Supplemental Data

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Supplemental Table S1. Primers for genotyping, ChIP, EMSA, qPCR, and recombinant vectors.

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Supplemental Figures



Supplemental Figure S1. Measured regions of the lateral vascular cambium. Relates to Figures 1, 2, 3, and 5.

The measured regions of the lateral vascular cambium are indicated by the red lines.



Supplemental Figure S2. The representative pictures of whole stem sections of different plants. Related to Figure 1, 2, 3, 5. Bars = $100 \ \mu m$.



Supplemental Figure S3. The hypocotyl phenotype of different plants. Related to Figure 1, 2, 3, 5.

(A) The representative pictures of whole hypocotyl sections of different plants. (B) The hypocotyl diameter of plants in (A). The hypocotyl (2 mm immediately below the shoot-hypocotyl junction) of 4-weeks-old plants (about 15 cm high) were immediately measured after being pulling out of soil. From 5 to 10 plants were measured three times each line. Bars = $200 \,\mu$ m.



Supplemental Figure S4. Expression patterns of the *BES1(L)* and *BES1(S)* in the secondary stem and vasculature. Related to Figure 1.

(A, B) GUS staining of the secondary stem (A) and its cross section (B) for the BES1(L)pro:GUS reporter line in the En-2 background. (C, D) GUS staining of the secondary stem and its cross section for the BES(S)pro:GUS reporter line in the En-2 background. Bars = 1 cm in (A) and (C), Bars = 20 µm in (B) and (D).



Supplemental Figure S5. Identification and phenotypes of the *max2-1 BES1*-RNAi line. Related to Figure 2.

(A) Representative photographs of 8-week-old Col-0, *BES1*-RNAi, *max2-1*, and *max2-1 BES1*-RNAi plants. (B) Representative photographs of basal stems and hypocotyls of 8-week-old Col-0, *BES1*-RNAi, *max2-1*, and *max2-1 BES1*-RNAi plants. Bar represents 1 cm. (C) Relative *BES1* mRNA levels in seedlings of Col-0, *BES1*-RNAi, *max2-1*, and *max2-1 BES1*-RNAi plants. Data are means \pm SD (*n* = 3). Bars represent 1 cm.



Supplemental Figure S6. Transient luciferase assay in *N. benthamiana* leaves demonstrates the co-expression with BES1 specifically repress *WOX4pro:LUC* activity. Related to Figure 3. The HsfA1d was used as the unrelated transcription factor control, GFP was the control effector. Effectors were BES1-FLAG and a stable-form BES1 (bes1-D), and *WOX4pro-LUC* was the reporter.



Supplemental Figure S7. Reduced *WOX4* transcript levels in the SL biosynthetic and signaling mutants *max3-1*, *max2-1*, and *Atd14-1*. Related to Figure 5.

The relative *WOX4* transcript level in Col-0 was set as "1". Data are means \pm SD (*n* = 3). *P* values were determined by Student's *t*-test, ***P* < 0.01.



Supplemental Figure S8. Phenotype of the *gWOX4-GFP/max2-1*. Related to Figure 5.

- (A) Representative photographs of 6-week-old max2-1 and gWOX4-GFP/max2-1 plants.
- (B) Representative photographs of 3-week-old max2-1 and gWOX4-GFP/max2-1 plants.
- (C) Representative photographs of basal stems and hypocotyls of max2-1 and
- gWOX4-GFP/max2-1 plants shown in (A). Bars represent 1 cm.



Supplemental Figure S9. Phenotype of the hypocotyl of *gWOX4-GFP/max2-1*. Related to Figure 5.

(A) Representative cross sections of the hypocotyl of *max2-1* plants and three independent *gWOX4-GFP/max2-1* lines (8-week-old plants). Bars = 50 μ m. (B) Quantification of the semidiameter of hypocotyl in *max2-1* plants and three independent *gWOX4-GFP/max2-1* lines shown in (A). Data are means \pm SD, n=15 (5 number were collected randomly in 3 plants/line). (C) Relative *WOX4* mRNA levels in seedlings of *max2-1* and three independent lines of *gWOX4-GFP/max2-1* in (A&B). Relative *WOX4* transcript levels were set to "1" in *max2-1*. Data are means \pm SD (*n* = 3).

Name	sequences 5'-3'
WOX4pro-CHIP-F1-F	TGATCACAGCTTGACACACGA
WOX4pro-CHIP-F1-R	ACCAGTGTAGGTTGATCATGACT
WOX4pro-CHIP-F2-F	AGTCATGATCAACCTACACTGGT
WOX4pro-CHIP-F2-R	GGTTTGGCAATGTCAAGCGT
WOX4pro-CHIP-F3-F	AGGTTCTGCAGTTCCCGATG
WOX4pro-CHIP-F3-R	GTCGAATGCTTTTCGGTGGC
WOX4pro-CHIP-F4-F	TGAGAGCCACCGAAAAGCAT
WOX4pro-CHIP-F4-R	GTCTGCATGGAGCACTACTT
WOX4pro-CHIP-F5-F	AGTGCTCCATGCAGACATGA
WOX4pro-CHIP-F5-R	TCCCAGATGAAGAAAACCCA
Probe a-F	CCATCTGATCACAGCTTGACACACGAATCAGGT
	TATCCATCGTGTGA
Probe a-R	TCACACGATGGATAACCTGATTCGTGTGTCAAG
	CTGTGATCAGATGG
Probe b-F	ATCATCTGTTCATTTTCCATTCTTTTTTCTTTTTCC
	TACCCAACTGGA
Dual a la D	TCCAGTTGGGTAGGAAAAAGAAAAGAATGGA
Probe b-K	AAATGAACAGATGAT
Duck	GTAACGAGAAAGGCATGCATAGCATTTGCTAGT
Probe C-F	TTTAACATATAGCA
Probe c-R	TGCTATATGTTAAAACTAGCAAATGCTATGCAT
	GCCTTTCTCGTTAC
WOX4-CDs-F	ATGAAGGTTCATGAGTTTTCGA
WOX4-CDs-R	TCTCCCTTCAGGATGGAGAG
<i>WOX4pro-</i> F	ACATATGAACAGTGGTAGAG
<i>WOX4pro-</i> R	CATTGCTATATGTTAAAA
LBP1.3	ATTTTGCCGATTTCGGAAC
<i>max2-1-</i> LP	TACATGCAAGCATGCAACTTC
<i>max2-1-</i> RP	AATAGGAACAAAATCGCCACC
wox4-1-LP	AGGTCTACCCCCTTTTCAACG
wox4-1-RP	AATGTGTGGGTTCAGTTGGAG
WOX4-RT-F	CGATCAAACCGGTCCGACAA
WOX4-RT-R	TTCTTGAGTCGGGTTCCACC
ACTINpro-CHIP-F	ACGAGGGAAAAGGCTGTCTG
ACTINpro-CHIP-R	GTCGCCGGAGATTCAAAACG
U-box-RT-F	TCTTCTTCTGCTACATCTACTCTC
U-box-RT-R	AGTGTGTGAACCCGTGAAC

Supplemental Table S1. Primers for genotyping, ChIP, EMSA, qPCR, and recombinant vectors.

Name	Plasmids	Restriction sites
WOX4pro:LUC	pCAMBIA1300 with LUC tag	EcoR I/Sac I
35Spro:BES1-FLAG	pCAMBIA1306 with FLAG tag	Kpn I/Xba I
WOX4pro:LUC-35Spro:REN	pGreenII 0800-LUC	Hind III/BamH I
gWOX4-GFP	pCAMBIA1302 with GFP tag	EcoR I/Sac I
35S:WOX4-HA	pCAMBIA1300 with HA tag	Kpn I/Xba I

Supplemental Table S2. Plasmids and restriction sites used for recombinant vectors.

Gene name	Accession number
ACTIN	AT3G18780
BES1	AT1G19350
D14	AT3G03990
HsfAld	AT1G32330
MAX2	AT2G42620
MAX3	AT2G44990
U-BOX	AT5G15400
WOX4	AT1G46480

Supplemental Table S3. Accession numbers of genes.