### **Supporting Information for**

## **Original article**

## [<sup>18</sup>F]MAGL-4-11 positron emission tomography molecular imaging of monoacylglycerol lipase changes in preclinical liver fibrosis models

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#### **Supplementary method**

# Radiosynthesis of [<sup>18</sup>F]MAGL-4-11 ((4-(3-(fluoro-<sup>18</sup>F)pyrrolidin-1-yl)benzoyl)azetidin-3-yl)piperazin-1-yl)(thiazol-2-yl)methanone)

The general labeling procedure for [<sup>18</sup>F]MAGL-4-11 formation was described previously<sup>1</sup>. The cyclotron-produced [<sup>18</sup>F]  $F^-$  was separated from H<sub>2</sub><sup>18</sup>O using the Sep-Pak Accell Plus QMA Plus Light cartridge (Waters; Milford, MA, USA). The produced  $[^{18}F]F^{-}$  was eluted from the cartridge with a mixture of aqueous K<sub>2</sub>CO<sub>3</sub> (4 mg in 200 DMSO) a of 4,7,13,16,21,24-hexaoxa-1,10μL and solution diazabicyclo[8,8,8]hexacosane (Kryptofix222; 7.5 mg) in CH<sub>3</sub>CN (200 µL), and transferred to a reaction vessel in the hot cell. After drying [<sup>18</sup>F]KF solution at 120 °C for 30 min to remove water and CH<sub>3</sub>CN, a solution of mesylate precursor (2.0 mg) in anhydrous DMSO (300 µL) was then added. The vessel was heated at 120 °C for 10 min, then diluted with HPCL mobile phase (500 µL), followed by injection into an HPLC column. HPLC purification was performed on an X Bridge Prep C18 column  $(10 \times 250 \text{ mm}, 5 \text{ }\mu\text{m})$  using a mobile phase of CH<sub>3</sub>CN/H<sub>2</sub>O/Et<sub>3</sub>N (30/70/0.1) at a flowrate of 5.0 mL/min. The radioactive fraction corresponding to the desired product was collected in a sterile flask, evaporated to dryness in vacuo, and reformulated in PBS containing 5% EtOH. The synthesis time was ca. 70 min from end of bombardment. Radiochemical and chemical purity were measured by an analytical HPLC (X Bridge Prep C18 column ( $4.6 \times 250$  mm, 5 µm) using a mobile phase of CH<sub>3</sub>CN / H<sub>2</sub>O + 0.1% Et<sub>3</sub>N (30/70) at a flow rate of 1.0 mL/min. The product identity was confirmed by the co-injection with unlabeled MAGL-4-11. Radiochemical yield was  $39.3 \pm 13.7\%$  (n = 5) decay-corrected based on  $[^{18}F]F^-$  with >99% radiochemical purity, and the molar activity was 111.8-328.7 GBq/µmol (3.03-8.87 Ci/µmol) (Fig. S3).

Pts ID	Fibrosis Grades	Age (yr)	Sex	Race	Ethnicity
Pts 1	F0	65	М	White	Non-Hispanic
Pts 2	F0	72	М	White	Non-Hispanic
Pts 3	F0	65	М	White	Non-Hispanic
Pts 4	F1	84	М	White	Non-Hispanic
Pts 5	F1	60	М	White	Non-Hispanic
Pts 6	F1	64	М	White	Non-Hispanic
Pts 7	F2	54	М	White	Non-Hispanic
Pts 8	F2	56	М	White	Non-Hispanic
Pts 9	F2	58	F	White	Non-Hispanic

### Table S1. Characteristics of the liver fibrosis patients

## **Table S2** Primers used for qPCR.

Gene name	Forward sequence 5'-> 3'	Reverse sequence 5'-> 3'		
CB1	CACCTTCCGCACCATCACCAC	GTCTCCCGCAGTCATCTTCTCTTG		
DAGLα	AGAATGTCACCCTCGGAATGG	GTGGCTCTCAGCTTGACAAAGG		
MAGL	CAAGGCCCTCATCTTTGTGT	ACGTGGAAGTCAGACACTAC		
FAAH	CCCAGATGGAACATTACAGG	CAGGATGACTGGTTTTCAGG		
NAPE-PLD	CACGGTAATGGTGGAAATGG	GTCCAGATGGTCATAGTGGTTG		
18s rRNA	GGGAGCCTGAGAAACGG	GGGTCGGGAGTGGGTAATTT		



**Figure S1** Hepatic panleukocyte (CD45) and macrophage (CD68) expression after CCl<sub>4</sub> (A) and BDL (B) treatment.



**Figure S2** Coronal and sagittal images of [<sup>18</sup>F]MAGL-4-11/PET-CT imaging hepatic MAGL expression with mild and severe CCl<sub>4</sub> (A) and BDL (B) mice.



Scheme S1 Synthesis of irreversible MAGL inhibitor (cold compound) MAGL-4-11. Conditions: (i) DIPEA, MeCN, 80 °C for 12 h; 85% yield; (ii) TFA, CH<sub>2</sub>Cl<sub>2</sub>, rt, 12 h; 99% yield; (iii) HOBT, EDC·HCl, Et<sub>3</sub>N, DMF, rt, 12 h; 78% yield; (iv) 1-chloroethyl chloroformate, CH<sub>2</sub>Cl<sub>2</sub>, rt, 2 h; then MeOH, 35 °C 2 h; 86% yield; (v) pyrrolidin-3-ol hydrochloride, K<sub>2</sub>CO<sub>3</sub>, DMSO, 120 °C for 24 h; 79% yield; (vi) MsCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, rt, overnight; 84% yield; (vii) TBAF, THF, 70 °C, 2 h; 38% yield; (viii) LiOH, THF/MeOH/H<sub>2</sub>O, 40 °C, 16 h; 92% yield; (ix) HOBT, EDC·HCl, Et<sub>3</sub>N, DMF, rt, 12 h; 30% yield; DIPEA = N,N-diisopropylethylamine; TFA = trifluoroacetic acid; HOBT = 1-hydroxybenzotriazole hydrate;  $EDC \cdot HCl = N \cdot (3 - dimethylaminopropyl) \cdot N'$ ethylcarbodiimide hydrochloride;  $Et_3N = triethylamine;$ DMF = N, Ndimethylformamide; DMSO = methyl sulfoxide; MsCl = methanesulfonyl chloride; TBAF = tetrabutylammonium fluoride.



Scheme S2 Synthesis of MAGL PET tracer [<sup>18</sup>F]MAGL-4-11. Conditions: (i) LiOH, THF/MeOH/H<sub>2</sub>O, 40 °C, overnight; 97% yield; (iI) HOBT, EDC•HCl, Et<sub>3</sub>N, DMF, rt, 12 h; 22% yield; (iii) MsCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, rt, overnight; 70% yield; (vi) [<sup>18</sup>F]KF, DMSO, 120 °C, 10 min, 39% decay-corrected RCY. HOBT = 1-hydroxybenzotriazole hydrate; EDC·HCl = N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride; DMF = N,N-dimethylformamide; DMSO = methyl sulfoxide; MsCl = methanesulfonyl chloride.

### Reference

1. Chen Z, Mori W, Deng X, Cheng R, Ogasawara D, Zhang G, et al. Design, synthesis, and evaluation of reversible and irreversible monoacylglycerol lipase positron emission tomography (PET) tracers using a "tail switching" strategy on a piperazinyl azetidine skeleton. *J Med Chem* 2019;**62**:3336-53.