

Strain-promoted azide-alkyne cycloaddition-based PSMA-targeting ligands for multimodal intraoperative tumor detection of prostate cancer

Authors: Yvonne H.W. Derks^{1*}, Mark Rijpkema¹, Helene I.V. Amaldjais-Groenen², Cato C. Loeff¹, Kim E. de Roode², Annemarie Kip¹, Peter Laverman¹, Susanne Lütje³, Sandra Heskamp¹ and Dennis W. P. M. Löwik²

¹Department of Medical Imaging, Nuclear Medicine, Radboud university medical center, Radboud Institute for Molecular Life Sciences, 6525GA Nijmegen, The Netherlands.

²Radboud University Nijmegen, Institute for Molecules and Materials, Organic Chemistry, 6525XZ Nijmegen, The Netherlands.

³Department of Nuclear Medicine, University Hospital Bonn, 53127 Bonn, Germany.

***Corresponding author:**

Yvonne Derks

Department of Medical Imaging, Nuclear Medicine

Radboud university medical center

Geert Grooteplein Zuid 10

6525GA Nijmegen

Phone: +31 (0)24 365 5340

E-mail: yvonne.derks@radboudumc.nl

ORCID: 0000-0002-8512-4103

Supporting Information

Supplementary materials and methods

DIPCDI coupling of protected amino acids:

Fmoc-protected amino acid (3.0 eq.), 1-hydroxybenzotriazole hydrate (HOBT, 1M in DMF, 3.6 eq.), N,N'-Diisopropylcarbodiimide (DIPCDI, 1M in DMF, 3.3 eq.) were added to the resin and agitated until the Kaiser test was negative (~45 minutes) after which the resin was capped with a mixture of Ac₂O (10 eq) and pyridine (10 eq) in DMF for 5 minutes and subsequently washed with DMF (3x10 mL).

HATU coupling of protected amino acids:

Fmoc-protected amino acid (3.0 eq.), 1-hydroxybenzotriazole hydrate (HOBT, 3.6 eq.), (1-[Bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxide hexafluorophosphate (HATU, 2.9 eq.) and N,N'-diisopropylethylamine (DIPEA, 6 eq.) were dissolved in DMF. The solution was pre-activated for 2 minutes before it was added to the resin. The mixture was agitated until the Kaiser test was negative (~1.5 hrs.) after which the resin was washed with DMF (3x10 mL) and DCM (3x10 mL).

Fmoc removal:

The resin was treated with 20% piperidine in DMF 3x6 minutes. The product was washed with DMF (3x10 mL).

Alloc removal:

Phenylsilane (25eq) and tetrakis(triphenylphosphine)palladium(0) (0.3eq) in DCM were added to the resin. The mixture was agitated for 20-30 minutes under a stream of argon (upon which the color changed from yellow to dark brown). Next the resin was washed with DCM, DMF and sodium diethyldithiocarbamate (0.5% in DMF) until the brown color had completely disappeared (generally 3 times).

Synthesis of the PSMA binding motif and linker

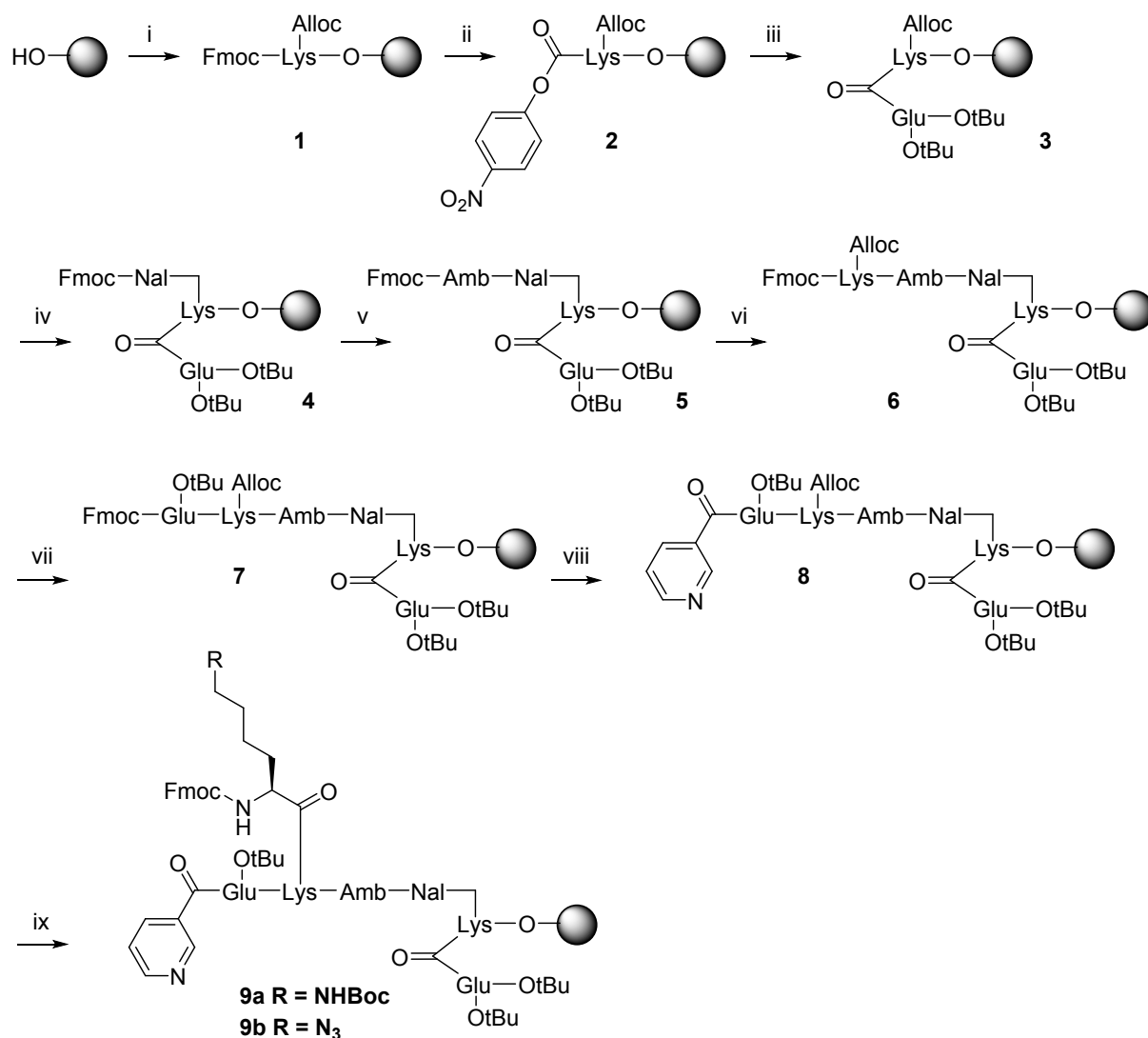


Fig. S1. Synthesis of the common PSMA binding motif and linker.

i) Wang resin (1 eq., 1.0 mmol/g, 1.00 g) was swollen in 10 mL DMF for 10 minutes. Fmoc-Lys(Alloc)-OH (3 eq., 3 mmol, 1.87 g), 4-dimethylaminopyridine (1 eq., 1 mmol, 122.2 mg), HOBt (3.6 eq., 3.6 mmol, 1M in DMF) and DIPCDI (3.3 eq., 3.3 mmol, 1M in DMF) were added to the resin and mixed on a bench roller for 20 hours. The reagents were removed from the resin by vacuum filtration. The resin was washed with DMF (3x10 mL) and DCM (3x10 mL). The Fmoc-loading was determined to be 0.5 mmol/g. Next, the resin was capped with a solution of pyridine (0.34 mL/g resin) and benzoyl chloride (0.34 mL/g resin) in DCM for 1 hour.

- ii) The resin was washed with DCM (3x10 mL) and DMF (3x10 mL) and after Fmoc removal DIPEA (0.52 mL, 3 eq., 3 mmol,), 4-nitrophenyl chloroformate (2 eq., 2.0 mmol, 402 mg) in 2 mL DCM were added to the H-Lys(Mtt)-resin (1eq, 0.5 mmol/g, 2 g) and the resin was agitated for 1 hour. Consecutively a Kaiser test was performed to check for completion (39).
- iii) Glutamic acid di-*tert*-butyl ester hydrochloride (3 eq., 3 mmol, 887.4 mg) and DIPEA (4 eq., 4 mmol, 0.70 mL) in DCM were added to the resin and the mixture was agitated for 1 hour. The resin was washed with DCM (3x10 mL) and DMF (3x10 mL).
- iv) After Alloc removal, Fmoc-3-(2-naphthyl)-L-alanine (Fmoc-Nal) was coupled using HATU.
- v) After Fmoc removal either Fmoc-(4-aminomethyl)benzoic acid (Fmoc-Amb) or trans-4-(aminomethyl)cyclohexane-1-carboxylic acid (Fmoc-Amc) was coupled using DIPCDI.
- vi) After Fmoc removal Fmoc-Lys(Alloc)-OH was coupled using DIPCDI.
- vii) After Fmoc removal Fmoc-Glu(OtBu)-OH was coupled using DIPCDI.
- viii) After Fmoc removal nicotinic acid was coupled using DIPCDI
- ix) After Alloc removal, Fmoc-Lys(Alloc) was coupled using DIPCDI.
- x) DIPEA (2 eq.) and DOTA-OSu were added to the resin in NMP and mixed on a bench roller at room temperature respectively for 6-8 hrs. Upon a negative Kaiser test the resin was washed with DMF (3x), DCM (3x), MeOH (3x) and diethyl ether (3x).
- xi) The peptide was cleaved from the resin with trifluoroacetic acid/H₂O (95:5, v/v) for two hours after which the resin was filtered off and the peptide was precipitated in diethyl ether. After drying in air the crude peptide was lyophilized from water.
- xii) After Fmoc removal, Fmoc-Gly-OH was coupled. This was repeated twice to couple two more glycines.
- xiii) After Fmoc removal N-succinimidyl S-acetylthioacetate (SATA, 3eq.) in DMF was added to the resin. Upon a negative Kaiser test (~45 minutes) the resin was washed with DMF (3x), DCM (3x), MeOH (3x) and diethyl ether (3x)

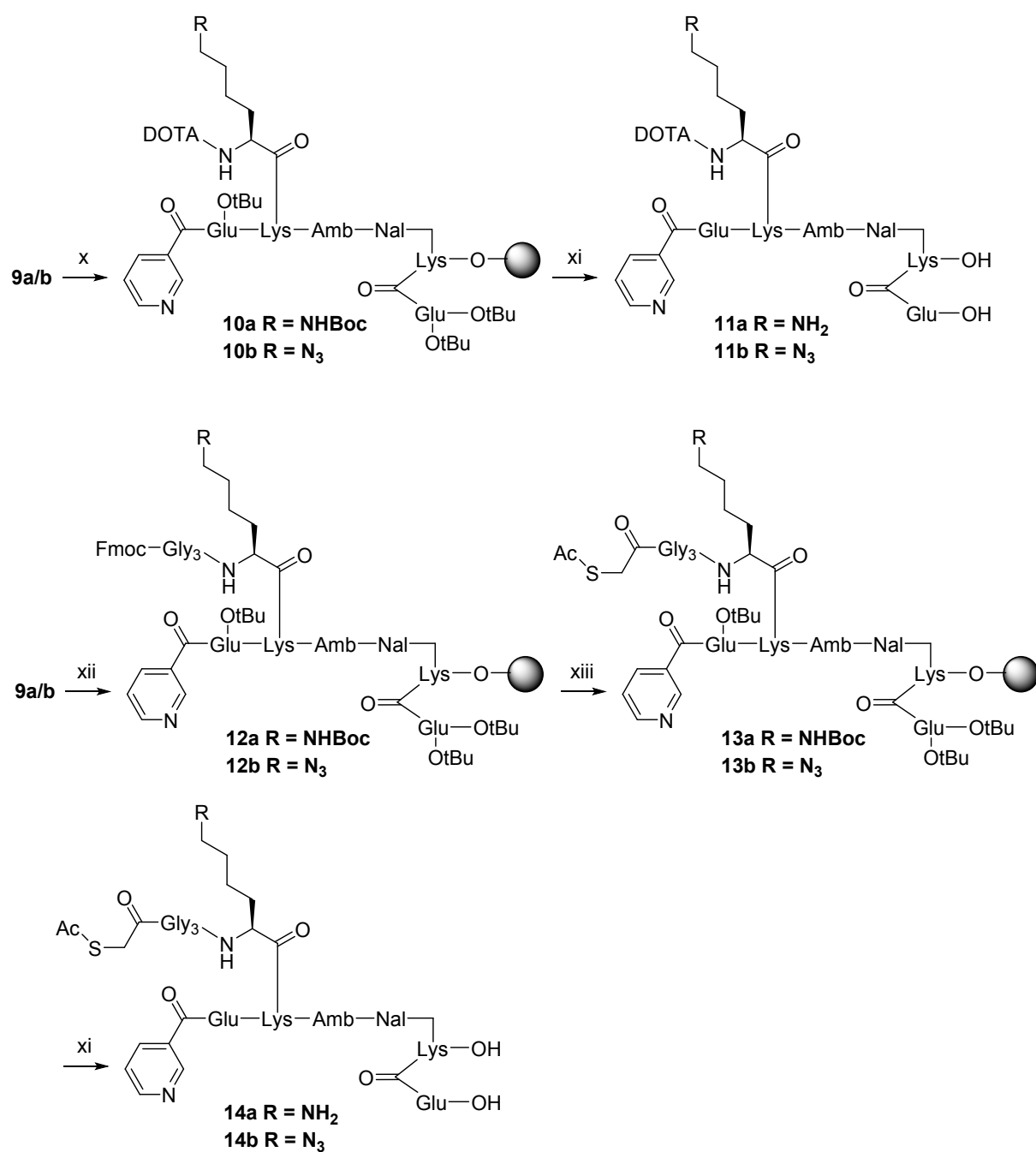


Fig. S2. Synthesis of peptide ligands without dye.

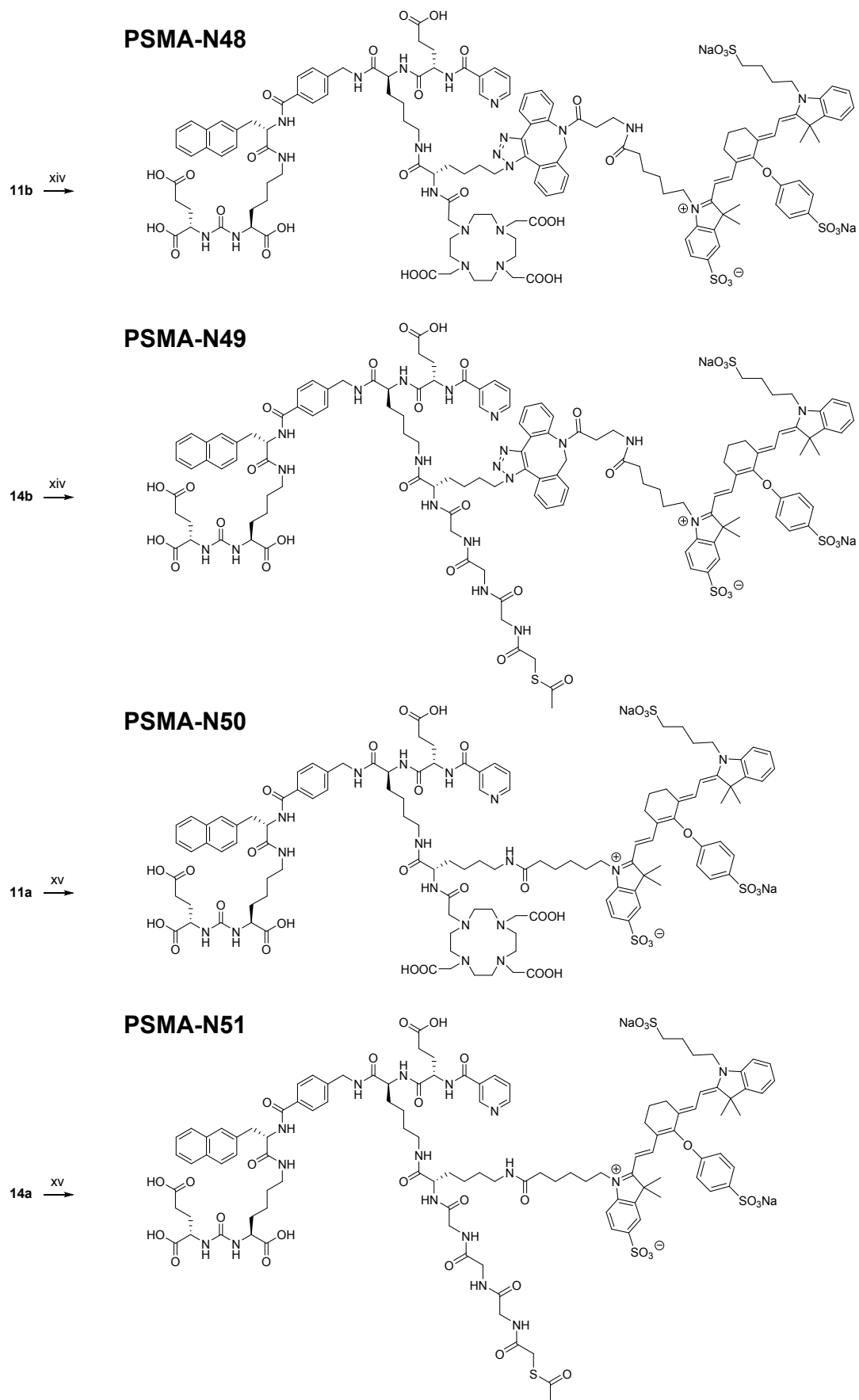


Fig. S3. Conjugation of IRDye800CW to the ligands.

Conjugation with IRDye800CW:

Peptide was dissolved in phosphate buffer (0.25 M, pH 8) after which the dye OSu ester (0.5-0.6 eq. in dry DMF) was added and shaken at rt for 4-6 hrs. The product was purified directly by preparative HPLC.

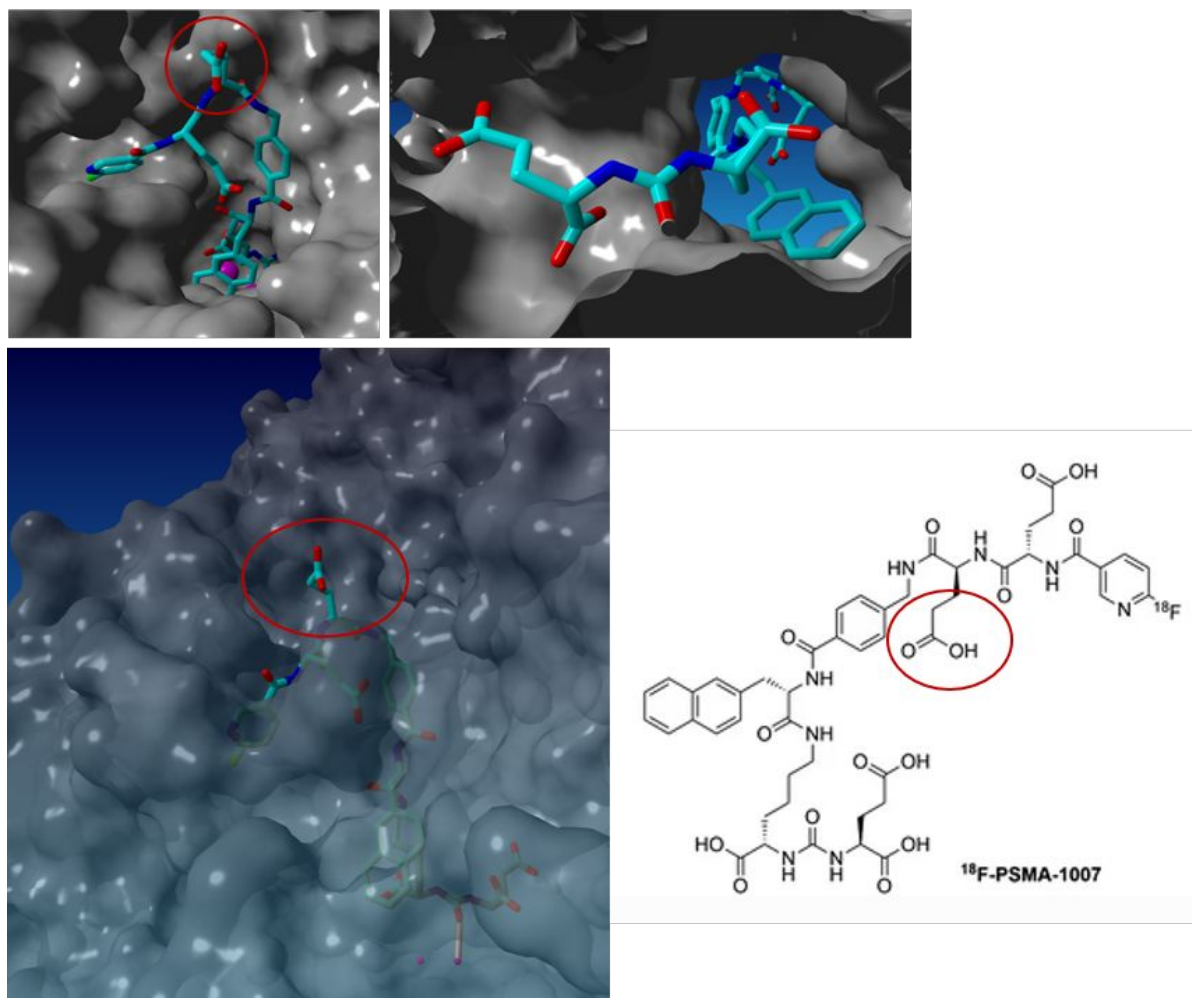
Analytical HPLC:

Compounds were analyzed on a Shimadzu LC-20A Prominence system with a dual UV-Vis detector (Shimadzu, 's Hertogenbosch, The Netherlands) equipped with a C18 Gemini-NX column, 150 × 3 mm, particle size 3 μm (Phenomenex, Utrecht, The Netherlands) Solvent A was 0.1% trifluoroacetic acid (TFA) in H₂O and solvent B was 0.1% TFA in acetonitrile (MeCN). A gradient of 5-100% acetonitrile (30 min.) was applied.

Preparative HPLC:

All compounds were purified on a Shimadzu dual-pump LC-20A Prominence system (Shimadzu, 's Hertogenbosch, The Netherlands) equipped with a C18 Gemini-NX column, 150 × 10 mm, particle size 10 μm (Phenomenex, Utrecht, The Netherlands), applying a gradient of 20-80% methanol in triethylammonium acetate buffer (10 mM, pH 7) for all IRDye containing compounds or a gradient of 5-100% acetonitrile in water (0.1% TFA) for all others.

Supplementary results



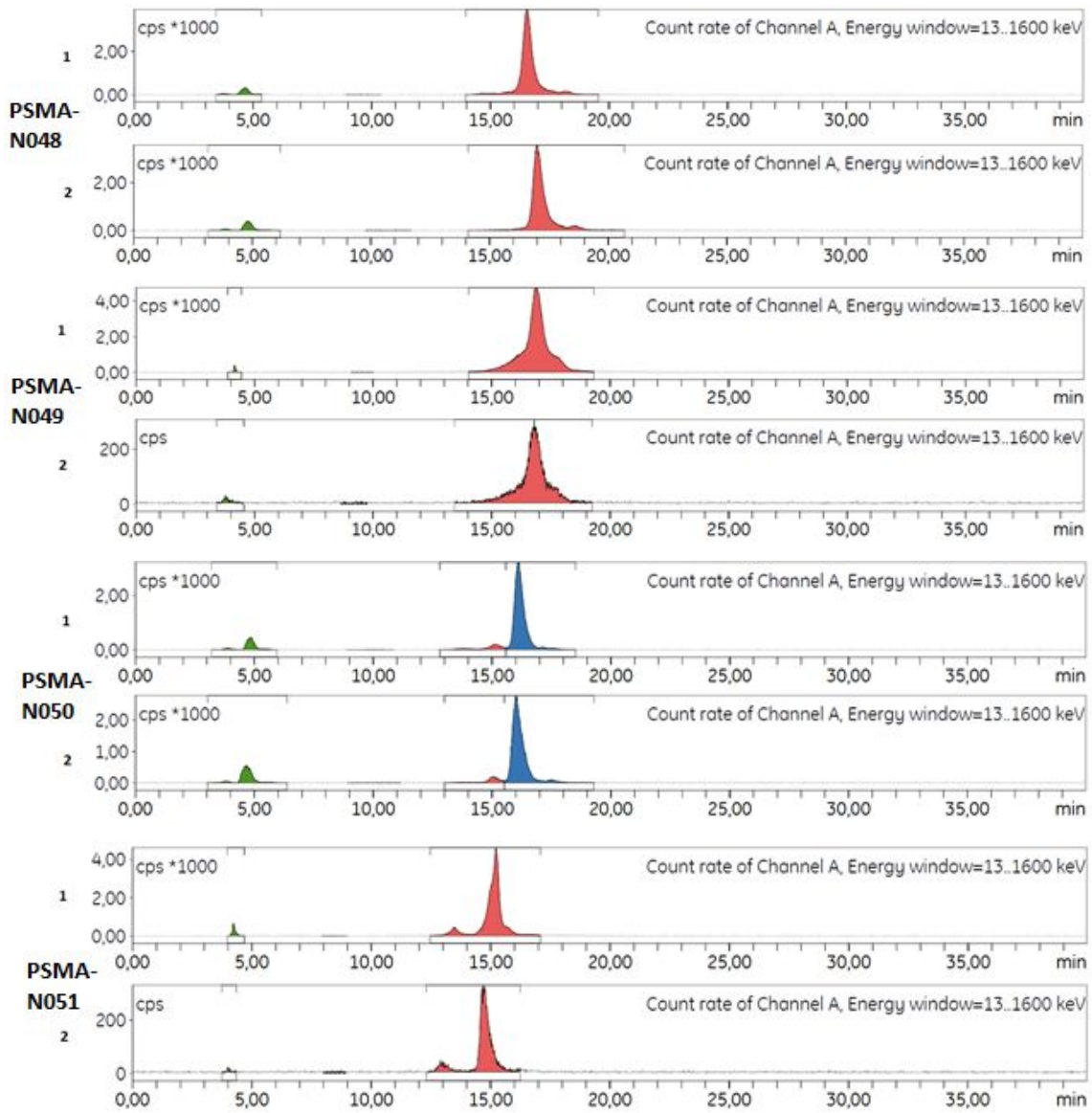


Fig. S5. Serum stability determination in human serum. Serum stability of four dual-labeled ligands was tested with use of RP-HPLC. Stability determination before (1) and after 2 hours of incubation in human serum (2).

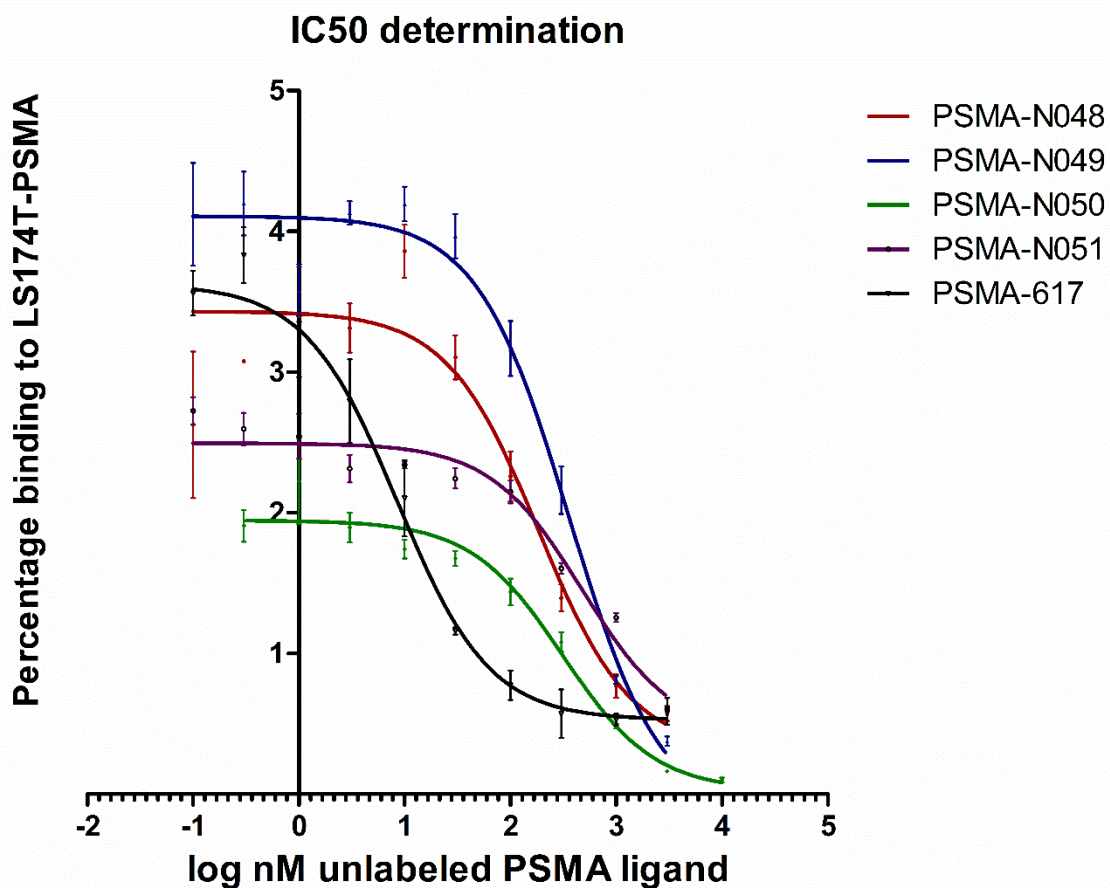


Fig. S6. IC₅₀ determination of PSMA-N048, PSMA-N049, PSMA-N050, PSMA-N051 and PSMA-617 in PSMA-positive LS174T cells. IC₅₀ graphs of PSMA-N048 (red), PSMA-N049 (blue), PSMA-N050 (green) and PSMA-N051 (purple). IC₅₀ is defined as the concentration required to inhibit the binding of clinically available ¹¹¹In-PSMA-617 by 50%. Data is expressed as percentage binding of PSMA-617 (y-axis) and the log scale of unlabeled ligand (cold, x-axis). PSMA-617 was used as a positive control (black).

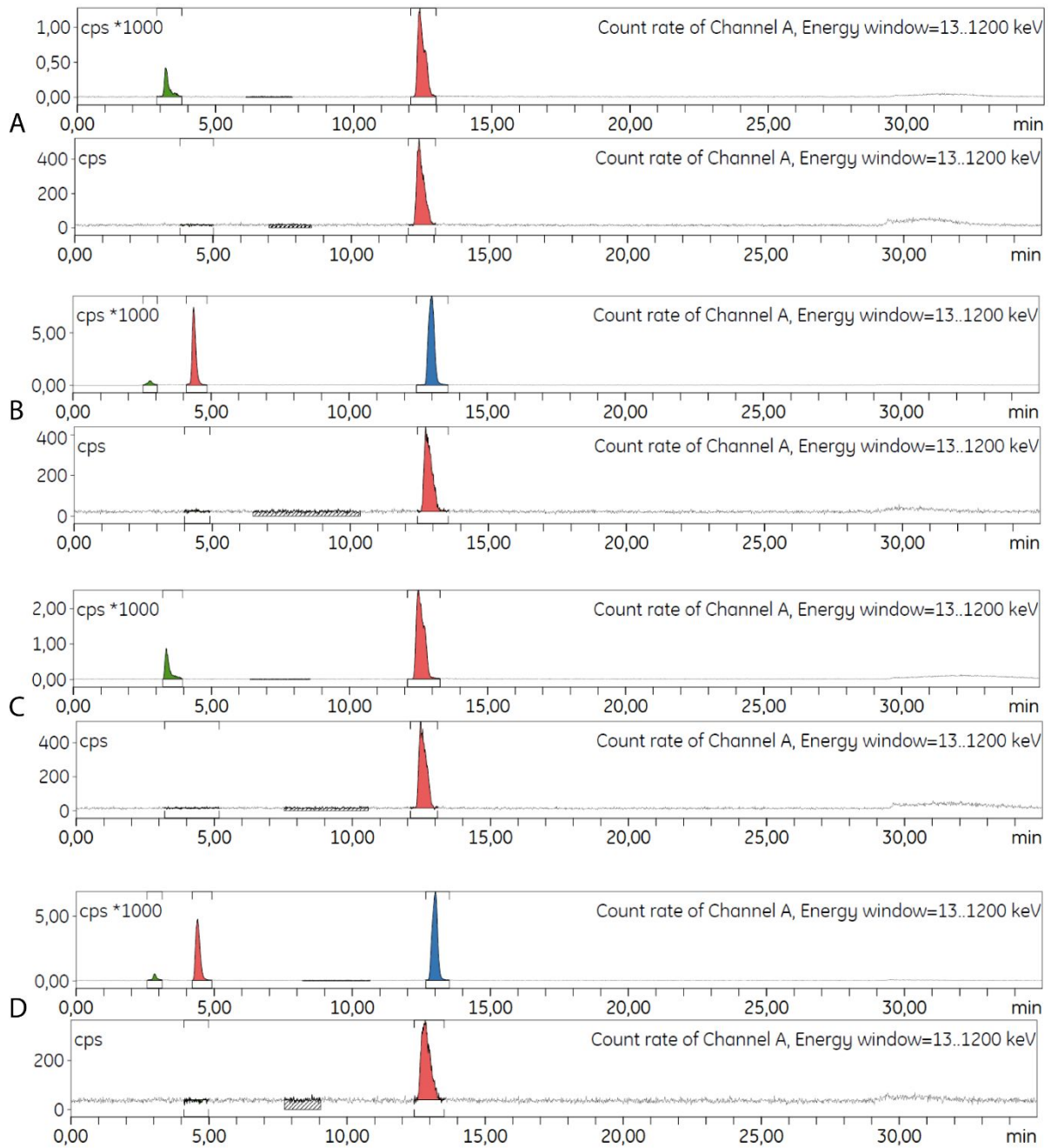


Fig. S7. Radio-HPLC before and after C18 purification. Radio-HPLC of (A) N48, (B) N49, (C) N50 and (D) N51 before and after Seppak C18 purification. Ligands were labeled with ^{111}In or $^{99\text{m}}\text{Tc}$ and purified before intravenous injection in mice.

Table S1. Pharmacokinetics of ^{99m}Tc-labeled PSMA-N049 and ¹¹¹In-labeled PSMA-N048 in mice bearing LS174T-PSMA and LS174T xenografts

	<i>PSMA-N049</i> 2h	<i>PSMA-N049</i> 4h	<i>PSMA-N049</i> 24h	<i>PSMA-N048</i> 2h	<i>PSMA-N048</i> 4h	<i>PSMA-N048</i> 24h
Biodistribution						
Blood	2.8 ± 0.5	1.6 ± 0.2	0.3 ± 0.1	1.8 ± 0.2	0.8 ± 0.1	0.1 ± 0.0
Muscle	0.6 ± 0.2	0.4 ± 0.1	0.1 ± 0.0	0.8 ± 0.1	0.3 ± 0.1	0.3 ± 0.0
Tumor- LS174T	1.7 ± 0.4	1.3 ± 0.2	0.6 ± 0.2	1.9 ± 0.4	1.4 ± 0.3	1.0 ± 0.4
Tumor+ LS174T-PSMA	9.0 ± 1.0	10.5 ± 1.7	8.9 ± 0.7	23.0 ± 4.2	22.8 ± 1.6	17.8 ± 1.6
Heart	1.2 ± 0.1	0.9 ± 0.1	0.2 ± 0.0	1.2 ± 0.3	0.8 ± 0.1	0.4 ± 0.1
Lung	2.8 ± 0.5	1.8 ± 0.2	0.5 ± 0.0	5.5 ± 1.6	2.4 ± 0.8	0.9 ± 0.4
Spleen	3.9 ± 0.4	3.3 ± 0.7	1.6 ± 0.4	3.8 ± 0.6	2.9 ± 0.5	2.0 ± 0.5
Pancreas	0.8 ± 0.4	0.6 ± 0.1	0.2 ± 0.0	0.8 ± 0.1	0.6 ± 0.1	0.3 ± 0.0
Liver	5.1 ± 0.5	4.4 ± 0.6	2.6 ± 1.3	2.8 ± 0.3	2.5 ± 0.2	2.2 ± 0.2
Stomach	1.5 ± 0.2	1.1 ± 0.1	0.4 ± 0.0	1.3 ± 0.2	0.8 ± 0.1	0.5 ± 0.0
Kidney	43.4 ± 2.3	60.5 ± 13.5	43.1 ± 7.2	103.4 ± 7.1	103.1 ± 5.6	70.0 ± 5.9
Adrenals	3.3 ± 0.6	2.9 ± 1.1	0.9 ± 0.3	3.7 ± 1.4	3.1 ± 0.3	1.9 ± 0.4
Duodenum	1.5 ± 0.3	1.0 ± 0.2	0.4 ± 0.0	1.1 ± 0.1	0.7 ± 0.1	0.4 ± 0.1
Prostate	1.1 ± 0.3	0.8 ± 0.8	0.4 ± 0.1	1.5 ± 0.4	0.9 ± 0.5	0.8 ± 0.2
Salivary glands	1.8 ± 0.1	1.4 ± 0.4	0.5 ± 0.1	1.9 ± 0.2	1.2 ± 0.1	1.1 ± 0.1
Bone marrow	1.8 ± 0.1	1.5 ± 0.3	1.1 ± 0.2	1.2 ± 0.4	1.0 ± 0.2	1.6 ± 0.4
Bone	1.0 ± 0.2	0.7 ± 0.1	0.4 ± 0.0	1.3 ± 0.3	0.8 ± 0.1	0.6 ± 0.0
Tumor/Blood	3.1 ± 0.4	6.1 ± 1.1	33.0 ± 8.7	12.8 ± 1.9	12.8 ± 1.9	262.4 ± 28.7
Tumor/Kidney	0.2 ± 0.0	0.2 ± 0.0	0.2 ± 0.0	0.2 ± 0.0	0.2 ± 0.0	0.3 ± 0.0
Tumor/ Tumor-	5.5 ± 1.2	7.4 ± 2.5	16.3 ± 4.7	14.3 ± 2.5	14.3 ± 2.5	20.7 ± 4.5
Tumor/Spleen	2.4 ± 0.4	3.3 ± 0.8	6.2 ± 2.2	6.2 ± 1.8	6.2 ± 1.8	9.8 ± 3.2
Tumor/Liver	1.7 ± 0.1	2.3 ± 0.5	3.8 ± 1.3	8.2 ± 1.0	8.2 ± 1.0	7.8 ± 1.3
Tumor/Salivary gland	4.6 ± 1.1	7.4 ± 1.9	17.8 ± 2.3	12.2 ± 2.2	12.2 ± 2.2	15.6 ± 2.0
Tumor/Prostate	9.2 ± 2.4	13.7 ± 4.2	24.7 ± 6.1	16.8 ± 4.2	16.8 ± 4.2	22.8 ± 5.6

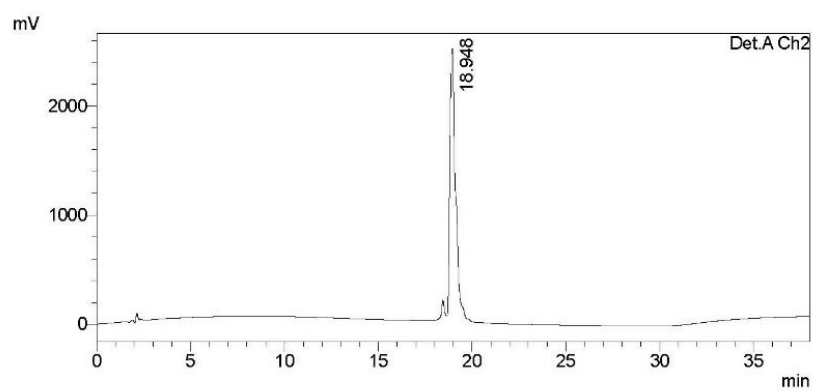
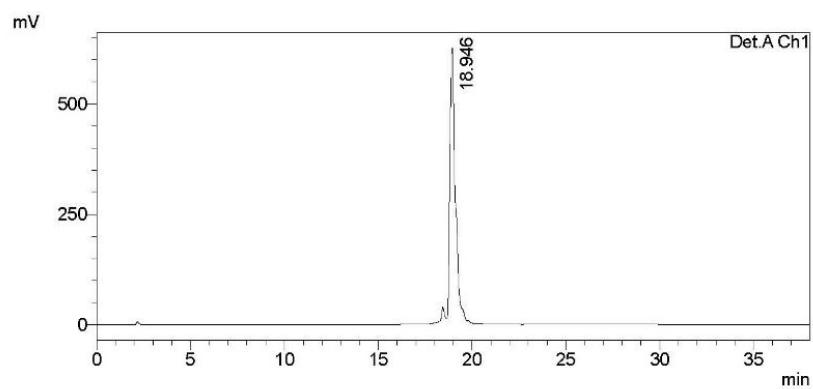
Table S2. Biodistribution of ¹¹¹In- or ^{99m}Tc labeled multimodal ligands in mice bearing LS174T-PSMA and LS174T xenografts 2 h p.i.

	<i>PSMA-N049</i>	<i>PSMA-N051</i>	<i>PSMA-N048</i>	<i>PSMA-N050</i>	<i>PSMA-617</i>
Biodistribution					
Blood	3.5 ± 1.6	2.1 ± 0.5	1.9 ± 0.2	1.1 ± 0.1	0.1 ± 0.02
Muscle	0.8 ± 0.1	0.7 ± 0.2	0.8 ± 0.2	0.4 ± 0.1	0.1 ± 0.1
Tumor- LS174T	1.9 ± 0.6	1.6 ± 1.4	2.1 ± 0.5	1.1 ± 0.3	0.2 ± 0.2
Tumor+ LS174T-PSMA	12.0 ± 1.4	17.7 ± 3.7	21.2 ± 1.2	25.3 ± 2.0	18.5 ± 1.9
Heart	1.6 ± 0.2	1.0 ± 0.2	1.0 ± 0.1	0.6 ± 0.1	0.03 ± 0.01
Lung	4.0 ± 0.4	1.9 ± 0.8	2.8 ± 0.4	1.6 ± 0.3	0.1 ± 0.1
Spleen	5.4 ± 0.6	5.8 ± 1.2	5.7 ± 0.8	5.5 ± 1.7	0.5 ± 0.2
Pancreas	1.0 ± 0.1	1.0 ± 0.6	0.8 ± 0.1	0.7 ± 0.2	0.1 ± 0.1
Liver	4.3 ± 0.3	1.7 ± 0.3	3.3 ± 0.3	1.2 ± 0.1	0.1 ± 0.02
Stomach	1.8 ± 0.2	1.0 ± 0.3	1.5 ± 0.2	0.8 ± 0.1	0.1 ± 0.1
Kidney	73.9 ± 5.6	81.6 ± 13.5	121.6 ± 8.2	129.9 ± 15.3	7.1 ± 2.6
Adrenals	5.8 ± 1.6	6.9 ± 2.7	8.7 ± 0.5	8.8 ± 2.4	1.4 ± 0.6
Duodenum	1.9 ± 0.2	1.1 ± 0.2	1.3 ± 0.3	0.8 ± 0.2	0.1 ± 0.1
Prostate	1.5 ± 0.3	1.3 ± 0.5	1.5 ± 0.6	1.4 ± 0.7	1.5 ± 3.1
Salivary glands	1.9 ± 0.1	1.4 ± 0.3	2.0 ± 0.2	1.0 ± 0.1	0.1 ± 0.02
Bone marrow	3.0 ± 1.7	2.2 ± 0.9	2.9 ± 1.0	1.4 ± 0.4	0.9 ± 1.3
Bone	1.3 ± 0.2	0.6 ± 0.1	1.6 ± 0.2	0.6 ± 0.1	0.3 ± 0.5
Tumor/Organ ratios					
Tumor/Blood	6.0 ± 7.0	8.4 ± 0.9	11.3 ± 0.8	22.8 ± 2.6	424.4 ± 173.9
Tumor/Kidney	0.16 ± 0.01	0.22 ± 0.04	0.18 ± 0.01	0.20 ± 0.02	3.0 ± 1.5
Tumor/Negative tumor	6.8 ± 2.1	15.2 ± 6.2	10.4 ± 2.4	24.3 ± 6.6	141.5 ± 52.8
Tumor/Spleen	2.2 ± 0.3	3.0 ± 0.1	3.8 ± 0.6	4.9 ± 1.3	43.5 ± 25.0
Tumor/Liver	2.8 ± 0.3	10.3 ± 1.4	6.4 ± 0.3	21.1 ± 1.6	266.6 ± 73.5
Tumor/Salivary gland	6.2 ± 0.6	12.9 ± 1.7	10.8 ± 1.1	24.6 ± 1.6	232.9 ± 68.5
Tumor/Prostate	8.3 ± 1.4	13.9 ± 2.5	16.7 ± 7.3	22.3 ± 10.2	140.0 ± 95.5

Fig. S8. HPLC and MALDI-ToF spectra of PSMA-N048, PSMA-N049, PSMA-N050 and PSMA-N051.

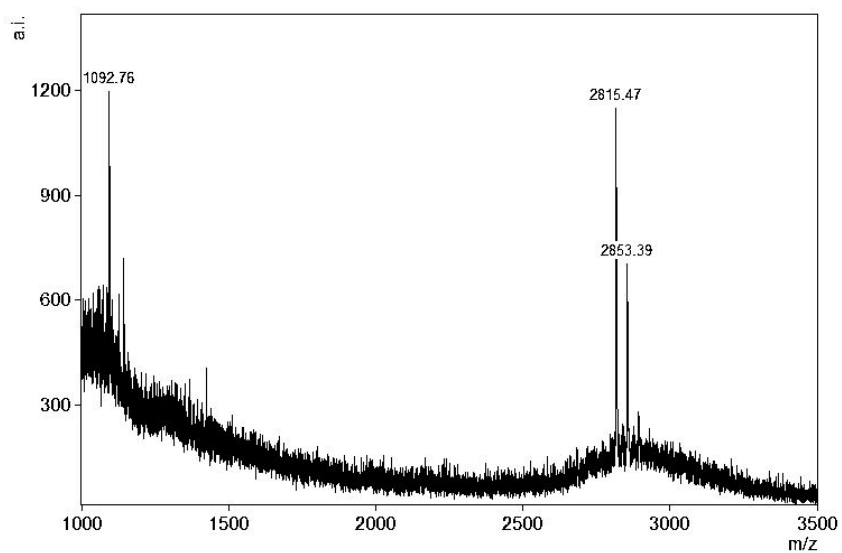
PSMA-N048

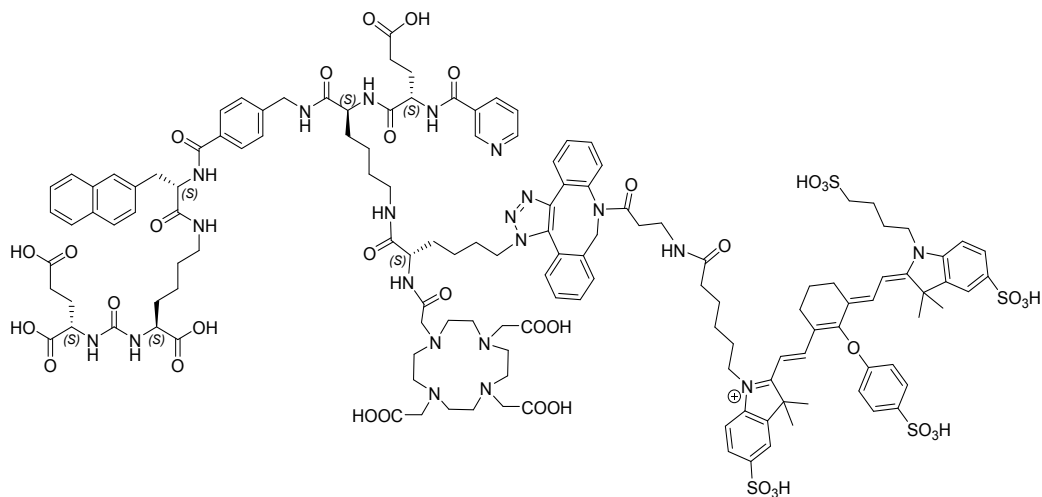
HPLC chromatogram



1 Det.A Ch1/260nm
2 Det.A Ch2/215nm

MALDI-ToF spectrum with matrix SA (Sinapinic acid)

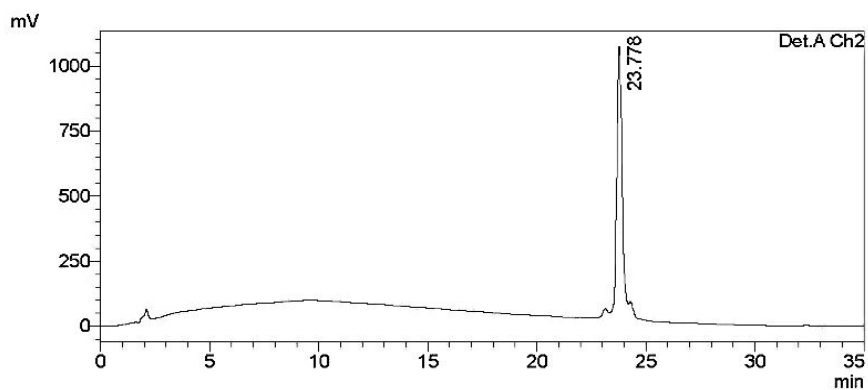
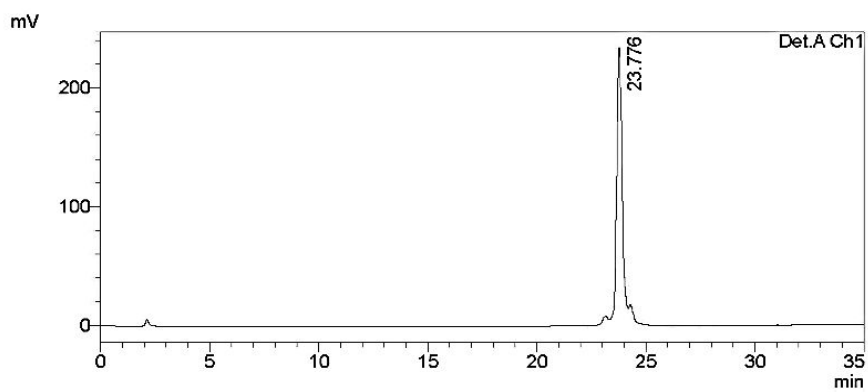




Chemical Formula: C₁₃₆H₁₆₆N₂₁O₃₇S₄⁺
 m/z: 2814.07 (100.0%), 2815.07 (73.0%), 2813.06 (68.0%), 2816.07 (24.5%)

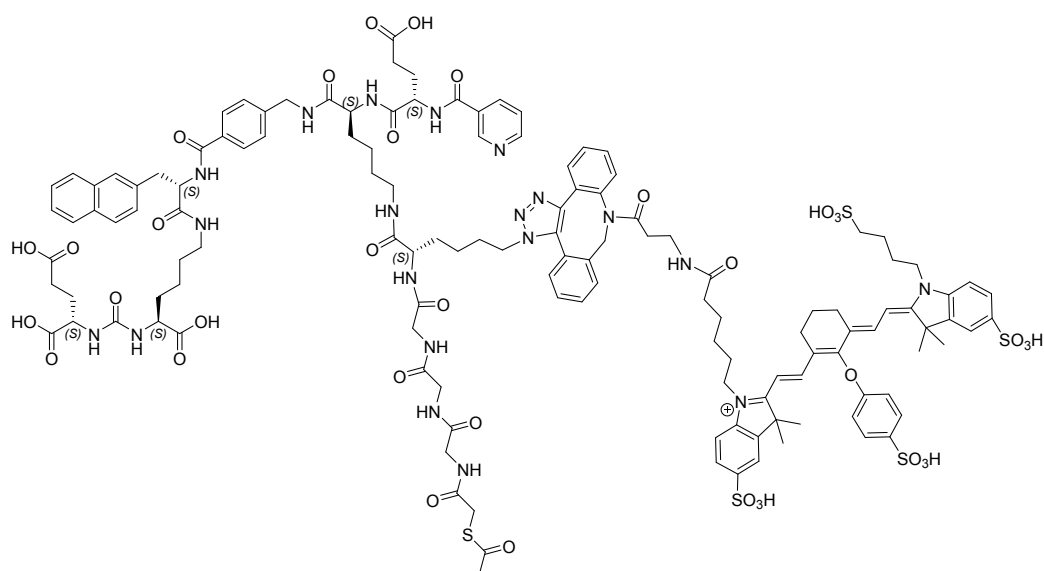
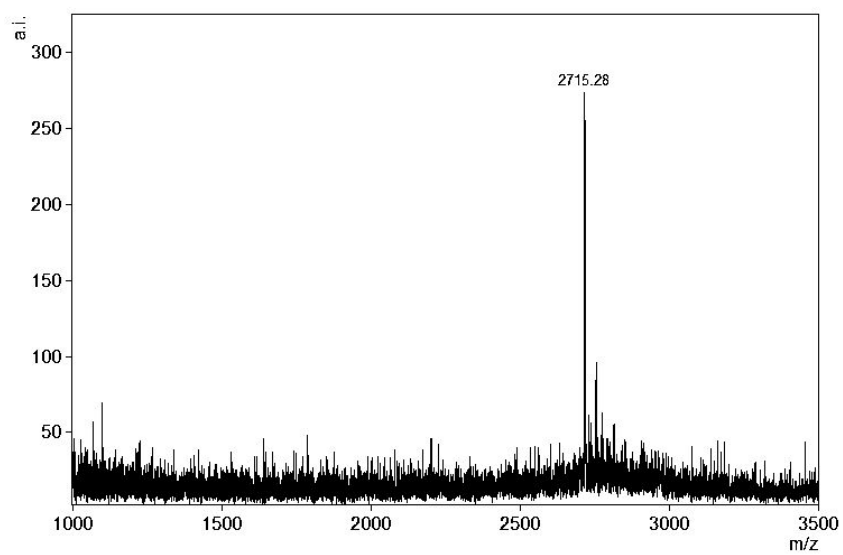
PSMA-N049

HPLC chromatogram



1 Det.A Ch1/260nm
 2 Det.A Ch2/215nm

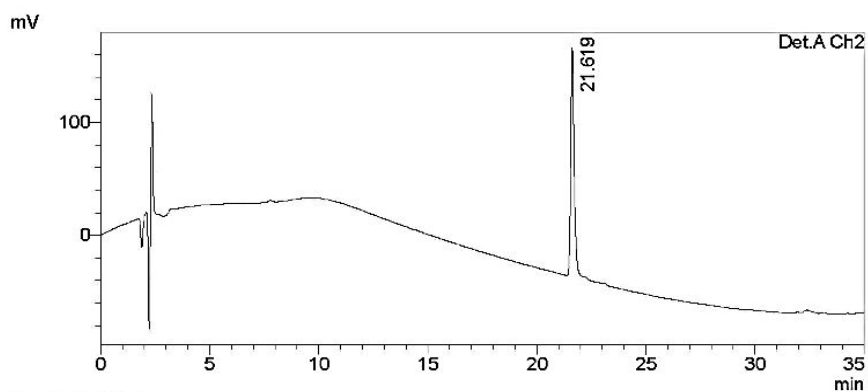
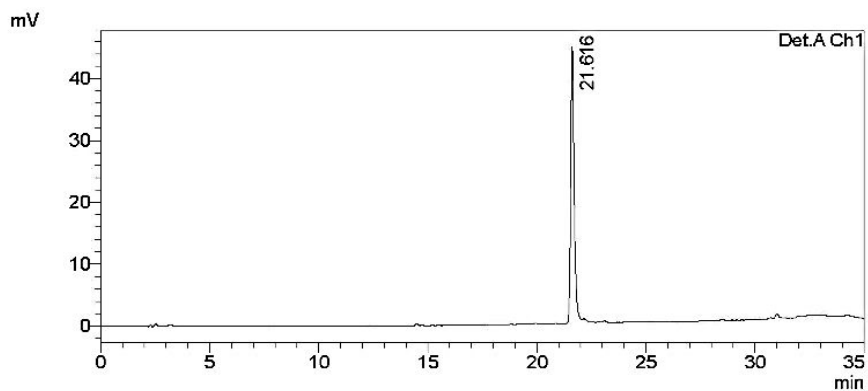
MALDI-ToF spectrum with matrix SA (Sinapinic acid)



Chemical Formula: C₁₃₀H₁₅₃N₂₀O₃₅S₅⁺
m/z: 2714.94 (100.0%), 2713.94 (71.1%), 2715.95 (39.4%), 2715.95 (30.4%), 2716.95 (23.7%), 2716.94 (22.6%)

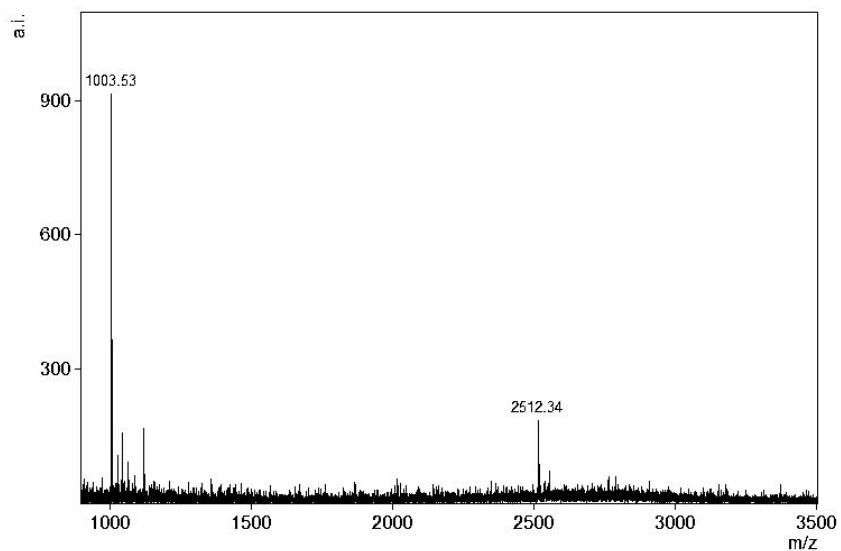
PSMA-N050

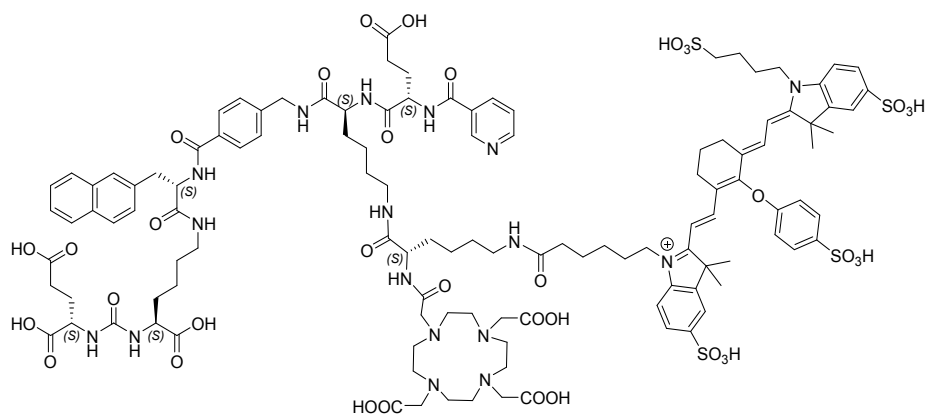
HPLC chromatogram



1 Det.A Ch1/260nm
2 Det.A Ch2/215nm

MALDI-ToF spectrum with matrix SA (Sinapinic acid)

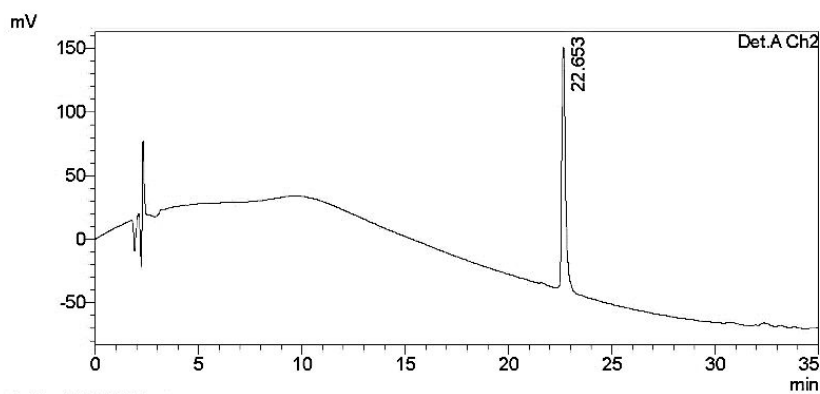
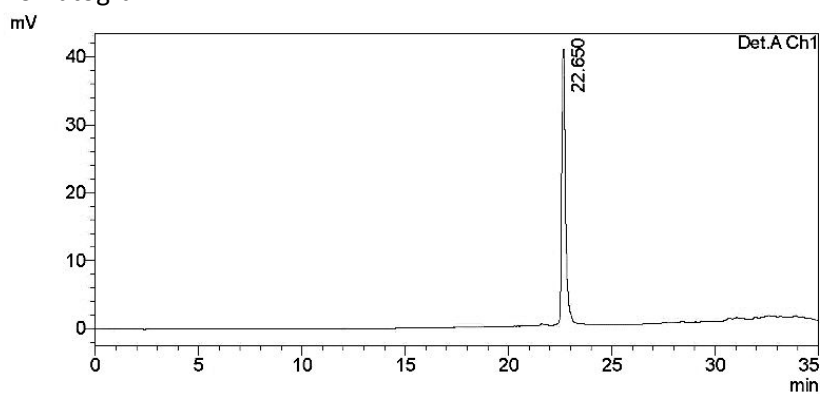




Chemical Formula: $C_{118}H_{152}N_{17}O_{36}S_4^+$
 m/z: 2511.95 (100.0%), 2510.95 (78.4%), 2512.95 (63.3%), 2513.95 (18.1%)

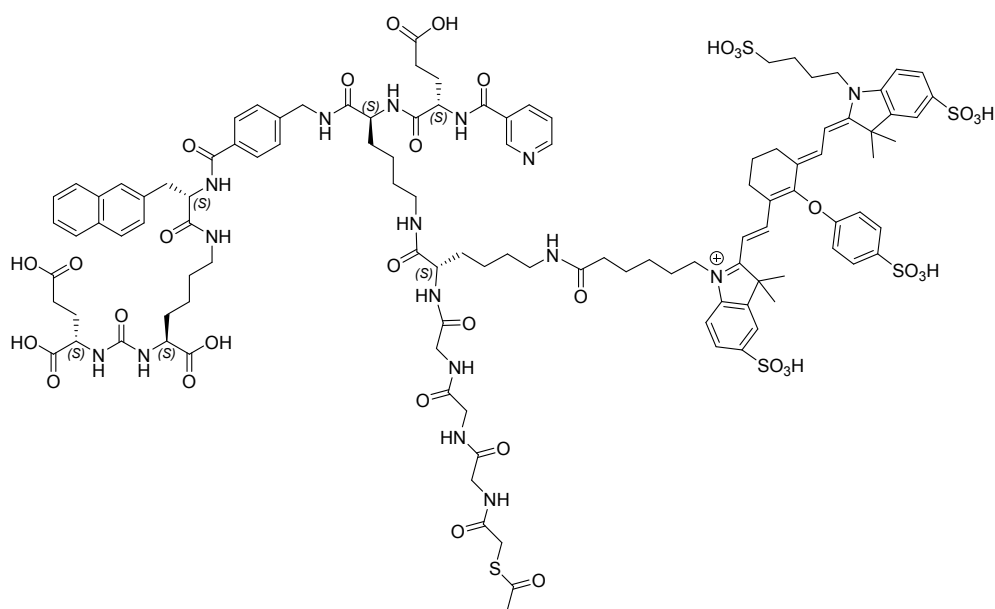
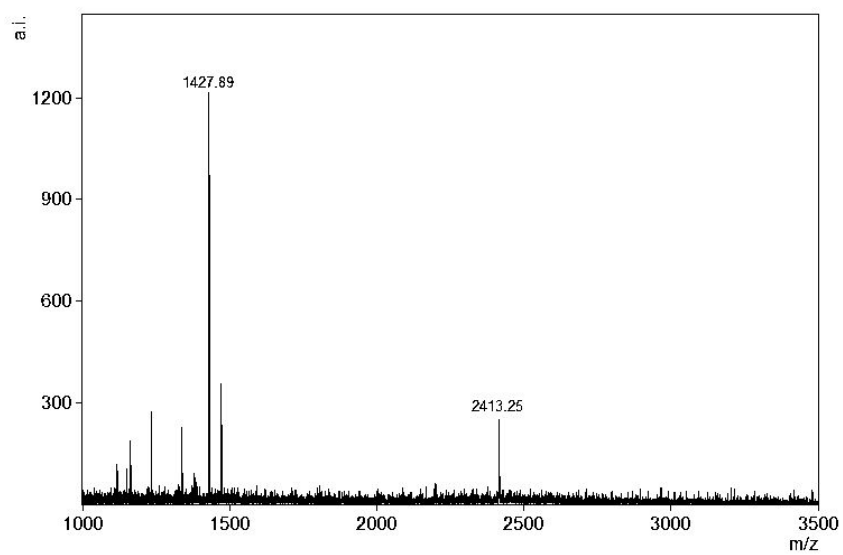
PSMA-N051

HPLC chromatogram



1 Det.A Ch1/260nm
 2 Det.A Ch2/215nm

MALDI-ToF spectrum with matrix SA (Sinapinic acid)



Chemical Formula: $C_{112}H_{139}N_{16}O_{34}S_5^+$
m/z: 2412.83 (100.0%), 2411.82 (82.6%), 2413.83 (35.3%), 2413.83 (24.7%), 2414.82 (22.6%)