Supplementary Information

Photocatalytic Defluoroalkylation and Hydrodefluorination of Trifluoromethyls using *o*-Phosphinophenolate

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1. Supplementary Methods

1-1 General Information

A. Materials:

All reactions were carried out in oven-dried Schlenk tubes under argon atmosphere (purity≥99.999%) unless otherwise mentioned. Commercial reagents were purchased from Adamas-beta, TCI and Aldrich. Organic solutions were concentrated under reduced pressure on Buchi rotary evaporator. The LED lamps were purchased from Kessil (PR160-427 nm, 440 nm, 456 nm, 467 nm). The Photo Reaction Setup was purchased from HepatoChem.



Supplementary Figure 1. The Photo Reaction Setup and violet LED lamps

B. Analytical Methods:

¹H-NMR, ¹⁹F-NMR and ¹³C-NMR spectra were recorded on a Bruker Avance 400 spectrometer at ambient temperature. Data for ¹H-NMR are reported as follows: chemical shift (ppm, scale), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet and/or multiplet resonances, br = broad), coupling constant (Hz), and integration. Data for ¹³C- NMR are reported in terms of chemical shift (ppm, scale), multiplicity, and coupling constant (Hz). HRMS analysis was performed on Finnigan LCQ advantage Max Series MS System. ESI-mass data were acquired using

a Thermo LTQ Orbitrap XL Instrument equipped with an ESI source and controlled by Xcalibur software. UV-Vis spectrum was measured by UV-3600. Flash column chromatographic purification of products was accomplished using forced-flow chromatography on Silica Gel (200-300 mesh).

1-2 Preparation of Catalysts

1-2-1 General Procedure for preparation of catalyst

PO catalysts **PO1**, **PO3**, **PO4**, **PO5**, **PO6**, **PO7** were known compounds and prepared according to the literature procedures.¹⁻⁶



Other **PO** catalysts used in this work were prepared by the following method:



The Catalyst *o*-Phosphinophenol (PO) was prepared according to a previous published protocol.⁷ *n*-Butyl lithium (1.6 M in hexane, 20 mmol) was added into a solution of *o*-bromophenol (10 mmol) in Et₂O (22 mL) at -78 °C, giving immediately a white suspension. The cold bath was removed and stirring was continued at room temperature for 30 mins. Chlorodiphenylphosphine (10 mmol) was added into this solution at -78 °C. After stirring for 3 h, the cold bath was removed and stirring was continued at 0 °C for 18 h. The mixture was extracted with aqueous solutions of NaH₂PO₄ (0.1 M), and the combined organic layers were washed with brine, dried over Na₂SO₄, filtered and concentrated in vacuo. The crude residue was purified by flash column chromatography on silica gel to give desired product.

2-(diphenylphosphaneyl)-6-methylphenol (PO2): Following the general procedure, obtained in 75% yield as white solid after silica gel chromatography. (2.19 g, eluent: petroleum ether/ethyl acetate = 10/1). Mp = 143 – 145 °C ¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.27 (m, 10H), 7.16 (dd, *J* = 7.1, 1.1 Hz, 1H), 6.94 – 6.71 (m, 2H), 6.41 (d, *J* = 7.9 Hz, 1H), 2.26 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 157.6 (d, *J* = 18.8 Hz), 135.1 (d, *J* = 3.8 Hz), 133.4 (d, *J* = 18.6 Hz), 132.9, 132.33 (d, *J* = 3.4 Hz), 129.0, 128.7 (d, *J* = 7.3 Hz), 124.5 (d, *J* = 1.8 Hz), 120.7 (d, *J* = 2.9 Hz), 120.0 (d, *J* = 3.0 Hz), 16.3 (d, *J* = 2.6 Hz). ³¹P NMR (162 MHz, CDCl₃) δ -29.9.

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₉H₁₈OP, 293.1090; found, 293.1092.

1-3 Preparation of Substrates

1-3-1 General Procedure for preparation of trifluoroacetamides and pentafluoropropionamides



General Procedure 1: Trifluoroacetamides and Pentafluoropanamides.⁸ To a solution of aniline and Et₃N (1.5 equiv) in CH_2Cl_2 was added trifluoroacetic anhydride (1.2 equiv) dropwise at 0 °C. After addition, the mixture was stirred at room temperature until TLC to indicate aniline disappeared. The reaction was then quenched with H₂O and extracted three times with CH_2Cl_2 . The combined organic layers were washed with brine, dried over Na₂SO₄, filtered and concentrated in vacuo. The crude residue was purified by flash column chromatography on silica gel to give desired products.

1-3-2 General Procedure for preparation of trifluoroacetates

$$\mathbb{R}^{\mathcal{O}H} \xrightarrow{\begin{array}{c} (CF_3CO)_2O \ (1.2 \text{ equiv}) \\ \underline{2,6\text{-lutidine} \ (1.5 \text{ equiv})} \\ Et_2O, 0 \ ^{\circ}C \end{array}} \mathbb{R}_{O} \xrightarrow{\begin{array}{c} O \\ CF_3 \end{array}}$$

General Procedure 2: Trifluoroacetates.⁸ To a solution of alcohol and 2,6-lutidine (1.5 equiv) in Et₂O was added trifluoroacetic anhydride (1.3 equiv) dropwise at 0 °C. After addition, the mixture was stirred at 0 °C until TLC indicated that alcohol disappeared. The reaction was then poured into H₂O and extracted with 1N HCl (×2), saturated aqueous NaHCO₃, brine, dried over Na₂SO₄, filtered, and concentrated in vacuo. The crude residue was purified by flash column chromatography on silica gel to give desired products.

2. Supplementary Discussion

2-1 Investigation of the Key Reaction Parameters

Supplementary Table 1: Parameters affecting defluoroalkylation of trifluoroacetamides

	$\begin{array}{c} \begin{array}{c} H \\ H \\ O \\ O \\ \end{array} + \\ \begin{array}{c} O \\ O \\ \end{array} \\ \begin{array}{c} H \\ O \\ \end{array} \\ \begin{array}{c} CF_3 \\ HCO_2K (2.0 \text{ equiv}) \\ \hline HCO_2K (2.0 \text{ equiv}) \\ \hline DMA (2.0 \text{ mL}), \text{ r.t., 24 h} \\ \hline \text{violet LEDs (427 nm)} \end{array} \end{array}$	N C C C C C C C C C C C C C C C C C C C
Entry	Variations from standard conditions	Yield (%) ^{<i>a</i>}
1	none	90
2	DMF instead of DMA	89
3	DMSO instead of DMA	91
4	NMP instead of DMA	80
5	MeCN instead of DMA	23
6	THF instead of DMA	4
7	Acetone instead of DMA	7
8	1,4-Dioxane instead of DMA	0
9	Toluene instead of DMA	0
10	DCM instead of DMA	0
11	PO2 instead of PO1	90
12	PO3 instead of PO1	87
13	PO4 instead of PO1	93
14	PO5 instead of PO1	65

15	PO6 instead of PO1	15
16	PO7 instead of PO1	< 5
17	CySH instead of 1-Adamantanethiol	75
18	DABCO instead of 1-Adamantanethiol	0
19	Triethylsilane instead of HCO ₂ K	30
20	HCO ₂ Na instead of HCO ₂ K	82
21	HCO ₂ Li instead of HCO ₂ K	92
22	HCO ₂ Cs instead of HCO ₂ K	70
23	2 (0.4 mmol) instead of 2 (0.3 mmol)	96
24	PO1 (2 mol%) instead of PO1 (10 mol%)	52
25	PO1 (5 mol%) instead of PO1 (10 mol%)	67
26	1-AdSH (5 mol%) instead of 1-AdSH (20 mol%)	59
27	1-AdSH (10 mol%) instead of 1-AdSH (20 mol%)	72
28	440 nm instead of 427 nm	70
29	456 nm instead of 427 nm	trace
30	467 nm instead of 427 nm	trace
31	PPh ₃ instead of PO1	trace
32	4-t-Bu-C ₆ H ₄ -OH instead of PO1	0
33	PPh ₃ + 4- <i>t</i> -Bu-C ₆ H ₄ -OH (1:1, 10 mol%) instead of PO1 (10 mol%)	10
34	<i>w/o</i> PO1	0
35	w/o 1-AdSH	trace
36	w/o HCO ₂ K	18
37	w/o light	0
38	Under air	trace

"Yield determined by ¹H-NMR using diphenylmethane as an internal standard.



Supplementary Table 2: Parameters affecting hydrodefluorination of trifluoroacetamides

$ \begin{array}{c} H \\ CF_{3} \\ O \\ 1, 0.2 \text{ mmol} \end{array} $	PO1 (10 mol %) Cs ₂ CO ₃ (30 mol %) 1-AdSH (20 mol %) HCO ₂ Cs (1.2 equiv) DMSO (2.0 mL), r.t., 24 h violet LEDs (427 nm)			
Entry Vari	ations from standard cond	litions	Yield 4 $(\%)^a$	Yield 4'(%) ^a
1	none		94	< 5

2	HCO ₂ Cs (2.0 equiv) instead of HCO ₂ Cs (1.2 equiv)	79	17
3	HCO ₂ Cs (3.0 equiv) instead of HCO ₂ Cs (1.2 equiv)	60	38
4	DMA instead of DMSO	65	trace
5	DMF instead of DMSO	50	trace
6	<i>w/o</i> 1-AdSH	12	n.d.
7	w/o HCO ₂ Cs	10	n.d.

"Yield determined by ¹H-NMR using diphenylmethane as internal standard.

2-2 Experimental Procedures and Spectral Data

2-2-1 General Procedure

General Procedure A: trifluoroacetamide (1.0 equiv, 0.2 mmol) (if solid), Alkene (1.5 equiv, 0.3 mmol) (if solid), PO1 (10 mol%), Cs_2CO_3 (30 mol%), 1-adamananethiol (20 mol%), HCO₂K (2.0 equiv, 0.4 mmol) were placed in a transparent Schlenk tube equipped with a stirring bar. The tube was evacuated and filled with argon (three times). To these solids, trifluoroacetamide (1.0 equiv, 0.2 mmol) (if liquid), alkene (1.5 equiv, 0.3 mmol) (if liquid), and anhydrous DMA (2 mL) were added via a gastight syringe under argon atmosphere. The reaction mixture was stirred under irradiation with violet LEDs (427 nm) in a photoreactor at room temperature for 24 h. The mixture was quenched with brine and extracted with ethyl acetate (3×10 mL). The organic layers were combined and concentrated under vacuo. The product was purified by flash column chromatography on silica gel.

General Procedure B: trifluoacatete (2.0 equiv, 0.4 mmol) (if solid), alkene (1.0 equiv, 0.2 mmol) (if solid), **PO1** (10 mol%), Cs_2CO_3 (30 mol%), 1-adamananethiol (20 mol%), HCO₂K (2.0 equiv, 0.4 mmol) were placed in a transparent Schlenk tube equipped with a stirring bar. The tube was evacuated and filled with argon (three times). To these solids, trifluoacetate (2.0 equiv, 0.4 mmol) (if liquid), alkene (1.0 equiv, 0.2 mmol) (if liquid), and anhydrous DMA (2 mL) were added via a gastight syringe under argon atmosphere. The reaction mixture was stirred under irradiation with violet LEDs (427 nm) in a photoreactor at room temperature for 24 h. The mixture was quenched with brine and extracted with ethyl acetate (3 × 10 mL). The organic layers were combined and concentrated under vacuo. The product was purified by flash column chromatography on silica.

General Procedure C: trifluoroacetamide (1.0 equiv, 0.2 mmol) (if solid), **PO1** (10 mol%), Cs_2CO_3 (30 mol%), 1-adamananethiol (20 mol%), HCO_2Cs (1.2 equiv, 0.24 mmol) were placed in a transparent Schlenk tube equipped with a stirring bar. The tube was evacuated and filled with argon (three times). To these solids,

trifluoroacetamide (1.0 equiv, 0.2 mmol) (if liquid) and anhydrous DMSO (2 mL) were added via a gastight syringe under argon atmosphere. The reaction mixture was stirred under irradiation with violet LEDs (427 nm) in a photoreactor at room temperature for 24 h. The mixture was quenched with brine and extracted with ethyl acetate (3×10 mL). The organic layers were combined and concentrated under vacuo. The product was purified by flash column chromatography on silica.

General Procedure D: trifluoromethyl (hetero) arenes (1.0 equiv, 0.2 mmol) (if solid), alkene (1.5 equiv, 0.3 mmol) (if solid), **PO1** (10 mol%), Cs_2CO_3 (30 mol%), 1-adamananethiol (20 mol%), HCO₂Li (2.0 equiv, 0.4 mmol) were placed in a transparent Schlenk tube equipped with a stirring bar. The tube was evacuated and filled with argon (three times). To these solids, trifluomethyl (hetero) arenes (1.0 equiv, 0.2 mmol) (if liquid), alkene (1.5 equiv, 0.3 mmol) (if liquid), and anhydrous DMSO (2 mL) were added via a gastight syringe under argon atmosphere. The reaction mixture was stirred under irradiation with violet LEDs (427 nm) in a photoreactor at room temperature for 24 h. The mixture was quenched with brine and extracted with ethyl acetate (3 × 10 mL). The organic layers were combined and concentrated under vacuo. The product was purified by flash column chromatography on silica.

2-2-2 Spectral Data

2,2-difluoro-6-hydroxy-*N***-phenylhexanamide (3):** Following the general procedure A, obtained in 86% yield as white solid after silica gel chromatography. (41.9 mg, eluent: petroleum ether/ethyl acetate = 2/1).

 $Mp = 61 - 63 \ ^{\circ}C$

¹H NMR (400 MHz, CDCl₃) δ 8.09 (s, 1H), 7.57 (d, *J* = 7.9 Hz, 2H), 7.37 (t, *J* = 7.9 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 3.66 (t, *J* = 5.7 Hz, 2H), 2.32 – 2.12 (m, 2H), 1.73 (s, 1H), 1.62 (dd, *J* = 7.0, 4.0 Hz, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 162.2 (t, J = 28.6 Hz), 136.0, 129.2, 125.6, 120.3, 118.3 (t, J = 253.6 Hz), 62.2, 33.5 (t, J = 23.2 Hz), 31.9, 18.1 (t, J = 4.5 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -105.5 (2F, td, J = 17.3, 2.9 Hz).

HRMS (ESI) *m*/*z*: [M+Na]⁺ Calcd. for C₁₂H₁₅F₂NO₂Na, 266.0963; found, 266.0962.

2,2-difluoro-*N***-phenylacetamide (4):** Following the general procedure C, obtained in 94% yield as white solid after silica gel chromatography. (32.1 mg, eluent: petroleum ether/ethyl acetate = 15/1). The compound data was in agreement with the literature (*Science* **2021**, *371*, 1232–1240).

¹H NMR (400 MHz, CDCl₃) δ 7.92 (s, 1H), 7.57 (d, *J* = 8.0 Hz, 2H), 7.38 (t, *J* = 7.9 Hz, 2H), 7.21 (t, *J* = 7.4 Hz, 1H), 6.02 (t, *J* = 54.4 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 160.3 (t, J = 24.1 Hz), 135.7, 129.3, 125.9, 120.3, 108.6 (t, J = 254.2 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -125.5 (2F, dd, J = 54.4, 2.3 Hz).

2,2-difluoro-6-hydroxy-*N***-(4-methoxyphenyl)hexanamide (5):** Following the general procedure A, obtained in 80% yield as white solid after silica gel chromatography. (43.5 mg, eluent: petroleum ether/ethyl acetate = 2/1).

 $Mp = 90 - 92 \ ^{\circ}C$

¹H NMR (400 MHz, *d*₆-DMSO) δ 10.38 (s, 1H), 7.59 (d, *J* = 9.1 Hz, 2H), 6.93 (d, *J* = 9.1 Hz, 2H), 3.74 (s, 3H), 3.41 (t, *J* = 5.7 Hz, 2H), 2.25 – 2.04 (m, 2H), 1.52 – 1.42 (m, 4H).

¹³C NMR (101 MHz, d_6 -DMSO) δ 162.3 (t, J = 29.4 Hz), 156.8, 130.7, 122.9, 118.7 (t, J = 251.5 Hz), 114.3, 60.7, 55.7, 34.1 (t, J = 23.3 Hz), 32.2, 18.6 (t, J = 4.3 Hz).

¹⁹F NMR (376 MHz, d_6 -DMSO) δ -104.1 (2F, t, J = 17.3 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₇F₂NO₃Na, 296.1069; found, 296.1072.



2,2-difluoro-6-hydroxy-*N***-(o-tolyl)hexanamide** (6): Following the general procedure A, obtained in 70% yield as colorless oil after silica gel chromatography. (36.0 mg, eluent: petroleum ether/ethyl acetate = 2/1).

¹H NMR (400 MHz, CDCl₃) δ 7.91 (s, 1H), 7.83 (d, *J* = 7.9 Hz, 1H), 7.23 (dd, *J* = 7.5, 3.6 Hz, 2H), 7.15 (t, *J* = 7.1 Hz, 1H), 3.67 (t, *J* = 5.7 Hz, 2H), 2.29 (s, 3H), 2.27 – 2.14 (m, 2H), 1.71 (s, 1H), 1.68 – 1.59 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 162.2 (t, J = 28.6 Hz), 133.8, 130.7, 129.4, 127.0, 126.3, 122.9, 118.5 (t, J = 253.4 Hz), 62.2, 33.5 (t, J = 23.3 Hz), 31.9, 18.1 (t, J = 4.5 Hz), 17.5.

¹⁹F NMR (376 MHz, CDCl₃) δ -105.4 (2F, td, J = 17.2, 3.2 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₇F₂NO₂Na, 280.1120; found, 280.1124.

N-([1,1'-biphenyl]-4-yl)-2,2-difluoro-6-hydroxyhexanamide (7): Following the general procedure A, obtained in 85% yield as white solid after silica gel chromatography. (54.3 mg, eluent: petroleum ether/ethyl acetate = 2/1).

 $Mp = 162 - 164 \text{ }^{\circ}\text{C}$

¹H NMR (400 MHz, Acetone-*d*₆) δ 9.79 (s, 1H), 7.88 (d, *J* = 8.3 Hz, 2H), 7.83 – 7.61 (m, 4H), 7.46 (dd, *J* = 7.9, 7.4 Hz, 2H), 7.42 – 7.22 (m, 1H), 3.58 (s, 2H), 2.36 – 2.11 (m, 2H), 1.71 – 1.48 (m, 4H).

¹³C NMR (101 MHz, Acetone- d_6) δ 162.2 (t, J = 28.6 Hz), 140.2, 137.5, 136.9, 128.9, 127.2, 127.2, 126.6, 120.9, 118.6 (t, J = 272.0 Hz), 61.0, 33.8 (t, J = 23.4 Hz), 32.2, 18.3 (t, J = 4.6 Hz).

¹⁹F NMR (376 MHz, Acetone- d_6) δ -106.1 (2F, td, J = 17.1, 1.4 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₈H₁₉F₂NO₂Na, 342.1276; found, 342.1277.



ethyl4-(2-fluoro-6-hydroxy-2-(4-hydroxybutyl)hexanamido)benzoate(8):Following the general procedure A, obtained in 63% yield as white solid after silicagel chromatography. (46.5 mg, eluent: petroleum ether/ethyl acetate = 1/1).

 $Mp = 90 - 92 \ ^{\circ}C$

¹H NMR (400 MHz, CDCl₃) δ 8.41 (d, *J* = 8.6 Hz, 1H), 8.02 (d, *J* = 8.7 Hz, 2H), 7.67 (d, *J* = 8.7 Hz, 2H), 4.36 (q, *J* = 7.1 Hz, 2H), 3.60 (t, *J* = 5.8 Hz, 4H), 2.15 – 1.96 (m, 4H), 1.94 – 1.72 (m, 2H), 1.69 – 1.49 (m, 6H), 1.39 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 170.6 (d, J = 20.4 Hz), 166.1, 140.8, 130.8, 126.6, 119.2, 101.1 (d, J = 188.8 Hz), 62.2, 61.0, 36.9 (d, J = 22.0 Hz), 32.4, 19.5 (d, J = 3.1 Hz), 14.3.

¹⁹F NMR (376 MHz, CDCl₃) δ -164.3 – -164.6 (1F, m).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₉H₂₈FNO₅Na, 392.1844; found, 392.1843.



N-(4-cyanophenyl)-2-fluoro-6-hydroxy-2-(4-hydroxybutyl)hexanamide(9):Following the general procedure A, obtained in 50% yield as colorless oil after silicagel chromatography. (32.2 mg, eluent: petroleum ether/ethyl acetate = 1/1).

¹H NMR (400 MHz, CDCl₃) δ 8.32 (d, J = 9.0 Hz, 1H), 7.73 (d, J = 8.9 Hz, 2H), 7.65 (d, J = 8.8 Hz, 2H), 3.79 – 3.54 (m, 4H), 2.16 – 1.96 (m, 3H), 1.96 – 1.77 (m, 3H), 1.62 – 1.53 (m, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 170.7 (d, J = 20.4 Hz), 140.7, 133.4, 119.8, 118.6, 107.9, 101.2 (d, J = 188.7 Hz), 62.4, 36.9 (d, J = 22.1 Hz), 32.4, 19.5 (d, J = 3.2 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -164.0 - -164.8 (1F, m).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₇H₂₄FN₂O₃, 323.1765; found, 323.1763.

2,2-difluoro-6-hydroxy-*N***-(pyridin-3-yl)hexanamide (10):** Following the general procedure A, obtained in 68% yield as colorless oil after silica gel chromatography. (33.2 mg, eluent: petroleum ether/ethyl acetate = 2/1).

¹H NMR (400 MHz, CDCl₃) δ 8.69 (s, 1H), 8.47 (s, 1H), 8.43 (d, J = 4.4 Hz, 1H), 8.19 (d, J = 8.4 Hz, 1H), 7.34 (dd, J = 8.4, 4.7 Hz, 1H), 3.68 (t, J = 5.7 Hz, 2H), 2.31 -2.15 (m, 2H), 1.95 (s, 1H), 1.68 -1.60 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 162.8 (t, J = 29.5 Hz), 146.4, 141.6, 133.3, 127.9, 123.9, 118.1 (t, J = 253.4 Hz), 62.1, 33.5 (t, J = 23.1 Hz), 31.9, 18.1 (t, J = 4.4 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -105.2 (2F, td, J = 17.3, 2.5 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₁H₁₅F₂N₂O₂, 245.1096; found, 245.1104.



2,2-difluoro-6-hydroxy-*N***-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)** hexanamide (11): Following the general procedure A, obtained in 88% yield as colorless oil after silica gel chromatography. (64.9 mg, eluent: petroleum ether/ethyl acetate = 2/1).

¹H NMR (400 MHz, CDCl₃) δ 8.08 (s, 1H), 7.81 (d, *J* = 8.5 Hz, 2H), 7.59 (d, *J* = 8.5 Hz, 2H), 3.67 (t, *J* = 5.9 Hz, 2H), 2.35 – 2.11 (m, 2H), 1.74 (s, 1H), 1.67 – 1.60 (m, 4H), 1.34 (s, 12H).

¹³C NMR (101 MHz, CDCl₃) δ 162.1 (t, J = 28.7 Hz), 138.6, 135.9, 119.1, 118.2 (t, J = 253.8 Hz), 83.9, 62.2, 33.4 (t, J = 23.1 Hz), 31.9, 24.9, 18.1 (t, J = 4.4 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -105.4 (2F, td, J = 17.3, 3.0 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₈H₂₆BF₂NO₄Na, 392.1815; found, 392.1818.



4-cyclohexyl-2,2-difluoro-*N***-phenylbutanamide** (15): Following the general procedure A, obtained in 84% yield as white solid after silica gel chromatography. (47.0 mg, eluent: petroleum ether/ethyl acetate = 30/1).

 $Mp = 70 - 72 \ ^{\circ}C$

¹H NMR (400 MHz, CDCl₃) δ 7.98 (s, 1H), 7.58 (d, *J* = 7.6 Hz, 2H), 7.37 (t, *J* = 7.9 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 2.35 – 2.05 (m, 2H), 1.68 (t, *J* = 17.5 Hz, 5H), 1.46 – 1.33 (m, 2H), 1.31 – 1.08 (m, 4H), 0.90 (dd, *J* = 21.8, 11.0 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 162.2 (t, J = 28.9 Hz), 136.1, 129.2, 125.5, 120.2, 118.7 (t, J = 253.4 Hz), 37.2, 33.0, 31.4 (t, J = 23.1 Hz), 28.8 (t, J = 3.8 Hz), 26.5, 26.2.

¹⁹F NMR (376 MHz, CDCl₃) δ -105.6 (2F, td, J = 17.4, 2.6 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₆H₂₁F₂NONa, 304.1483; found, 304.1490.



2,2-difluoro-*N***-phenyldecanamide (16):** Following the general procedure A, obtained in 83% yield as white solid after silica gel chromatography. (46.1 mg, eluent: petroleum ether/ethyl acetate = 30/1).

 $Mp = 45 - 47 \ ^{\circ}C$

¹H NMR (400 MHz, CDCl₃) δ 7.97 (s, 1H), 7.58 (d, *J* = 8.0 Hz, 2H), 7.37 (t, *J* = 7.9 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 2.28 – 2.04 (m, 2H), 1.57 – 1.46 (m, 2H), 1.39 – 1.21 (m, 10H), 0.87 (t, *J* = 6.7 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 162.1 (t, J = 28.5 Hz), 136.1, 129.2, 125.5, 120.2, 118.5 (t, J = 253.3 Hz), 33.8 (t, J = 23.1 Hz), 31.8, 29.2, 29.14, 29.07, 22.6, 21.6 (t, J = 4.3 Hz), 14.1.

¹⁹F NMR (376 MHz, CDCl₃) δ -105.5 (2F, td, J = 17.5, 3.0 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₆H₂₄F₂NO, 284.1820; found, 284.1829.



2,2-difluoro-3,4-dimethyl-*N***-phenylpentanamide (17):** Following the general procedure A, obtained in 85% yield as colorless oil after silica gel chromatography. (41.0 mg, eluent: petroleum ether/ethyl acetate = 30/1).

¹H NMR (400 MHz, CDCl₃) δ 8.07 (s, 1H), 7.58 (dd, J = 8.6, 1.0 Hz, 2H), 7.43 – 7.30 (m, 2H), 7.24 – 7.13 (m, 1H), 2.55 – 2.31 (m, 1H), 2.19 – 1.99 (m, 1H), 1.05 – 0.96 (m, 6H), 0.92 (d, J = 6.8 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 162.6 (t, J = 28.9 Hz), 136.1, 129.2, 125.5, 120.3, 120.2 (t, J = 257.2 Hz), 41.4 (t, J = 20.6 Hz), 26.3 (t, J = 2.5 Hz), 21.9, 17.4 (t, J = 1.6 Hz), 6.8 (dd, J = 5.6, 4.2 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -109.8 (1F, dd, *J* = 252.6, 19.4 Hz), -111.4 (1F, dd, *J* = 252.6, 16.9 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₃H₁₈F₂NO, 242.1351; found, 242.1357.



diethyl 2-(4,4-difluoro-5-oxo-5-(phenylamino)pentyl)malonate (18): Following the general procedure A, obtained in 80% yield as colorless oil after silica gel chromatography. (59.5 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 8.01 (s, 1H), 7.62 – 7.54 (m, 2H), 7.37 (t, J = 8.0 Hz, 2H), 7.19 (t, J = 7.4 Hz, 1H), 4.19 (qd, J = 7.1, 1.5 Hz, 4H), 3.34 (t, J = 7.4 Hz, 1H), 2.22 (ddd, J = 25.4, 17.2, 8.0 Hz, 2H), 1.98 (dd, J = 15.9, 7.6 Hz, 2H), 1.64 – 1.54 (m, 2H), 1.25 (t, J = 7.1 Hz, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 169.1, 161.9 (t, J = 28.6 Hz), 136.0, 129.2, 125.6, 120.2, 118.0 (t, J = 253.7 Hz), 61.5, 51.7, 33.3 (t, J = 23.4 Hz), 28.1, 19.5 (t, J = 4.4 Hz), 14.0.

¹⁹F NMR (376 MHz, CDCl3) δ -105.5 (2F, td, J = 17.3, 2.9 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₈H₂₃F₂NO₅Na, 394.1437; found, 394.1443.

ethyl 6,6-difluoro-7-oxo-7-(phenylamino)heptanoate (19): Following the general procedure A, obtained in 85% yield as colorless oil after silica gel chromatography. (52.6 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 8.02 (s, 1H), 7.58 (d, *J* = 8.2 Hz, 2H), 7.37 (t, *J* = 7.8 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 4.12 (q, *J* = 7.1 Hz, 2H), 2.33 (t, *J* = 7.3 Hz, 2H), 2.28 – 2.13 (m, 2H), 1.77 – 1.65 (m, 2H), 1.63 – 1.49 (m, 2H), 1.24 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 173.2, 162.0 (t, J = 28.8 Hz), 136.0, 129.2, 125.6, 120.2, 118.2 (t, J = 253.7 Hz), 60.4, 33.9, 33.5 (t, J = 23.3 Hz), 24.4, 21.2 (t, J = 4.4 Hz), 14.2.

¹⁹F NMR (376 MHz, CDCl₃) δ -105.5 (2F, td, *J* = 17.4, 3.1 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₅H₁₉F₂NO₃Na, 322.1225; found, 322.1231.



2-cyclooctyl-2,2-difluoro-*N***-phenylacetamide (20):** Following the general procedure A, obtained in 80% yield as white solid after silica gel chromatography. (47.0 mg, eluent: petroleum ether/ethyl acetate = 30/1).

 $Mp = 52 - 54 \ ^{o}C$

¹H NMR (400 MHz, CDCl₃) δ 8.03 (s, 1H), 7.58 (d, *J* = 8.0 Hz, 2H), 7.37 (t, *J* = 7.8 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 2.89 – 2.39 (m, 1H), 1.99 – 1.69 (m, 4H), 1.67 – 1.36 (m, 10H).

¹³C NMR (101 MHz, CDCl₃) δ 162.5 (t, J = 29.1 Hz), 136.1, 129.2, 125.5, 120.23 (t, J = 253.4 Hz), 120.21, 40.6 (t, J = 20.5 Hz), 26.5, 26.2, 25.5 (t, J = 3.7 Hz), 25.4. ¹⁹F NMR (376 MHz, CDCl₃) δ -112.21 (2F, dd, J = 17.5, 2.1 Hz). HRMS (ESI) m/z: [M+H]⁺ Calcd. for C₁₆H₂₂F₂NO, 282.1664; found, 282.1670.



2,2-difluoro-3-(4-isopropylcyclohex-1-en-1-yl)-*N*-phenylpropanamide (21):

Following the general procedure A, obtained in 87% yield as white solid after silica

gel chromatography. (53.5 mg, eluent: petroleum ether/ethyl acetate = 30/1). The compound data was in agreement with the literature (*Science* **2021**, *371*, 1232–1240). ¹H NMR (400 MHz, CDCl₃) δ 7.95 (s, 1H), 7.54 (d, *J* = 7.9 Hz, 2H), 7.36 (t, *J* = 7.9 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 5.67 (s, 1H), 2.82 (t, *J* = 17.5 Hz, 2H), 2.23 – 1.94 (m, 3H), 1.87 – 1.67 (m, 2H), 1.44 (dd, *J* = 13.1, 6.5 Hz, 1H), 1.35 – 1.08 (m, 2H), 0.85 (t, *J* = 6.5 Hz, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 162.2 (t, *J* = 28.6 Hz), 136.0, 129.2, 129.1, 128.1, 125.6, 120.3, 117.7 (t, *J* = 249.5 Hz), 41.7 (t, *J* = 23.2 Hz), 39.5, 32.1, 30.1, 29.2, 26.4, 19.9, 19.6.

¹⁹F NMR (376 MHz, CDCl₃) δ -103.9 (2F, td, J = 17.4, 2.5 Hz).



2,2-difluoro-*N***-phenyl-5-(trimethylsilyl)pentanamide (22):** Following the general procedure A, obtained in 85% yield as colorless oil after silica gel chromatography. (48.3 mg, eluent: petroleum ether/ethyl acetate = 30/1).

¹H NMR (400 MHz, CDCl₃) δ 8.21 (s, 1H), 7.73 (d, *J* = 7.8 Hz, 2H), 7.52 (t, *J* = 7.9 Hz, 2H), 7.34 (t, *J* = 7.4 Hz, 1H), 2.46 – 2.21 (m, 2H), 1.87 – 1.57 (m, 2H), 0.84 – 0.59 (m, 2H), 0.15 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 164.3 (t, J = 28.8 Hz), 138.1, 131.2, 127.6, 122.3, 120.2 (t, J = 253.4 Hz), 39.5 (t, J = 22.7 Hz), 18.5, 18.3 (t, J = 4.5 Hz), 0.3. ¹⁹F NMR (376 MHz, CDCl₃) δ -105.5 (2F, td, J = 17.5, 3.0 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₄H₂₂F₂NOSi, 286.1433; found, 286.1432.



2,2-difluoro-N-phenyl-5-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)pentanami

de (23): Following the general procedure A, obtained in 73% yield as colorless oil after silica gel chromatography. (49.5 mg, eluent: petroleum ether/ethyl acetate = 20/1).

¹H NMR (400 MHz, CDCl₃) δ 8.02 (s, 1H), 7.58 (d, *J* = 7.8 Hz, 2H), 7.37 (t, *J* = 7.2 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 2.37 – 2.10 (m, 2H), 1.64 (dt, *J* = 15.7, 7.8 Hz, 2H), 1.24 (s, 12H), 0.86 (t, *J* = 7.7 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 162.2 (t, J = 28.6 Hz), 136.1, 129.2, 125.5, 120.2, 118.3 (t, J = 253.3 Hz), 83.2, 36.0 (t, J = 22.9 Hz), 24.8, 16.3 (t, J = 4.6 Hz), 10.8. ¹⁹F NMR (376 MHz, CDCl₃) δ -105.3 (2F, td, J = 17.4, 3.0 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₇H₂₄BF₂NO₃Na, 362.1710; found, 362.1705.



tert-butyl (4,4-difluoro-5-oxo-5-(phenylamino)pentyl)carbamate (24): Following the general procedure A, obtained in 83% yield as colorless oil after silica gel chromatography. (54.5 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 8.11 (s, 1H), 7.58 (d, *J* = 7.8 Hz, 2H), 7.36 (t, *J* = 7.4 Hz, 2H), 7.19 (t, *J* = 7.3 Hz, 1H), 4.66 (s, 1H), 3.19 (d, *J* = 4.9 Hz, 2H), 2.50 – 2.02 (m, 2H), 1.84 – 1.67 (m, 2H), 1.43 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 161.9 (t, *J* = 28.7 Hz), 155.9, 136.0, 129.2, 125.6, 120.3, 118.1 (t, *J* = 253.7 Hz), 79.4, 39.8, 31.1 (t, *J* = 23.7 Hz), 28.4, 22.5.

¹⁹F NMR (376 MHz, CDCl₃) δ -105.3 (2F, t, *J* = 17.2 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₆H₂₂F₂N₂O₅Na, 351.1491; found, 351.1500.



4,4-difluoro-5-oxo-5-(phenylamino)pentyl acetate (25): Following the general procedure A, obtained in 72% yield as colorless oil after silica gel chromatography. (39.1 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 8.05 (s, 1H), 7.58 (d, *J* = 7.9 Hz, 2H), 7.37 (t, *J* = 7.9 Hz, 2H), 7.20 (t, *J* = 7.4 Hz, 1H), 4.13 (t, *J* = 6.4 Hz, 2H), 2.39 – 2.19 (m, 2H), 2.06 (s, 3H), 1.98 – 1.82 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 171.0, 161.8 (t, J = 28.7 Hz), 136.0, 129.3, 125.7, 120.2, 118.0 (t, J = 253.9 Hz), 63.2, 30.6 (t, J = 23.7 Hz), 21.1 (t, J = 4.4 Hz), 20.9. ¹⁹F NMR (376 MHz, CDCl₃) δ -105.7 (2F, td, J = 17.4, 2.6 Hz). HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₅F₂NO₃Na, 294.0912; found, 294.0919.



3,3-difluoro-4-oxo-4-(phenylamino)butyl acetate (26): Following the general procedure A, obtained in 69% yield as colorless oil after silica gel chromatography. (35.5 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 8.06 (s, 1H), 7.59 (d, J = 7.6 Hz, 2H), 7.37 (t, J = 8.0 Hz, 2H), 7.20 (t, J = 7.4 Hz, 1H), 4.33 (t, J = 6.3 Hz, 2H), 2.68 – 2.49 (m, 2H), 1.97 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 170.7, 161.5 (t, J = 28.1 Hz), 136.0, 129.3, 125.6, 120.2, 117.5 (t, J = 253.7 Hz), 57.5 (t, J = 5.9 Hz), 33.2 (t, J = 23.8 Hz), 20.7.

¹⁹F NMR (376 MHz, CDCl₃) δ -105.5 (2F, td, J = 17.5, 3.0 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₂H₁₃F₂NO₃Na, 280.0756; found, 280.0763.



4,4-difluoro-5-oxo-5-(phenylamino)pentan-2-yl acetate (27): Following the general procedure A, obtained in 58% yield as colorless oil after silica gel chromatography. (31.4 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 8.13 (s, 1H), 7.59 (d, *J* = 8.0 Hz, 2H), 7.36 (t, *J* = 7.9 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 5.27 (ddq, *J* = 12.6, 6.3, 3.1 Hz, 1H), 2.75 – 2.52 (m, 1H), 2.37 (dtd, *J* = 17.9, 14.7, 3.3 Hz, 1H), 1.89 (s, 3H), 1.32 (d, *J* = 6.4 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 170.5, 161.6 (t, *J* = 28.1 Hz), 136.1, 129.2, 125.6,

120.1, 116.7 (t, J = 243.9 Hz), 64.9 (t, J = 5.2 Hz), 39.7 (t, J = 23.3 Hz), 21.0, 20.8. ¹⁹F NMR (376 MHz, CDCl₃) δ -102.9 - -105.9 (2F, m).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₅F₂NO₃Na, 294.0912; found, 294.0913.

2,2-difluoro-*N*-**phenyl-4-(trimethylsilyl)butanamide (28):** Following the general procedure A, obtained in 90% yield as colorless oil after silica gel chromatography. (48.7 mg, eluent: petroleum ether/ethyl acetate = 30/1).

¹H NMR (400 MHz, CDCl₃) δ 8.03 (s, 1H), 7.59 (d, *J* = 7.9 Hz, 2H), 7.37 (t, *J* = 7.9 Hz, 2H), 7.19 (t, *J* = 7.4 Hz, 1H), 2.30 – 2.02 (m, 2H), 0.78 – 0.56 (m, 2H), 0.03 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 164.6 (t, J = 28.9 Hz), 138.4, 131.5, 127.8, 122.5, 121.2 (t, J = 253.7 Hz), 31.0 (t, J = 24.3 Hz), 10.0 (t, J = 2.7 Hz), 0.3.

¹⁹F NMR (376 MHz, CDCl₃) δ -107.1 (2F, td, J = 16.7, 2.9 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₉F₂NOSiNa, 294.1096; found, 294.1100.



4-ethoxy-2,2-difluoro-*N***-phenylbutanamide (29):** Following the general procedure A, obtained in 76% yield as colorless oil after silica gel chromatography. (36.9 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 8.06 (s, 1H), 7.57 (d, *J* = 8.1 Hz, 2H), 7.36 (t, *J* = 7.0 Hz, 2H), 7.18 (t, *J* = 7.2 Hz, 1H), 3.66 (t, *J* = 6.3 Hz, 2H), 3.46 (q, *J* = 7.0 Hz, 2H), 2.61 – 2.40 (m, 2H), 1.10 (t, *J* = 7.0 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 161.9 (t, J = 28.4 Hz), 136.2, 129.2, 125.4, 120.1, 161.8 (t, J = 253.7 Hz), 66.5, 63.6 (t, J = 6.0 Hz), 34.3 (t, J = 23.5 Hz), 14.9.

¹⁹F NMR (376 MHz, CDCl₃) δ -104.4 (2F, td, J = 16.1, 2.8 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₂H₁₅F₂NO₂Na, 266.0963; found, 266.0970.



2,2-difluoro-4-(*N*-methylacetamido)-*N*-phenylbutanamide (30): Following the general procedure A, obtained in 85% yield as colorless oil after silica gel chromatography. (45.9 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 8.96 (s, 1H), 7.66 (dd, *J* = 8.6, 1.0 Hz, 2H), 7.39 – 7.32 (m, 2H), 7.17 (t, *J* = 7.4 Hz, 1H), 3.65 (t, *J* = 6.5 Hz, 2H), 3.01 (s, 3H), 2.51 – 2.30 (m, 2H), 2.00 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.5, 161.8 (t, J = 28.5 Hz), 136.5, 129.1, 125.3, 120.4, 117.3 (t, J = 253.7 Hz), 41.2 (t, J = 5.6 Hz), 36.0, 31.4 (t, J = 23.7 Hz), 21.8. ¹⁹F NMR (376 MHz, CDCl₃) δ -104.7 (2F, td, J = 16.0, 1.9 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₆F₂N₂O₂Na, 293.1072; found, 293.1075.



4-(9H-carbazol-9-yl)-2,2-difluoro-*N***-phenylbutanamide (31):** Following the general procedure A, obtained in 73% yield as white solid after silica gel chromatography. (53.1 mg, eluent: petroleum ether/ethyl acetate = 10/1).

Mp = 155 - 157 °C

¹H NMR (400 MHz, CDCl₃) δ 8.07 (d, *J* = 7.7 Hz, 2H), 7.88 (s, 1H), 7.54 – 7.41 (m, 6H), 7.36 (t, *J* = 8.0 Hz, 2H), 7.26 – 7.17 (m, 3H), 4.69 – 4.60 (m, 2H), 2.91 – 2.61 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 161.2 (t, J = 27.9 Hz), 139.8, 135.7, 129.2, 125.9, 125.8, 123.2, 120.5, 120.3, 119.4, 117.2 (t, J = 254.8 Hz), 108.4, 36.2 (t, J = 5.8 Hz), 32.6 (t, J = 23.0 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -104.64 (2F, td, J = 17.0, 2.8 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₂₂H₁₉F₂N₂O, 365.1460; found, 365.1460.



1,1-diethyl 5-(4-methoxyphenethyl) 5,5-difluoropentane-1,1,5-tricarboxylate (32): Following the general procedure B, obtained in 86% yield as colorless oil after silica gel chromatography. (73.9 mg, eluent: petroleum ether/ethyl acetate = 20/1). ¹H NMR (400 MHz, CDCl₃) δ 7.14 (d, *J* = 8.5 Hz, 2H), 6.85 (d, *J* = 8.5 Hz, 2H), 4.42 (t, *J* = 7.0 Hz, 2H), 4.20 (q, *J* = 7.0 Hz, 4H), 3.79 (s, 3H), 3.29 (t, *J* = 7.5 Hz, 1H), 2.95 (t, *J* = 7.0 Hz, 2H), 2.12 – 1.96 (m, 2H), 1.91 (dd, *J* = 15.8, 7.7 Hz, 2H), 1.60 – 1.37 (m, 2H), 1.27 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 169.0, 164.0 (t, J = 33.0 Hz), 158.5, 129.9, 128.7, 115.9 (t, J = 250.4 Hz), 114.0, 67.2, 61.5, 55.2, 51.6, 34.1 (t, J = 23.4 Hz), 33.9, 28.0, 19.4 (t, J = 4.4 Hz), 14.0.

¹⁹F NMR (376 MHz, CDCl₃) δ -106.0 (2F, t, J = 16.7 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₂₁H₂₉F₂O₇, 431.1876; found, 431.1875.



ethyl 4-(dimethyl(phenyl)silyl)-2,2-difluorobutanoate (33): Following the general procedure B, obtained in 83% yield as colorless oil after silica gel chromatography. (47.5 mg, eluent: petroleum ether).

¹H NMR (400 MHz, CDCl₃) δ 7.46 – 7.36 (m, 2H), 7.35 – 7.22 (m, 3H), 4.22 (q, *J* = 7.1 Hz, 2H), 2.10 – 1.74 (m, 2H), 1.25 (t, *J* = 7.1 Hz, 3H), 0.94 – 0.70 (m, 2H), 0.23 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 164.4 (t, J = 33.2 Hz), 137.6, 133.5, 129.3, 127.9, 116.8 (t, J = 250.2 Hz), 62.7, 29.4 (t, J = 24.3 Hz), 13.9, 6.9 (t, J = 2.6 Hz), -3.4. ¹⁹F NMR (376 MHz, CDCl₃) δ -107.4 (2F, t, J = 16.1 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₄H₂₀F₂O₂Si, 309.1093; found, 309.1090.



ethyl 4-(benzyloxy)-2,2-difluorobutanoate (34): Following the general procedure B, obtained in 85% yield as colorless oil after silica gel chromatography. (43.9 mg, eluent: petroleum ether/ethyl acetate = 30/1).

¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.26 (m, 5H), 4.46 (s, 2H), 4.17 (q, *J* = 7.1 Hz, 2H), 3.65 (t, *J* = 6.1 Hz, 2H), 2.52 – 2.33 (m, 2H), 1.24 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 164.0 (t, J = 34.1 Hz), 137.7, 128.4, 127.8, 127.7, 115.3 (t, J = 249.9 Hz), 73.3, 63.4 (t, J = 6.3 Hz), 62.7, 35.3 (t, J = 23.6 Hz), 13.8. ¹⁹F NMR (376 MHz, CDCl₃) δ -105.7 (2F, t, J = 15.3 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₆F₂O₃Si, 281.0960; found, 281.0969.



ethyl 4-(9H-carbazol-9-yl)-2,2-difluorobutanoate (35): Following the general procedure B, obtained in 89% yield as colorless oil after silica gel chromatography. (56.4 mg, eluent: petroleum ether/ethyl acetate = 30/1).

¹H NMR (400 MHz, CDCl₃) δ 8.07 (d, *J* = 7.8 Hz, 2H), 7.47 (ddd, *J* = 8.2, 7.1, 1.2 Hz, 2H), 7.38 (d, *J* = 8.2 Hz, 2H), 7.31 – 7.20 (m, 2H), 4.65 – 4.39 (m, 2H), 4.12 (q, *J* = 7.1 Hz, 2H), 2.83 – 2.43 (m, 2H), 1.20 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 163.4 (t, J = 32.3 Hz), 139.8, 126.0, 123.2, 120.6, 119.5, 115.1 (t, J = 250.9 Hz), 108.4, 63.2, 35.9 (t, J = 5.9 Hz), 33.3 (t, J = 23.2 Hz), 13.8.

¹⁹F NMR (376 MHz, CDCl₃) δ -105.9 (2F, t, *J* = 16.6 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₈H₁₈F₂NO₂, 318.1300; found, 318.1313.

N-([1,1'-biphenyl]-4-yl)-2,2-difluoroacetamide (36): Following the general procedure C, obtained in 80% yield as white solid after silica gel chromatography. (39.4 mg, eluent: petroleum ether/ethyl acetate = 10/1).

 $Mp = 163 - 165 \ ^{\circ}C$

¹H NMR (400 MHz, d_6 -DMSO) δ 10.85 (s, 1H), 7.83 – 7.75 (m, 2H), 7.73 – 7.62 (m, 4H), 7.52 – 7.42 (m, 2H), 7.41 – 7.21 (m, 1H), (m, 42) (m, 42) (m, 44) (m, 44)

4H), 7.52 - 7.42 (m, 2H), 7.41 - 7.31 (m, 1H), 6.42 (t, J = 53.7 Hz, 1H).

¹³C NMR (101 MHz, *d*₆-DMSO) δ 165.8 (t, *J* = 25.9 Hz), 144.6, 141.9, 141.6, 134.2, 132.5, 132.3, 131.6, 125.8, 113.6 (t, *J* = 246.6 Hz).

¹⁹F NMR (376 MHz, d_6 -DMSO) δ -125.1 (2F, d, J = 53.6 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₄H₁₁F₂NONa, 270.0701; found, 270.0707.

2,2-difluoro-N-(4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)acetamide (37): Following the general procedure C, obtained in 75% yield as colorless oil after silica gel chromatography. (44.5 mg, eluent: petroleum ether/ethyl acetate = 10/1). ¹H NMR (400 MHz, CDCl₃) δ 8.00 (s, 1H), 7.82 (d, *J* = 8.5 Hz, 2H), 7.59 (d, *J* = 8.5 Hz, 2H), 6.01 (t, *J* = 54.4 Hz, 1H), 1.34 (s, 12H). ¹³C NMR (101 MHz, CDCl₃) δ 160.3 (t, *J* = 24.9 Hz), 138.2, 135.9, 119.2, 108.5 (t, *J*

= 254.3 Hz), 83.9, 24.8.

¹⁹F NMR (376 MHz, CDCl₃) δ -125.5 (2F, d, J = 54.3 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₄H₁₉BF₂NO₃, 298.1421; found, 298.1415.

2,2-difluoro-*N***-(4-methoxyphenyl)acetamide (38):** Following the general procedure C, obtained in 90% yield as white solid after silica gel chromatography. (36.2 mg, eluent: petroleum ether/ethyl acetate = 15/1). The compound data was in agreement with the literature (*Science* **2021**, *371*, 1232–1240).

¹H NMR (400 MHz, CDCl₃) δ 7.93 (s, 1H), 7.47 (d, *J* = 8.9 Hz, 2H), 6.89 (d, *J* = 9.0 Hz, 2H), 6.01 (t, *J* = 54.4 Hz, 1H), 3.80 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 160.2 (t, J = 24.4 Hz), 157.4, 128.6, 122.2, 114.4, 108.6 (t, J = 253.8 Hz), 55.5.

¹⁹F NMR (376 MHz, CDCl₃) δ -125.5 (2F, dd, *J* = 54.4, 2.0 Hz).

2,2-difluoro-*N***-(o-tolyl)acetamide (39):** Following the general procedure C, obtained in 83% yield as white solid after silica gel chromatography. (30.7 mg, eluent: petroleum ether/ethyl acetate = 15/1).

$$Mp = 64 - 66 \ ^{\circ}C$$

¹H NMR (400 MHz, CDCl₃) δ 7.83 (d, *J* = 7.9 Hz, 1H), 7.77 (s, 1H), 7.25 – 7.20 (m, 2H), 7.16 (t, *J* = 7.4 Hz, 1H), 6.04 (t, *J* = 54.4 Hz, 1H), 2.29 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 160.4 (t, J = 24.2 Hz), 133.4, 130.8, 129.5, 127.1, 126.5, 122.9, 108.8 (t, J = 254.0 Hz), 17.4.

¹⁹F NMR (376 MHz, CDCl₃) δ -125.4 (2F, dd, J = 54.4, 2.5 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₉H₁₀F₂NO, 186.0725; found, 186.0721.

ethyl 4-acetamidobenzoate (40): Following the general procedure C, obtained in 96% yield as white solid after silica gel chromatography. (39.8 mg, eluent: petroleum ether/ethyl acetate = 1/1). The compound data was in agreement with the literature (*Angew. Chem., Int. Ed.* 2016, 55, 3823–3827).

¹H NMR (400 MHz, CDCl₃) δ 7.99 (d, *J* = 8.5 Hz, 2H), 7.94 (s, 1H), 7.61 (d, *J* = 8.3 Hz, 2H), 4.36 (q, *J* = 7.1 Hz, 2H), 2.20 (s, 3H), 1.38 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 168.8, 166.3, 142.2, 130.7, 125.8, 118.8, 60.9, 24.7, 14.3.

2,3,3,3-tetrafluoro-*N***-phenylpropanamide (41):** Following the general procedure C, obtained in 90% yield as white solid after silica gel chromatography. (39.8 mg, eluent: petroleum ether/ethyl acetate = 15/1). The compound data was in agreement with the literature (*Science* **2021**, *371*, 1232–1240).

¹H NMR (400 MHz, CDCl₃) δ 7.96 (s, 1H), 7.57 (dd, *J* = 8.5, 1.0 Hz, 2H), 7.39 (dd, *J* = 10.8, 5.1 Hz, 2H), 7.22 (t, *J* = 7.4 Hz, 1H), 5.21 (dq, *J* = 46.5, 6.4 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 158.9 (d, *J* = 18.2 Hz), 135.7, 129.3, 125.9, 120.5 (qd, *J* = 280.4, 25.7 Hz), 120.3, 85.7 (dq, *J* = 205.4, 34.0 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -75.7 (3F, d, J = 11.0 Hz), -200.1 (1F, q, J = 11.0 Hz).



2-fluoro-6-hydroxy-*N***-phenyl-2-(trifluoromethyl)hexanamide (43):** Following the general procedure C, obtained in 80% yield as white solid after silica gel chromatography. (46.9 mg, eluent: petroleum ether/ethyl acetate = 1/1). Mp = 65– 67 °C ¹H NMR (400 MHz, CDCl₃) δ 8.16 (s, 1H), 7.57 (dd, *J* = 8.5, 0.9 Hz, 2H), 7.37 (t, *J* = 8.0 Hz, 2H), 7.20 (t, *J* = 7.4 Hz, 1H), 3.65 (td, *J* = 6.1, 1.6 Hz, 2H), 2.53 – 2.27 (m, 1H), 2.16 – 2.00 (m, 1H), 1.70 (s, 1H), 1.68 – 1.57 (m, 3H), 1.56 – 1.41 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 162.1 (d, *J* = 19.5 Hz), 136.0, 129.2, 125.7, 121.8 (qd, *J* = 285.1, 28.5 Hz), 120.5, 95.9 (dq, *J* = 202.7, 30.3 Hz), 61.8, 31.9, 30.12 (d, *J* = 20.2 Hz), 18.5 (d, *J* = 2.5 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -78.3 (3F, d, *J* = 6.7 Hz), -169.8 – -183.4 (1F, m). HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₃H₁₆F₄NO₂, 294.1112; found, 294.1116.



5,5-difluoro-5-(2-(trifluoromethyl)phenyl)pentan-1-ol (44): Following the general procedure D, obtained in 81% yield as colorless oil after silica gel chromatography. (43.4 mg, eluent: petroleum ether/ethyl acetate = 2/1). The compound data was in agreement with the literature (*J. Am. Chem. Soc.* **2018**, *140*, 163–166).

¹H NMR (400 MHz, CDCl₃) δ 7.78 (d, *J* = 7.9 Hz, 1H), 7.67 – 7.56 (m, 2H), 7.52 (t, *J* = 7.3 Hz, 1H), 3.63 (t, *J* = 6.1 Hz, 2H), 2.40 – 2.10 (m, 2H), 1.96 (s, 1H), 1.70 – 1.55 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 136.1 (t, *J* = 28.0 Hz), 131.8, 129.8, 128.1 (t, *J* = 9.1 Hz), 127.5 (q, *J* = 6.5 Hz), 127.1 (q, *J* = 32.4 Hz), 123.6 (q, J = 273.5 Hz), 122.5 (t, J = 244.8 Hz), 62.5, 39.3 (t, *J* = 26.9 Hz), 32.1, 18.7 (t, *J* = 3.6 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -57.7 (3F, t, *J* = 16.1 Hz), -93.3 (2F, h, *J* = 16.5 Hz).



5,5-difluoro-5-(3-(trifluoromethyl)phenyl)pentan-1-ol (45): Following the general procedure D, obtained in 80% yield as colorless oil after silica gel chromatography.

(42.9 mg, eluent: petroleum ether/ethyl acetate = 2/1). The compound data was in agreement with the literature (*J. Am. Chem. Soc.* **2018**, *140*, 163–166). ¹H NMR (400 MHz, CDCl₃) δ 7.63 (s, 1H), 7.60 – 7.51 (m, 2H), 7.44 (t, *J* = 7.8 Hz, 1H), 3.49 (t, *J* = 6.1 Hz, 2H), 2.37 – 1.74 (m, 2H), 1.72 – 1.19 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 138.4 (t, *J* = 27.5 Hz), 130.9 (q, *J* = 32.7 Hz), 129.1, 128.4 (t, *J* = 5.6 Hz), 126.7 – 126.4 (m), 123.8 (q, *J* = 272.3 Hz), 122.3 (t, *J* = 242.8 Hz), 122.1 – 121.7 (m), 62.1, 38.7 (t, *J* = 27.2 Hz), 31.9, 18.8 (t, *J* = 4.1 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -62.8 (3F, s), -96.2 (2F, t, *J* = 16.4 Hz).



5,5-difluoro-5-(4-(trifluoromethyl)phenyl)pentan-1-ol (46): Following the general procedure D, obtained in 78% yield as colorless oil after silica gel chromatography. (41.8 mg, eluent: petroleum ether/ethyl acetate = 2/1). The compound data was in agreement with the literature (*J. Am. Chem. Soc.* **2018**, *140*, 163–166).

¹H NMR (400 MHz, CDCl₃) δ 7.68 (d, J = 8.1 Hz, 2H), 7.59 (d, J = 8.1 Hz, 2H), 3.60 (t, J = 6.1 Hz, 2H), 2.30 – 2.00 (m, 2H), 1.80 – 1.38 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 140.9 (t, J = 27.4 Hz), 131.8 (q, J = 32.7 Hz), 125.7 – 125.4 (m, 2xC), 123.8 (q, J = 272.3 Hz), 122.4 (t, J = 242.9 Hz), 62.2, 38.7 (t, J = 27.1 Hz), 31.9, 18.9 (t, J = 4.1 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -62.9 (3F, s), -96.5 (2F, t, J = 16.3 Hz).

5-(3-amino-5-(trifluoromethyl)phenyl)-5,5-difluoropentan-1-ol (47): Following the general procedure D, obtained in 88% yield as yellow oil after silica gel chromatography. (49.8 mg, eluent: petroleum ether/ethyl acetate = 2/1). ¹H NMR (400 MHz, CDCl₃) δ 7.05 (s, 1H), 6.92 (s, 1H), 6.90 (s, 1H), 3.62 (t, *J* = 6.2 Hz, 2H), 3.12 (s, 2H), 2.20 – 2.02 (m, 2H), 1.74 – 1.40 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 146.9, 139.5 (t, J = 27.1 Hz), 132.0 (q, J = 32.4 Hz), 123.8 (q, J = 272.5 Hz), 122.4 (t, J = 243.0 Hz), 114.4 (t, J = 5.9 Hz), 112.6 – 112.3 (m), 111.8 – 111.3 (m), 62.4, 38.6 (t, J = 27.3 Hz), 32.1, 18.9 (t, J = 4.1 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -62.9 (3F, s), -96.1 (2F, t, J = 16.3 Hz). HRMS (ESI) m/z: [M+Na]⁺ Calcd. for C₁₂H₁₄F₅NONa, 306.0888; found, 306.0894.



5,5-difluoro-5-(3-(hydroxymethyl)-5-(trifluoromethyl)phenyl)pentan-1-ol (48): Following the general procedure D, obtained in 85% yield as colorless oil after silica gel chromatography. (50.6 mg, eluent: petroleum ether/ethyl acetate = 2/1). ¹H NMR (400 MHz, CDCl₃) δ 7.67 (s, 1H), 7.64 (d, J = 7.0 Hz, 2H), 4.77 (s, 2H), 3.60 (t, J = 5.9 Hz, 2H), 2.26 (s, 1H), 2.21 – 2.07 (m, 2H), 1.62 – 1.40 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ 142.6, 138.6 (t, J = 27.5 Hz), 131.3 (q, J = 32.7 Hz), 126.6 (t, J = 5.6 Hz), 124.9 – 124.5 (m), 123.7 (q, J = 272.5 Hz), 122.4 (t, J = 243.0Hz), 121.3 – 120.7 (m), 63.9, 62.3, 38.6 (t, J = 27.1 Hz), 31.9, 18.9 (t, J = 4.1 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -62.7 (3F, s), -95.7 (2F, t, J = 16.3 Hz). HRMS (ESI) m/z: [M+Na]⁺ Calcd. for C₁₃H₁₅F₅O₂Na, 321.0884; found, 321.0893.



5,5-difluoro-5-(3-methoxy-5-(trifluoromethyl)phenyl)pentan-1-ol (49): Following the general procedure D, obtained in 73% yield as colorless oil after silica gel chromatography. (43.5 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 7.30 (s, 1H), 7.17 (s, 2H), 3.88 (s, 3H), 3.64 (t, *J* = 6.1 Hz, 2H), 2.20 – 2.02 (m, 2H), 1.77 – 1.11 (m, 4H).

¹³C NMR (101 MHz, CDCl₃) δ 159.9, 139.9 (t, J = 27.5 Hz), 132.3 (q, J = 32.7 Hz), 123.5 (q, J = 272.6 Hz), 122.1 (t, J = 243.3 Hz), 114.4 (t, J = 5.8 Hz), 114.3 – 113.5 (m), 112.1–111.7 (m), 62.4, 55.7, 38.7 (t, J = 27.2 Hz), 32.1, 18.9 (t, J = 4.1 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -62.8 (3F, s), -96.1 (2F, t, J = 16.4 Hz). HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₅F₅O₂Na, 321.0884; found, 321.0880.



5,5-difluoro-5-(3-methoxyphenyl)pentan-1-ol (50): Following the general procedure D, obtained in 50% yield as colorless oil after silica gel chromatography. (23.0 mg, eluent: petroleum ether/ethyl acetate = 5/1).

¹H NMR (400 MHz, CDCl₃) δ 7.33 (t, J = 7.9 Hz, 1H), 7.04 (d, J = 7.6 Hz, 1H), 6.99 (s, 1H), 6.95 (d, J = 8.2 Hz, 1H), 3.83 (s, 3H), 3.63 (t, J = 6.2 Hz, 2H), 2.45 – 1.98 (m, 2H), 1.65 – 1.56 (m, 2H), 1.55 – 1.47 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 159.6, 138.8 (t, J = 26.6 Hz), 129.6, 122.8 (t, J = 242.5 Hz), 117.2 (t, J = 6.2 Hz), 115.1, 110.7 (t, J = 6.6 Hz), 62.5, 55.4, 38.8 (t, J = 27.6 Hz), 32.2, 19.0 (t, J = 4.2 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -95.4 (2F, t, J = 16.1 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₂H₁₆F₂O₂Na, 253.1011; found, 253.1009.



5,5-difluoro-5-(3-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)phenyl)pentan-1-ol

(51): Following the general procedure D, obtained in 78% yield as colorless oil after silica gel chromatography. (50.9 mg, eluent: petroleum ether/ethyl acetate = 2/1). ¹H NMR (400 MHz, CDCl₃) δ 7.88 (s, 1H), 7.84 (d, *J* = 7.3 Hz, 1H), 7.55 (d, *J* = 7.7 Hz, 1H), 7.41 (t, *J* = 7.5 Hz, 1H), 3.60 (t, *J* = 6.1 Hz, 2H), 2.32 – 2.04 (m, 2H), 1.69 – 1.41 (m, 4H), 1.34 (s, 12H).

¹³C NMR (101 MHz, CDCl₃) δ 136.7 (t, J = 26.5 Hz), 136.0, 131.1 (t, J = 6.0 Hz), 127.8, 127.7 (t, J = 6.0 Hz), 123.1 (t, J = 242.3 Hz), 84.1, 62.4, 38.9 (t, J = 27.5 Hz), 32.2, 24.9, 18.9 (t, J = 4.1 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -95.6 (2F, t, J = 16.3 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₇H₂₅BF₂O₃Na, 349.1757; found, 349.1787.



5,5-difluoro-5-(2-methoxypyridin-3-yl)pentan-1-ol (52): Following the general procedure D, obtained in 90% yield as colorless oil after silica gel chromatography. (41.6 mg, eluent: petroleum ether/ethyl acetate = 2/1).

¹H NMR (400 MHz, CDCl₃) δ 8.22 (d, J = 4.5 Hz, 1H), 7.78 (d, J = 7.4 Hz, 1H), 6.94 (dd, J = 7.4, 5.0 Hz, 1H), 4.00 (s, 3H), 3.63 (t, J = 6.4 Hz, 2H), 2.51 – 2.11 (m, 2H), 1.66 – 1.55 (m, 2H), 1.51 – 1.37 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 160.5 (t, J = 4.4 Hz), 148.3, 135.8 (t, J = 8.3 Hz), 121.8 (t, J = 227.8 Hz), 119.4 (t, J = 27.4 Hz), 116.4, 62.5, 53.7, 36.1 (t, J = 26.1 Hz), 32.2, 19.0 (t, J = 4.2 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -96.3 (2F, t, *J* = 16.9 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₁H₁₆F₂NO₂, 232.1144; found, 232.1147.



5-(6-aminopyridin-2-yl)-5,5-difluoropentan-1-ol (53): Following the general procedure D, obtained in 57% yield as colorless oil after silica gel chromatography. (24.6 mg, eluent: petroleum ether/ethyl acetate = 2/1).

¹H NMR (400 MHz, CDCl₃) δ 7.50 (t, *J* = 7.8 Hz, 1H), 6.93 (d, *J* = 7.4 Hz, 1H), 6.53 (d, *J* = 8.3 Hz, 1H), 4.71 (s, 2H), 3.64 (t, *J* = 6.2 Hz, 2H), 2.26 (ddd, *J* = 24.3, 16.4, 7.9 Hz, 2H), 1.60 (dt, *J* = 12.9, 6.6 Hz, 2H), 1.56 – 1.45 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 158.2, 152.9 (t, J = 28.9 Hz), 138.6, 121.6 (t, J = 241.7 Hz), 109.9, 109.8 (t, J = 5.3 Hz), 62.3, 35.9 (t, J = 25.7 Hz), 32.0, 18.6 (t, J = 4.3 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ -99.7 (2F, t, *J* = 16.6 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₀H₁₅F₂N₂O, 217.1147; found, 217.1146.



3-(1,1-difluoro-3-(trimethylsilyl)propyl)-5-(trifluoromethyl)aniline (54):

Following the general procedure D, obtained in 92% yield as yellow oil after silica gel chromatography. (57.3 mg, eluent: petroleum ether/ethyl acetate = 20/1).

¹H NMR (400 MHz, CDCl₃) δ 7.21 (s, 1H), 7.07 (s, 1H), 7.05 (s, 1H), 3.96 (s, 2H), 2.31 – 1.94 (m, 2H), 0.87 – 0.57 (m, 2H), 0.15 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 149.2, 141.9 (t, J = 27.4 Hz), 134.3 (q, J = 32.3 Hz), 126.1 (q, J = 272.4 Hz), 125.2 (t, J = 243.1 Hz), 116.9 (t, J = 5.9 Hz), 114.7 – 114.5 (m), 114.4 – 113.6 (m), 36.0 (t, J = 28.5 Hz), 10.9 (t, J = 2.6 Hz), 0.2. ¹⁹F NMR (376 MHz, CDCl₃) δ -62.9 (3F, s), -97.9 (2F, t, J = 15.9 Hz). HRMS (ESI) m/z: [M+H]⁺ Calcd. for C₁₃H₁₉F₅NSi, 312.1201; found, 312.1210.



3-(4,4-diethoxy-1,1-difluorobutyl)-5-(trifluoromethyl)aniline (55): Following the general procedure D, obtained in 76% yield as yellow oil after silica gel chromatography. (51.8 mg, eluent: petroleum ether/ethyl acetate = 9/1).

¹H NMR (400 MHz, CDCl₃) δ 7.06 (s, 1H), 6.92 (s, 1H), 6.90 (s, 1H), 4.49 (t, *J* = 5.6 Hz, 1H), 3.62 (dq, *J* = 9.4, 7.1 Hz, 2H), 3.47 (dq, *J* = 9.4, 7.1 Hz, 2H), 2.33 – 1.98 (m, 2H), 1.91 – 1.71 (m, 2H), 1.19 (t, *J* = 7.1 Hz, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 147.0, 139.4 (t, *J* = 27.1 Hz), 132.1 (q, *J* = 32.4 Hz), 123.8 (q, *J* = 272.4 Hz), 122.3 (t, *J* = 242.8 Hz), 114.3 (t, *J* = 6.0 Hz), 112.5 – 112.2 (m), 111.7 – 111.2 (m), 101.8, 61.5, 34.0 (t, *J* = 27.7 Hz), 26.8 (t, *J* = 3.7 Hz), 15.3. ¹⁹F NMR (376 MHz, CDCl₃) δ -63.0 (3F, s), -96.4 (2F, t, *J* = 16.6 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₅H₂₀F₅NO₂Na, 364.1306; found, 364.1313.



4-(3-(3-amino-5-(trifluoromethyl)phenyl)-3,3-difluoropropyl)-1,3-dioxolan-2-one (56): Following the general procedure D, obtained in 72% yield as yellow oil after silica gel chromatography. (46.8 mg, eluent: petroleum ether/ethyl acetate = 2/1). ¹H NMR (400 MHz, CDCl₃) δ 7.05 (s, 1H), 6.95 (s, 1H), 6.90 (s, 1H), 4.90 – 4.70 (m, 1H), 4.57 (t, *J* = 8.2 Hz, 1H), 4.25 – 3.97 (m, 1H), 2.48 – 2.11 (m, 2H), 1.96 (dd, *J* = 14.9, 7.3 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 154.6, 147.3, 138.7 (t, J = 26.8 Hz), 132.3 (q, J = 32.6 Hz), 123.7 (q, J = 272.2 Hz), 121.5 (t, J = 243.5 Hz), 114.0 (t, J = 5.9 Hz), 112.9 – 112.7 (m), 111.9 – 109.5 (m), 75.9, 69.2, 34.4 (t, J = 28.2 Hz), 27.3 (t, J = 3.6 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -63.1 (3F, s), -96.7 (1F, ddd, J = 245.5, 21.0, 11.7 Hz), -97.8 (1F, ddd, J = 245.5, 20.5, 12.6 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₃H₁₃F₅NO₃, 326.0810; found, 326.0815.



ethyl6-(3-amino-5-(trifluoromethyl)phenyl)-6,6-difluorohexanoate(57):Following the general procedure D, obtained in 84% yield as yellow oil after silica gelchromatography. (57.0 mg, eluent: petroleum ether/ethyl acetate = 9/1).

¹H NMR (400 MHz, CDCl₃) δ 7.04 (s, 1H), 6.92 (s, 1H), 6.90 (s, 1H), 4.11 (q, *J* = 7.1 Hz, 2H), 3.63 (s, 2H), 2.30 (t, *J* = 7.4 Hz, 2H), 2.21 – 1.98 (m, 2H), 1.66 (dt, *J* = 15.2, 7.5 Hz, 2H), 1.53 – 1.40 (m, 2H), 1.24 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 173.4, 146.9, 139.5 (t, *J* = 27.2 Hz), 132.1 (q, *J* = 32.4 Hz), 123.8 (q, *J* = 272.5 Hz), 122.2 (t, *J* = 243.0 Hz), 114.4 (t, *J* = 5.9 Hz), 112.8 – 112.2 (m), 111.9 – 111.2 (m), 60.4, 38.6 (t, *J* = 27.4 Hz), 34.0, 24.5, 21.9 (t, *J* = 4.0 Hz), 14.2.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.0 (3F, s), -96.2 (2F, t, J = 16.3 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₅H₁₈F₅NO₂Na, 362.1150; found, 362.1150.



1-(3-(3-amino-5-(trifluoromethyl)phenyl)-3,3-difluoropropyl)pyrrolidin-2-one

(58): Following the general procedure D, obtained in 88% yield as yellow oil after silica gel chromatography. (56.7 mg, eluent: petroleum ether/ethyl acetate = 2/1). ¹H NMR (400 MHz, CDCl₃) δ 7.05 (s, 1H), 6.93 (s, 2H), 3.53 – 3.44 (m, 2H), 3.34 (t, *J* = 7.0 Hz, 2H), 2.46 – 2.34 (m, 2H), 2.34 – 2.28 (m, 2H), 1.95 (dt, *J* = 15.4, 7.5 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 175.2, 147.1, 138.7 (t, J = 26.7 Hz), 132.1 (q, J = 32.5 Hz), 123.7 (q, J = 272.6 Hz), 121.3 (t, J = 243.5 Hz), 114.5 (t, J = 6.1 Hz), 112.8 – 112.6 (m), 111.4 – 110.9 (m), 47.3, 36.8 (t, J = 5.0 Hz), 36.1 (t, J = 27.4 Hz), 30.9, 17.9.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.0 (3F, s), -96.1 (2F, t, *J* = 16.3 Hz). HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₁₄H₁₆F₅N₂O, 323.1177; found, 323.1183.



3-(1,1-difluoro-3-((tetrahydro-2H-pyran-2-yl)oxy)propyl)-5-(trifluoromethyl)anil ine (59): Following the general procedure D, obtained in 82% yield as yellow oil after silica gel chromatography. (55.6 mg, eluent: petroleum ether/ethyl acetate = 9/1). ¹H NMR (400 MHz, CDCl₃) δ 7.09 (s, 1H), 6.92 (s, 2H), 4.52 (s, 1H), 3.88 (dt, *J* = 10.2, 6.9 Hz, 1H), 3.77 (ddd, *J* = 11.1, 7.9, 3.0 Hz, 1H), 3.59 – 3.37 (m, 2H), 2.56 – 2.35 (m, 2H), 1.77 – 1.39 (m, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 147.0, 139.2 (t, *J* = 26.8 Hz), 132.0 (q, *J* = 32.4 Hz), 123.8 (q, *J* = 272.5 Hz), 121.5 (t, *J* = 243.1 Hz), 114.3 (t, *J* = 6.3 Hz), 112.5 – 112.3 (m), 112.1 – 111.3 (m), 98.9, 62.2, 61.3 (t, *J* = 5.2 Hz), 39.0 (t, *J* = 27.1 Hz), 30.5, 25.3, 19.3.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.0 (3F, s), -94.5 (2F, dt, J = 39.8, 16.1 Hz). HRMS (ESI) m/z: [M+H]⁺ Calcd. for C₁₅H₁₉F₅NO₂, 340.1330; found, 340.1325.



2-(3-(dimethyl(phenyl)silyl)-1,1-difluoropropyl)-10-(3-(4-methylpiperazin-1-yl)pr opyl)-10H-phenothiazine (60): Following the general procedure D, obtained in 70% yield as colorless oil after silica gel chromatography. (77.2 mg, eluent: dichloromethane /methanol = 10/1). ¹H NMR (400 MHz, CDCl₃) δ 7.50 – 7.41 (m, 2H), 7.33 (dd, J = 5.6, 1.4 Hz, 3H), 7.19 – 7.07 (m, 3H), 6.91 (t, J = 8.2 Hz, 4H), 3.91 (t, J = 6.8 Hz, 2H), 2.47 – 2.42 (m, 10H), 2.25 (s, 3H), 2.11 – 1.99 (m, 2H), 1.96 – 1.81 (m, 2H), 0.92 – 0.71 (m, 2H), 0.26 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 145.4, 144.8, 138.0, 136.5 (t, J = 26.9 Hz), 133.5, 129.2, 127.9, 127.5, 127.4, 127.2, 127.0, 124.6, 123.5 (t, J = 243.1 Hz), 122.8, 119.2 (t, J = 6.0 Hz), 115.8, 112.2 (t, J = 6.4 Hz), 55.5, 55.1, 53.2, 46.0, 45.3, 33.8 (t, J = 29.2 Hz), 24.3, 8.1 (t, J = 2.0 Hz), -3.3.

¹⁹F NMR (376 MHz, CDCl₃) δ -96.5 (2F, t, J = 15.6 Hz).

HRMS (ESI) *m/z*: [M+H]⁺ Calcd. for C₃₁H₄₀F₂N₃SSi, 552.2675; found, 552.2668.



2-(3-(difluoromethyl)phenyl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (61):

Following the general procedure D, obtained in 70% yield as colorless oil after silica gel chromatography. (35.5 mg, eluent: petroleum ether/ethyl acetate = 15/1).

¹H NMR (400 MHz, CDCl₃) δ 8.02 – 7.86 (m, 2H), 7.61 (d, J = 7.7 Hz, 1H), 7.46 (t, J

= 7.6 Hz, 1H), 6.64 (t, *J* = 56.4 Hz, 1H), 1.35 (s, 12H).

¹³C NMR (101 MHz, CDCl₃) δ 137.1 (t, J = 1.8 Hz), 133.7 (t, J = 22.3 Hz), 131.9 (t, J

= 6.1 Hz), 128.2 (t, *J* = 6.0 Hz), 128.1, 114.9 (t, *J* = 238.6 Hz), 84.2, 24.9.

¹⁹F NMR (376 MHz, CDCl₃) δ -110.3 (2F, d, J = 56.4 Hz).

HRMS (ESI) *m/z*: [M+Na]⁺ Calcd. for C₁₃H₁₇BF₂O₂Na, 277.1182; found, 277.1184.

2-3 Experimental Studies on Mechanism

2-3-1 Measurement of quantum yield

According to the procedure of Glorius⁹ and Melchiorre¹⁰, the photon flux of the LEDs ($\lambda_{max} = 427 \text{ nm}$) was determined by standard ferrioxalate actinometry. A 0.15 M solution of ferrioxalate was prepared by dissolving potassium ferrioxalate hydrate (0.737 g) in 10 mL H₂SO₄ (0.05 M). A buffered solution of 1,10-phenanthroline was prepared by dissolving sodium acetate (5.63 g) and phenanthroline (25 mg) in 25 mL H₂SO₄ (0.5 M) at the same time. Both solutions were stored in the dark. To determine the photon flux of the LED, the ferrioxalate solution (1.0 mL) was placed in a cuvette and irradiated for 900 seconds at $\lambda_{max} = 427$ nm. After irradiation, the phenanthroline solution (0.175 mL) was added to the cuvette and the mixture was stirred in the dark for 1 hour to allow the ferrous ions to completely coordinate to the phenanthroline. The absorbance of the solution was measured at 510 nm. A non-irradiated sample was also prepared and the absorbance at 510 nm was measured. Using then the Beer's Law, the number of moles of Fe²⁺ produced by light irradiation is obtained by:

$$Fe^{2+} = \frac{v_1 v_3 \Delta A(510 nm)}{10^3 v_2 l \varepsilon (510 nm)}$$

where v_1 is the irradiated volume (1 mL), v_2 is the aliquot of the irradiated solution taken for the determination of the ferrous ions (1 mL), v_3 is the final volume after complexation with phenanthroline (10 mL), 1 is the optical path-length of the irradiation cell (1 cm), $\Delta A(510 \text{ nm})$ the optical difference in absorbance between the irradiated solution and the one stored in the dark, $\epsilon(510 \text{ nm})$ is that of the complex Fe(phen)₃²⁺(11100 L mol⁻¹cm⁻¹).

The photon flux (F) is obtained by using the following equation:

$$\phi(\lambda) = \frac{mol(Fe^{2+})}{F(1-10^{-A(\lambda)})}$$

Wher Φ (λ) = The quantum yield for Fe²⁺ formation at 420 nm is 1.1. A(λ) = ferrioxalate actinometer absorbance at 420 nm, which was measured placing 1 mL of
the solution in a cuvette of pathlength 1 cm by UV/Vis spectrophotometry. We obtained an absorbance value of 3.5. The photon flux (**F**) is 3.1×10^{-9} einsteins/s.



To obtain the quantum yield (Φ). The number of moles of the product **3** were determined by ¹H-NMR analysis using diphenylmethane as internal standard. As such, a photocatalytic reaction was performed under the set of optimized reaction conditions under visible light irradiation of 427 nm blue LEDs. After 900 s of light irradiation, 2.0 x 10⁻⁶ moles of **3** were obtained. The quantum yield of this reaction was calculated using the following equation:

$$\phi(427 nm) = \frac{moles of product}{F(1-10^{-A(427 nm)})t}$$

Where A(427 nm) = is the absorbance at 427 nm of the photocatalytic reaction whichwas measured placing 1 mL of the solution in a cuvette of path length 1 cm by UV/Vis spectrophotometry.

t = is the reaction time.

The quantum yield (Φ) of the reaction is 4.4.

2-3-2 UV-vis absorption spectroscopic measurements



Stock solutions of 1, 2, PO1, AdSH, HCO_2K were prepared with the same concentration used in the reaction. Cs_2CO_3 was used in 150 mol% to ensure generation of PO anion under measurement condition. The solutions were prepared in the presence of air using DMA as solvent.



Supplementary Figure 2. UV/vis absorption spectra of catalyst, substrate, and reaction mixture.



Supplementary Figure 3. UV/vis absorption spectra of 4-tert-Butylphenol anion and PO1 anion.

2-3-3 Stern-Volmer Quenching Study

To evaluate the role of **PO1** anion in this process, we conducted Stern-Volmer fluorescence quenching experiments.¹¹ The samples were prepared mixing the **PO1** anion (5×10^{-4} M, freshly prepared in situ by the deprotonation of **PO1** with Cs₂CO₃) with the required amount of **1** in a total volume of 1 mL of dry DMA in a 10 × 10 mm light path quartz fluorescence cuvette under an argon atmosphere. The excitation

wavelength was fixed at 384 nm, the emission light was acquired from 400 nm to 700 nm. Quenching was not observed in similar experiments using **2** and 1-adamantanethiol.



Supplementary Figure 4. Quenching of the PO1 anion emission (5 \times 10⁻⁴ M in DMA) in the presence of increasing amounts of 1.



Supplementary Figure 5. Stern-Volmer quenching plot of substrate I.

Other reaction components:



Supplementary Figure 6. Stern-Volmer quenching plot of 2.



Supplementary Figure 7. Stern-Volmer quenching plot of 1-adamantanethiol.

2-3-4 Electrochemical Measurements

2-3-4-1 Cyclic Voltammogram of Catalyst

Tetrabutylammonium hexafluorophosphate (1161 mg, 3.0 mmol) was added to a 0.01 M solution of the **PO1** anion catalyst (generated in situ by the deprotonation of the **PO1** catalyst with 1.3 equiv Cs_2CO_3) in 30 mL of dry DMSO and the solution was vigorously bubbled with Ar for 5 minutes prior to the measurement. The oxidation/reduction potential was measured using platinum disc working electrode, a

platinum wire counter electrode, and a saturated calomel electrode (SCE) at 0.1 V/s scan rate.



Supplementary Figure 8. The cyclic voltammogram of the **PO1** anions vs SCE in DMSO at 0.1V/s.

With this data in hand we calculated the redox potential of the excited **PO1** anion employing the following equation:¹²

$$E_{p/2}(PO1^{-}/PO1^{-*}) = E_{p/2}(PO1^{-}/PO1^{-}) - E_{0-0}(PO1^{-*}/PO1^{-})$$

 $E_{p/2}(PO1^{-}/PO1^{-}) = 0.06 \text{ V vs. SCE}$, In the absence of vibrational structures, E_{0-0} can be roughly estimated from the absorption spectrum.¹³ This corresponds to 420 nm, which translates into an $E_{0-0}(PO1^{-*}/PO1^{-})$ of 2.95 eV for the PO1 anion.

 $E_{p/2}(PO1^{-}/PO1^{-*}) = E_{p/2}(PO1^{-}/PO1^{-}) - E_{0-0}(PO1^{-*}/PO1^{-}) = 0.06 - 2.95 = -2.89 \text{ V}$ vs. SCE



2-3-4-2 Cyclic Voltammogram of Substrate

Electrochemical potentials were obtained with a standard set of conditions to main internal consistency. Samples were prepared with 0.3 mmol of substrate in 30 mL of 0.1 M tetrabutylammonium hexafluorophosphate in dry, degassed acetonitrile. Measurements employed a glassy carbon working electrode, platinum wire counter electrode, saturated calomel reference electrode, and a scan rate of 0.1 V/s. Reductions were measured by scanning potentials in the negative direction; the glassy carbon electrode was polished between each scan.



Supplementary Figure 9. Cyclic voltammogram of 2,2,2-trifluoro-N-phenylacetamide.



Supplementary Figure 10. Cyclic voltammogram of N-benzyl-2,2,2-trifluoroacetamide.



Supplementary Figure 11. Cyclic voltammogram of 2,2,2-trifluoroacetamide.



Supplementary Figure 12. Cyclic voltammogram of ethyl 2,2,2-trifluoroacetate.





Supplementary Figure 13. ¹H NMR (400 MHz, CDCl₃) spectrum of PO2



¹³C NMR (101 MHz, CDCl₃) spectrum of **2-(diphenylphosphaneyl)-6-methylphenol (PO2)**

Supplementary Figure 14. ¹³C NMR (101 MHz, CDCl₃) spectrum of PO2



Supplementary Figure 15. ³¹P NMR (162 MHz, CDCl₃) spectrum of PO2



Supplementary Figure 16. ¹H NMR (400 MHz, CDCl₃) spectrum of 3



Supplementary Figure 17. ¹³C NMR (101 MHz, CDCl₃) spectrum of 3



Supplementary Figure 18. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 3



Supplementary Figure 19. ¹H NMR (400 MHz, CDCl₃) spectrum of 4



Supplementary Figure 20. ¹³C NMR (101 MHz, CDCl₃) spectrum of 4



Supplementary Figure 21. ¹⁹F NMR (101 MHz, CDCl₃) spectrum of 4



Supplementary Figure 22. ¹H NMR (400 MHz, *d*₆-DMSO) spectrum of 5



Supplementary Figure 23. ¹³C NMR (101 MHz, *d*₆-DMSO) spectrum of 5



Supplementary Figure 24. ¹⁹F NMR (376 MHz, *d*₆-DMSO) spectrum of 5



Supplementary Figure 25. ¹H NMR (400 MHz, CDCl₃) spectrum of 6



Supplementary Figure 26. ¹³C NMR (101 MHz, CDCl₃) spectrum of 6



Supplementary Figure 27. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 6



Supplementary Figure 28. ¹H NMR (400 MHz, Acetone-*d*₆) spectrum of 7



Supplementary Figure 29. ¹³C NMR (101 MHz, Acetone-*d*₆) spectrum of 7



Supplementary Figure 30. ¹⁹F NMR (376 MHz, Acetone-*d*₆) spectrum of 7



Supplementary Figure 31. ¹H NMR (400 MHz, CDCl₃) spectrum of 8



Supplementary Figure 32. ¹³C NMR (101 MHz, CDCl₃) spectrum of 8



Supplementary Figure 33. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 8







Supplementary Figure 35. ¹³C NMR (101 MHz, CDCl₃) spectrum of 9



Supplementary Figure 36. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 9



Supplementary Figure 37. ¹H NMR (400 MHz, CDCl₃) spectrum of 10



Supplementary Figure 38. ¹³C NMR (101 MHz, CDCl₃) spectrum of 10



Supplementary Figure 39. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 10



Supplementary Figure 40. ¹H NMR (400 MHz, CDCl₃) spectrum of 11



Supplementary Figure 41. ¹³C NMR (101 MHz, CDCl₃) spectrum of 11



Supplementary Figure 42. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 11



Supplementary Figure 43. ¹H NMR (400 MHz, CDCl₃) spectrum of 15



Supplementary Figure 44. ¹³C NMR (101 MHz, CDCl₃) spectrum of 15



Supplementary Figure 45. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 15



Supplementary Figure 46. ¹H NMR (400 MHz, CDCl₃) spectrum of 16



Supplementary Figure 47. ¹³C NMR (101 MHz, CDCl₃) spectrum of 16



Supplementary Figure 48. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 16



Supplementary Figure 49. ¹H NMR (400 MHz, CDCl₃) spectrum of 17



Supplementary Figure 50. ¹³C NMR (101 MHz, CDCl₃) spectrum of 17



Supplementary Figure 51. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 17



Supplementary Figure 52. ¹H NMR (400 MHz, CDCl₃) spectrum of 18



Supplementary Figure 54. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 18



Supplementary Figure 55. ¹H NMR (400 MHz, CDCl₃) spectrum of 19



Supplementary Figure 56. ¹³C NMR (101 MHz, CDCl₃) spectrum of 19



Supplementary Figure 57. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 19







Supplementary Figure 59. ¹³C NMR (101 MHz, CDCl₃) spectrum of 20



Supplementary Figure 60. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 20



Supplementary Figure 61. ¹H NMR (400 MHz, CDCl₃) spectrum of 21



Supplementary Figure 62. ¹³C NMR (101 MHz, CDCl₃) spectrum of 21



Supplementary Figure 63. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 21



Supplementary Figure 64. ¹H NMR (400 MHz, CDCl₃) spectrum of 22



Supplementary Figure 65. ¹³C NMR (101 MHz, CDCl₃) spectrum of 22



Supplementary Figure 66. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 22



Supplementary Figure 67. ¹H NMR (400 MHz, CDCl₃) spectrum of 23



Supplementary Figure 68. ¹³C NMR (101 MHz, CDCl₃) spectrum of 23



Supplementary Figure 69. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 23



Supplementary Figure 70. ¹H NMR (400 MHz, CDCl₃) spectrum of 24


Supplementary Figure 71. ¹³C NMR (101 MHz, CDCl₃) spectrum of 24



Supplementary Figure 72. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 24



Supplementary Figure 73. ¹H NMR (400 MHz, CDCl₃) spectrum of 25



Supplementary Figure 74. ¹³C NMR (101 MHz, CDCl₃) spectrum of 25



Supplementary Figure 75. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 25







Supplementary Figure 77. ¹³C NMR (101 MHz, CDCl₃) spectrum of 26



Supplementary Figure 78. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 26



Supplementary Figure 79. ¹H NMR (400 MHz, CDCl₃) spectrum of 27



Supplementary Figure 80. ¹³C NMR (101 MHz, CDCl₃) spectrum of 27



Supplementary Figure 81. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 27



Supplementary Figure 82. ¹H NMR (400 MHz, CDCl₃) spectrum of 28



Supplementary Figure 83. ¹³C NMR (101 MHz, CDCl₃) spectrum of 28



Supplementary Figure 84. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 28



Supplementary Figure 85. ¹H NMR (400 MHz, CDCl₃) spectrum of 29



Supplementary Figure 86. ¹³C NMR (101 MHz, CDCl₃) spectrum of 29



Supplementary Figure 87. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 29



Supplementary Figure 88. ¹H NMR (400 MHz, CDCl₃) spectrum of 30



Supplementary Figure 89. ¹³C NMR (101 MHz, CDCl₃) spectrum of 30



Supplementary Figure 90. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 30



Supplementary Figure 91. ¹H NMR (400 MHz, CDCl₃) spectrum of 31



Supplementary Figure 92. ¹³C NMR (101 MHz, CDCl₃) spectrum of 31







Supplementary Figure 94. ¹H NMR (400 MHz, CDCl₃) spectrum of 32





Supplementary Figure 96. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 32



Supplementary Figure 97. ¹H NMR (400 MHz, CDCl₃) spectrum of 33



Supplementary Figure 98. ¹³C NMR (101 MHz, CDCl₃) spectrum of 33







Supplementary Figure 100. ¹H NMR (400 MHz, CDCl₃) spectrum of 34



¹³C NMR (101 MHz, CDCl₃) spectrum of ethyl 4-(benzyloxy)-2,2-difluorobutanoate (34)

Supplementary Figure 101. ¹³C NMR (101 MHz, CDCl₃) spectrum of 34



Supplementary Figure 102. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 34



Supplementary Figure 103. ¹H NMR (400 MHz, CDCl₃) spectrum of 35



Supplementary Figure 104. ¹³C NMR (101 MHz, CDCl₃) spectrum of 35











Supplementary Figure 108. ¹⁹F NMR (376 MHz, *d*₆-DMSO) spectrum of 36



Supplementary Figure 109. ¹H NMR (400 MHz, CDCl₃) spectrum of 37



Supplementary Figure 110. ¹³C NMR (101 MHz, CDCl₃) spectrum of 37



Supplementary Figure 111. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 37



Supplementary Figure 112. ¹H NMR (400 MHz, CDCl₃) spectrum of 38





Supplementary Figure 114. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 38



Supplementary Figure 115. ¹H NMR (400 MHz, CDCl₃) spectrum of 39



Supplementary Figure 116. ¹³C NMR (101 MHz, CDCl₃) spectrum of 39



Supplementary Figure 117. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 39



Supplementary Figure 118. ¹H NMR (400 MHz, CDCl₃) spectrum of 40





Supplementary Figure 120. ¹H NMR (400 MHz, CDCl₃) spectrum of 41





Supplementary Figure 122. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 41



Supplementary Figure 123. ¹H NMR (400 MHz, CDCl₃) spectrum of 43



Supplementary Figure 124. ¹³C NMR (101 MHz, CDCl₃) spectrum of 43







Supplementary Figure 126. ¹H NMR (400 MHz, CDCl₃) spectrum of 44





Supplementary Figure 128. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 44









Supplementary Figure 131. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 45



Supplementary Figure 132. ¹H NMR (400 MHz, CDCl₃) spectrum of 46





Supplementary Figure 134. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 46





Supplementary Figure 136. ¹³C NMR (101 MHz, CDCl₃) spectrum of 47



Supplementary Figure 137. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 47



Supplementary Figure 138. ¹H NMR (400 MHz, CDCl₃) spectrum of 48





Supplementary Figure 140. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 48



Supplementary Figure 141. ¹H NMR (400 MHz, CDCl₃) spectrum of 49






Supplementary Figure 143. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 49



Supplementary Figure 144. ¹H NMR (400 MHz, CDCl₃) spectrum of 50





Supplementary Figure 146. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 50



Supplementary Figure 147. ¹H NMR (400 MHz, CDCl₃) spectrum of 51



Supplementary Figure 148. ¹³C NMR (101 MHz, CDCl₃) spectrum of 51





Supplementary Figure 150. ¹H NMR (400 MHz, CDCl₃) spectrum of 52





Supplementary Figure 152. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 52



Supplementary Figure 153. ¹H NMR (400 MHz, CDCl₃) spectrum of 53



Supplementary Figure 154. ¹³C NMR (101 MHz, CDCl₃) spectrum of 53



Supplementary Figure 155. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 53



Supplementary Figure 156. ¹H NMR (400 MHz, CDCl₃) spectrum of 54





Supplementary Figure 158. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 54



Supplementary Figure 159. ¹H NMR (400 MHz, CDCl₃) spectrum of 55



Supplementary Figure 160. ¹³C NMR (101 MHz, CDCl₃) spectrum of 55



Supplementary Figure 161. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 55



Supplementary Figure 162. ¹H NMR (400 MHz, CDCl₃) spectrum of 56





Supplementary Figure 164. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 56



Supplementary Figure 165. ¹H NMR (400 MHz, CDCl₃) spectrum of 57







Supplementary Figure 167. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 57



Supplementary Figure 168. ¹H NMR (400 MHz, CDCl₃) spectrum of 58





Supplementary Figure 170. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 58



Supplementary Figure 171. ¹H NMR (400 MHz, CDCl₃) spectrum of 59



Supplementary Figure 172. ¹³C NMR (101 MHz, CDCl₃) spectrum of 59



Supplementary Figure 173. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 59



Supplementary Figure 174. ¹H NMR (400 MHz, CDCl₃) spectrum of 60





Supplementary Figure 176. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of 60



Supplementary Figure 178. ¹³C NMR (101 MHz, CDCl₃) spectrum of 61



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