# nature portfolio

Corresponding author(s):	Erwin J. G. Peterman
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## **Reporting Summary**

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

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For	all st	atistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Cor	nfirmed
	x	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
x		A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
×		The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
X		A description of all covariates tested
x		A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	x	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
×		For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
×		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
x		For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
x		Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
	'	Our web collection on statistics for biologists contains articles on many of the points above.

### Software and code

Policy information about availability of computer code

Data collection We used a custom written LabVIEW program (LV 2011, as used previously in Sarlós, K. et al., Nat. Struct. Mol. Biol. 25, 868 (2018)) to record FD-curves and MicroManager to record fluorescence images.

Data analysis

We used a custom written LabVIEW program (LV 2011, see above) to extract FD-curves, OriginPro 2016 to analyze FD-curves, a custom written MATLAB program (ML 2015b, provided in the Supplementary Information) to determine single gate opening step sizes and ImageJ codes publicly available on https://imagej.nih.gov/ij/ to analyze fluorescence data.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

#### Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The minimum data set required to reproduce this work are in the main manuscript and Supplementary Information. For all relevant, representative data, we additionally provide three independent examples in the Supplementary Information. Source data are provided with this paper.

Field-spe	ecific reporting
Please select the o	one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.
<b>x</b> Life sciences	Behavioural & social sciences Ecological, evolutionary & environmental sciences
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Life scie	nces study design
All studies must d	isclose on these points even when the disclosure is negative.
Sample size	Minimum sample size is stated in every caption and was determined based on the number of tethered DNA molecules successfully obtained.
Data exclusions	No data were excluded.
Replication	Every experiment, from which a main text or SI figure was derived, was repeated at least on three separate measurement days (buffers and samples were prepared freshly on each day), spread over a minimum of 2 weeks. All reported data were reproducible.
Randomization	This does not apply in our case, since data in single molecule studies are inherently randomized.
Blinding	This does not apply in our case, since -other than a step-fitting algorithm- we did not fit any data, but instead use differences of the raw data

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems		Methods	
n/a Inv	volved in the study	n/a	Involved in the study
x	Antibodies	×	ChIP-seq
x	Eukaryotic cell lines	×	Flow cytometry
x	Palaeontology and archaeology	×	MRI-based neuroimaging
x	Animals and other organisms		
x	Human research participants		
x	Clinical data		
x	Dual use research of concern		

in the presence of absence of certain reagents to draw conclusions.