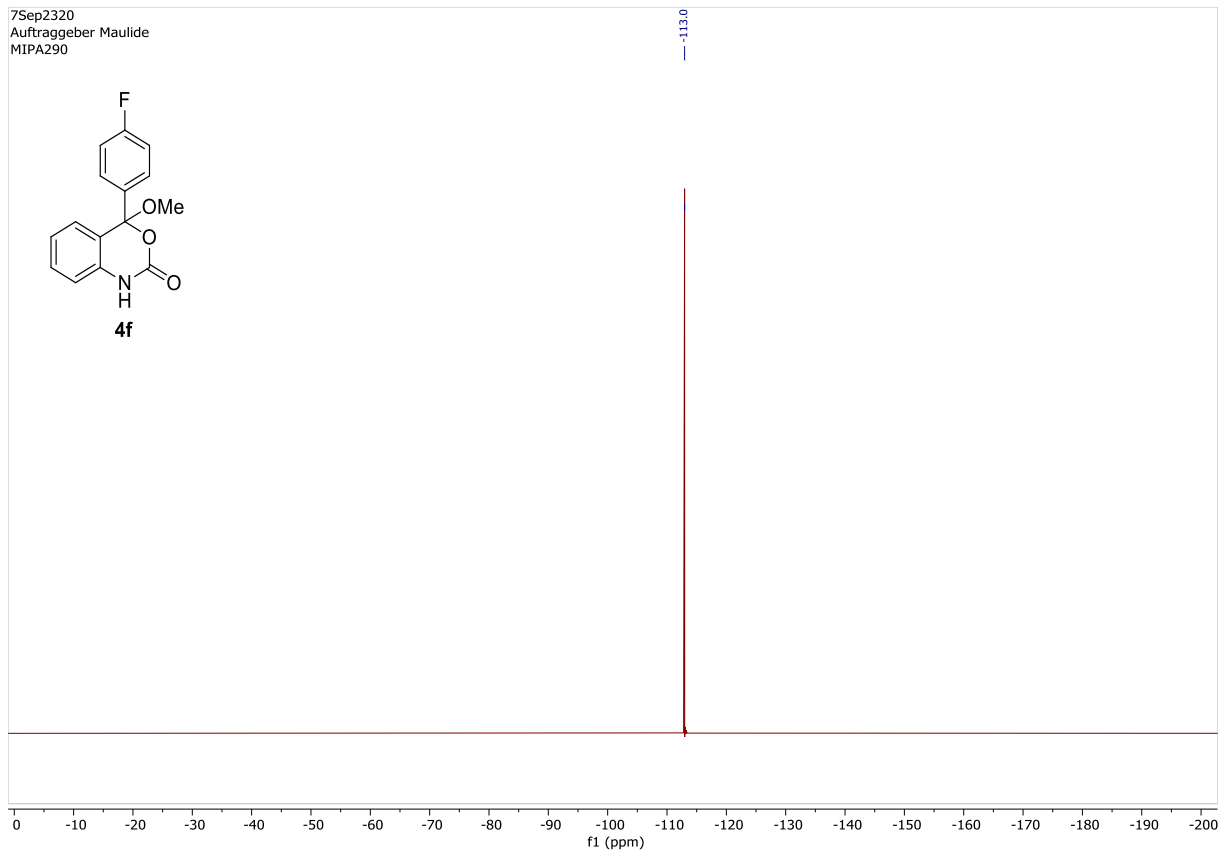
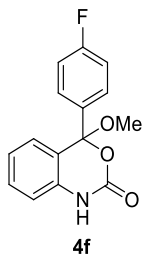


$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 150 MHz

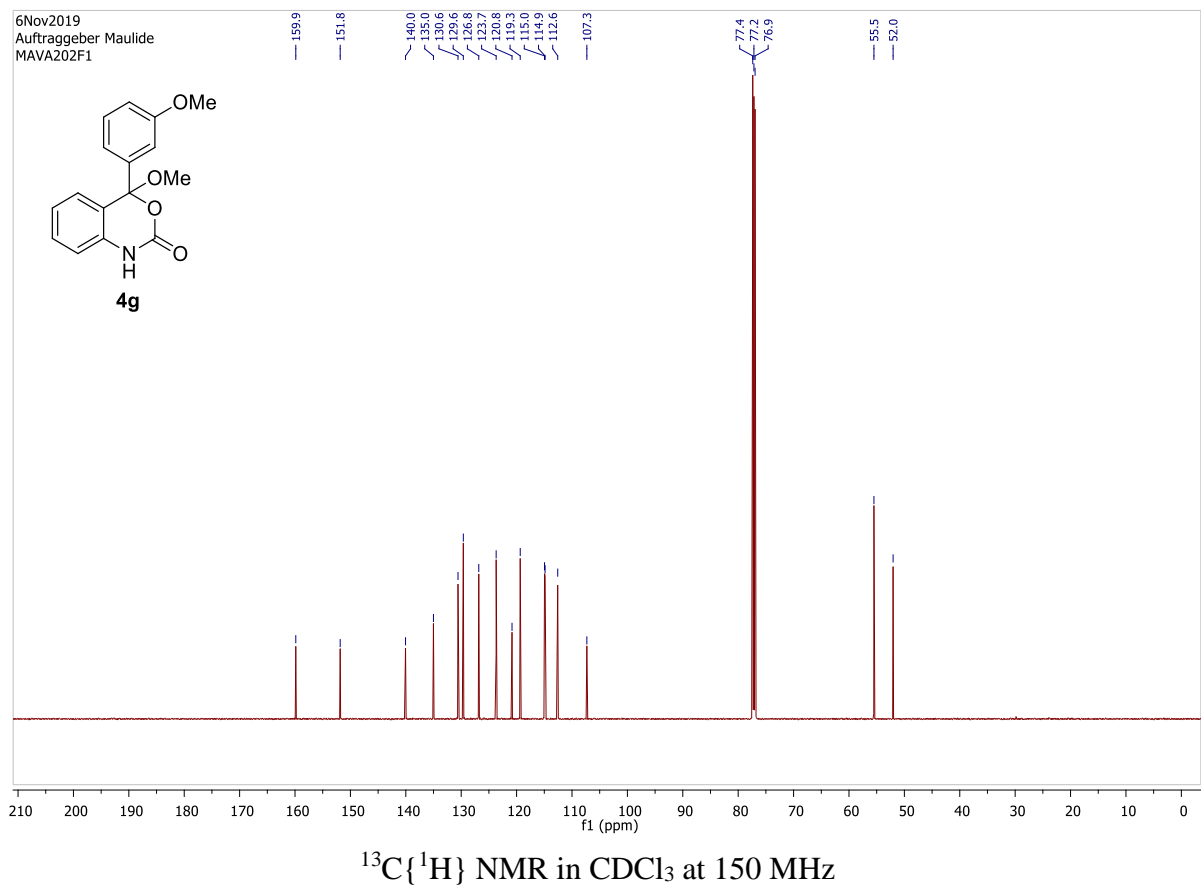
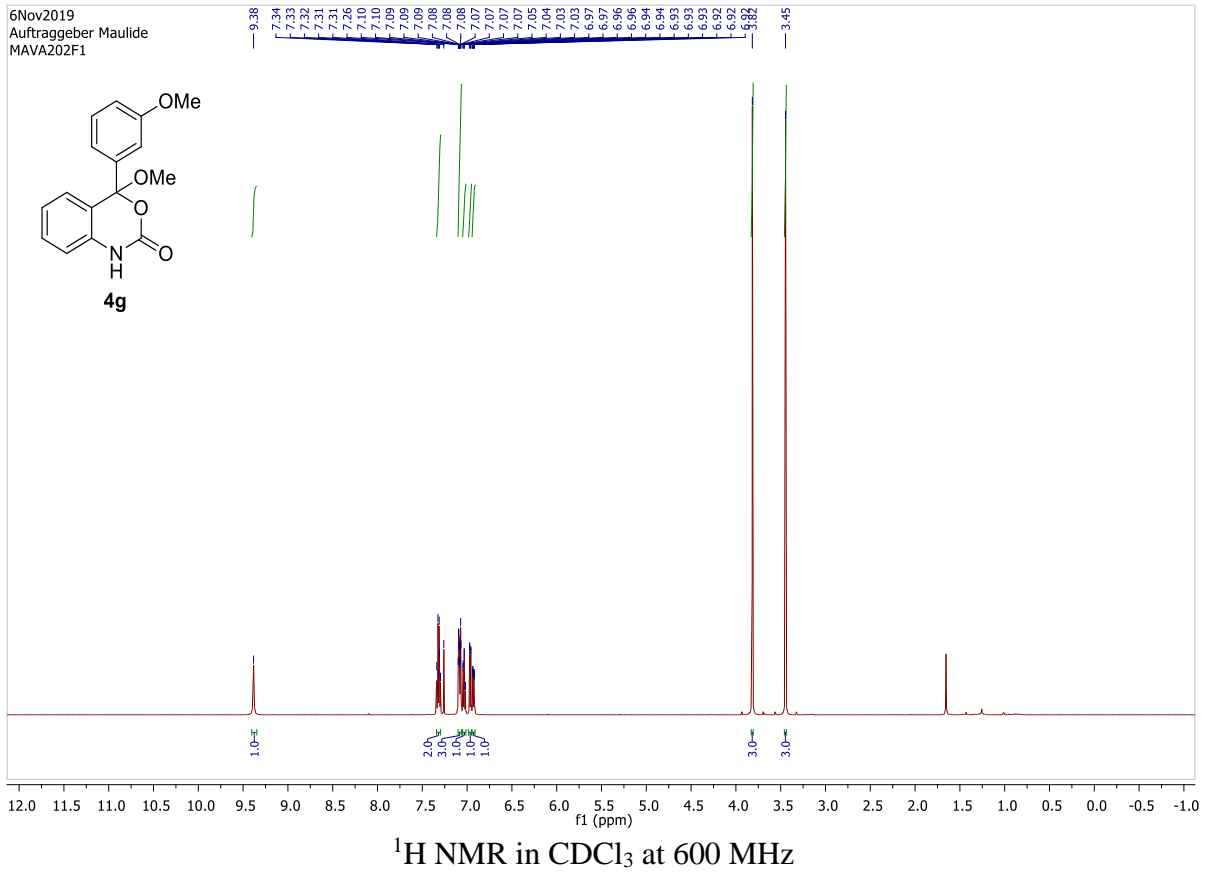


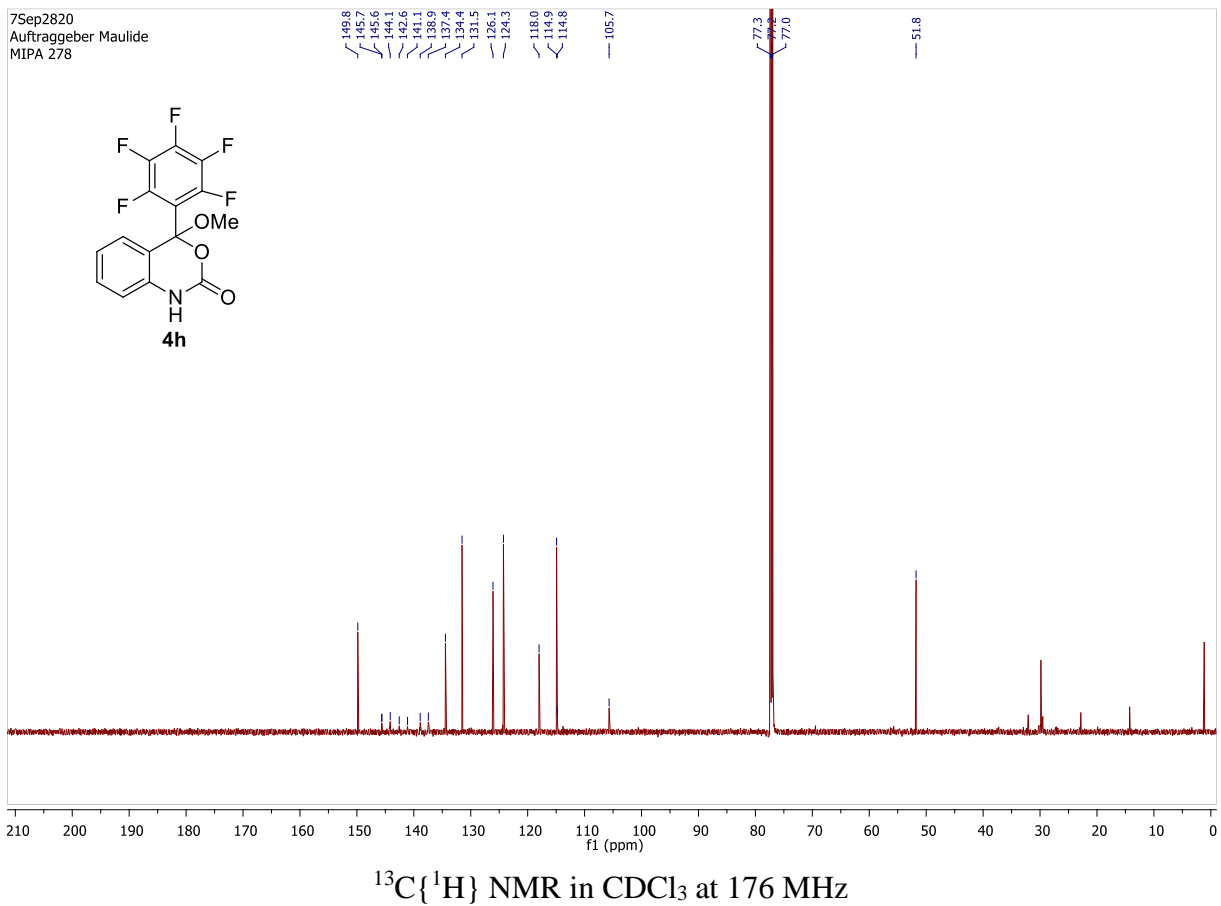
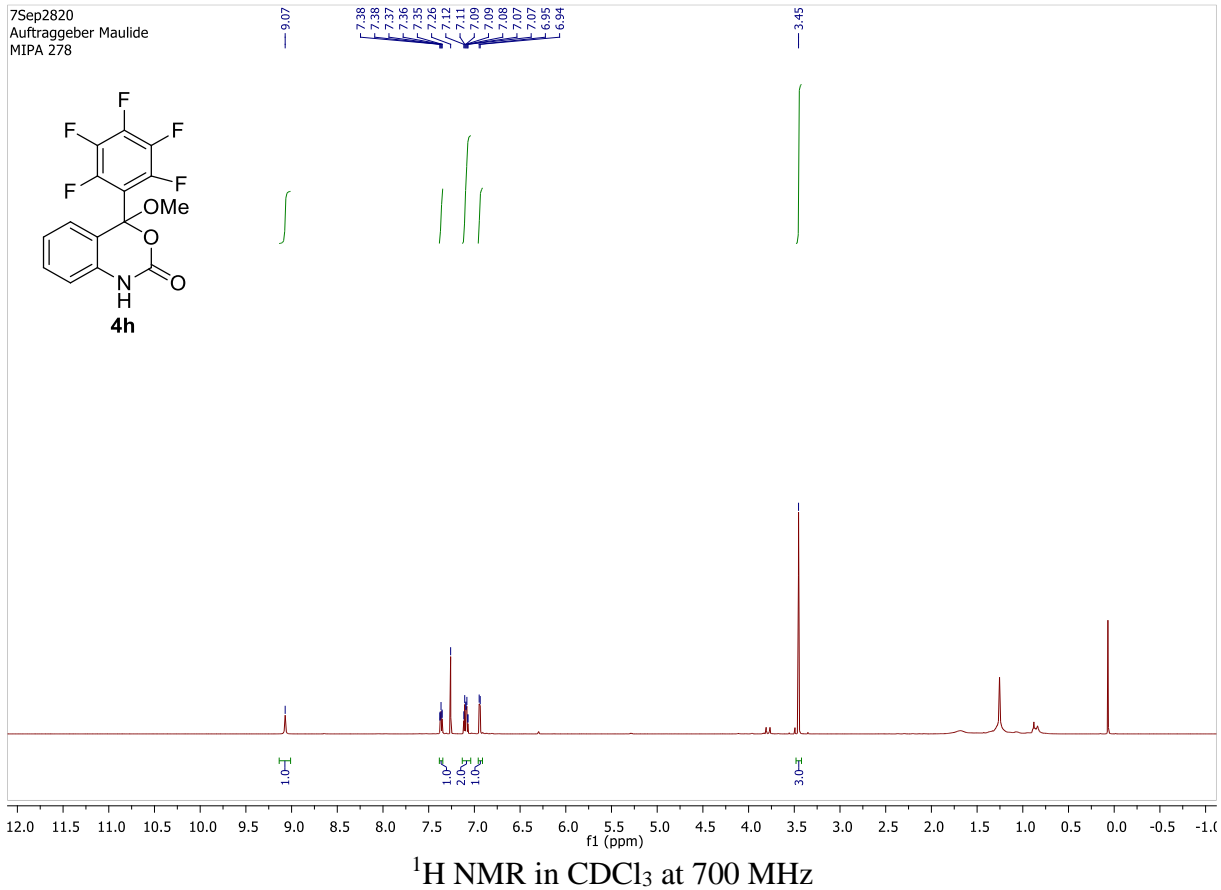


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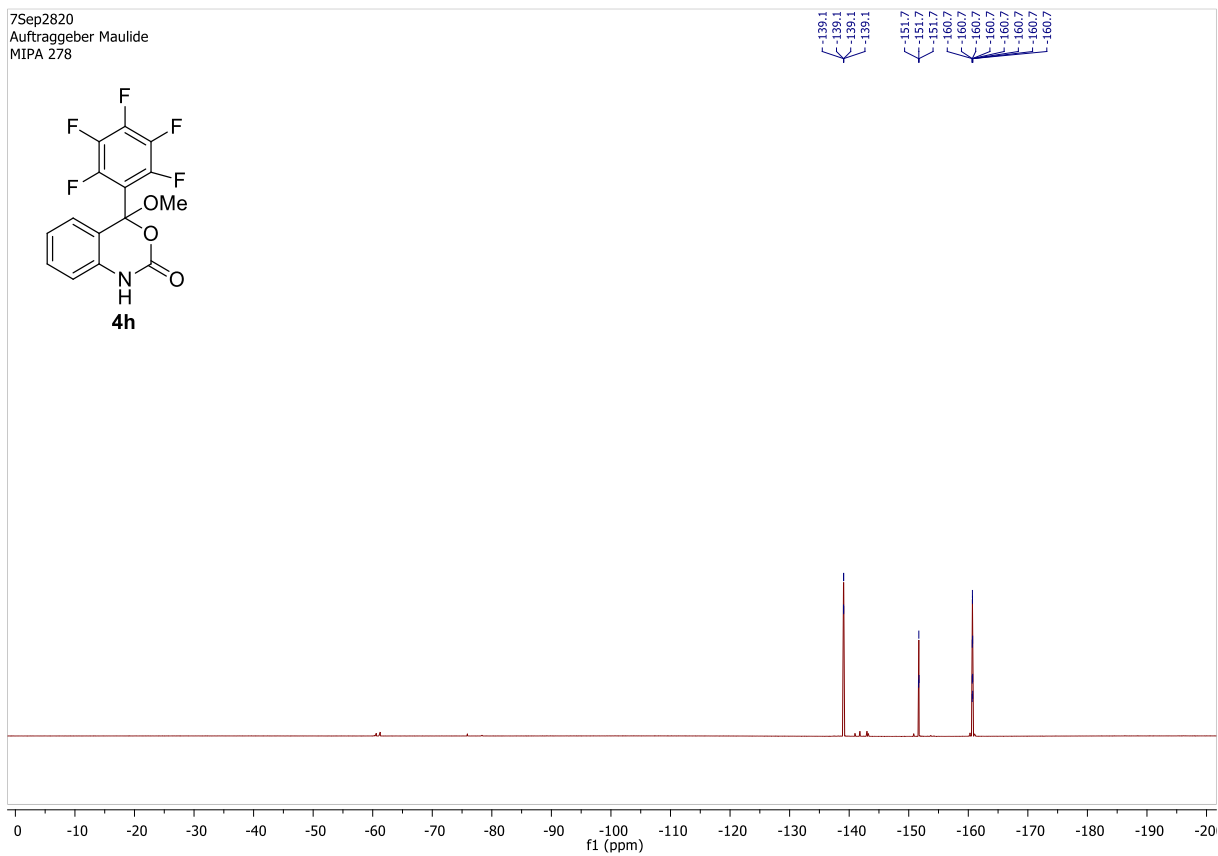
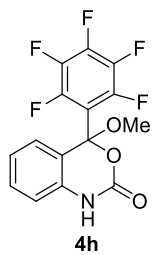


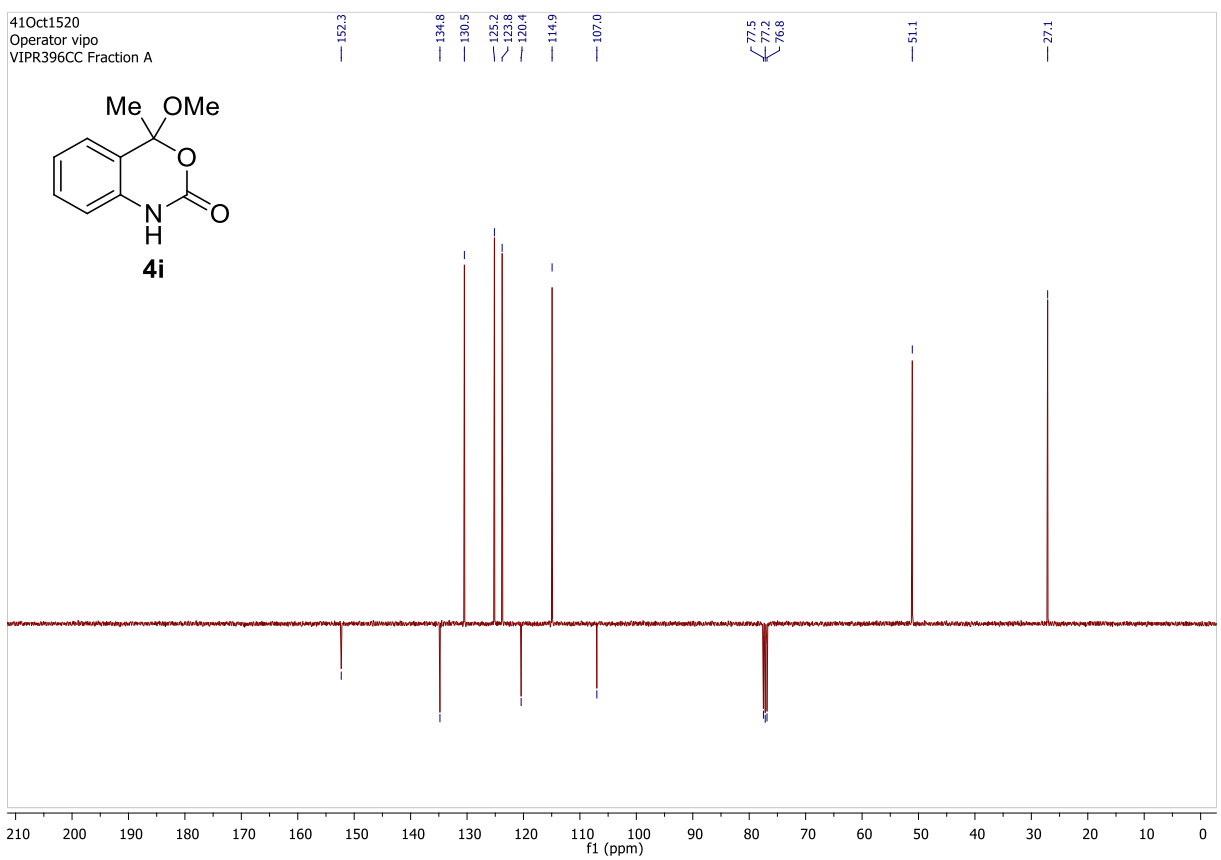
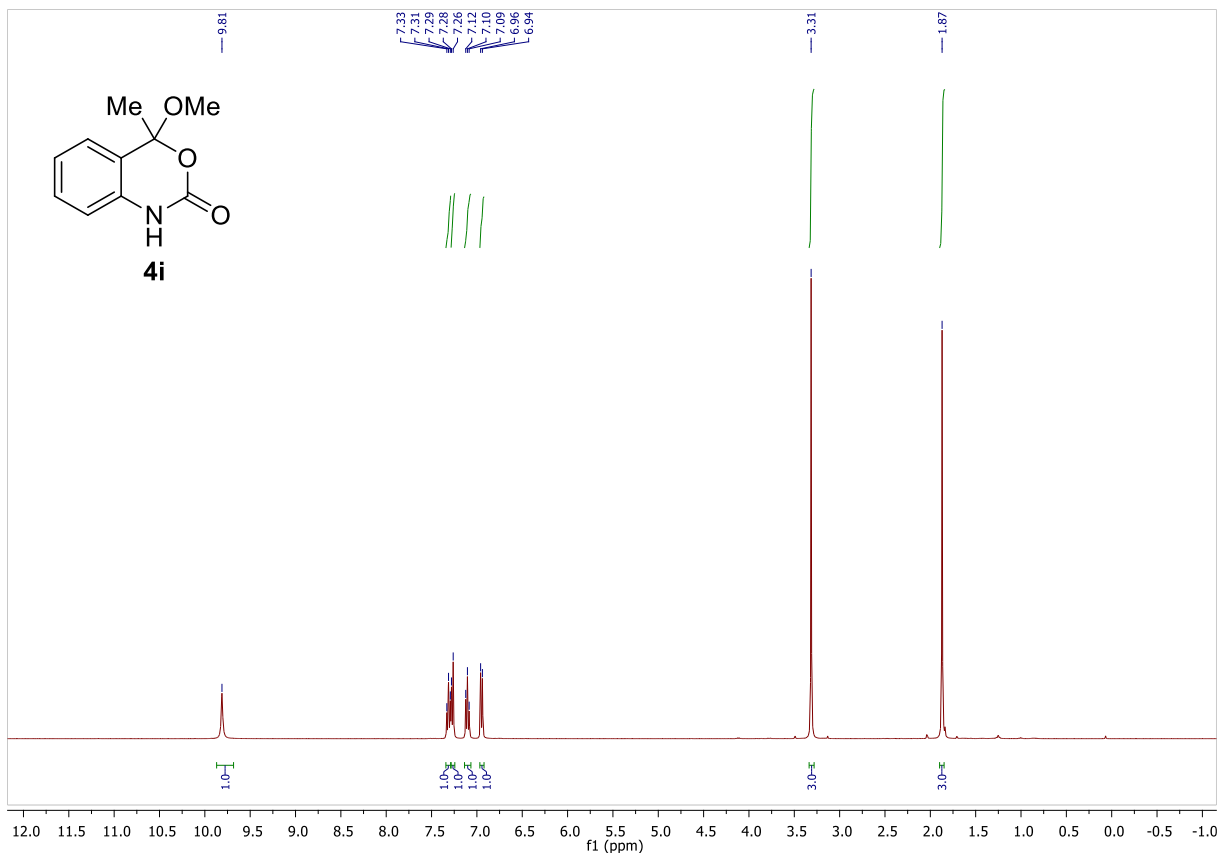
^{19}F NMR in $\text{DMSO-}d_6$ at 659 MHz

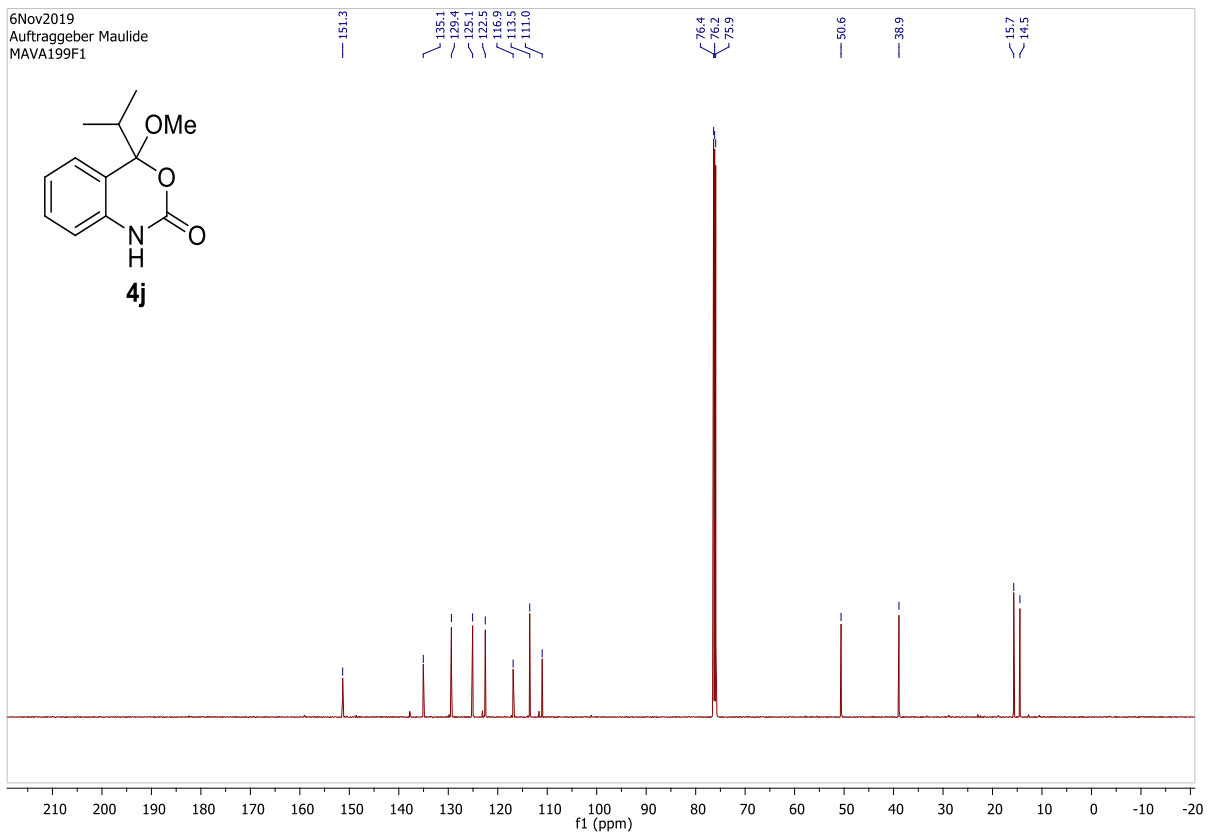
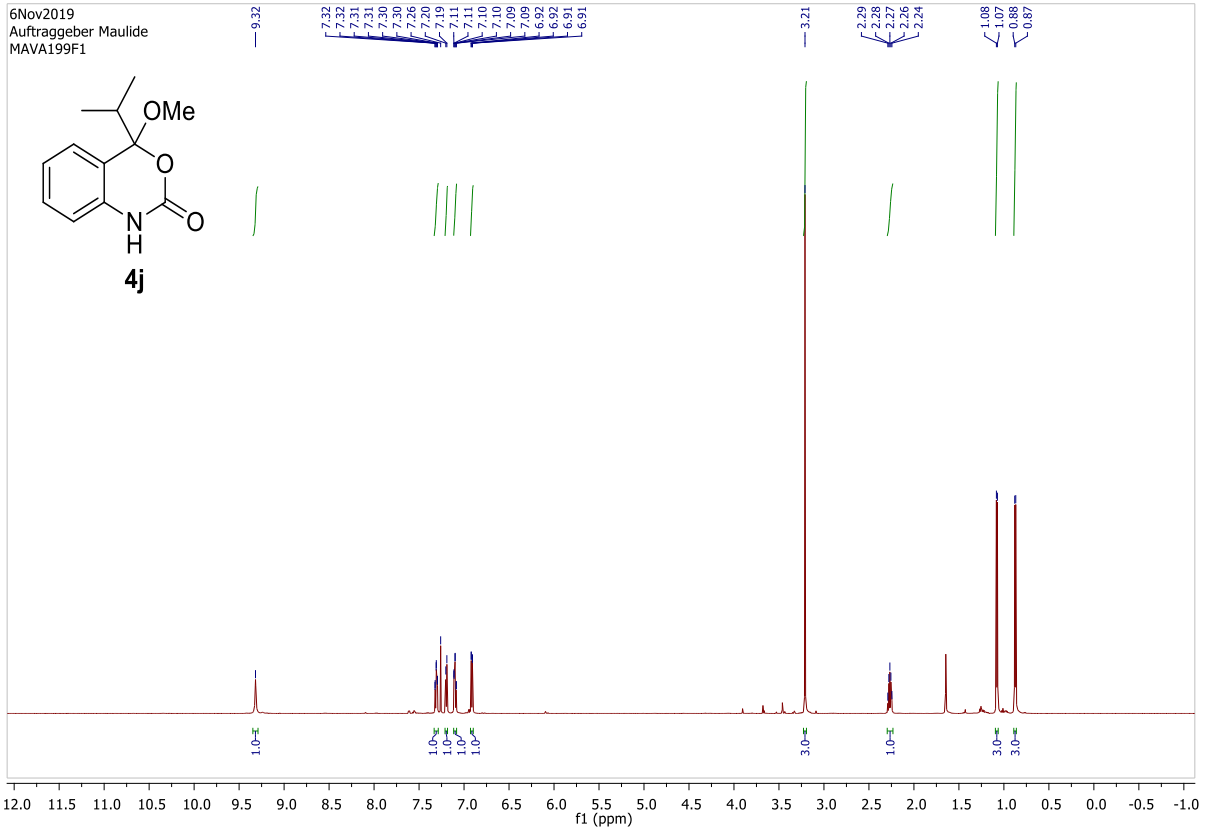


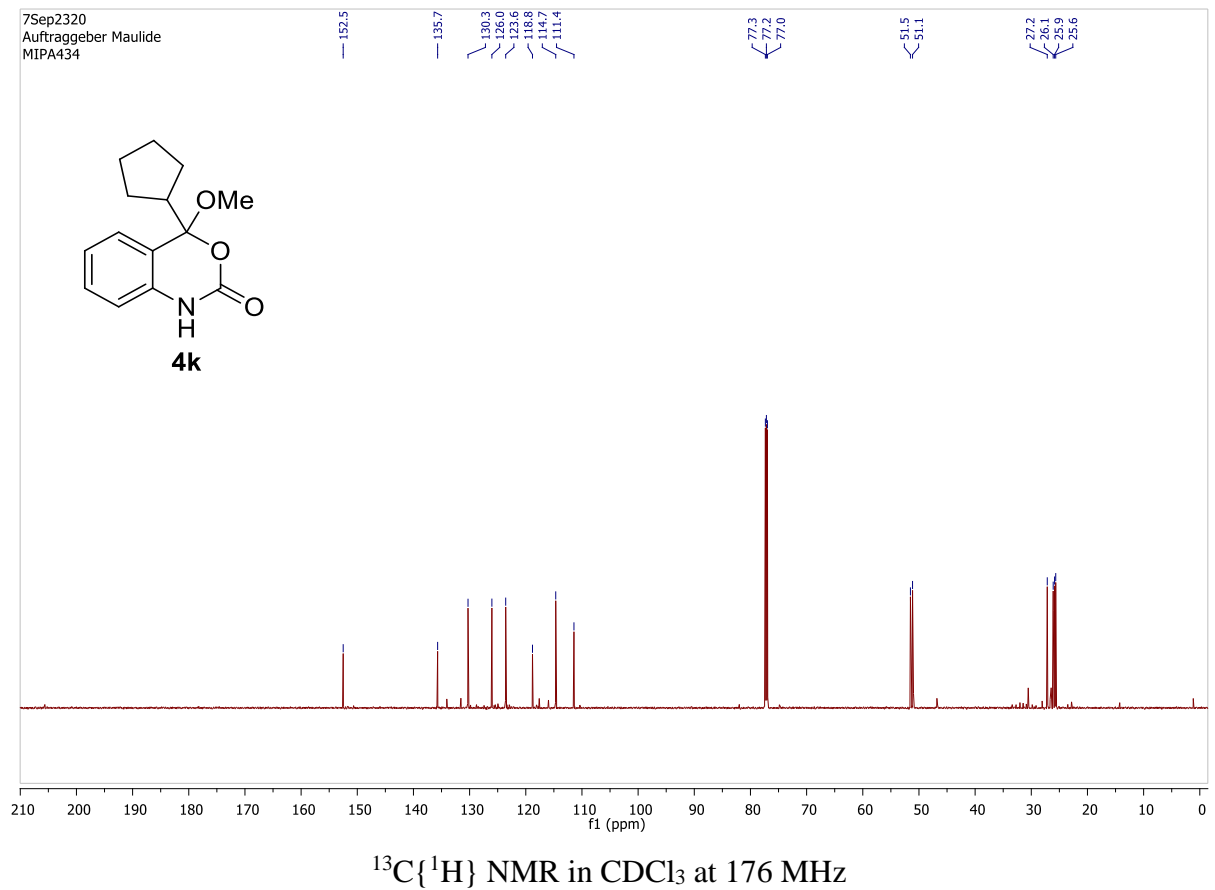
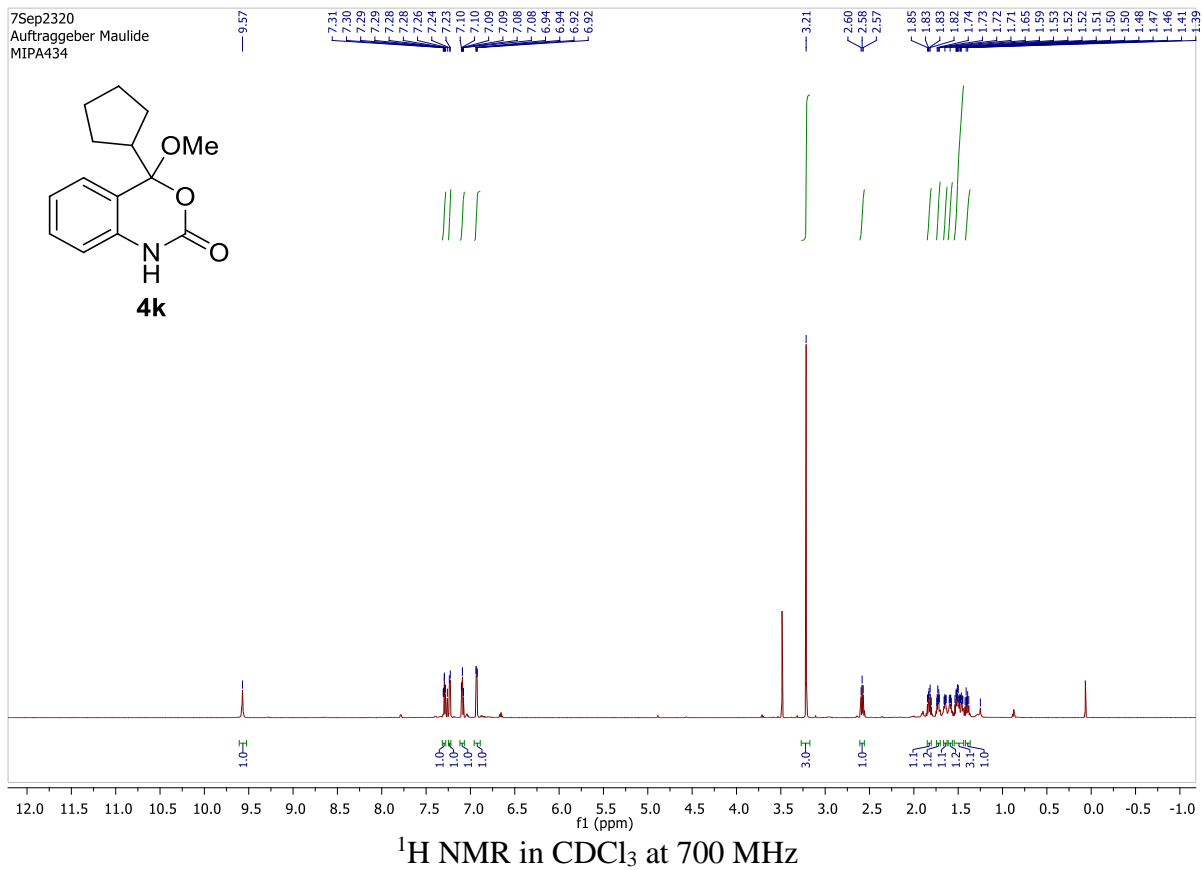


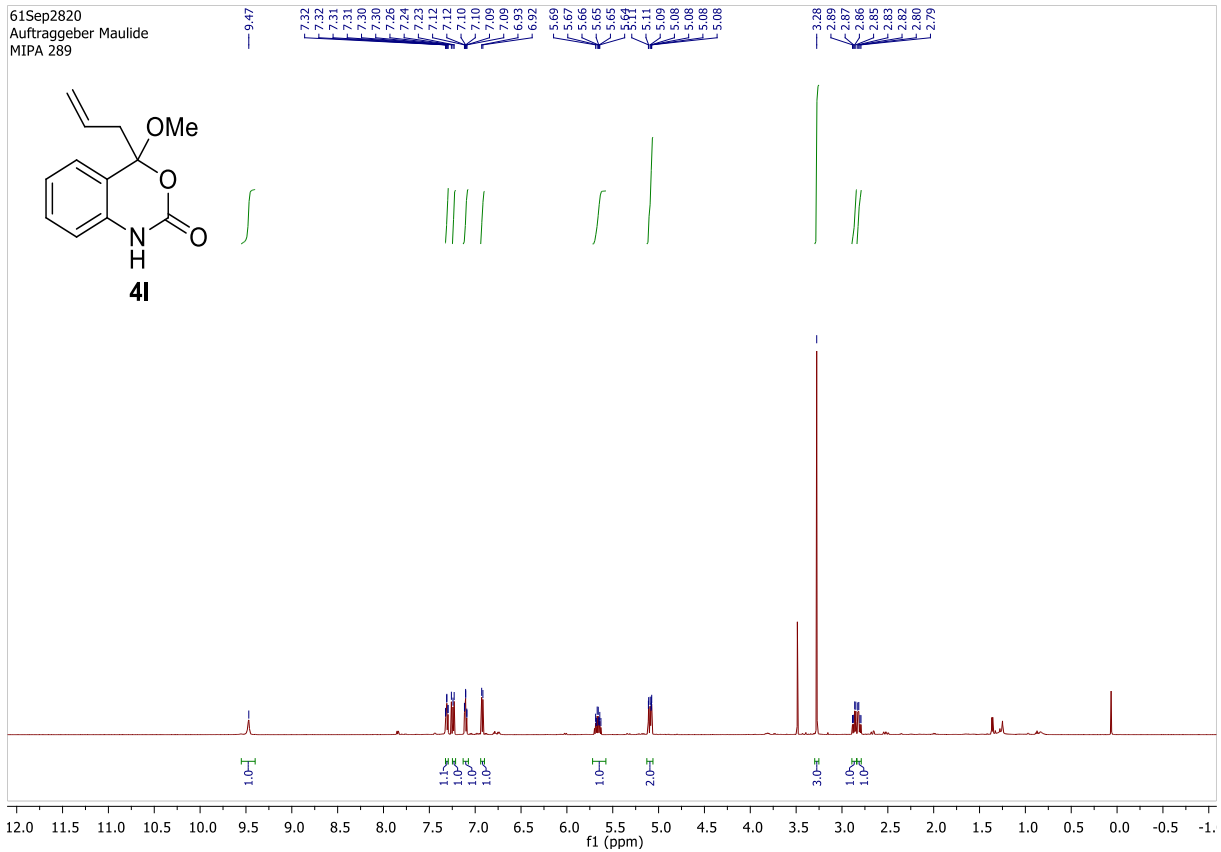
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MIPA 278



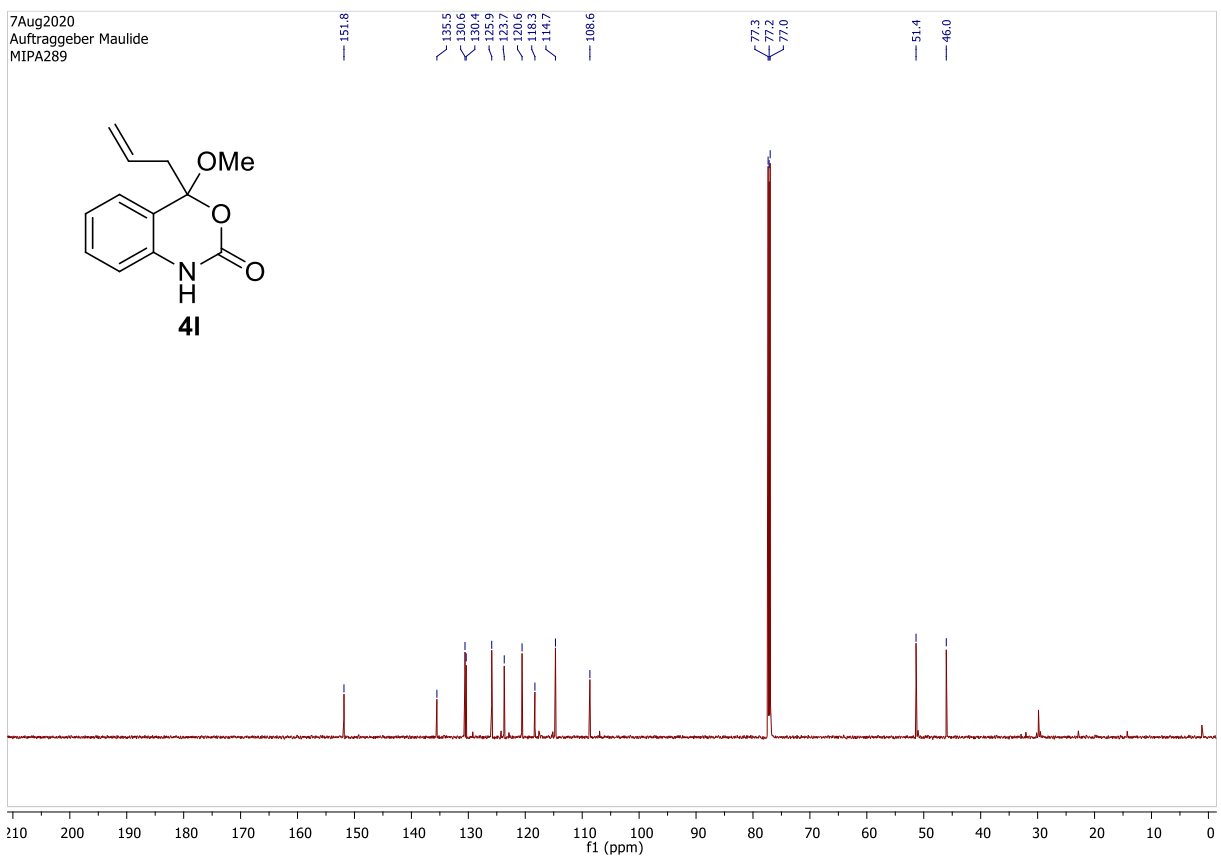




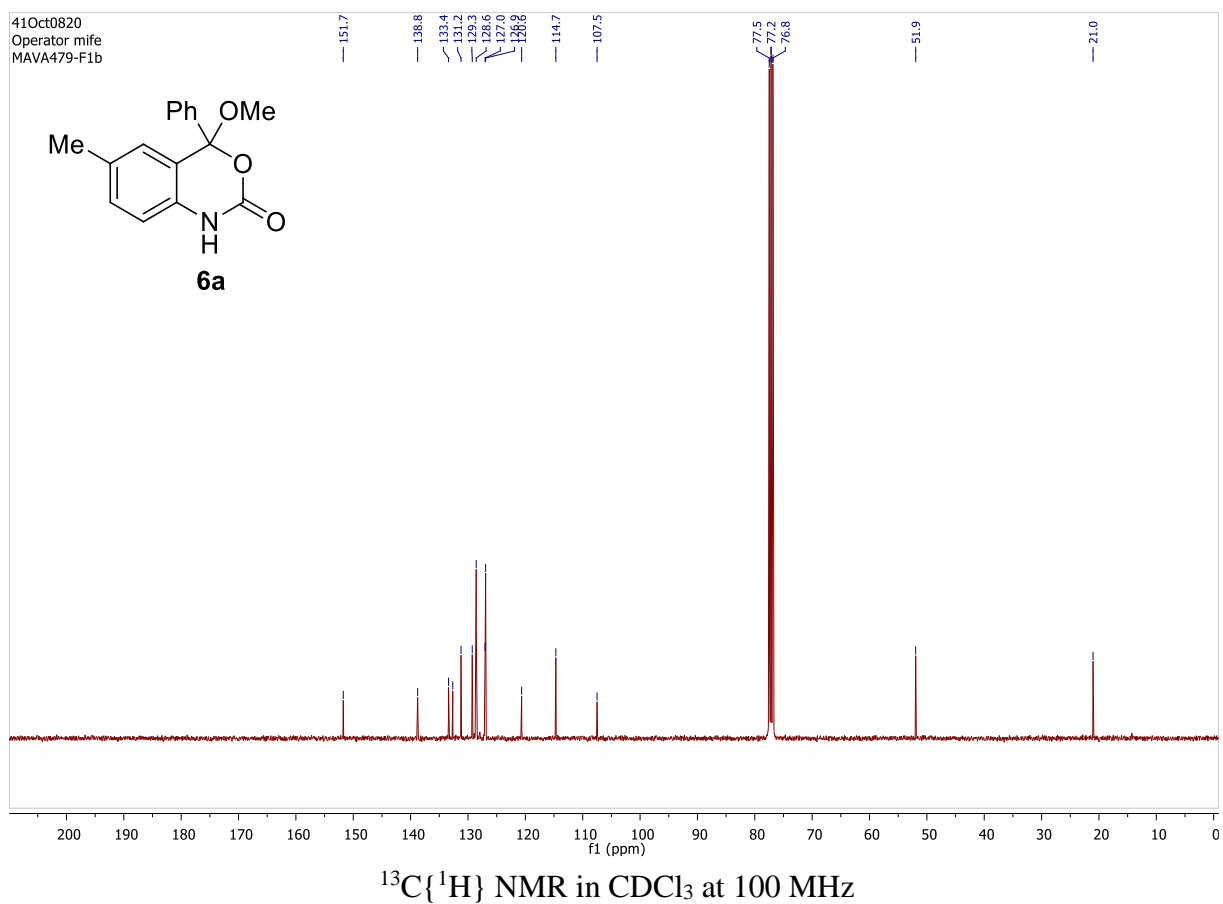
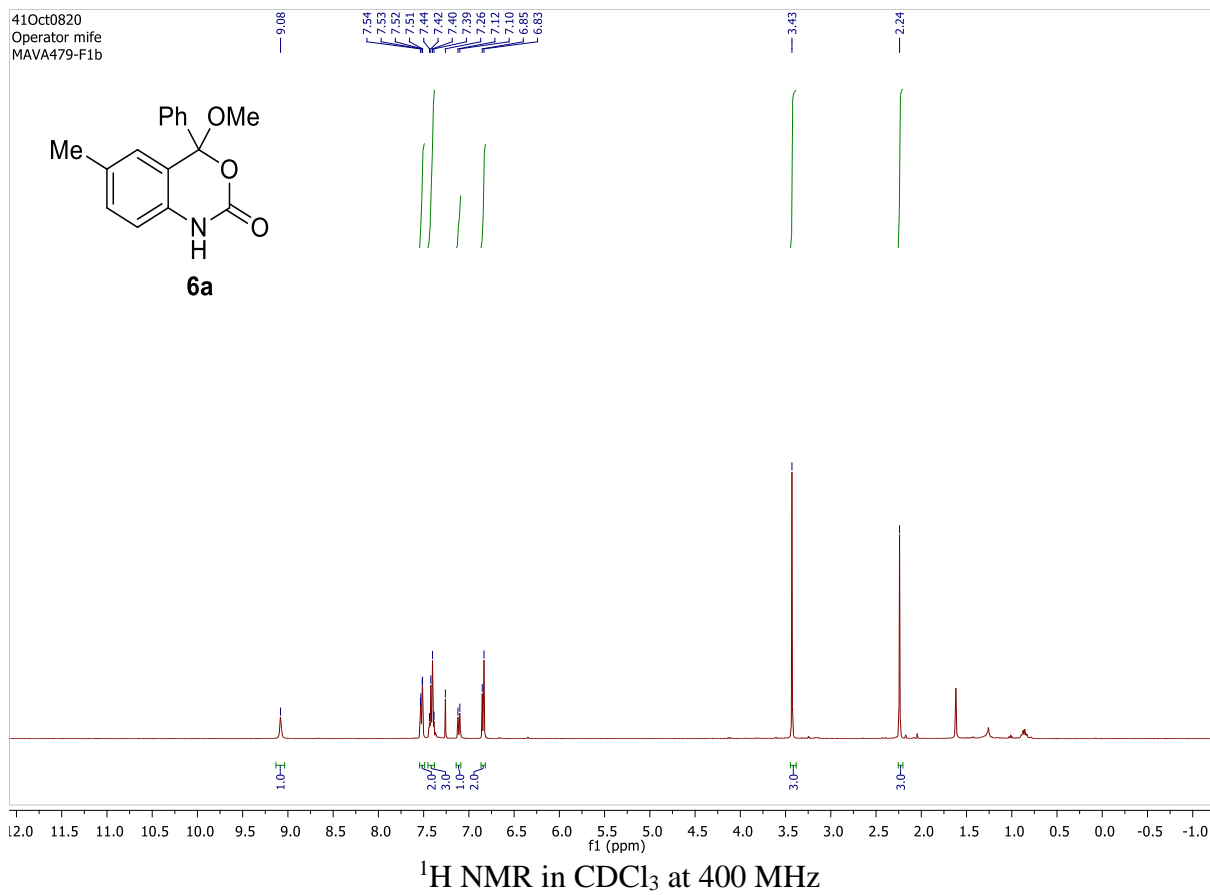


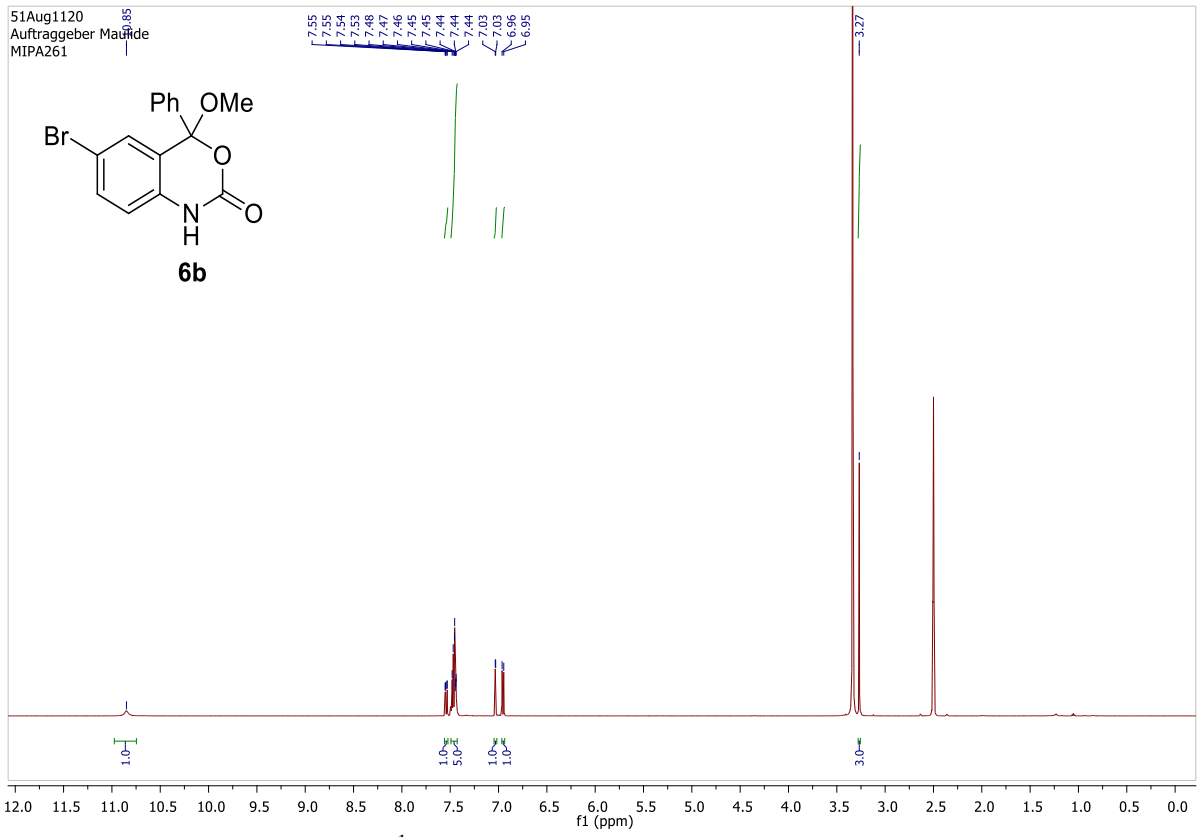


^1H NMR in CDCl_3 at 700 MHz

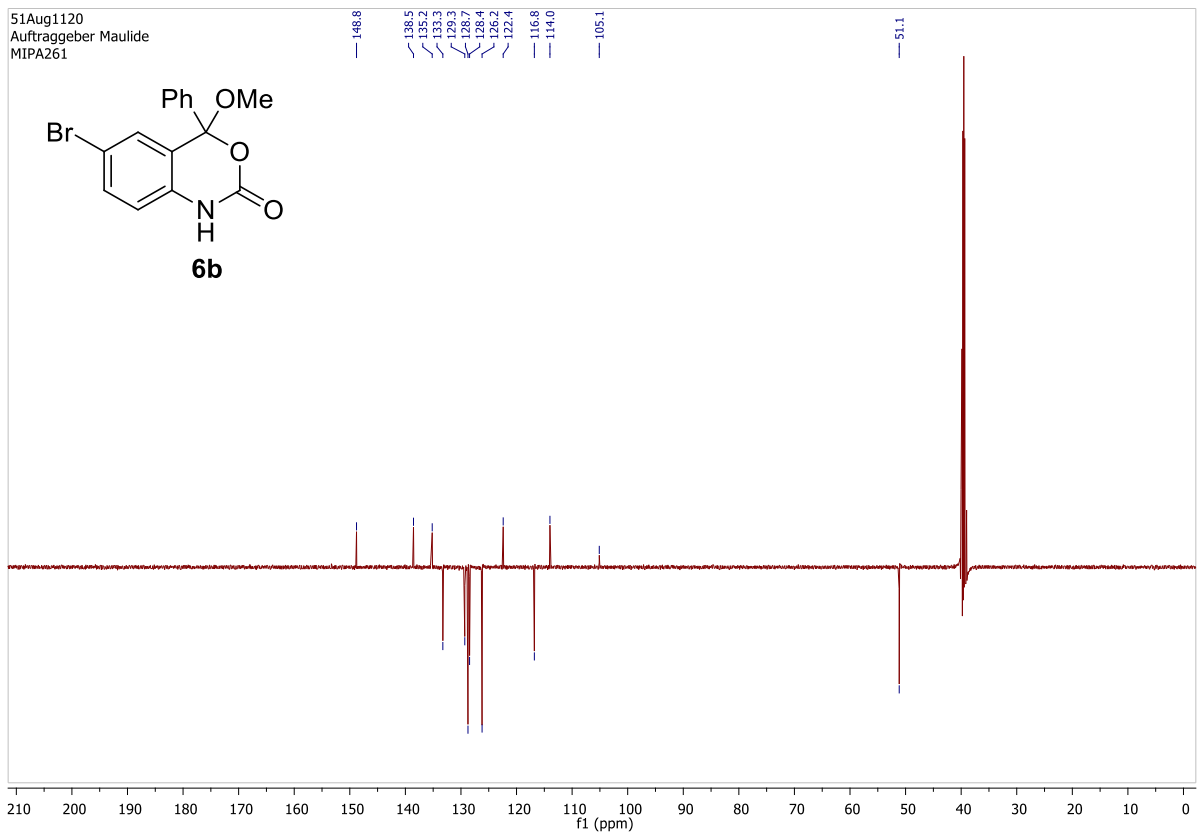


$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 176 MHz

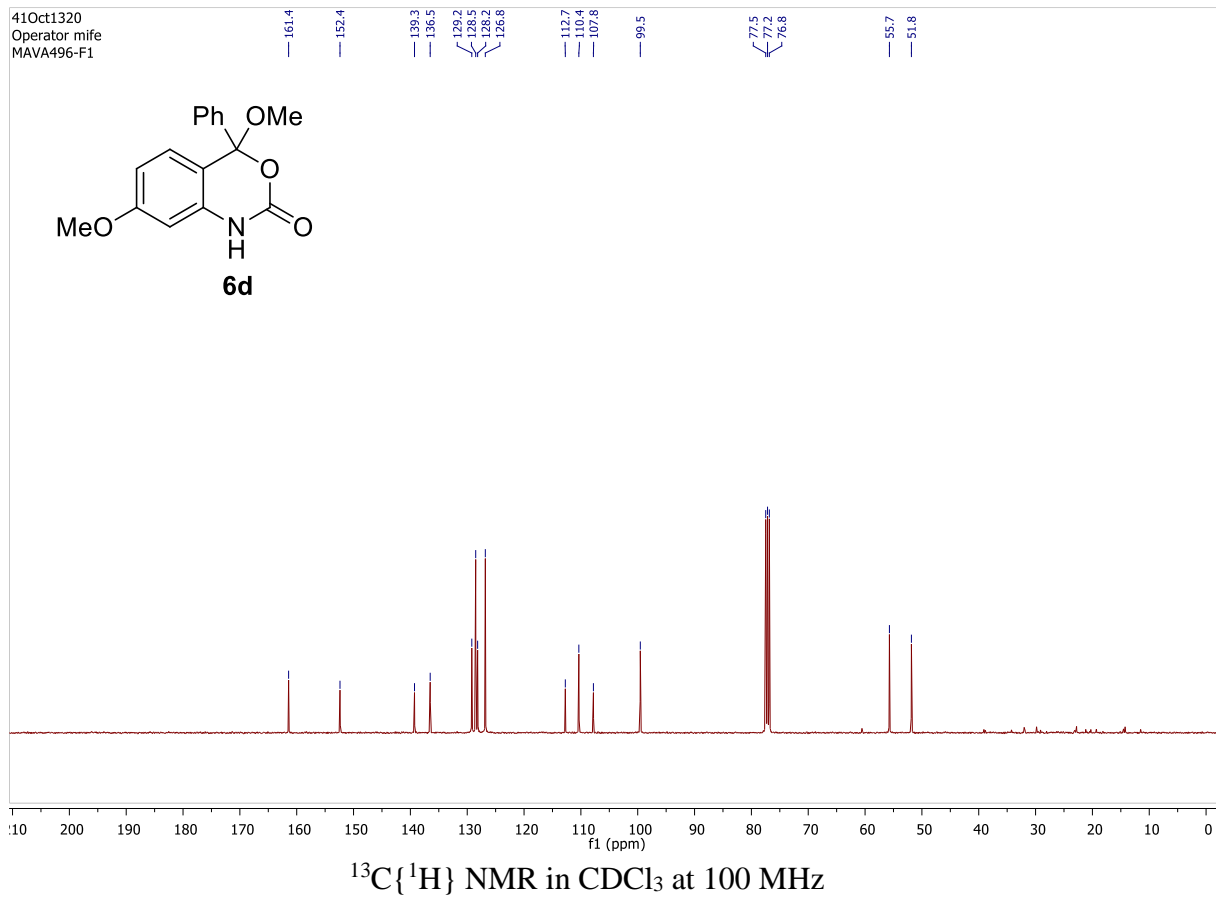
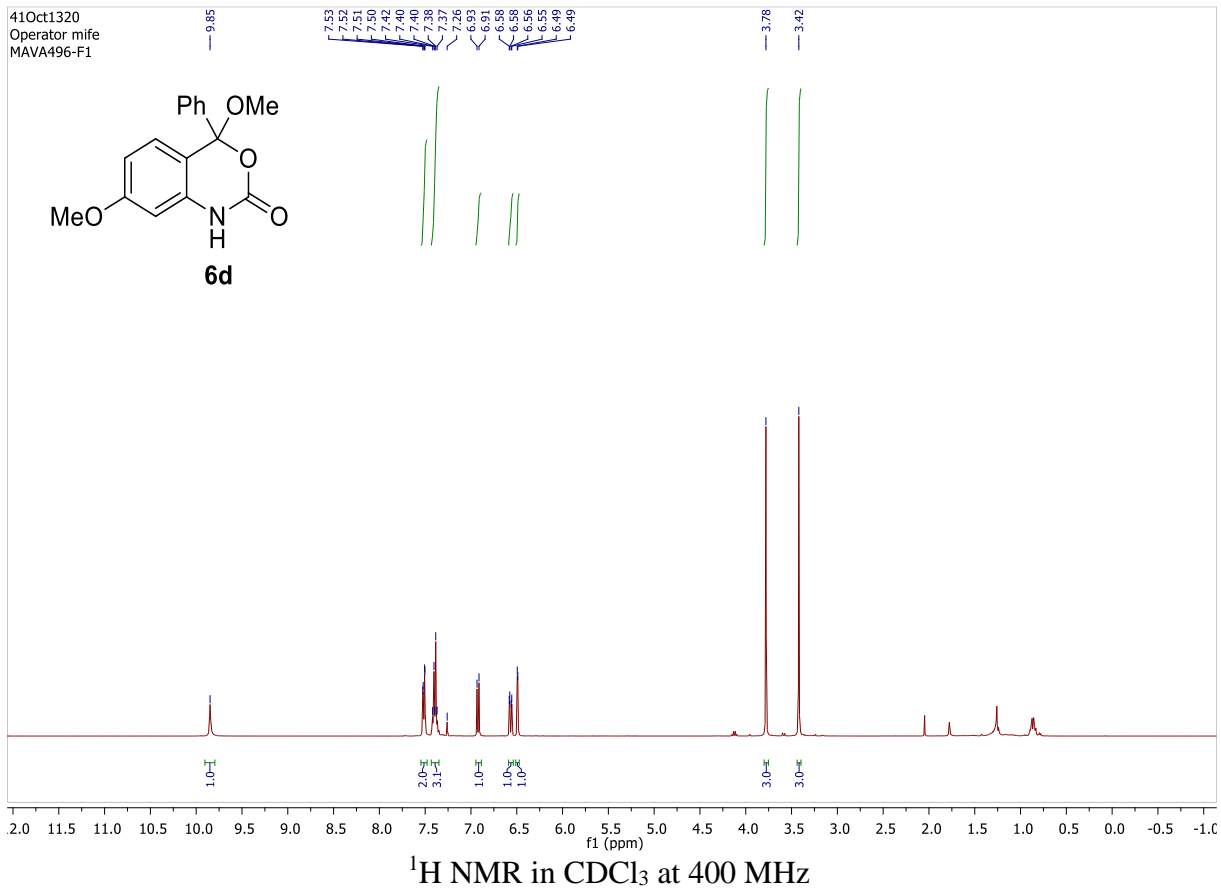


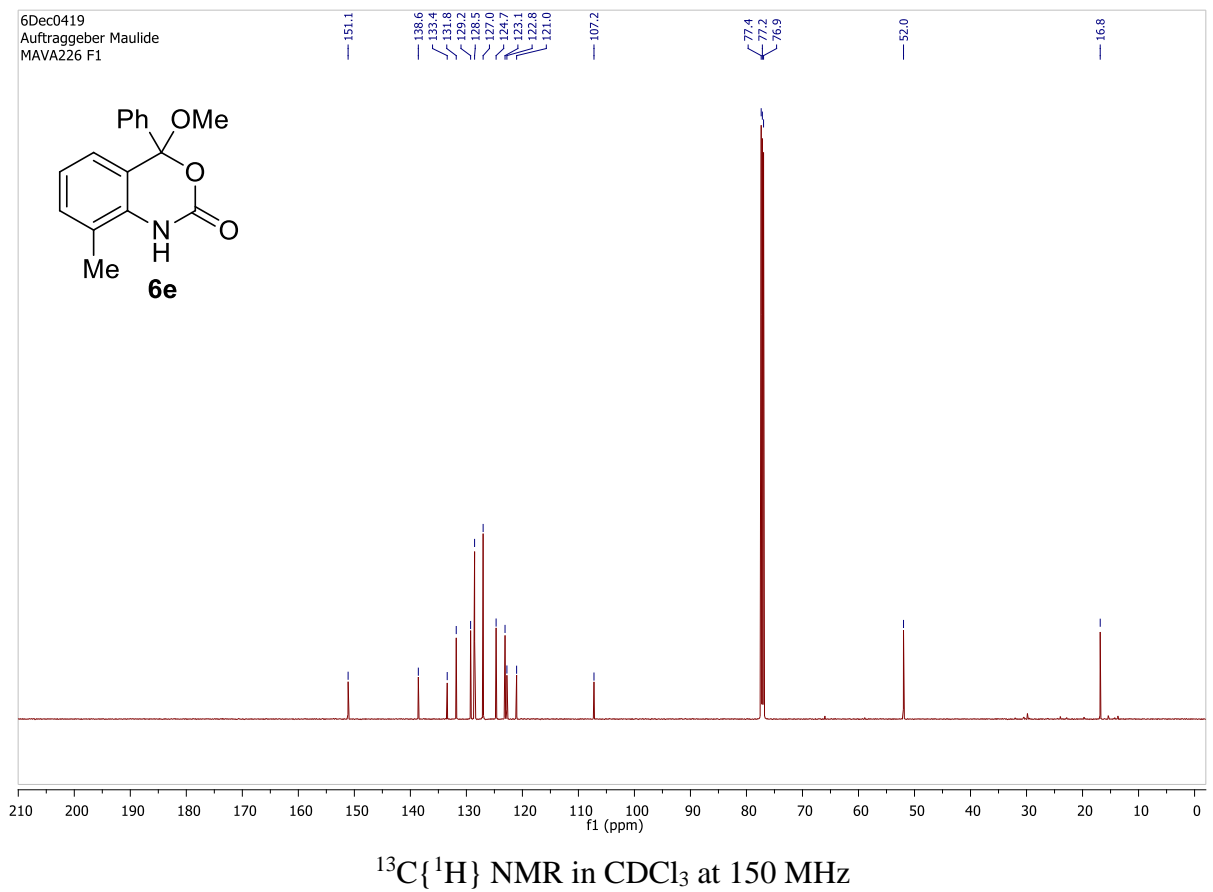
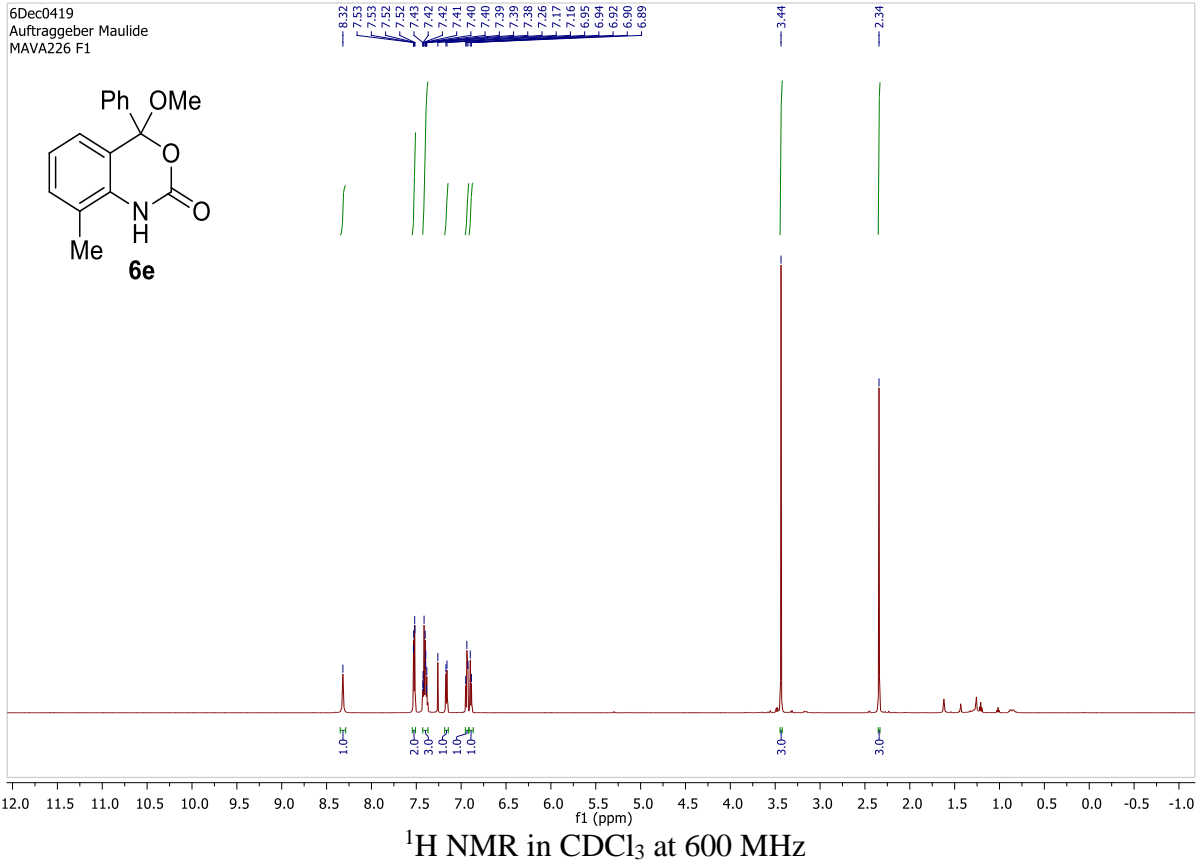


^1H NMR in CDCl_3 at 500 MHz

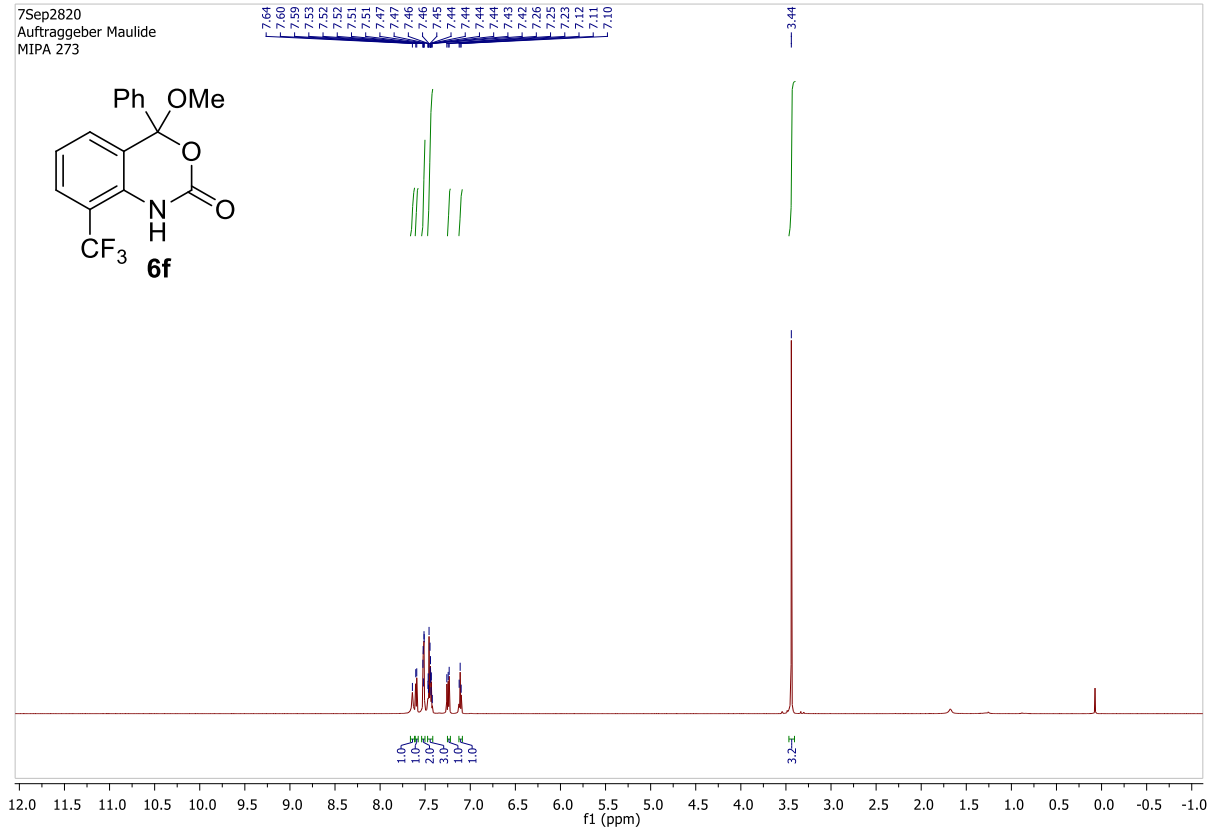
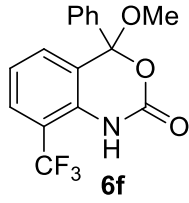


$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 126 MHz



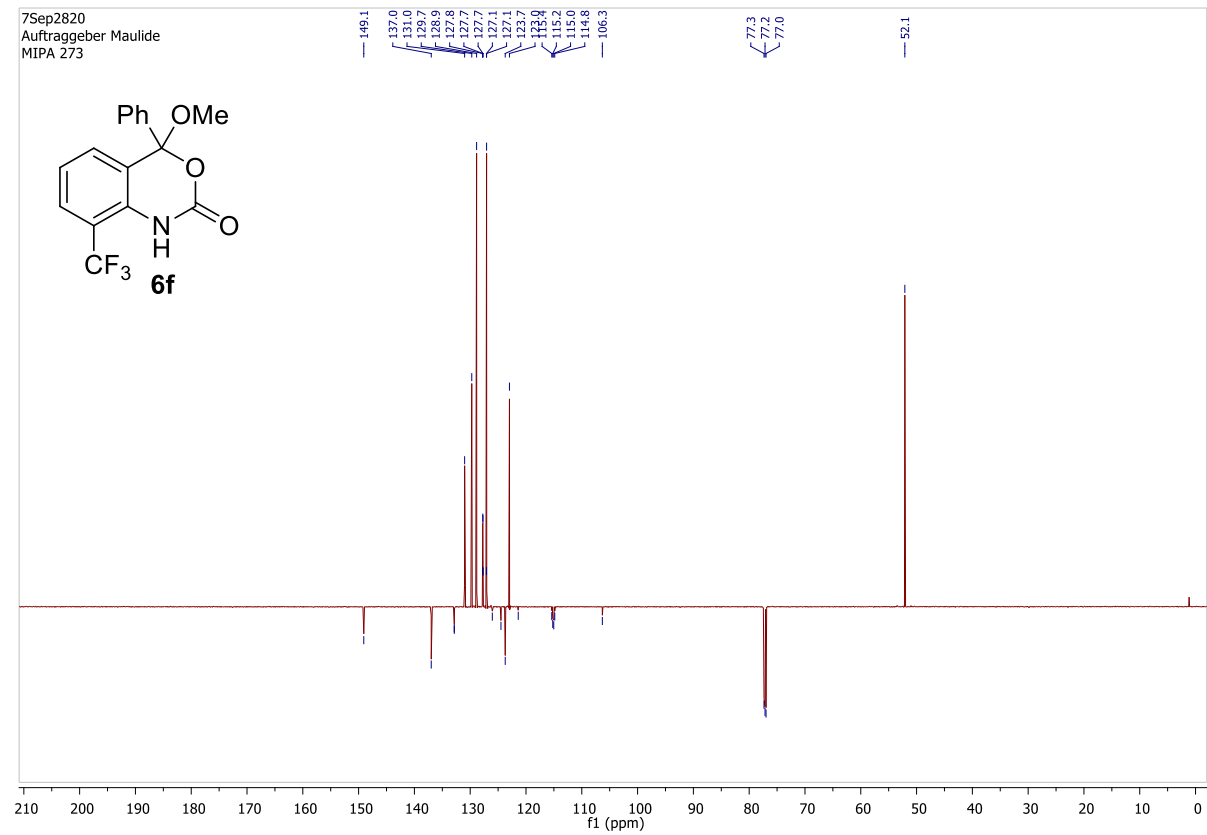
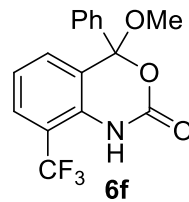


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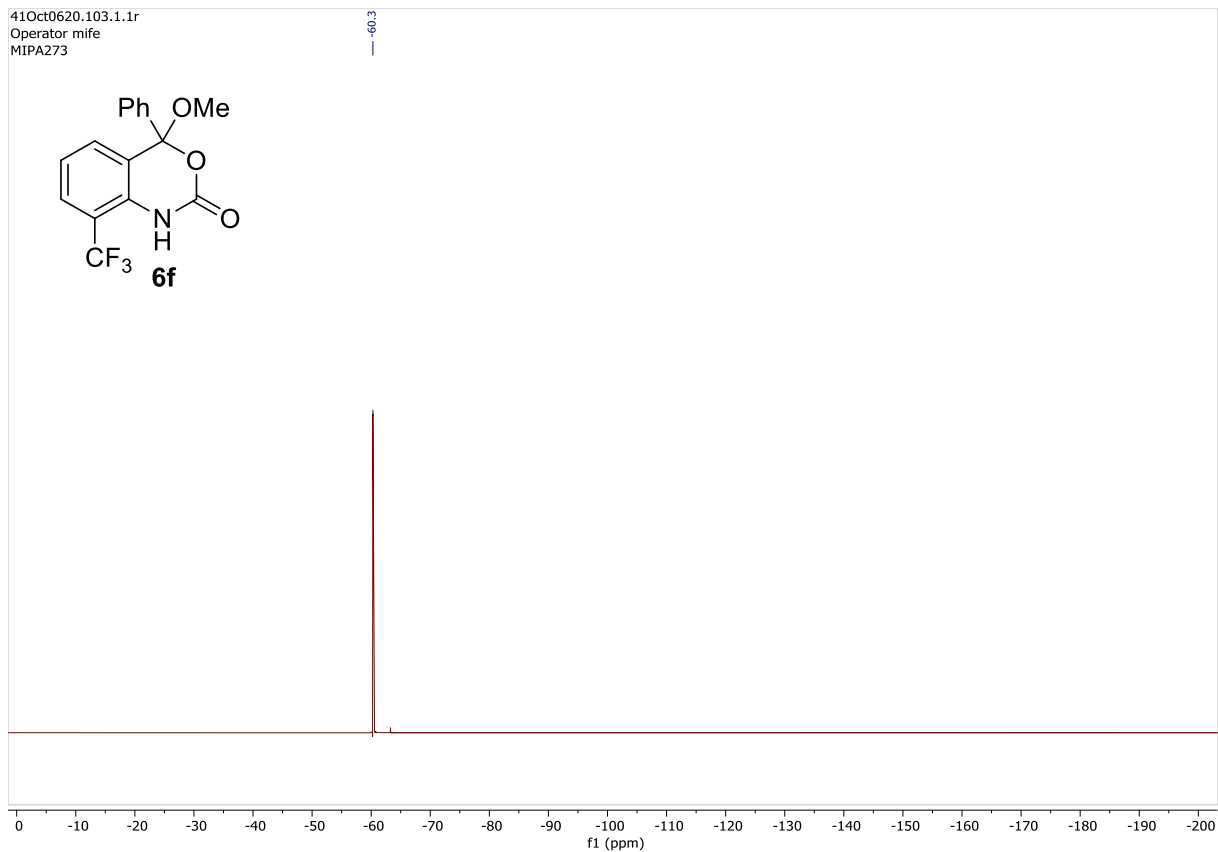
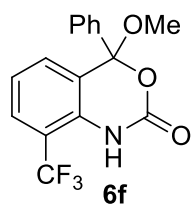
^1H NMR in CDCl_3 at 700 MHz

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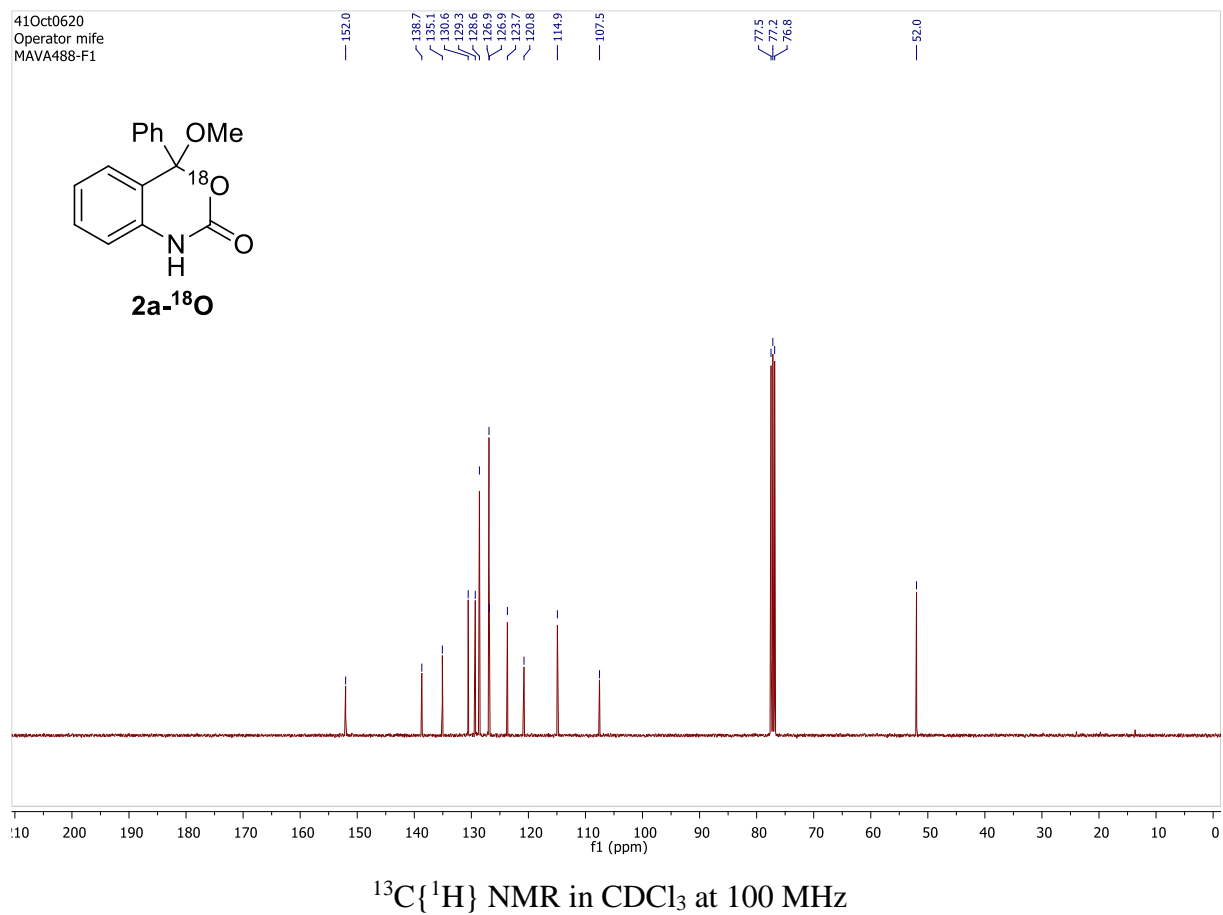
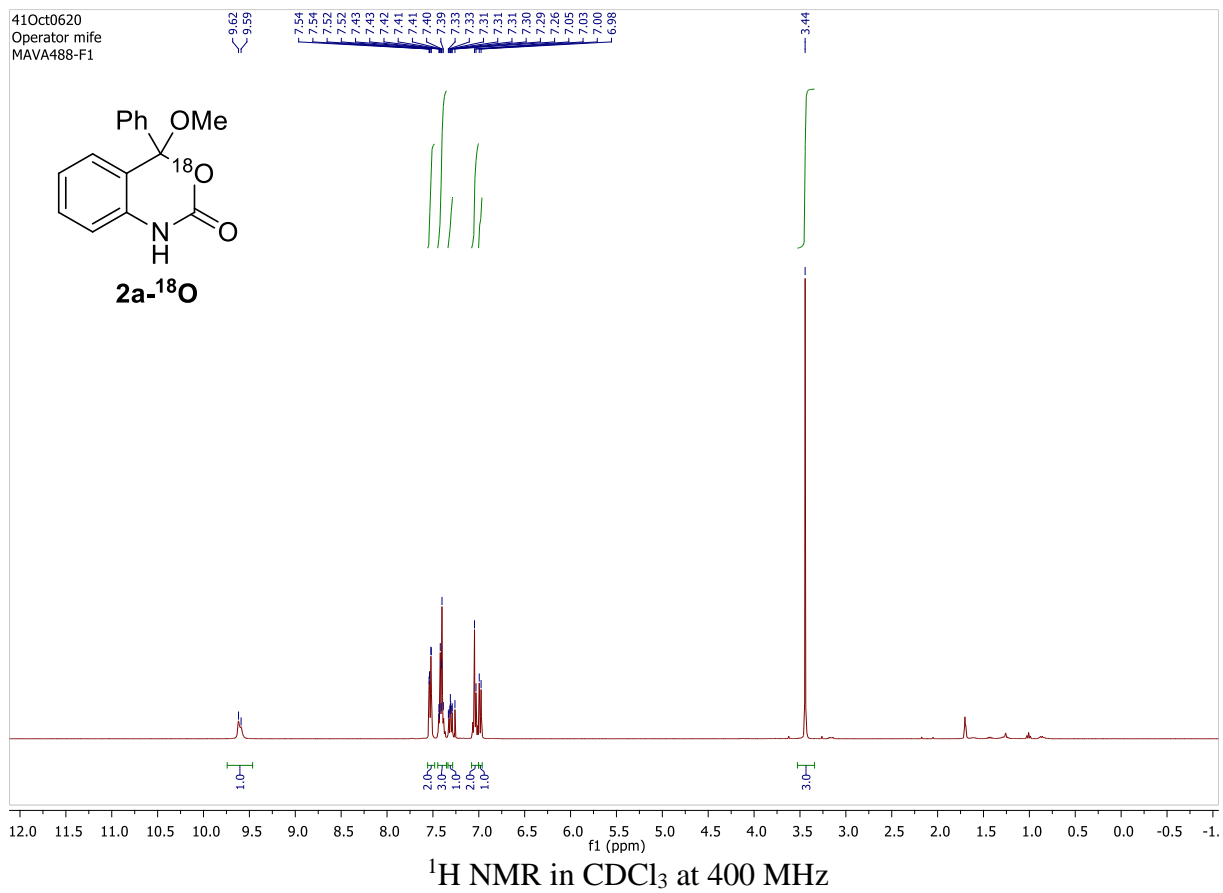


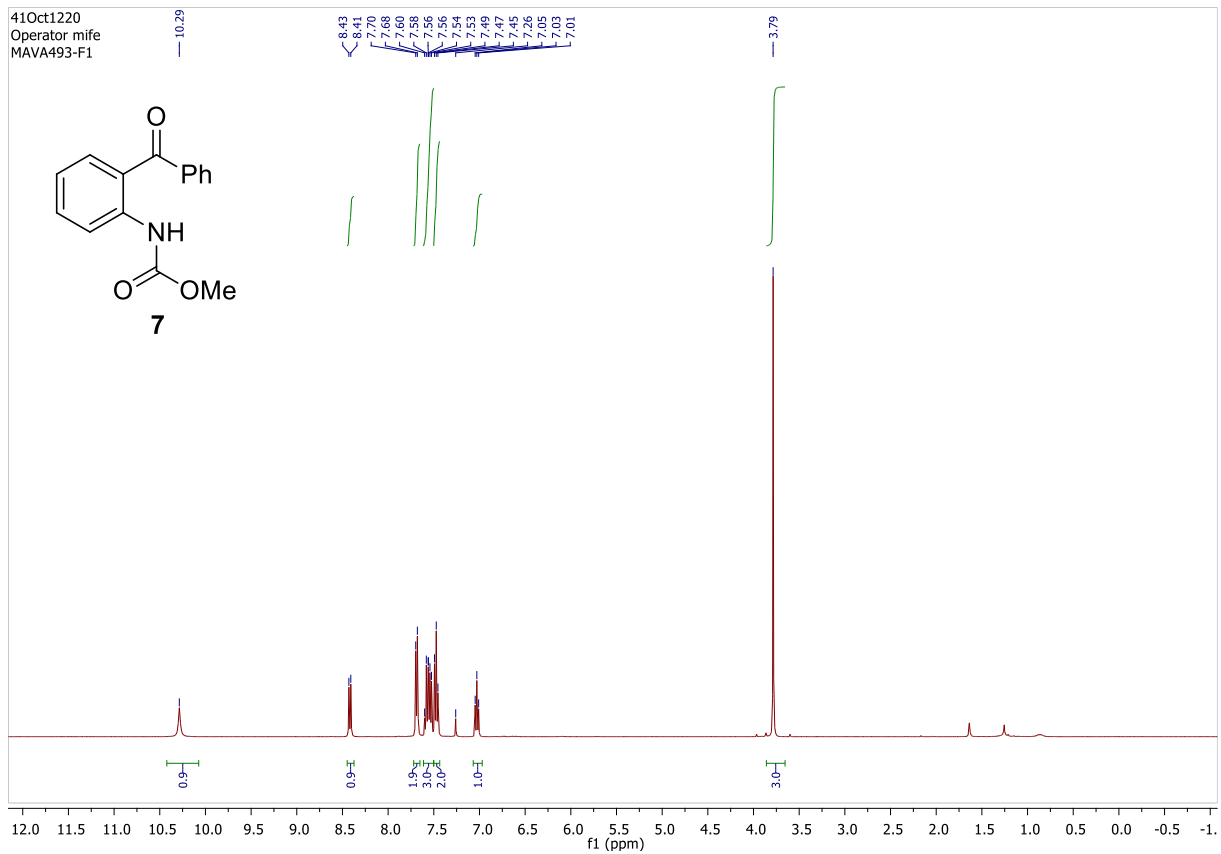
$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 175 MHz

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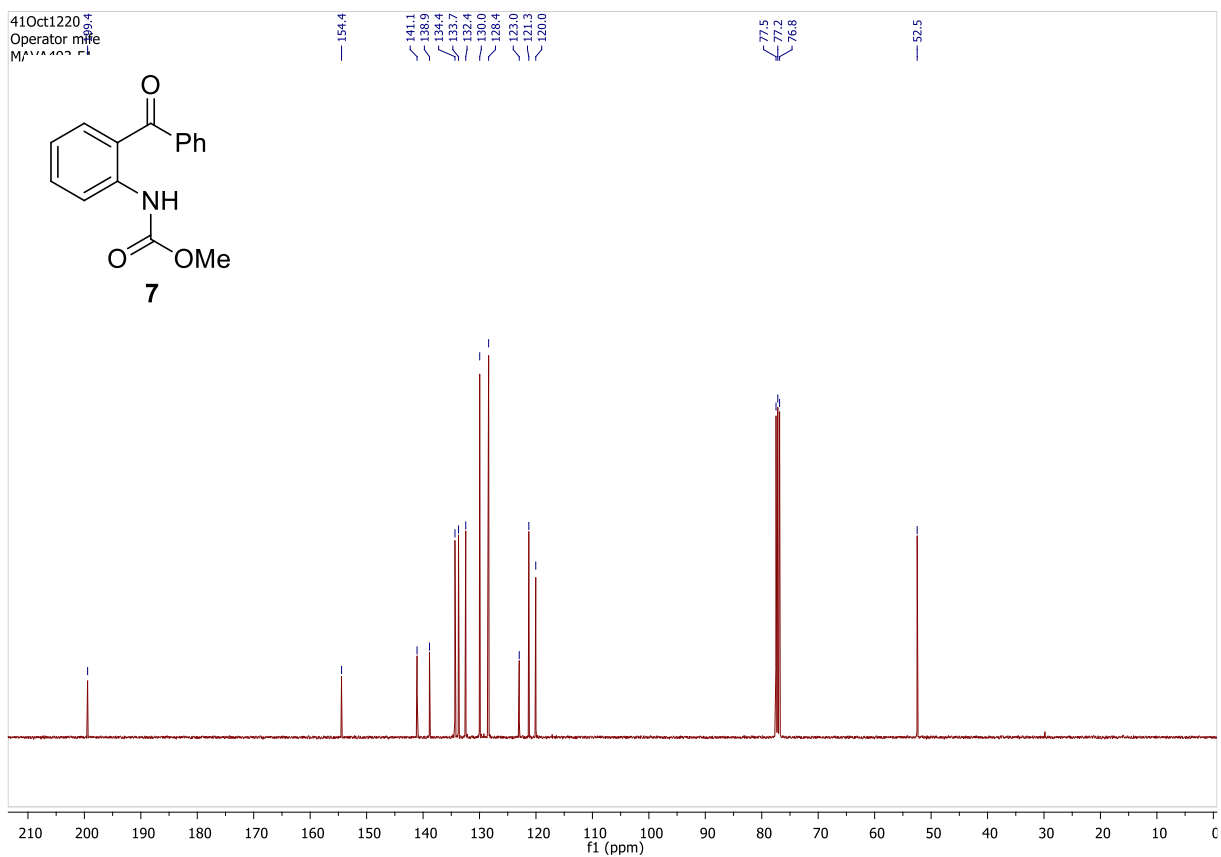


^{19}F NMR in $\text{DMSO-}d_6$ at 376 MHz

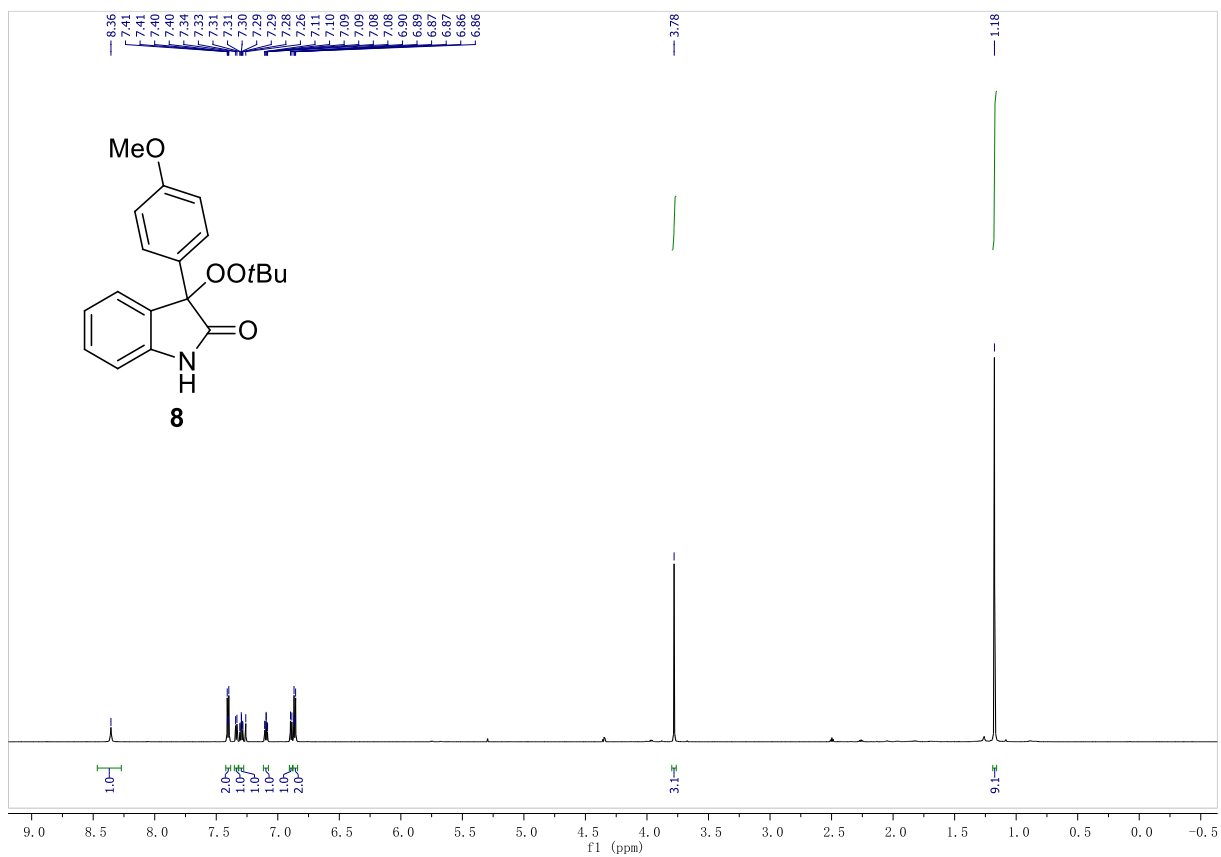




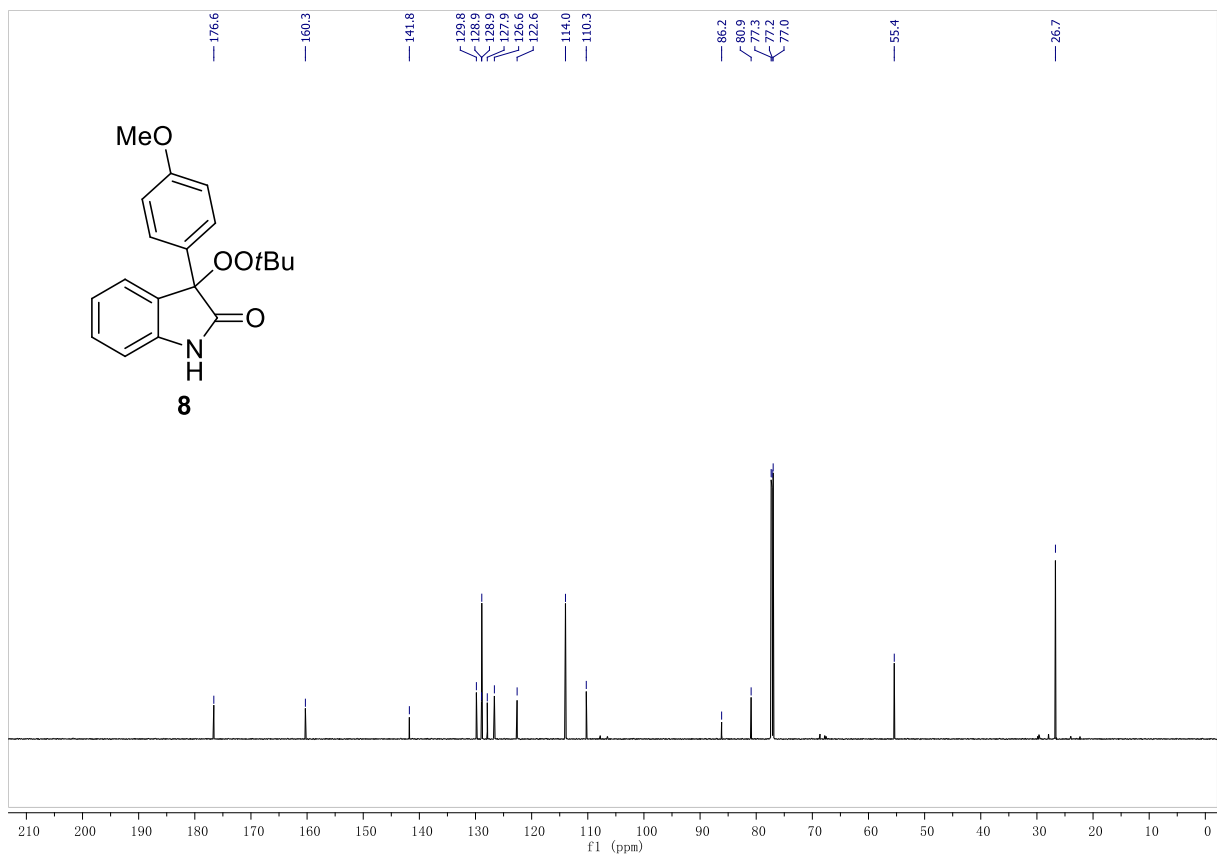
^1H NMR in CDCl_3 at 400 MHz



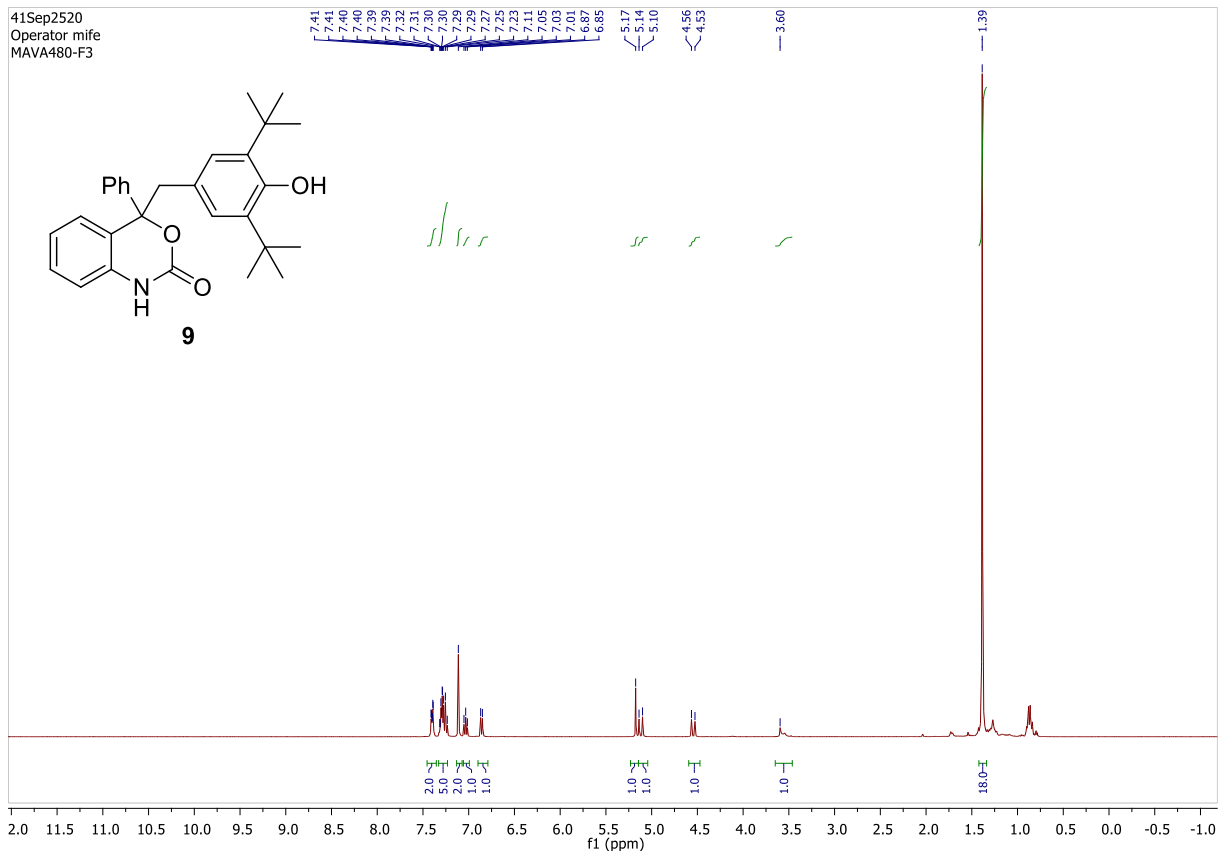
$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 100 MHz



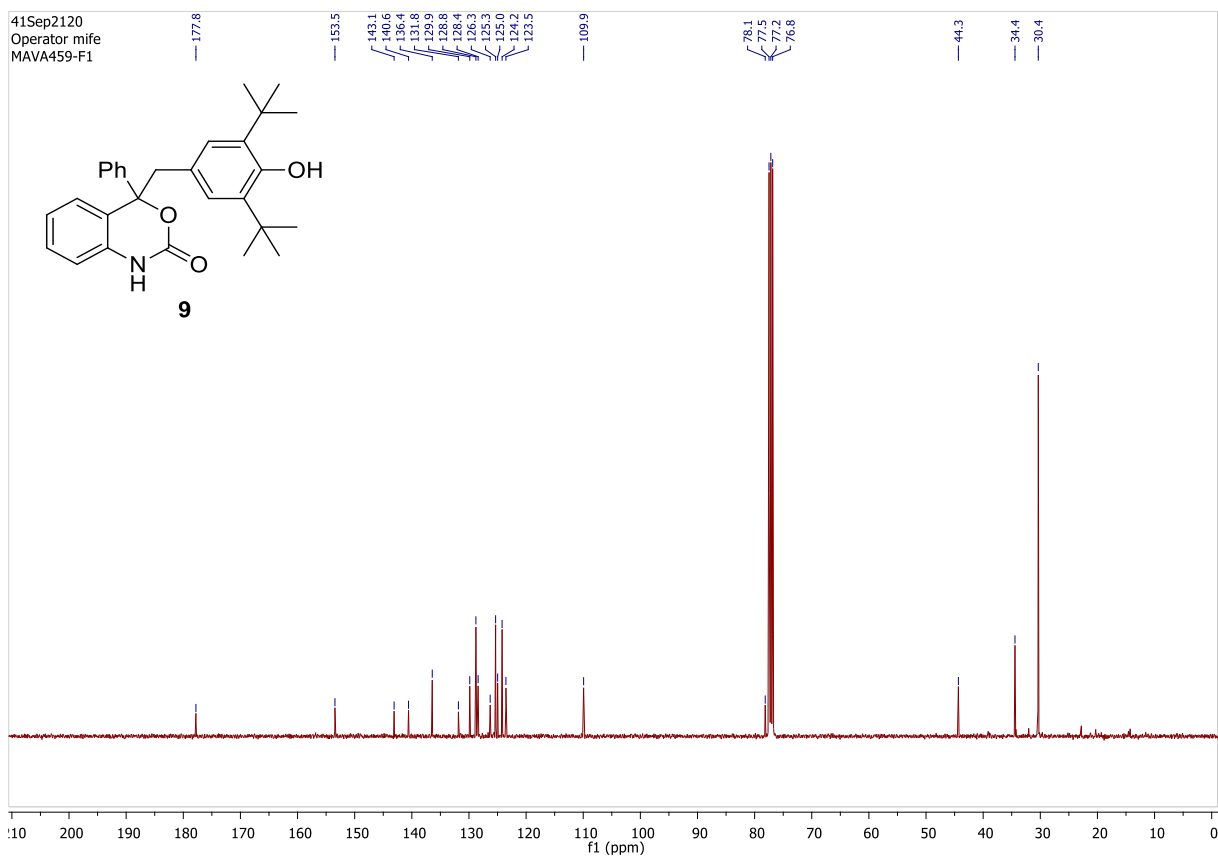
¹H NMR in CDCl₃ at 700 MHz



¹³C NMR in CDCl₃ at 176 MHz



^1H NMR in CDCl_3 at 400 MHz



$^{13}\text{C}\{^1\text{H}\}$ NMR in CDCl_3 at 100 MHz

8 References

- ¹ Ma, S.; Han, X.; Krishnan, S.; Virgil, S. C.; Stoltz, B. M. Catalytic Enantioselective Stereoablative Alkylation of 3-Haloindoles: Facile Access to Oxindoles with C3 All-Carbon Quaternary Stereocenters. *Angew. Chem. Int. Ed.* **2009**, *48*, 8037–8041.
- ² Toullec, P. Y.; Jagt, R. B. C.; de Vries, J. G.; Feringa, B. L.; Minnaard, A. J. Rhodium-Catalyzed Addition of Arylboronic Acids to Isatins: An Entry to Diversity in 3-Aryl-3-Hydroxyoxindoles. *Org. Lett.* **2006**, *8*, 2715–2718.
- ³ Gade, A. B.; Bagle, P. N.; Shinde, P. S.; Bhardwaj, V.; Banerjee, S.; Chande, A.; Patil, N. T. Catalytic Enantioselective 1,3-Alkyl Shift in Alkyl Aryl Ethers: Efficient Synthesis of Optically Active 3,3'-Diaryloxindoles. *Angew. Chem. Int. Ed.* **2018**, *57*, 5735–5739.
- ⁴ Xiao, Z. -K.; Yin, H. -Y.; Shao, L. -X. N-Heterocyclic Carbene-Palladium(II)-1-Methylimidazole Complex Catalyzed α -Arylation of Oxindoles with Aryl Chlorides and Aerobic Oxidation of the Products in a One-Pot Procedure. *Org. Lett.* **2013**, *15*, 1254–1257.
- ⁵ Ghosh, S.; Kintada, L. K.; Bhunia, S.; Bisai, A. Lewis acid-catalyzed Friedel–Crafts alkylations of 3-hydroxy-2-oxindole: an efficient approach to the core structure of azonazine. *Chem. Commun.* **2012**, *48*, 10132–10134.
- ⁶ Chaudhari, M. B.; Sutar, Y.; Malpathak, S.; Hazra, A.; Gnanaprakasam, B. Transition-Metal-Free C–H Hydroxylation of Carbonyl Compounds. *Org. Lett.* **2017**, *19*, 3628–3631.
- ⁷ Yin, L.; Kanai, M.; Shibasaki, M. A Facile Pathway to Enantiomerically Enriched 3-Hydroxy-2-Oxindoles: Asymmetric Intramolecular Arylation of α -Keto Amides Catalyzed by a Palladium–DifluorPhos Complex. *Angew. Chem. Int. Ed.* **2011**, *50*, 7620–7623.
- ⁸ Verschueren, K.; Cobbaut, M.; Demaerel, J.; Saadah, L.; Voet, A. R. D.; Van Lint, J.; De Borggraeve, W. M. Discovery of a potent protein kinase D inhibitor: insights in the binding mode of pyrazolo[3,4-d]pyrimidine analogues. *Med. Chem. Commun.* **2017**, *8*, 640–646.
- ⁹ Parrick, J.; Yahya, A.; Ijaz, A. S.; Yizun, J. Convenient preparation of 3,3-dibromo-1,3-dihydroindol-2-ones and indole-2,3-diones (isatins) from indoles. *J. Chem. Soc., Perkin Trans. 1* **1989**, 2009–2015.
- ¹⁰ Klare, H. F. T.; Goldberg, A. F. G.; Duquette, D. C.; Stoltz, B. M. Oxidative Fragmentations and Skeletal Rearrangements of Oxindole Derivatives. *Org. Lett.* **2017**, *19*, 988–991.
- ¹¹ Bruker SAINT v8.38B Copyright © 2005–2019 Bruker AXS.
- ¹² Sheldrick, G. M. (1996). *SADABS*. University of Göttingen, Germany.
- ¹³ Dolomanov, O. V.; Bourhis, L. J.; Gildea, R. J.; Howard, J. A. K.; Puschmann, H. OLEX2, *J. Appl. Cryst.* **2009**, *42*, 339–341.
- ¹⁴ Huebschle, C. B.; Sheldrick, G. M.; Dittrich, B. ShelXle: a Qt graphical user interface for SHELXL, *J. Appl. Cryst.* **2011**, *44*, 1281–1284.
- ¹⁵ Sheldrick, G. M. (2015). *SHELXS v 2016/4* University of Göttingen, Germany.
- ¹⁶ Spek, A. L. Structure validation in chemical crystallography. *Acta Cryst.* **2009**, D65, 148–155.