Putrescine and its metabolic precursor arginine promote biofilm and c-di-GMP synthesis in

Pseudomonas aeruginosa

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## **Supplemental Figures and Tables**



Figure S1. Mutants in predicted polyamine catabolism and biosynthesis genes have reduced levels of putrescine and spermidine. *P. aeruginosa* PAO1  $\Delta spuC$  (putrescine catabolism),  $\Delta speD$  (spermidine biosynthesis), and  $\Delta speAC$  (putrescine biosynthesis) mutants were grown in M63 media without exogenous arginine or putrescine. Targeted metabolomics was used to quantify putrescine or spermidine levels. All 3 mutants had significant reductions in both spermidine and putrescine compared to wildtype cells. Mean +/- sd is shown; letters indicate genotypes with significantly different levels of polyamines by ANOVA and Tukey's HSD.



Figure S2. L-arginine robustly promotes biofilm formation independent of SpeA and SpeC. A) PAO1 or the *speAC* mutant were treated with 20 mM L-arginine or 20 mM of KCl as a control for the chloride ions in L-arginine HCl salt. \* indicates p < 0.0001 by student's t-test. Error bars represent standard deviation. Data points show all technical replicates from 3 biological replicates. B) Wild-type *P. aeruginosa* PAO1 and  $\Delta speA$ ,  $\Delta speC$ , and  $\Delta speAC$  mutants were treated with 2.5 mM L-arginine hydrochloride. At this concentration, L-arginine does not induce biofilm formation in any of the genetic backgrounds, including the L-arginine-accumulating  $\Delta speA$  and  $\Delta speAC$  strains. Error bars represent standard deviation. Data points show all technical replicates from 4 biological replicates.



Figure S3. Addition of exogenous L-arginine leads to a slight increase in bacterial density in wild-type but not in the  $\triangle eps::FRT$  strain. A-B) Growth curves of *P. aeruginosa* PAO1 WT and  $\triangle eps::FRT$  strain in M63 supplemented with 20 mM L-arginine HCl, 20 mM KCl, or equal volume H<sub>2</sub>O (error bands represent 95% confidence intervals, n $\ge$ 17 from 3 biological replicates). C) L-arginine HCl supplementation does not affect the maximum growth rate of either wild-type or the  $\triangle eps::FRT$  strain. D) While the L-arginine HCl supplementation increases the maximum of OD<sub>600</sub> at 8 hr (D), this increase is dependent on exopolysaccharide biosynthesis (\*p<0.05 by student's t-test, n $\ge$ 17 from 3 biological replicates). E) The addition of L-arginine HCl or KCl does not significantly alter the maximum growth rate of either the WT or  $\triangle eps::FRT$  strain (p=0.2477 by one-way ANOVA, n $\ge$ 17 from 3 biological replicates).

## Table S1 Primers used in this study

Names	<b>Restriction sites</b>	Sequences $(5' \rightarrow 3')$
speA-UpF	HindIII	TTTAAAAGCTTCGCCTGTCGGCGACG
speA-UpRc		GCTAGCCAGGCGCGGTGATCTC
speA-DnF		GCGCCTGGCTAGCCCGTCG
speA-DnRc	XbaI	CAATTTCTAGAGGCCCTGGTGGCGTTC
speC-UpF	HindIII	TTTAAAAGCTTCGCCCAGGTGACCCAG
speC-UpRc		CGACTGCGGGTTGGGACTCCCAATG
speC-DnF		CAACCCGCAGTCGCCTCTGCTAC
speC-DnRc	Xba I	CAATTTCTAGAACGGGTTGTAGGCAATTTCCC
speD-UpF	Xba I	CTTAATCTAGAGCCCAAGGTGTTCACGAAG
speD-UpRc		CGTGTGCGACGTGGGGGAACTCTC
speD-DnF		TTCCCCACGTCGCACACGAGGAAG
speD-DnRc	Hind III	TTAAAAGCTTAGGCGCTGTACCAGGGC
speE-UpF	Hind III	CTTAAAAGCTTGGCGGCCACCAGC
speE-UpRc		GGTGAAGCGGGGCCGGGATCTCCC
speE-DnF		GATCCCGGCCCCGCTTCACCAAGAAG
speE-DnRc	Xba I	CTTAATCTAGATCGCGATGCCGTCG
spuC-UpF	Xba I	CTTAATCTAGAAGTGCTGCCGCTGTTC
spuC-UpRc		CTCAGGGACGTCACACCTCTTCTATTCAAG
spuC-DnF		GGTGTGACGTCCCTGAGCGGACTTTTG
spuC-DnRc	Hind III	CTTAAAAGCTTGTAGCCGATGCCGATGG
spuD-UpF	Hind III	CTTAAAAGCTTCCTGGAGAACATCCGCATC
spuD-UpRc		TCGCGGAGCGGGGTAGCTCC
spuD-DnF		ACCCCGCTCCGCGAGGAGCC
spuD-DnRc	Xba I	CAATTTCTAGATCTTCTTCTCCGCCTGCAC
GFPmut3-F		ATGTCTAAAGGTGAAGAATTATTC
GFPmut3-Rc		TTATTTGTACAATTCATCCATACC