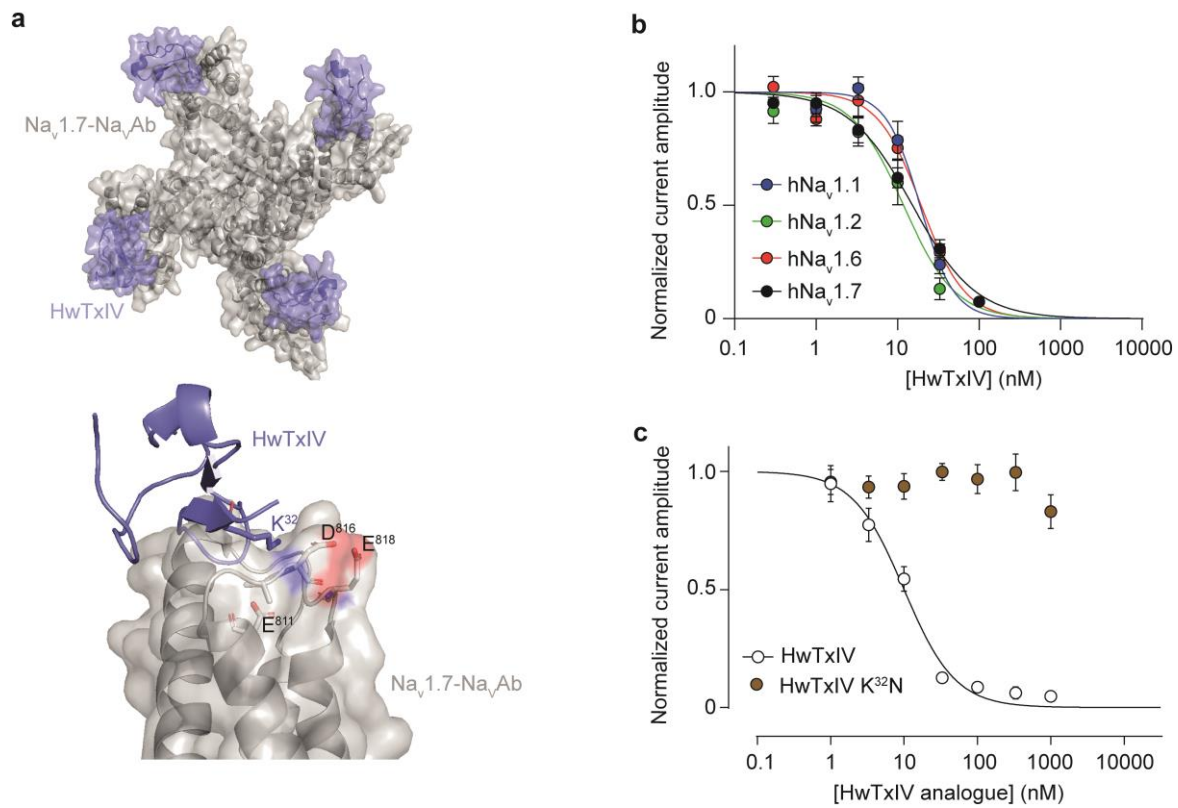
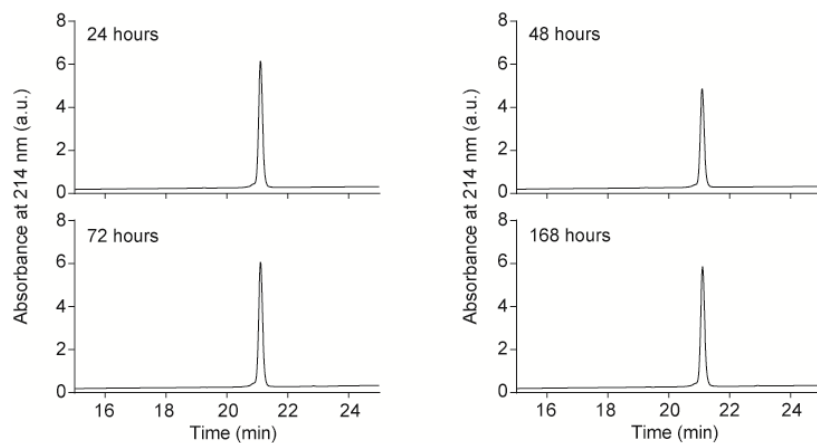


**In vivo spatiotemporal control of voltage-gated ion channels by using
photoactivatable peptidic toxins**

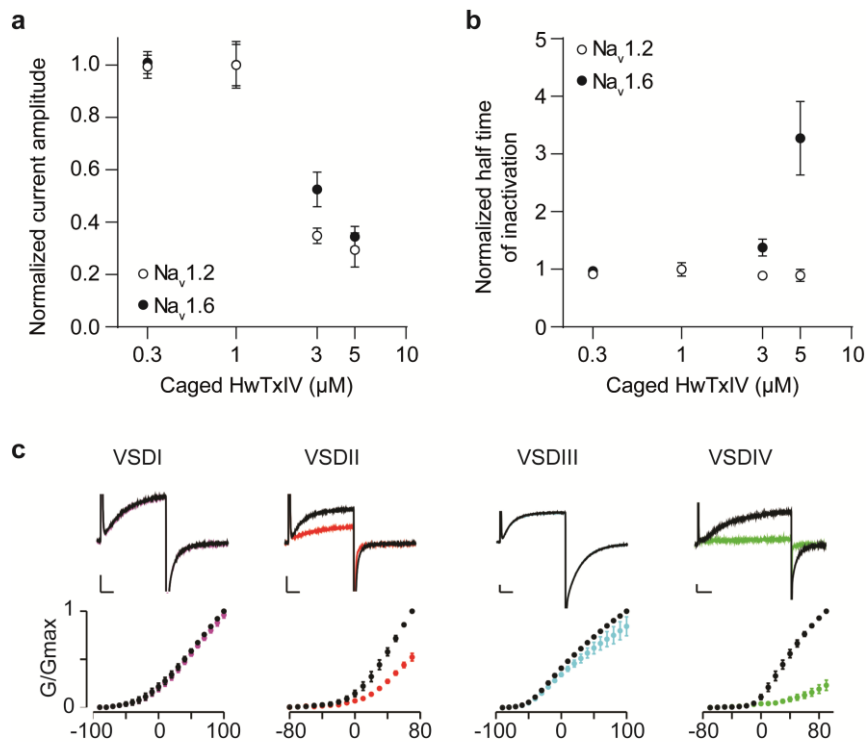
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Meudal⁵, Aude Lafoux⁶, Sébastien Nicolas^{1,2}, Duong Chu⁷, Cécile Caumes⁴, Rémy
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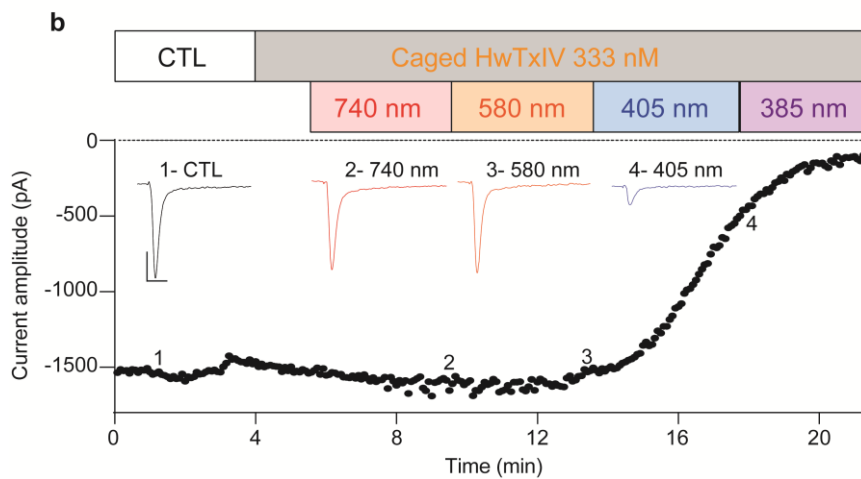
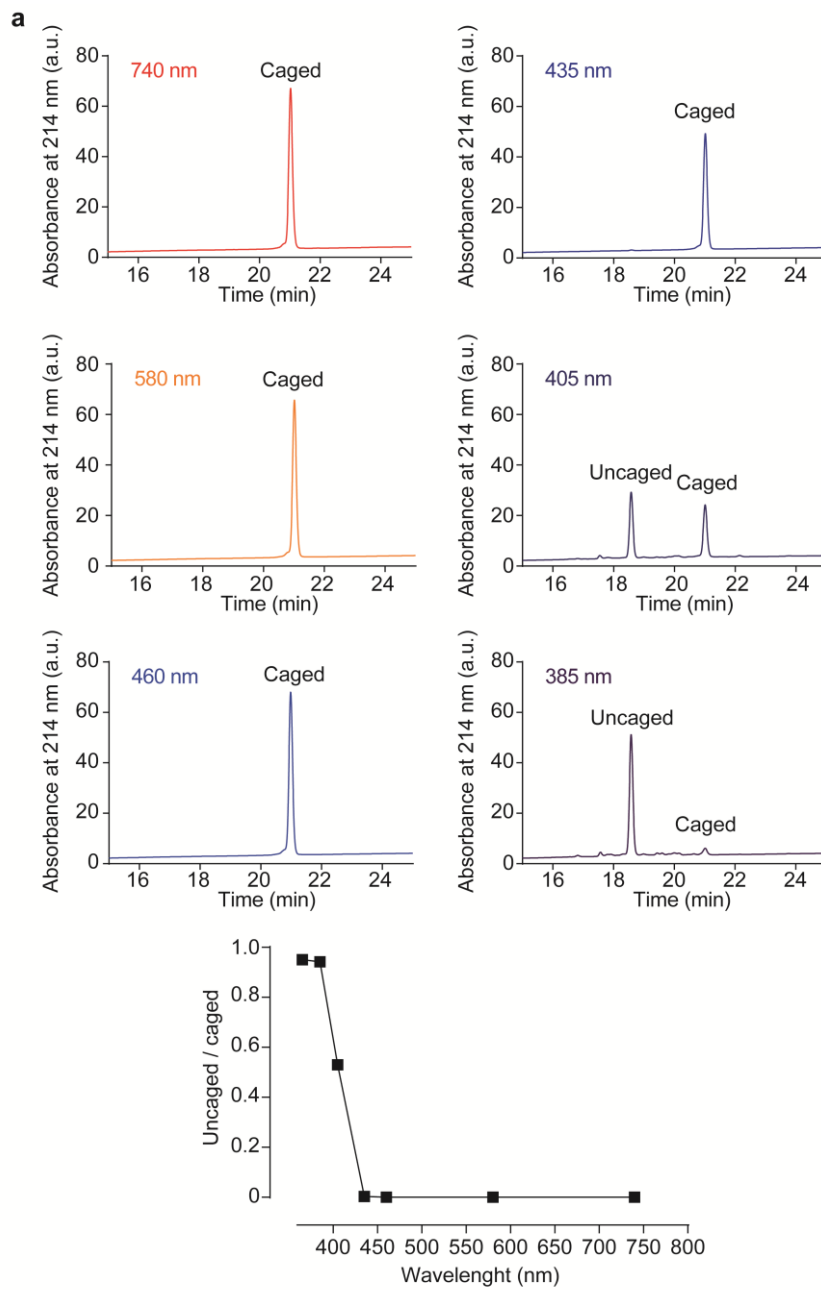
Supplementary Figure 1. (a) Molecular structure of the HwTxIV / NaV1.7-NavAb chimera complex (PDB code 7K48). (b) Average dose-response curves (mean \pm SEM) for inhibition of hNav1.1 (blue), hNav1.2 (orange), hNav1.6 (red) and hNav1.7 (black) currents by non-caged HwTxIV. The data were fitted according to a Hill equation study (hNav1.1: 18.9 ± 1.2 nM, n=29; hNav1.2: 11.9 ± 1.1 nM, n=38; hNav1.6: 19.5 ± 1.3 nM, n=55; hNav1.7: 15.2 ± 1.1 nM, n=81). (c) Average dose-response curves for hNav1.7 current inhibition by the nHwTxIV (black) and nHwTxIV-K32N mutant (brown). The data were fitted according to a Hill equation ($IC_{50} > 1 \mu M$, n=44 for nHwTxIV-N32 versus $IC_{50} = 9.9 \text{ nM} \pm 0.9 \text{ nM}$, n=45 for nHwTxIV).



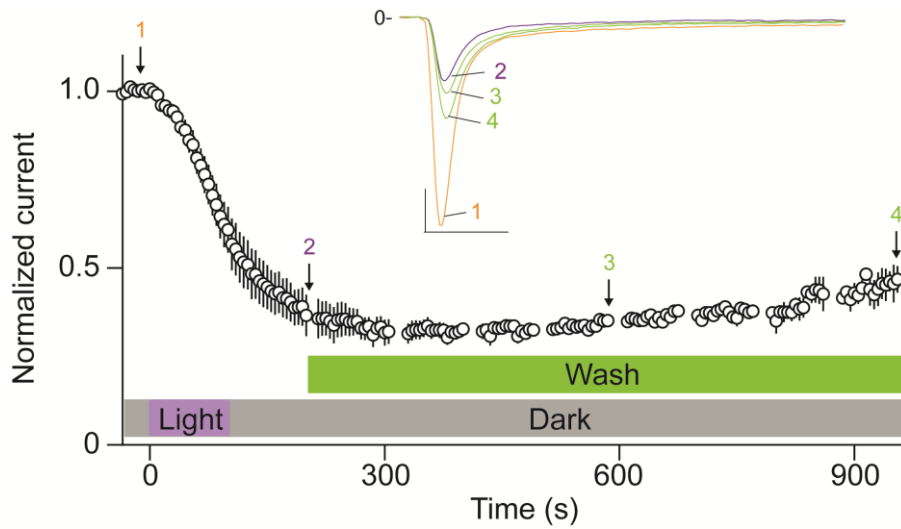
Supplementary Figure 2. Analytical RP-HPLC profiles of HwTxIV-Nvoc after different times in the dark demonstrating perfect stability of the caged peptide.



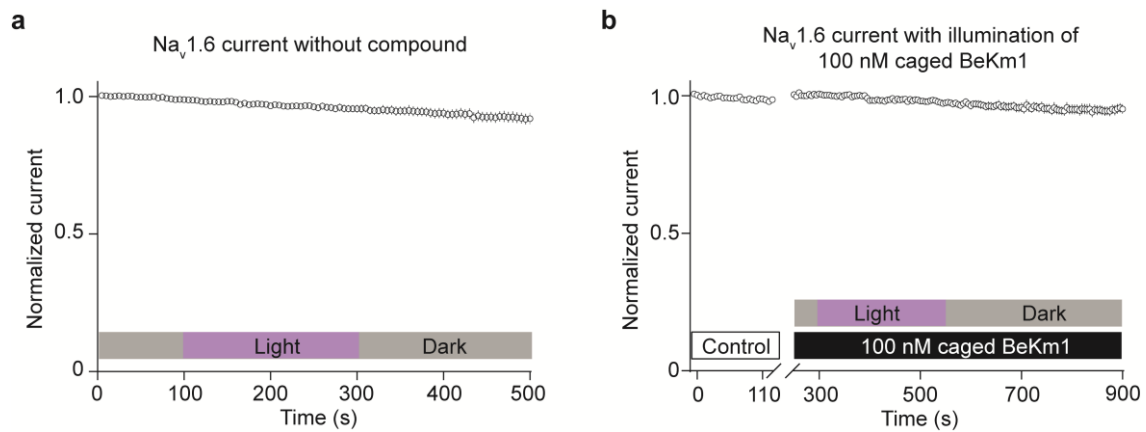
Supplementary Figure 3. (a) Average dose-response curves for hNav_v1.2 (black circles, n=11) and hNav_v1.6 (open circles, n=12) peak current inhibition by high concentrations of HwTxIV-Nvoc. (b) Average dose-response curves for hNav_v1.2 (black circles, n=11) and hNav_v1.6 (open circles, n=12) half-time of inactivation by high concentrations of HwTxIV-Nvoc. (c) Top: K⁺ currents from hNav_v1.6/rK_v2.1 chimeras harboring the VS domains of hNav_v1.6. Currents were elicited by depolarizations to +40 mV. Currents are shown before (black) and in the presence (colored traces) of 5 μM caged HwTxIV. Colors denote the domain from which the hNav_v1.6 paddle motif was transferred: VSDI in magenta; VSDII in red; VSDIII in cyan; VSDIV in green. Scale bars: 20 ms (abscissa) for VSDI-III, 40 ms for VSDIV; and 0.5 μA (ordinate axis). Bottom: Tail current conductance-activation (G-V) relationships for each of the hNav_v1.6/rK_v2.1 chimeras (n = 4). The holding voltage was -90 mV and the tail voltage was -70 mV (or -90 mV for VSDIII).



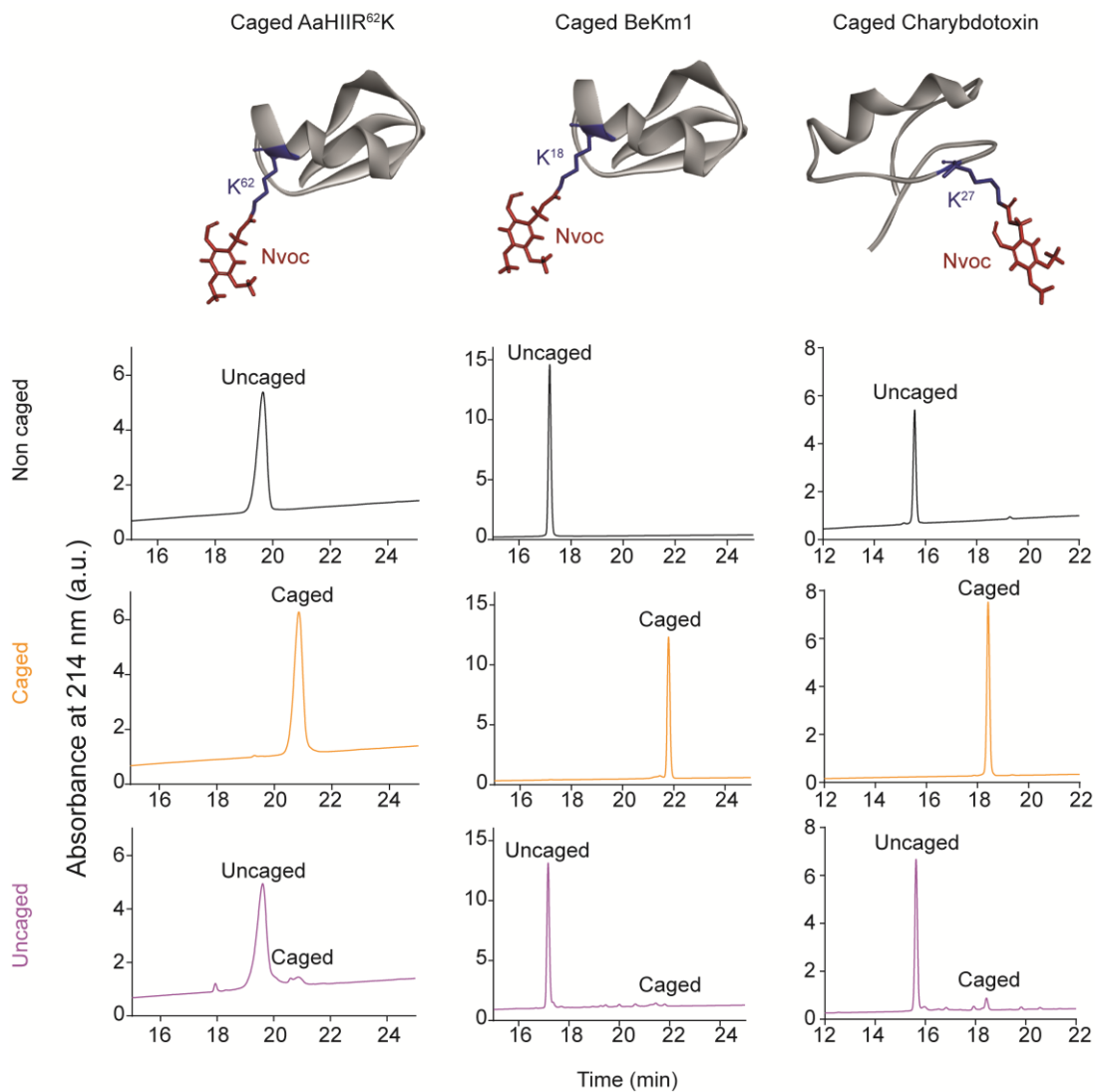
Supplementary Figure 4. (a) Top: Analytical RP-HPLC profiles of photolysis of HwTxIV-Nvoc at different wavelengths (10 min, power > 18 mW/cm²). Bottom: Uncaged/Caged ratio of RP-HPLC chromatogram peak area at different wavelengths. (b) Representative current traces and time course of hNa_v1.6 current inhibition by HwTxIV-Nvoc illuminated at different wavelengths. Scales: 2 ms, 500 pA.



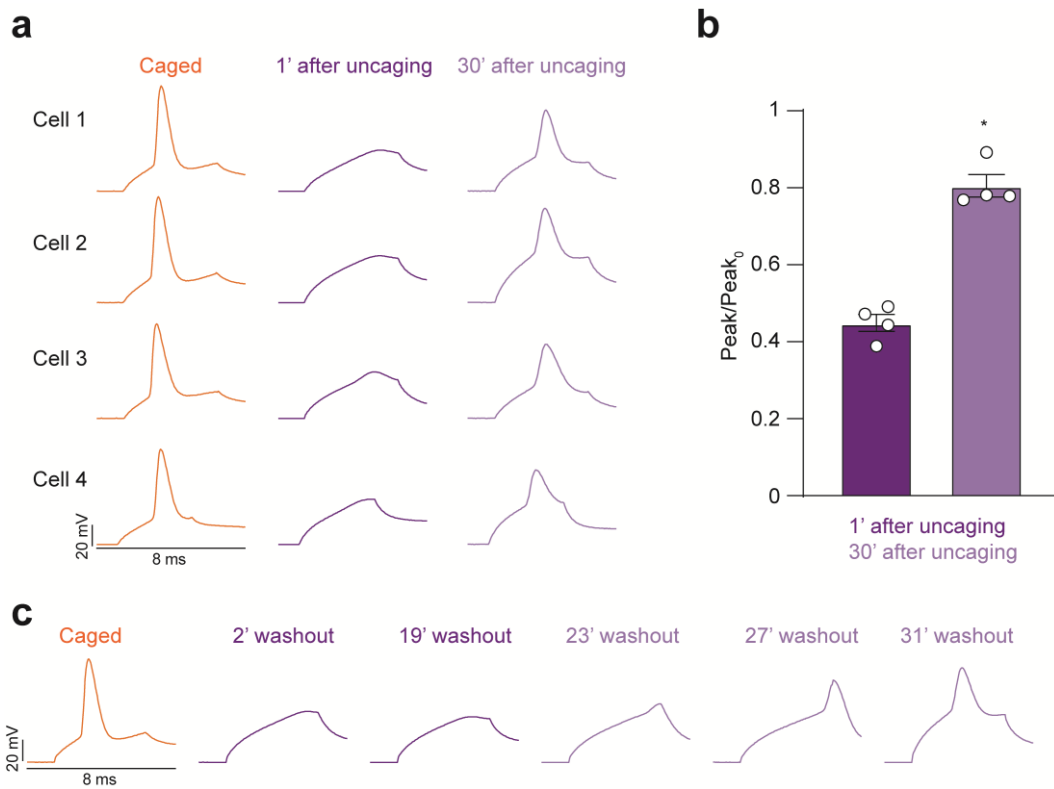
Supplementary Figure 5. Reversibility of hNav1.6 inhibition after HwTxIV-Nvoc uncaging. Representative current traces (inset) and average normalized time course of hNav_v1.6 current inhibition by HwTxIV-Nvoc illuminated at 365 nm 45 mW/cm². Wash of uncaged HwTxIV induces a partial 16% current recovery after 11 min of wash (n=6, mean ± SEM). Scales: 2 ms, 1 nA.



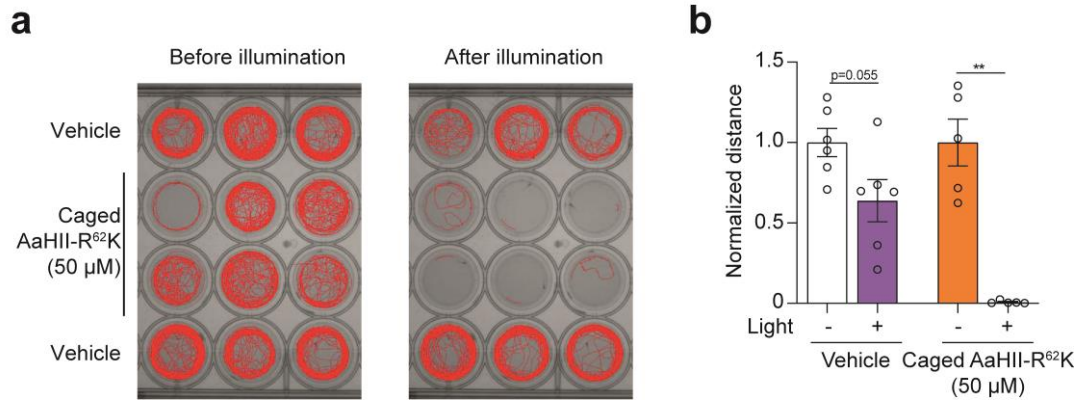
Supplementary Figure 6. (a) Average normalized time courses of hNa_v1.6 peak current inhibition by illumination (4 min, 365 nm, 45 mW/cm²) without any compound (n=22, mean ± SEM). (b) Average normalized time courses of hNa_v1.6 peak current inhibition by illumination (4 min, 365 nm, 45 mW/cm²) of 100 nM BeKm1-Nvoc (n=14, mean ± SEM). BeKm1 has no reported inhibitory effects on hNa_v1.6 peak current inhibition.



Supplementary Figure 7. Top: 3D structure of other Nvoc-protected toxins. Bottom: Analytical RP-HPLC profiles of non-caged (black), caged (orange) and uncaged (purple) AaHII-R⁶²K (left, PDB code 6NT4), BeKm1 (middle, PDB code 1J5J) and charybdotoxin (right, PDB code 4JTA).



Supplementary Figure 8. HwTxIV reversibility in L5 pyramidal neurons from mouse brain slices. (a) APs in the presence of 2.5 μ M HwTxIV-Nvoc (orange traces), and 1 and 30 minutes after uncaging the toxin for 4 different cells. (b) mean \pm SEM (n = 4 cells) of the depolarization peak, normalized to the caged values, 1 minute (0.45 ± 0.02) and 30 minutes after photolysis (0.81 ± 0.03). "*" indicates a significant increase of the peak ($p < 0.05$, paired t-test). (c) Example of kinetics of recovery of AP amplitude after photolysis of HwTxIV-Nvoc.



Supplementary Figure 9. In vivo photocontrol of caged AaHII-R⁶²K induces paralysis of zebrafish larvae. (a) Representative traces of zebrafish larvae movements in larvae injected with vehicle (top and bottom) or AaHII-R⁶²K-Nvoc (50 μM) prior or after 365 nm illumination. (b) Normalized distance traveled by larvae prior or after illumination after vehicle or AaHII-R⁶²K-Nvoc injection. (n=6 in each group, mean ± SEM; ** p<0.01 versus vehicle, paired t-test).

Supplementary Table 1. IC₅₀ values of native HwTxIV (nHwTxIV) and HwTxIVG^{1G⁴K³⁶} (HwTxIV) on hNav_{1.1}, hNav_{1.2}, hNav_{1.6} and hNav_{1.7} ion channels.

	nHwTxIV (nM)	HwTxIVG ^{1G⁴K³⁶} (nM)
hNav _{1.1}	50.7 ± 1.2	18.9 ± 1.2
hNav _{1.2}	8.3 ± 1.2	11.9 ± 1.1
hNav _{1.6}	154.3 ± 1.2	19.5 ± 1.3
hNav _{1.7}	9.9 ± 0.9	15.2 ± 1.1

Supplementary Data 1. Table of ¹H NMR chemical shifts of the non-caged HwTxIV.