

Appendix

Proteome-scale mapping of binding sites in the unstructured regions of the human proteome

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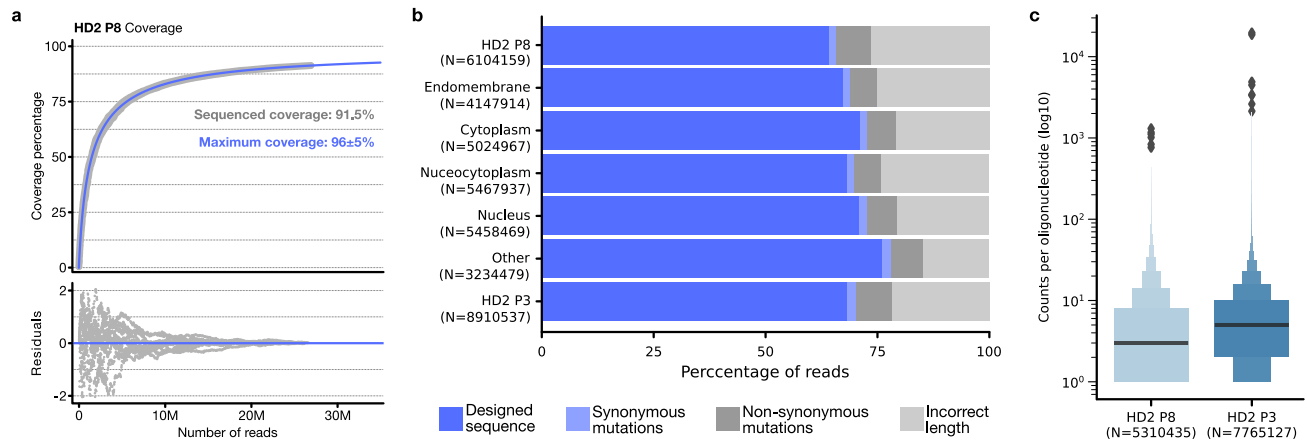


Figure S1. Non-design components and count distribution of the libraries. (a) Coverage of the HD2 P8 library as a function of sequencing depth, fitting of data to a double hyperbolic model (in blue) and residuals are shown. The contribution of reads of all sub-libraries were considered for calculating the coverage of the HD2 P8 library, as it is the result of the balanced combination of all compartment-specific sub-libraries. (b) Percentages of reads obtained by NGS of the libraries generated in this work associated with sequences that matched the designed sequences, those that presented mutations either silent or non-silent, and those that due to frame-shifting, insertion or deletion mutations did not match the expected oligonucleotide length. (c) Distribution of NGS counts for the designed oligonucleotides in the HD2 P8 and HD2 P3 libraries.

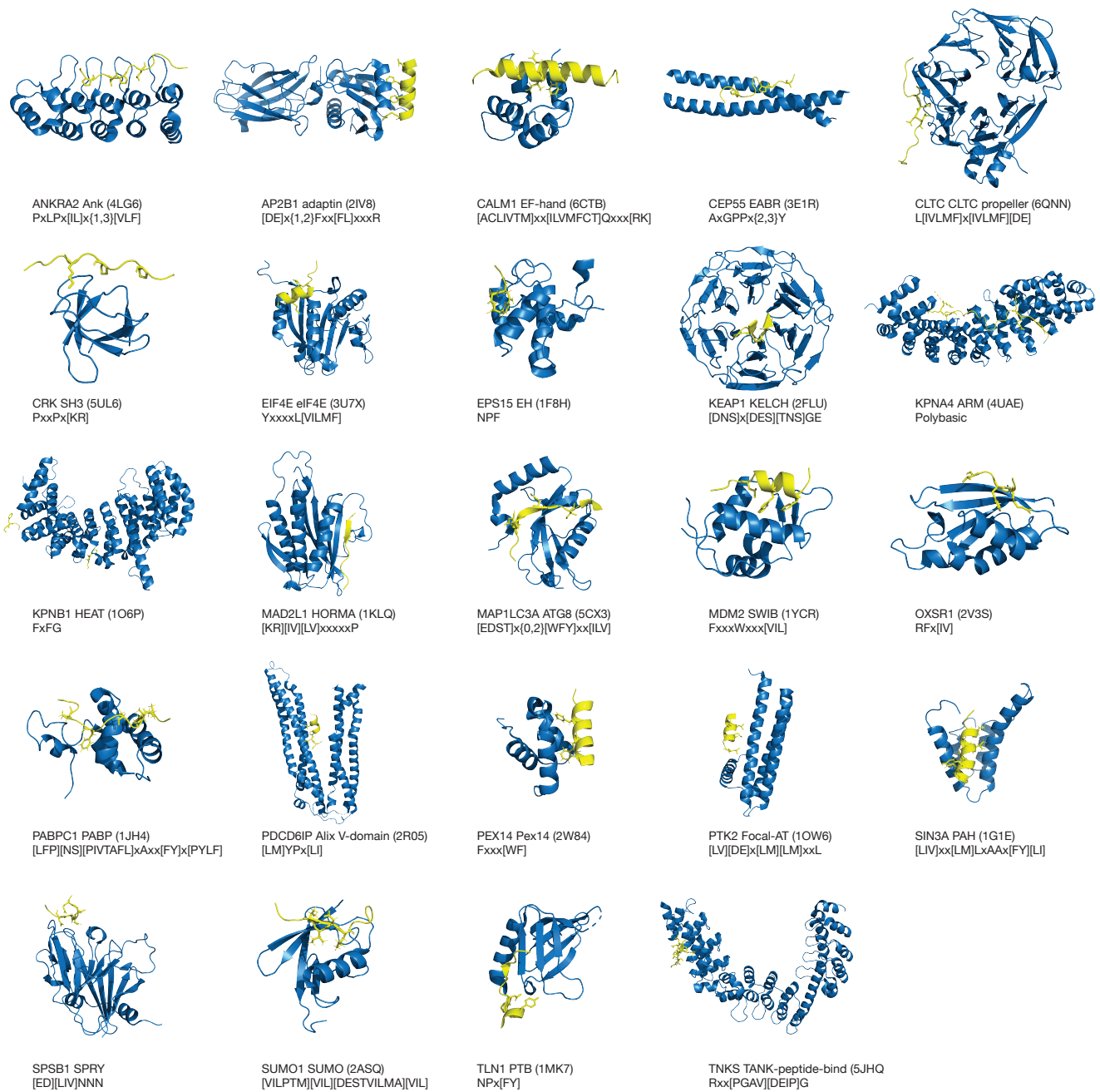


Figure S2. Representative structures of the bait proteins domains (blue) with bound ligands (yellow) and indicated binding motifs. PDB codes are indicated in the figure.

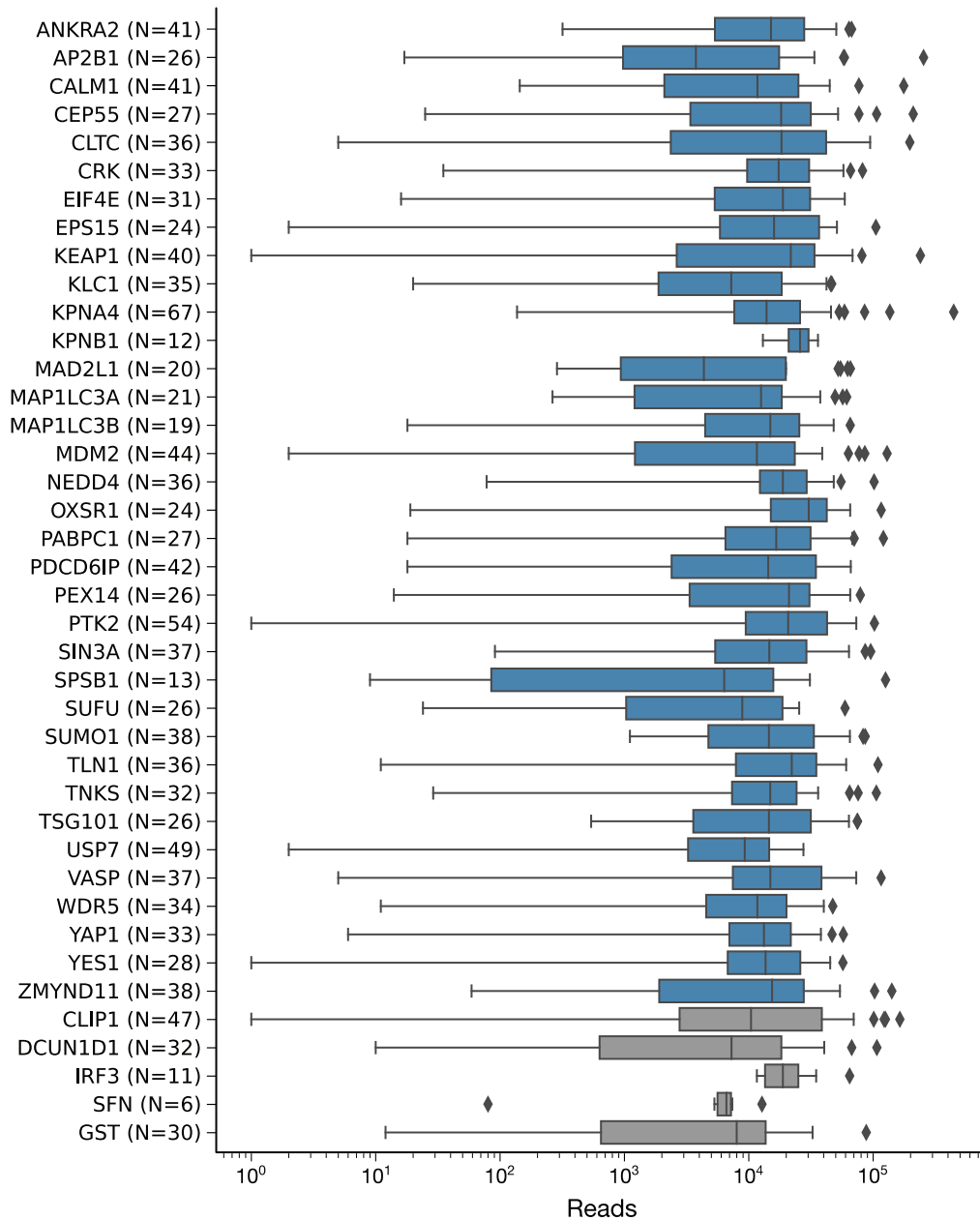


Figure S3. Distribution of NGS reads for binding enriched phage pools. Distribution of the NGS reads that matched the peptides encoded by the library are shown grouped by bait. Control baits are shown in gray. N = number of barcoded phage pools included in the analysis per bait.

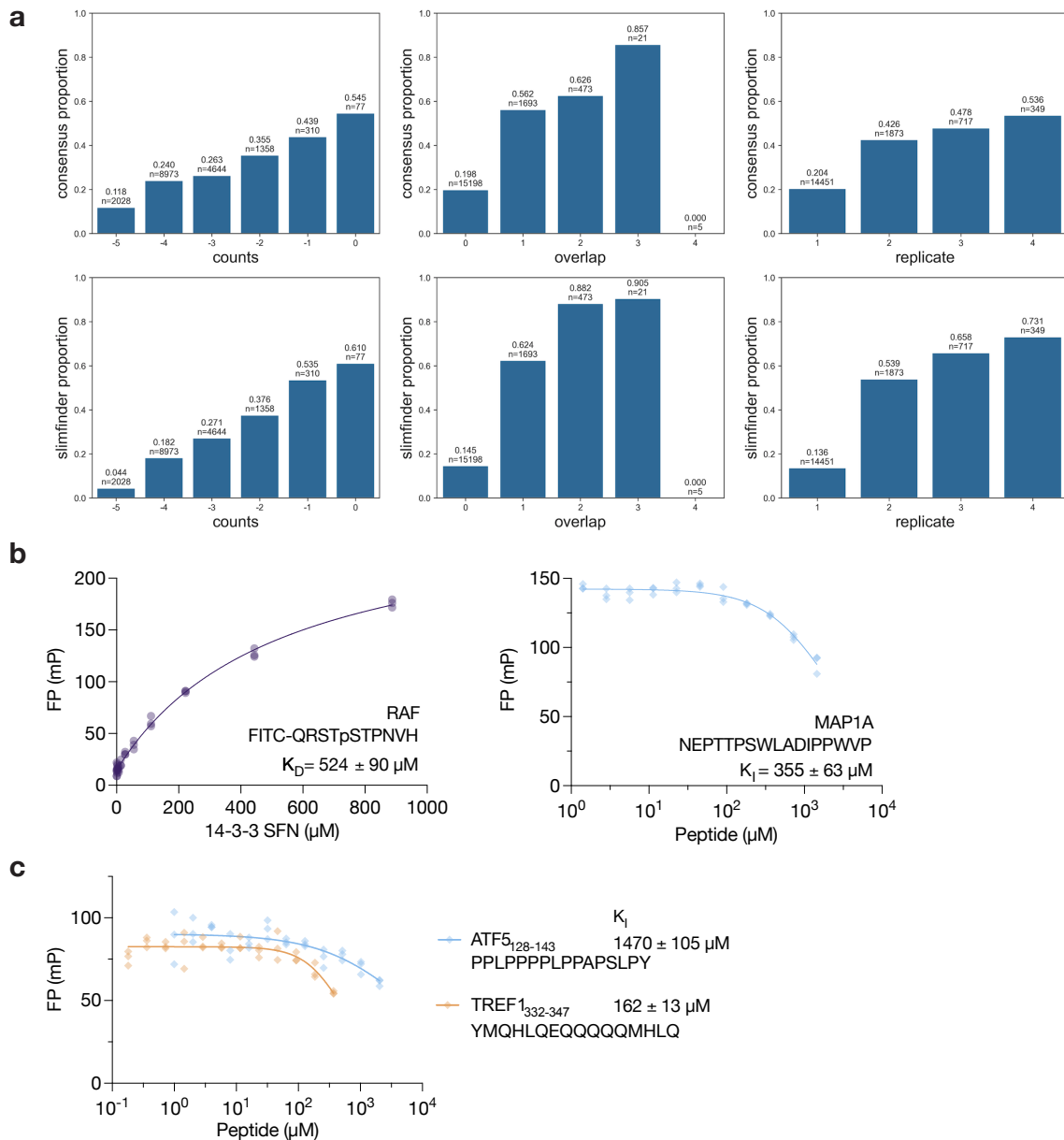


Figure S4. Analysis of the proportion of motif-containing peptides (a) and validation of low affinity interactions of peptides lacking consensus motifs (b-c).

(a) Proportion of peptides returned for a bait containing the expected ELM consensus for the bait (consensus proportion) or the SLIMFinder consensus enriched in the peptides in the screen of that bait (slimfinder proportion). The plots show the data grouped by three peptide metric statistics: count - \log^{10} of the normalized counts, overlap - the number of overlapping peptides returned in the screen for that bait, replicates - the number of replicates that the peptide was returned. (b) FP affinity measurements of the interactions between 14-3-3 SFN and the probe peptide phospho-RAF1255-264 (FITC-QRSTpSTPNVH; left) and the competition with unlabeled MAP1A₁₈₃₆₋₁₈₅₁ (right). (c) FP competition experiment for the Keap1 KELCH domain between the FITC-labeled probe peptide NFE1L1₂₂₆₋₂₄₃ and the unlabeled peptides ATF₁₂₈₋₁₄₃ and TREF₂₂₃₋₃₄₇ that lack Keap1 KELCH binding motif.

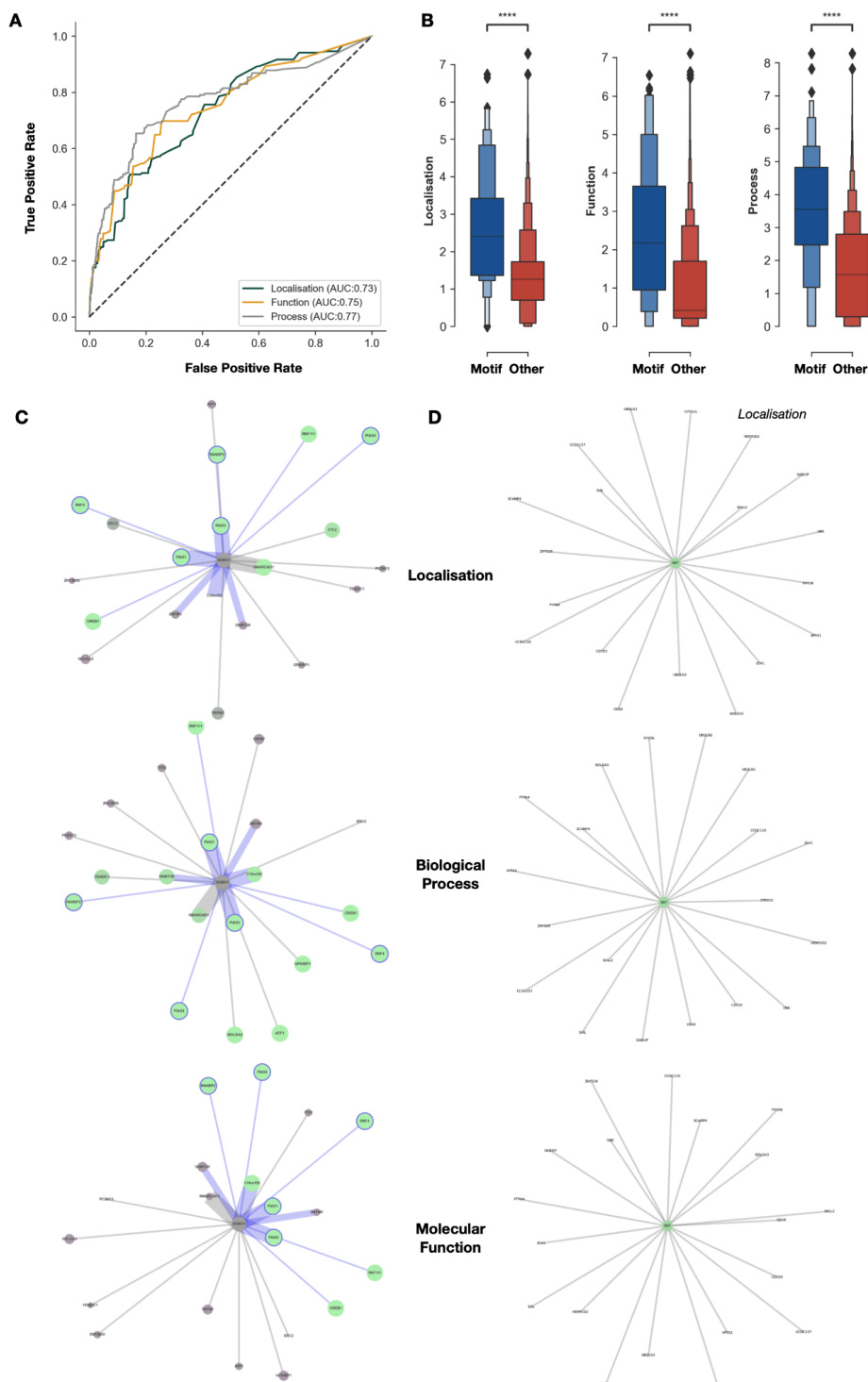


Figure S5. Overview of GO term enrichment analysis. (A) ROC curve showing the discriminator power of the shared GO terms analysis metrics for validated motifs (B) Box plots of Localisation, Biological Process and Molecular Function GO term enrichment in validated motif containing peptides and other returned peptides. (C-D) Network of representation of the SUMO (C) and GST (D, control) bait interactions with high/medium confidence peptide-containing proteins. Network is overlaid with data from a shared GO terms analysis. Three networks are displayed for each bait: Localisation, Biological Process and Molecular Function. Node colour and size is the p-value of the best shared ontology term (greener and larger nodes are more enriched). Edge weights and lengths represent the best peptide confidence for the protein (more confident peptides are closer to the bait with thicker edges). The edges of known interactors are shown in blue. Nodes representing validated motif-containing proteins are ringed in blue.

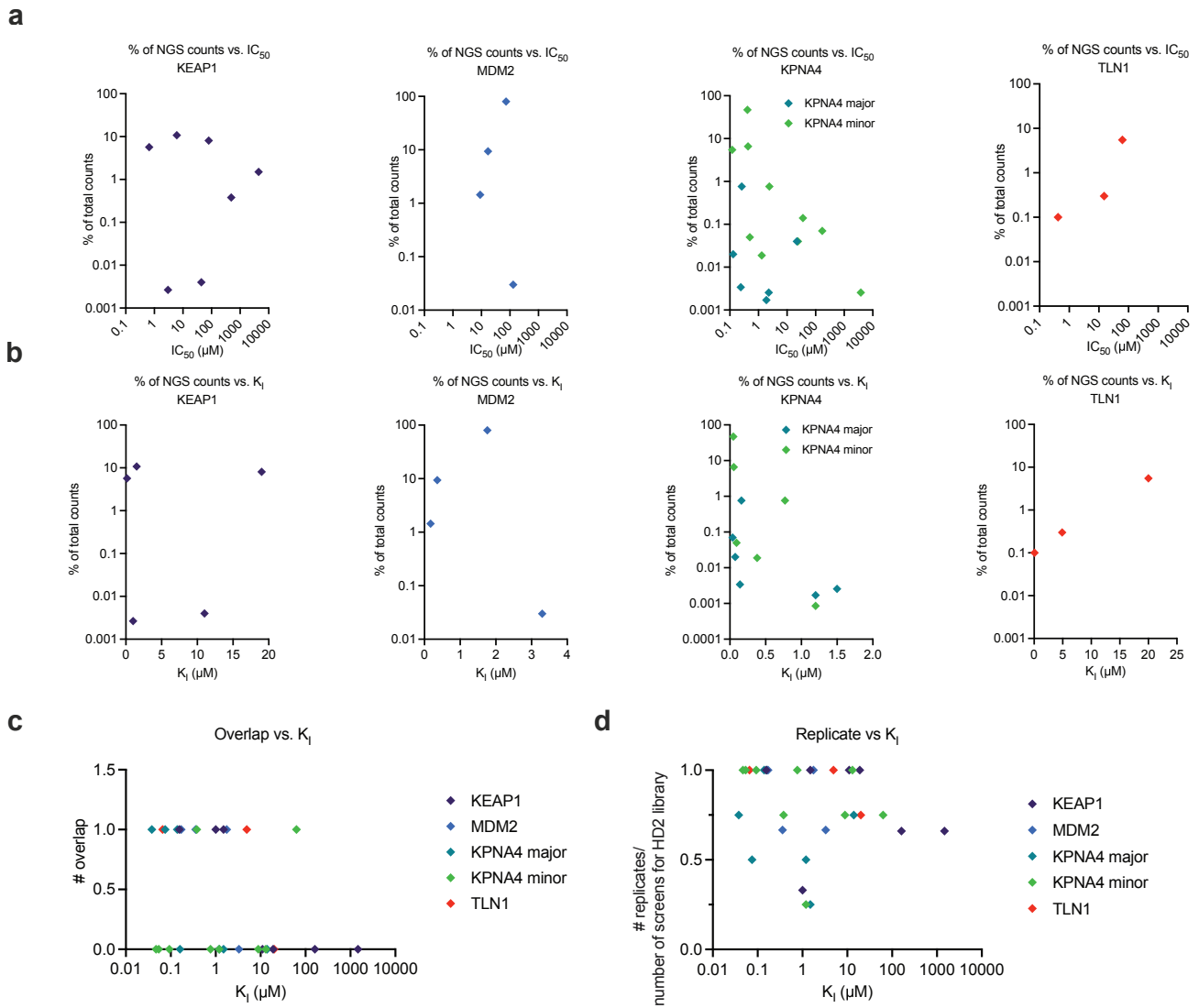


Fig S6. Analysis of correlation between affinity and ProP-PD results. The % NGS counts per were plotted against (a) IC_{50} values, and (b) K_d values. The affinities (K_d values) were further plotted against (c) the observed overlaps of the peptides in selections, and (d) the occurrence of peptides in replicate selections. A KPNA4 outlier was removed for better visualization.

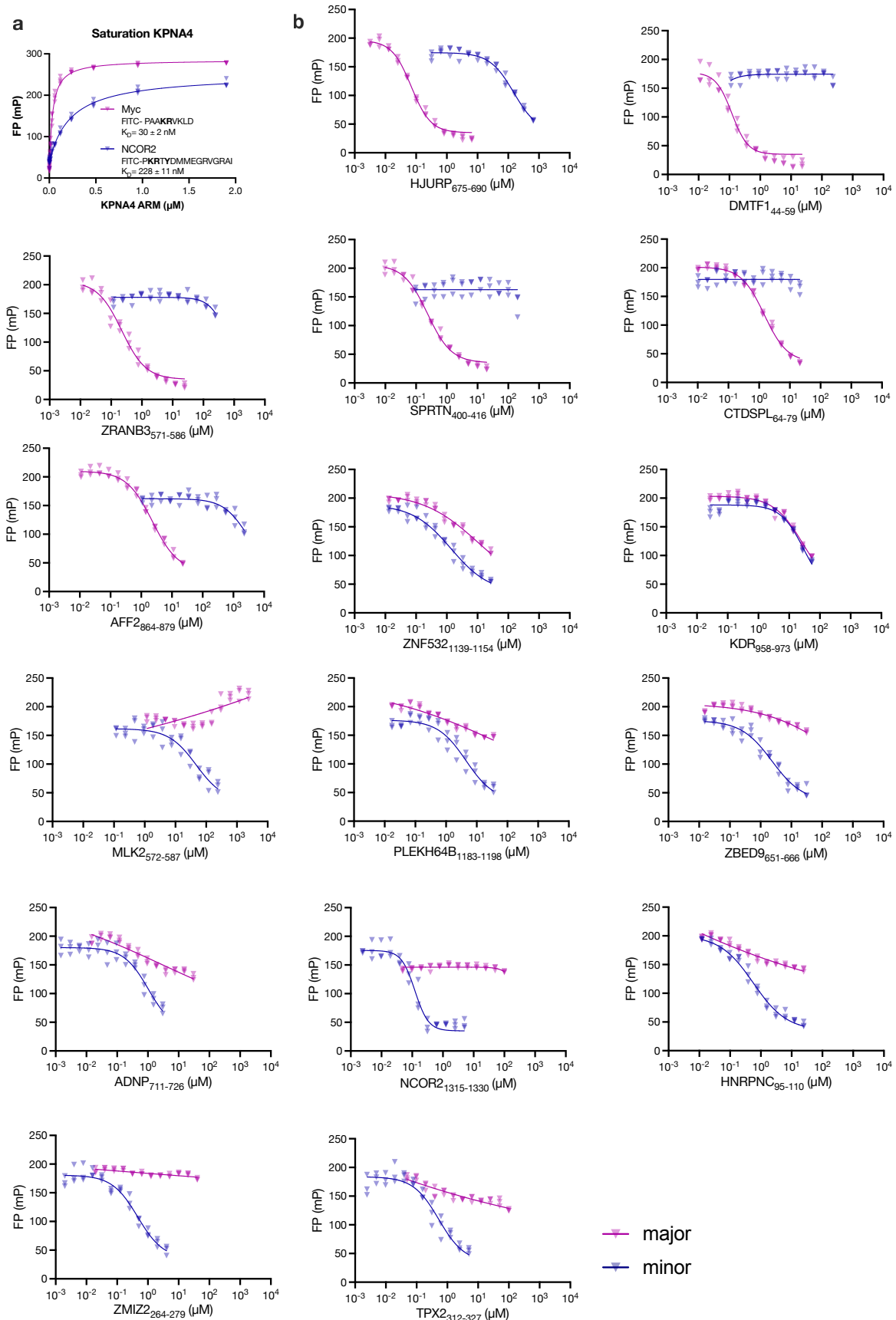


Fig S7. FP affinity measurements of KPNA4. (a) Saturation curves with probes for the for the major groove (Myc₃₂₀₋₃₂₈ FITC-PAAKRVKLD; purple) and minor groove (NCOR2₁₃₀₇₋₁₃₂₂; FITC-PKRTYDMMEGRVGRAI (blue)). (b) FP competition binding experiment of 18 peptides against the major groove probe FITC-Myc₃₂₀₋₃₂₈ (purple) and the minor groove FITC-NCOR2₁₃₀₇₋₁₃₂₂ (blue). Unlabeled peptides used for competition: HJURP₆₇₅₋₆₉₀, DMTF₁₄₄₋₅₉, ZRANB3₅₇₁₋₅₈₆, SPRTN₄₀₀₋₄₁₆, CTDSPL₆₄₋₇₉, AFF2₈₆₄₋₈₇₉, ZNF532₁₁₃₉₋₁₁₅₄, KDR₉₅₈₋₉₇₃, MLK2₅₇₂₋₅₈₇, PLEKH64B₁₁₈₃₋₁₁₉₈, ZBED9₆₅₁₋₆₆₆, ADNP₇₁₁₋₇₂₆, NCOR2₁₃₀₇₋₁₃₂₂, HNRPN₉₅₋₁₁₀, ZMIZ2₂₆₄₋₂₇₉, TPX2₃₁₂₋₃₂₇.

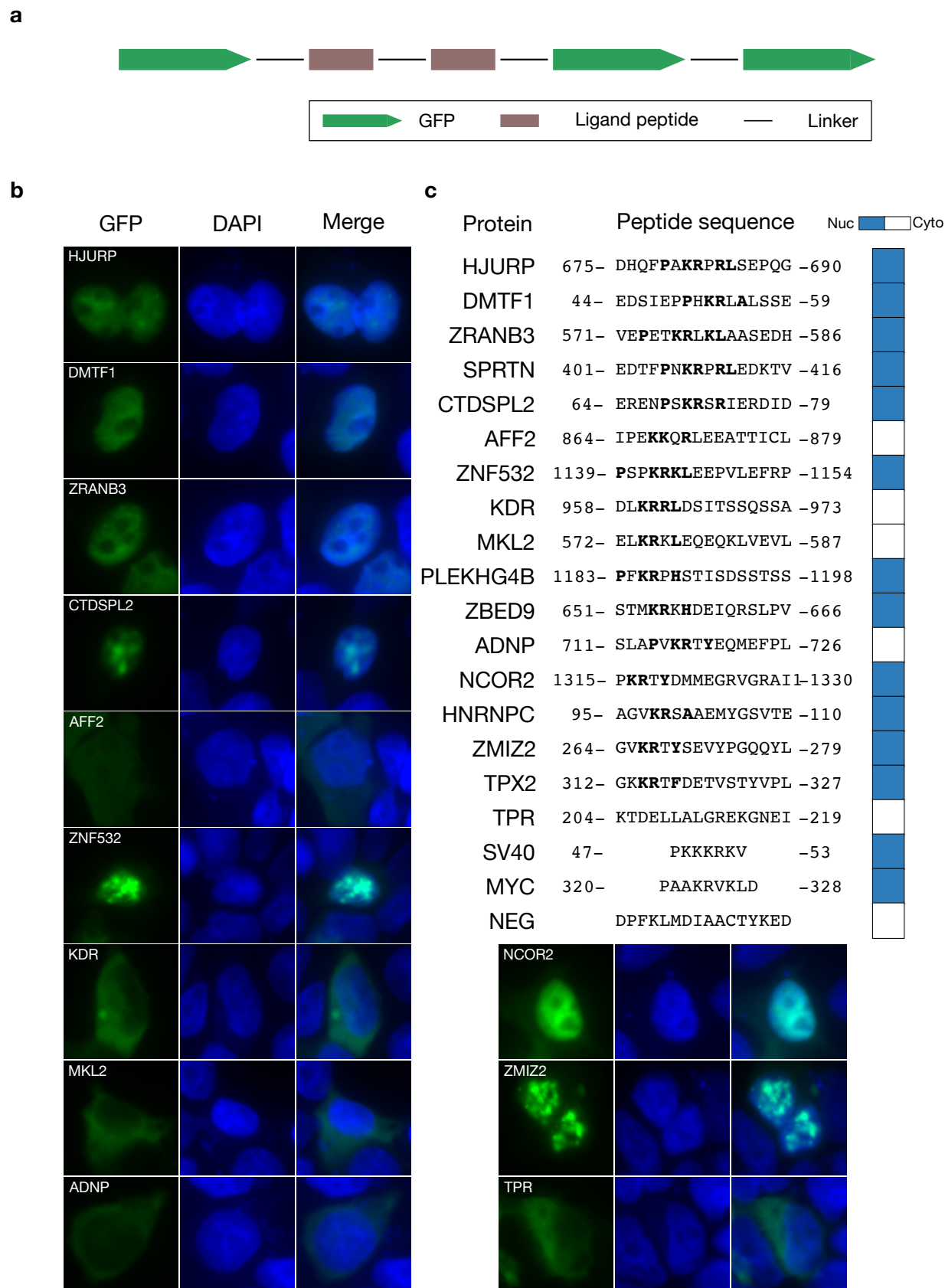
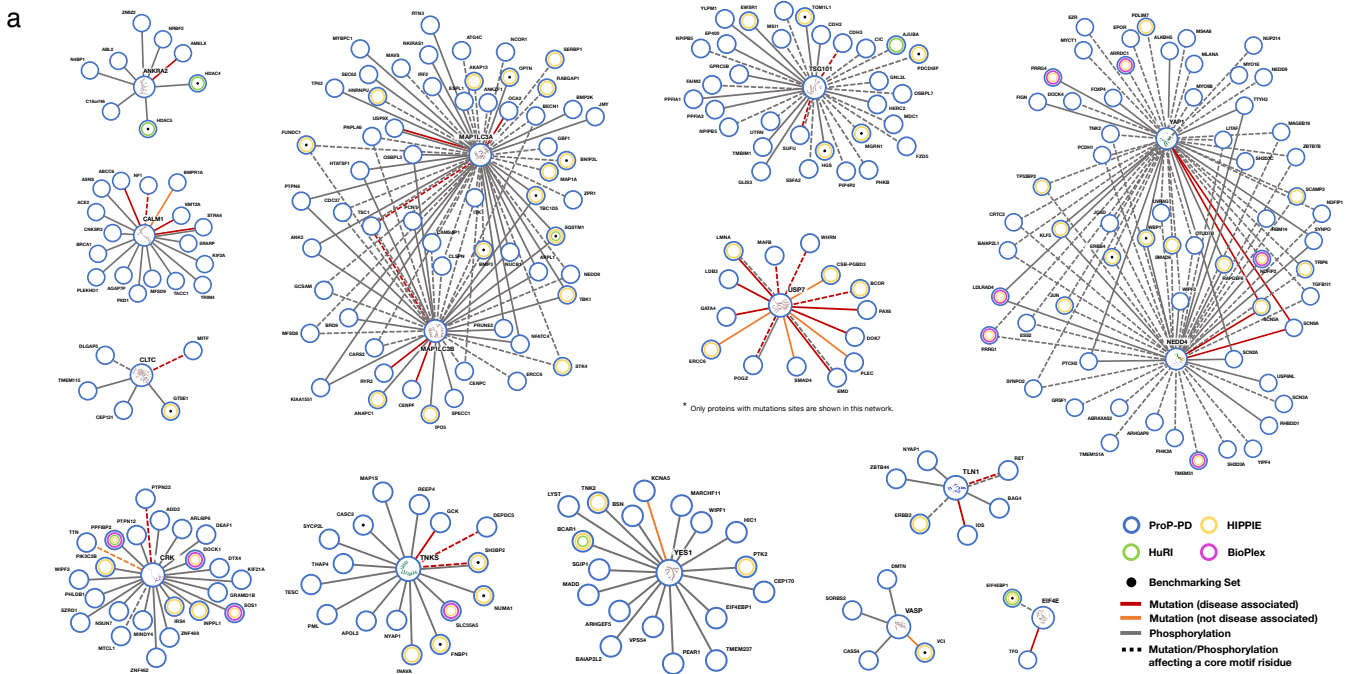


Fig S8. Validation of KPNA4 binding sequences as NLSs. (a) Schematic of the construct used as NLS sensor. (b) Cellular localization of NLS sensor with different peptides inserted. HEK293 cells were transiently transfected with the NLS sensor and fixed 36 h after transfection, and imaged using epifluorescence microscopy. The nucleus was stained with DAPI. (c) Overview of the peptide sequences fused to the NLS sensor, with the resulting cellular localization indicated.



b

ANKRA2	MAP1LC3A/B	TLN1	YAP1/NEDD4
AMELX HDAC4 HDAC5 C16orf46 NABP1 ABL2 ZMIZ2 NRBF2	MAP1LC3A binders AKAP13 ANKZF1 BMP2K BNIP3L ESPL1 GPI1 MAP1A USP9X OCA2 ZPR1 HNR1NP NKIRAS1 SEC62 TRF3 IRF2 MAVS OPTN RABGAP1 JNY MYBPC1 SERP1 NCOR1 ATG4C RTN3	RET ERBB2 IDS NYAP1 ZBTB44 BAG4 TNKS APOL2 CASC3 DEPDC5 FNBP1 GCK INAVA MAP1S NUMA1 NUP1 PML REEP4 SH3BP2 SLC35A5 SYCP2 TESC THAP4 TSG101	YAP1 binders MS4A8 MYC11 MLANA DOCK4 FOXP4 ARRDC1 MYO9B MYO1E ALKH5 NEDD9 PRRG4 EPOR PDLIM7 EZR FGH NUP214 YAP1 / NEDD4 binders PRRG1 BAIAP2L1 CRIC3 ERBB2 ESS2 JCAD KUN JLFBAD4 LDBR4 OTUD7B RABGF6 RBM14 SCAMP3 SCN5A SCN9A SH2D3C SMAD3 SYNPO TFEB11 TPS20 TRIP6 UVRAG WBP1 WIP2 ZBTB7B NDH1P PTCH2 SCN2A LITAF MAGEB1B PCDH1 SYNPO2 TNK2 TTYH3
CLTM1 ABC6 ASNS ACE2 NF1 BMPR1A KMT2A STR6A SRARP MFSN9 PKD1 AGAP7P KIF2A BRCA1 CKNKR3 PLEKH01 TACC1 TRIM4	MAP1LC3A / MAP1LC3B binders FUNDC1 BECN1 MFSN8 NFATC4 STK4 CARS2 KIAA1551 CENPF CDC37 ANK2 SQSTM1 HTASFS1 BNIP3 ITK CAMSAP1 PCNT TSC1 TBC1D5 APPL1 PNPLA6 PTPN6 NEDD9 NUC81 CLSNP PRUNE2 GCSAM OSBP1L3	CBX6 CDH2 CDH3 CIC FZD5 GNL3L GPCR5B HERC2 MDC1 MGRN1 MSH1 PIAP2 SUFU TM6IM1 UTRN VLM1 AJUBA ENP40 FAIM2 HSS NIPBP5 OSBP1L7 TOM1L1 GLI3 PHK3 CDWR1P ECC81 PPP1A PPP2A	YAP1 binders (continued) BRAXAS2 GRS1 ARHGAP5 PIK4C SH2D3A RHBD1D SCN3A TMEM151A TMEM51 YIPF4 USP6L YEN1 KCNAS WIP1 PTK2 BSN EIF4EBP1 PEAR1 VPS4 ARHGFE5 MADD BCAR1 March11 HIC CEP170 BAIAP2L2 TMEM237 LYST SGIP1 DLGAP2 TNK2
CEP131 DLGAP5 MITF GTSE1 TMEM115	CRK ADD3 ARLBP6 DEAF1 DOCK1 DTX4 GRAMD1B INPP1A IRS4 KIF21A MINDY4 MTCL1 NSUN7 PHLDB1 PKIC2B PPP1B2 PTPN12 PTPN22 SOS1 SZR1 TTN WIP2 ZNF462 ZNF469	USP7 * BCOR CSP-PGG03 DOK7 EMD ERCC3 GATA4 LDB3 LMNA MAFB PAK6 PLEC POGZ SMAD4 WHRN VASP VCL DMTN SORBS2 CASSA	NEDD4 binders ABRAXAS2 GRS1 ARHGAP5 PIK4C SH2D3A RHBD1D SCN3A TMEM151A TMEM51 YIPF4 USP6L YEN1 KCNAS WIP1 PTK2 BSN EIF4EBP1 PEAR1 VPS4 ARHGFE5 MADD BCAR1 March11 HIC CEP170 BAIAP2L2 TMEM237 LYST SGIP1 DLGAP2 TNK2
EIF4E	CENPF RYR2 SPECC1 ANAPC1 ERCC6 IPO5		
TFG EIF4EBP1			

Fig S9. Additional PPI networks based on peptides with reported disease-associated mutations or phosphosites. (a) Networks showing interactions of reproducibly selected high/medium confidence peptides with overlapping disease-associated mutations or phosphorylation. Peptides with mutations or phosphorylation residues occurring in the core motif or in the flanking regions (+/- 2 residues) are shown. Mutations associated with diseases are colored in red (orange if they are not disease associated). Phosphorylated sites are colored in gray. Dashed-edge lines represent core motif residues holding a mutation or a phosphorylation site. (b) Sequences of high/medium confidence peptides with disease-associated mutations in the binding motif. Motif-containing regions are highlighted with blue background, core motifs are indicated in bold letters, phosphorylation sites within or in vicinity of the motifs are indicated by a box, and disease associated mutations are indicated in red bold letters (mutation of core residue).