

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection RT-qPCR data was collected with Applied Biosystems 7500
ELISPOT data was collected with CTL S6 Fluorocore (Immunospot)
Luminescence/Absorbance was collected with SoftMax Pro Software

Data analysis RT-qPCR data was analyzed with Applied Biosystems 7500
ELISPOT data was analyzed with Fluoro-X™ FluoroSpot (Immunospot)
GraphPad Prism 8.3.0 and Microsoft Excel were used for data and statistical analyses

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

All relevant data of this study are available from the corresponding authors upon request.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	Sample sizes included (a) n=6 hamsters/group and (b) n=6 agent-vaccinated NHP, n=3 empty vector-vaccinated NHP and n=3 mock-vaccinated NHP. There is not sufficient information about protective efficacy with these vaccine constructs to conduct a formal power analysis. For the hamster study, n=6 was sufficient to evaluate COH04S1 immunogenicity in a small animal model, based on previous studies in mice using the same vaccine (Chiuppesi 2020). For the AGM study, the group size of 3 and 6 was determined based on similar studies conducted by others with RSV (Le Nouen 2014; Cheng 2001; Taylor 2017). This number of animals represents the minimal number of animals necessary to evaluate vaccine replication relative to RSV and vaccine efficacy following SARS-CoV-2 challenge. An N of 3 AGM per group was used by Woolsey et al. (2020) to establish the suitability of the model for SARS-CoV-2 infection.
Data exclusions	No data were excluded. Day 5 nasal swab sgRNA and gRNA copies in COH04S1 vaccinated AGM (Fig. 5d-e) were not available at the time of submission.
Replication	Viral loads, pseudovirus neutralization were tested in triplicates. ELISPOT, ELISA and PRNT were tested in duplicates. Technical replicates presented minimal differences and were averaged.
Randomization	Sex was balanced when possible. Otherwise animals were randomly assigned to the groups.
Blinding	Animals were assigned a number and assays were performed with blinding when possible. In some assays, blinding to investigator and operators was not possible given the involvement in both sample preparation and analysis. Histopathology analysis was performed by a blinded (hamsters) or semi-blinded (NHP, mock-vaccinated animals were unblinded) board-certified Pathologist.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involvement in the study
<input type="checkbox"/>	<input checked="" type="checkbox"/> Antibodies
<input type="checkbox"/>	<input checked="" type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input type="checkbox"/>	<input checked="" type="checkbox"/> Animals and other organisms
<input type="checkbox"/>	<input checked="" type="checkbox"/> Human research participants
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern

Methods

n/a	Involvement in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Antibodies

Antibodies used	anti-Hamster IgG(H+L) IgG3 (Southern Biotech 6061-05), anti-Hamster IgG1 IgG3 (Southern Biotech 1940-05), anti-HamsterIgG2/IgG3 IgG3 (Southern Biotech 1935-05), Goat anti-Monkey IgG (H+L) s (Thermo Fisher PA1-84631)
Validation	Southern Biotech 6061-05, 1940-05 and 1935-05 were validated by the vendor by ELISA.

Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)	Vero E6 cells (ATCC, CRL-1586), HEK293T/17 cells (ATCC CRL11268), HEK293T-ACE2 cells (J.D.Bloom lab), Vero TMPRSS2 cells (Vaccine Research Center-NIAID), Vero E6-hACE2 cells (BEI Resources NR-53726)
Authentication	None of the cell lines were validated in-house. ACE2 expression on ACE2+ cells was periodically verified by FACS.

Mycoplasma contamination

Cell lines were not tested for Mycoplasma contamination in house.

Commonly misidentified lines
(See [ICLAC](#) register)

No commonly misidentified lines were used in the study.

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals

15 male and 15 female Syrian golden hamsters (Envigo), 6-8 weeks old. 4 male and 20 female research naive, adult, African green monkeys, 3-7Kg purchased from a Bioqual's approved vendor.

Wild animals

No wild animals were used in the study

Field-collected samples

The study did not involve field-collected samples

Ethics oversight

The study was approved by: Bioqual (protocols 20-163 and 20-120) and City of Hope (protocols 20087 and 20075) Institutional Animal Care and Use Committees (IACUC)

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Human research participants

Policy information about [studies involving human research participants](#)

Population characteristics

Male and female healthcare workers >18 years of age were enrolled in a observational study to evaluate humoral and cellular immunity to COVID-19 EUA vaccines

Recruitment

Volunteers signed informed consent

Ethics oversight

Protocol 20720 was approved by by the City of Hope (COH) Institutional Review Board (IRB)

Note that full information on the approval of the study protocol must also be provided in the manuscript.