Induction of tier-2 neutralizing antibodies in mice with a DNA-encoded HIV envelope native like trimer

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Supplementary Figure 1. Full antigenic profile of *in vitro* expressed MD39 vs BG505.gp120_foldon (related to Figure 1). (A) Binding of MD39 to HIV bNAbs PGT145, PGT151, VRC34.01, 3BNC117, 10-1074 or non-NAbs F425, 3074, 4025, 17B. (B) Binding of BG505.gp120_foldon to HIV bNAbs and non-NAbs as in (A). (C) Quantification of binding of MD39 or BG505.gp120_foldon to HIV bNAbs or non-NAbs in terms of Area Under Curve (AUC). (D and E) Binding of *in vivo* produced MD39 (blue) and GP120-foldon (red) to bNAbs PGT145, 3BNC117, PGT151, 10-1074 and VRC34.01 (D) and non-NAbs 3074, F425, 4025 and 17b (E). (F) Normalized area under the curve (ratio of *in vitro* produced pMD39 binding versus pGP120-foldon binding) for binding to HIV bNAbs and non-NAbs.



Supplementary Figure 2. ICS Gating scheme. Gating scheme used to determine cytokine positive CD4⁺ or CD8⁺ T cells following peptide stimulation (related to Figure 2 and 3).



Supplementary Figure 3. DNA-encoded BG505.MD39 but not protein BG505.MD39 induced polyfunctional T-cell responses in BALB/c mice (related to Figure 2). Mice were immunized with 25ug of either DNA-encoded BG505.MD39 or RIBI-co-formulated protein BG505.MD39 at weeks 0, 3, 6 and euthanized at week 8 for cellular analysis. (A - B) ICS analyses of co-expression of IFN γ , TNF α and IL-2 cytokines in CD4⁺ (A) or CD8⁺ (B) T cells following stimulation with BG505 Envelope specific peptide pools. Two independent experiments were performed for each panel in the figure with similar findings. N=10 mice/group. Error bar represents standard deviation. Center of the error bar represents the mean. Source data are provided as a Source Data file.



Supplementary Figure 4. DNA-encoded BG505.MD39 vaccination induced both Tfh and GC B cell responses in BALB/c mice (related to Figure 2). Mice were immunized with 25ug of DNA-encoded BG505.MD39 on Day 0 and euthanized sequentially on Days 4, 7 and 10 for analyses of Tfh and GC B cells. (A) Comparison of flow plots of CXCR5+PD1+ cells amongst CD4+CD44+ cells in the iliac, inguinal, popliteal and spleen of naïve mice versus mice immunized with DNA-encoded BG505.MD39 10 d.p.i. (B) Kinetics of Tfh and GC B cell responses in the draining lymph nodes or spleens of mice vaccinated with DNA-encoded

BG505.MD39. One independent experiment was performed for each panel in the figure. Each line represents an animal. N=3 mice/group. Source data are provided as a Source Data file.



Supplementary Figure 5. Long immunization scheme with DNA-encoded BG505.MD39 improved polyfunctional CD4⁺ T-cell responses, while short immunization scheme improved polyfunctional CD8⁺ T-cell responses in mice (related to Figure 3). Mice in this panel were immunized either with a short scheme (Wks 0, 3, 6) or a long scheme (Wks 0, 3, and 16) with 25ug DNA used at each vaccination. The mice were euthanized 2 weeks post the final vaccination for cellular analyses. (A - B) ICS analyses of co-expression of IFN γ , TNF α and IL-2 cytokines in CD4⁺ (A) or CD8⁺ (B) T cells following stimulation with BG505 Envelope specific peptide pools in mice immunized with DNA-encoded BG505.MD39 using either the short or the long scheme. Two independent experiments were performed for each panel in the figure with similar findings. N=5 mice/group. Error bar represents standard deviation. Center of the error bar represents the mean. Source data are provided as a Source Data file.



Supplementary Figure 6. Sequence features of recovered murine antibody clones (related to Figure 5). (A) Demonstration of the barcode frequencies by clones (Left) or IGH HCDR3 identities (Right). (B) HCDR3 features of the 25 recovered MAb clones (left) or the 6 clones with IGHV6-6 germline VH gene usage (Right).





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Supplementary Figure 7. Further characterizations of BG505.MD39-specific murine antibody clones (related to Figure 6). (A-C) Binding of all isolated murine mAb clones to BG505.MD39 (A) or gp120_foldon (B) as determined by ELISA, and respective measured EC₅₀ values (C). (D) Reducing SDS PAGE analysis of BG505.MD39-specific MAb clones C05, C22, C24, C33, and C37 in comparison with PGT128 or backbone pVAX vector transfection supernatants. (E) Determination of the antibody concentrations from the transfection supernatants based on BG505.MD39 binding in comparison with purified protein standards. (F) Neutralization of autologous BG505.T332N virus by PGT128 (positive control), C33, and murine IgG2A antibody TA99 (negative control) starting at 50ug/mL. (G) Comparison of neutralization potential of PGT128 with BG505.T332N pseudo-virus versus BG505.T332N.T465N pseudo-virus. For (D), the experiments were repeated twice.



Supplementary Figure 8. Additional sequence features of recovered murine antibody clones (related to Figure 6). Comparisons of the heavy and light chain sequences of the 5 BG505.MD39-specific MAb clones by number of mutations (A) and positions for the mutations (B).

C24 Competition ELISA



C33 Competition ELISA



Supplementary Figure 9. Assessment of epitope specificity and affinities of C24 and C33 by competition ELISA and SPR (related to Figure 6). (A) Degree of binding of biotinylated C24 to MD39 in the presence of varying concentrations of non-biotinylated C05, C22, C24, C33, C37 and control buffer. (B) Degree of blocking of biotinylated C24 binding to MD39 by each of the aforementioned antibodies, as calculated

from (A). (C) Degree of binding of biotinylated C33 to MD39 in the presence of varying concentrations of non-biotinylated C05, C22, C24, C33, C37 and control buffer. (D) Degree of blocking of biotinylated C33 binding to MD39 by each of the aforementioned antibodies, as calculated from (C). (E and F) SPR trace to determine the binding and dissociation pattern of C24 (E) and C33 (F) to GP120 (to preserve 1:1 binding ratio). For (A-D), two technical replicates were determined for each experimental condition; error bar represents standard deviation. Source data are provided as a Source Data file.



Supplementary Figure 10. Cryo-EM data processing flow diagram resulting in a density map at 3.8Å global resolution (FSC 0.143 consistency between two independently refined data half sets) (related to Figure 7). Flow diagram comprises panels of representative map densities with corresponding atomic models superimposed as well as a standard Fourier shell correlation plot (FSC plot). MD39 is shown in grey; C05 Fab is shown in dark red.



Supplementary Figure 11. Computational modeling of murine mAbs bound to NLT. Comparison of the angle of approach in terms of binding to NLT between murine and NHP V5-directed mAbs (related to Figure 7).

Supplementary Table 1. Comparison of neutralizing antibody titers in mice immunized with DNAencoded NLT versus protein MD39. BG505.T332N or MLV ID50 neutralization titers in the sera of animals immunized with 25ug RIBI co-formulated protein BG505.MD39 or DNA-encoded BG505.MD39 at Weeks 0, 3, 6 two weeks post the final vaccination.

	Mouse ID	BG505.T332N	MLV
Protein	1	<45	<45
	2	<45	<45
	3	<45	<45
	4	<45	<45
	5	<45	<45
	6	<45	<45
	7	<45	<45
	8	<45	<45
	9	<45	<45
	10	<45	<45
DNA	1	<45	<45
	2	<45	<45
	3	232	<45
	4	<45	<45
	5	<45	<45
	6	73	<45
	7	50	<45
	8	<45	<45
	9	<45	<45
	10	<45	<45

Supplementary Table 2. Comparison of neutralizing antibody titers in mice immunized with DNAencoded NLT using a short versus a long boost scheme. BG505.T332N or MLV ID50 neutralization titers in the sera of naïve mice or animals immunized with 25ug DNA-encoded BG505.MD39 at Weeks 0, 3, 6 (short scheme) or Weeks 0, 3, 16 (long scheme) two weeks post the final vaccination.

	Mouse ID	BG505.T332N	MLV
Naïve	1	<45	<45
	2	<45	<45
	3	<45	<45
	4	<45	<45
	5	<45	<45
Short	1	<45	<45
	2	<45	<45
	3	<45	<45
	4	>1215	<45
	5	83	<45
	6	132	<45
	7	<45	<45
	8	<45	<45
	9	<45	<45
	10	<45	<45
Long	1	>1215	<45
	2	48	<45
	3	>1215	<45
	4	824	<45
	5	<45	<45
	6	194	<45
	7	53	<45
	8	<45	<45
	9	46	<45
	10	<45	<45

BG505.MD39TS/C05 Map PDB code 7SQ1 EMDB code 25376 Data collection **FEI Titan Krios** Microscope 300 Voltage (kV) Detector Gatan K3 Recording mode Counting Magnification (incl. post-magnification; EFTEM) 105,000 Movie micrograph pixelsize (Å) 0.84 Dose rate (e/[(camera pixel)*s]) 16.372 Number of frames per movie micrograph 60 Frame exposure time (ms) 50 2.59 Movie micrograph exposure time (s) Total dose $(e^{-}/Å^{2})$ 60.6 Defocus range (µm) 0.4-3.3 EM data processing Number of movie micrographs 5,485 Number of molecular projection images in map 121,929 C1 Symmetry Map resolution (FSC 0.143; Å) 3.8 Map sharpening B-factor $(Å^2)$ -75 Structure Building and Validation Number of atoms in deposited model 18,364 3,962 gp120 gp41 1,025 glycans 1,373 MolProbity score 1.147 (99%) Clashscore 3.57 EMRinger score 2.37 model to map correlation (FSC 0.5; Å) 3.8 Deviations from ideal Bond length outliers (RMSD; Å, #) 0.011 (0) Bond angles outliers (RMSD; °, #) 1.499 (28) Ramachandran plot Favored (%) 98.77 Allowed (%) 1.18 Outliers (%) 0.04

Supplementary Table 3. Key statistics of cryo-EM data processing, model building and validation.

Supplementary Note 1

Nucleic acid sequence: C05_HC_VH

Nucleic acid sequence: C05_Kappa

GACATTGTGCTGACCCAGTCTCCAGCTTCTTTGGCTGTGTCTCCAGGTCAGAGGGCCACCATCTCCTGCAGACCCA GCGAAAGTGTTGATAATTACGGCATTAGCTTTATGAACTGGTTCCAACAGAAACCAGGACAGCCACCCAAACTCCT CATCTATGCTGCATCCAACCGAGGATCCGGGGTCCCTGCCAGGTTTACTGGCAGTGGGTCTGGGACAGACTTCAG CCTCAACATCCATCCTATGGAGGAGGATGATATTGCAATGTATTTCTGTCAGCAAAGTAAGGAGGTTCCGTATACG TTCGGAGGGGGGACCAAACTGGAAATAAAA