Supplementary methods table 1. Strains used on this study

Strain	Relevant genotype	Description	Source or reference
S. epidermidis RP62a	*crispr*$^{+}$, neomycin^{R}, erythromycin^{R}	Used as the wild type *S. epidermidis* strain with a functional native type III-A CRISPR-Cas system	[1]
S. epidermidis LAM104	Δ*crispr,* neomycin^{R}, erythromycin^{R}	Isogenic strain of *S. epidermidis* RP62a with a deletion of the CRISPR spacer array.	[2]
S. aureus TB4 pWJ245 (gp14)	*crispr*$^{+}$	*S. aureus* TB4 transformed with a plasmid carrying the type III-A CRISPR-Cas system of *S. epidermidis* RP62a with a targeting spacer towards the gp14 gene of ϕNM1γ6	This study
S. aureus TB4 pGG-BsaI	*Δspacer*	*S. aureus* TB4 transformed with a plasmid carrying the type III-A CRISPR-Cas system of *S. epidermidis* RP62a with non-targeting spacer	This study
S. aureus TB4 pWJ248 (gp14, dCsm3/dCsm6)	*crispr*$^{+}$,*csm3*(D32A), *csm6*(R364A,H369A)	*S. aureus* TB4 pWJ245, but with catalytically inactive Csm3 and Csm6	This study
S. aureus TB4 pJTR122 (gp14, dHD/dCsm3)	*crispr*$^{+}$, *cas10*(H14A, D15A), *csm3*(D32A)	*S. aureus* TB4 pWJ245, but with catalytically inactive HD domain of Cas10	This study
S. aureus JAV6 pWJ245 (gp14, *lexA* K168A)	*crispr*$^{+}$, *lexA* K168A	SOS-deficient strain of *S. aureus* TB4 strain with pWJ245	This study
S. aureus TB4 pWJ191 (gp43)	*crispr*$^{+}$	*S. aureus* TB4 transformed with a plasmid carrying the type III-A CRISPR-Cas system of *S. epidermidis* RP62a with a targeting spacer towards the gp43 gene of ϕNM1γ6	This study
S. aureus TB4 pJTR109 (gp43, dHD)	*crispr*$^{+}$, *cas10*(H14A, D15A)	*S. aureus* TB4 pWJ191, but with catalytically inactive HD domain of Cas10	This study
S. aureus RN4220 pWJ242 (gp43, dCsm3/dCsm6), pJTR162	*crispr*$^{+}$, *csm3*(D32A), *csm6*(R364A,H369A), P_{tet}-Target	*S. aureus* RN4220 co-transformed with a dCsm3/dCsm6 type III-A CRISPR-Cas system and an inducible target plasmid	This study
S. aureus RN4220 pWJ242 (gp43, dCsm3/dCsm6), pWJ153	*crispr*$^{+}$, *csm3*(D32A), *csm6*(R364A,H369A), P_{tet}-NonTarget	*S. aureus* RN4220 co-transformed with a dCsm3/dCsm6 type III-A CRISPR-Cas system and an inducible plasmid lacking the target sequence	This study
S. aureus RN4220 pJTR127 (gp43, dHD/dCsm3/dCsm6), pJTR162	*crispr*$^{+}$, *cas10*(H14A, D15A), *csm3*(D32A), *csm6*(R364A,H369A), P_{tet}-Target	*S. aureus* RN4220 co-transformed with a dHD/dCsm3/dCsm6 type III-A CRISPR-Cas system and an inducible target plasmid	This study
S. aureus RN4220 pJTR127 (gp43, dHD/dCsm3/dCsm6), pWJ153	*crispr*$^{+}$, *cas10*(H14A, D15A), *csm3*(D32A), *csm6*(R364A,H369A), P_{tet}-NonTarget	*S. aureus* RN4220 co-transformed with a dHD/dCsm3/dCsm6 type III-A CRISPR-Cas system and an inducible plasmid lacking the target sequence	This study
S. aureus TB4 pDVB51 (pSauCas9)	*crispr*$^{+}$	*S. aureus* TB4 transformed with a plasmid carrying the type II-A CRISPR-Cas system	This study

		from *S. aureus*, with a spacer targeting ϕNM1γ6	
S. aureus TB4 pWJ245, pAV22 (gp14)	*crispr*$^{+}$	*S. aureus* TB4 transformed pWJ245 and the SOS reporter plasmid	This study
S. aureus JAV6 pWJ245, pAV22 (gp14, *lexA* K168A)	*crispr*$^{+}$, *lexA* K168A	SOS-deficient strain of *S. aureus* TB4 strain with pWJ245 and the SOS reporter plasmid	This study

References

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Supplementary methods table 2. Oligos used in this study

Plasmid	Description	Sequence
CYM-Pm128	Forward primer to amplify the *rpoB* locus from the *S. aureus* genome	CTTGTCCCGGGTAACTAGTAATTAATATGCTTATGGTATTTAGCTAAAAGC
CYM-Pm129	Reverse primer to amplify the *rpoB* locus from the *S. aureus* genome	TGCGAGCTAGCATCTGCAGTTTAATCAGTAACTTCTTTTTGTGTTTCAGGAGC ATC
CYM-Pm733	Forward primer to amplify the spacer array of the type III-A CRISPR-Cas locus of *S. epidermidis*	ATCATATAATCTTGTACTAGTGATTGTC
CYM-Pm734	Reverse primer to amplify spacer array of the type III-A CRISPR-Cas locus of *S. epidermidis*	TTTTTCCATCCCCTAGAAATTAATCAATGCG
CYM-MWS51	Sequencing adapter oligo from Kennedy et al. *Nature Protocols*. 2014	AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCC GATCT
CYM-MWS55	Sequencing adapter oligo with randomized barcodes from Kennedy et al. *Nature Protocols*. 2014	TCTTCTACAGTCANNNNNNNNNNNNAGATCGGAAGAGCACACGTCTGAACTCC AGTCAC
CYM-MWS13	Forward primer to amplify adaptor-ligated genomic fragments, from Kennedy et al. *Nature Protocols*. 2014	AATGATACGGCGACCACCGAG
CYM-MWS21-1	Reverse primer to amplify adaptor-ligated genomic fragment, with TruSeq index sequence 1	CAAGCAGAAGACGGCATACGAGATATCACGGTGACTGGAGTTCAGACGTGTGC
CYM-MWS21-2	Reverse primer to amplify adaptor-ligated genomic fragment, with TruSeq index sequence 2	CAAGCAGAAGACGGCATACGAGATCGATGTGTGACTGGAGTTCAGACGTGTGC
CYM-MWS21-3	Reverse primer to amplify adaptor-ligated genomic fragment, with TruSeq index sequence 3	CAAGCAGAAGACGGCATACGAGATTTAGGCGTGACTGGAGTTCAGACGTGTGC
CYM-MWS21-4	Reverse primer to amplify adaptor-ligated genomic fragment, with TruSeq index sequence 4	CAAGCAGAAGACGGCATACGAGATTGACCAGTGACTGGAGTTCAGACGTGTGC
GG424	For cloning pJTR127 (amplifies *cas* genes from pJTR126, R)	CATATTGCCTGATGAAGTGAATAG
GG425	For cloning pJTR127 (amplifies gp43 spacer from pWJ191, R)	CTATTCACTTCATCAGGCAATATG
W614	For cloning pJTR127 (amplifies *cas* genes from pJTR126, F)	GGTTATACTAAAAGTCGTTTGTTGG
W852	For cloning pJTR127 (amplifies gp43 spacer from pWJ191, F)	CCAACAAACGACTTTTAGTATAACC
W915	Forward primer for qPCR amplification of *S. aureus rho* gene	GTCAATGACCATAACGCAGAAG
W916	Reverse primer for qPCR amplification of *S. aureus rho* gene	CAATCGGTGTTACTAAATCCATG
W1051	For cloning gp14 spacer into pJTR116 backbone	GAACCTACGTCCGTAATGCTAGGATTTGCAAATTTCTTA
W1052	For cloning gp14 spacer into pJTR116 backbone	GATCTAAGAAATTTGCAAATCCTAGCATTACGGACGTAG
NP112	Primer to amplify the GFP coding sequence from pCN54	TCCAGATCTTCCTTCAGGTTATGACCATGTCACTTTGCTTGATATATGAG
NP114	Primer to amplify pLZ12spec	TGGTCATAACCTGAAGGAAGATCTG
AV119	Primer to amplify pLZ12spec	CAGGAAACAGCTATGACCATGATTAC
AV129	Primer to amplify the promoter from the SOS-responsive gene 4-oxalocrotonate tautomerase	ATGGTCATAGCTGTTTCCTGTCATTATTAATTTGAGTATATCATGTAATAC
AV130	Primer to amplify the promoter from the SOS-responsive gene 4-oxalocrotonate tautomerase	AGTTCTTCTCCTTTACTCATTGTTATTCCTCCCTTACTTGAATC
AV131	Primer to amplify the GFP coding sequence from pCN54	ATGAGTAAAGGAGAAGAACTTTTC
oDVB146	BsaI Cloning in pDVB47 Sau type II-A CRISPR, spacer targeting phiNM1 gp12	AGACGCGAAATCATACGCAAAAATATTCATGTTAG
oDVB147	BsaI Cloning in pDVB47 Sau type II-A CRISPR, spacer targeting phiNM1 gp12	AAAACTAACATGAATATTTTTGCGTATGATTTCGC

JTR1008	Forward primer for qPCR of *S. aureus umuC* gene	CGAGTGTTTCTTGTATTGAAAAGGGGC
JTR1009	Reverse primer for qPCR of *S. aureus umuC* gene	GTGGTATTTCAAACAATCGCGACCC
B585	Amplification of pDB114 backbone for construction of pDB236	GGAGAGGCGTATCATCAAACGAAAATTGGATAAAGTGGG
B586	Amplification of pDB114 backbone for construction of pDB236	TATTTTATTATGCATCGTTTGTTGAACTAATGGGTGC
B587	Amplification of type II-A CRISPR-cas locus of *S. aureus* M06/0171 into pDB236	TTAGTTCAACAAACGATGCATAATAAAATACAAGGTCC
B588	Amplification of type II-A CRISPR-cas locus of *S. aureus* M06/0171 into pDB236	TCCAATTTTCGTTTGATGATACGCCTCTCCCTATCAGTG
oDVB96	Amplification of pDB236 fragment to replace the CRISPR array for a BsaI cloning site	AGCATTTTCAGGTATAGGTGTTTTGGGAAACAATTTCCCCGAACCATTATATTTCTCTAC
oDVB97	Amplification of pDB236 fragment to replace the CRISPR array for a BsaI cloning site	TTTCCCAAAACACCTATACCTGAAAATGCTTTTTCTCTTTCTATTATTCCATGGACTTCA
oDVB126	Amplification of pDB236 fragment to replace the CRISPR array for a BsaI cloning site	TGAAAAAATTAAGCTGCACGATATGCAAGAAGGGAAATGTTTATACTCGTTAGAAGCAAT
oDVB127	Amplification of pDB236 fragment to replace the CRISPR array for a BsaI cloning site	GAGACCTTTGAGCTTCCGAGACTGGTCTCAGTCTACCTCATACCTAAAATTACAGAGTACTAAAACCTCTAGCATCATTATAAATCCTCA
oDVB128	Amplification of pDB236 fragment to replace the CRISPR array for a BsaI cloning site	TGAGACCAGTCTCGGAAGCTCAAAGGTCTCGTTTTAGTACTCTGTAATTTTAGGTATGAGGTAGACAGCAAAATCACTGATAGGGAGAGG
oDVB129	Amplification of pDB236 fragment to replace the CRISPR array for a BsaI cloning site	TCTTGCATATCGTGCAGCTTAATTTTTTCAATTAAATATTTAGCATTTTCTTTACCTGTC

Supplementary methods table 3. Plasmids used in this study

Plasmid	Description	Source or reference
pGG-BsaI (Δspc)	type III-A CRISPR-Cas with a BsaI cloning site inserted between the first and second repeats, used as a non-targeting spacer control	[1]
pWJ245 (gp14)	type III-A CRISPR-Cas with a spacer targeting the *gp14* gene of ϕNM1γ6	[2]
pWJ248 (gp14, dCsm3/dCsm6)	type III-A CRISPR-Cas with a spacer targeting the *gp14* gene of ϕNM1γ6 and *csm3*(D32A), *csm6*(R364A,H369A) mutations	[2]
pJTR122 (gp14, dHD/dCsm3)	type III-A CRISPR-Cas with a spacer targeting the *gp14* gene of ϕNM1γ6 and *cas10*(H14A, D15A) and *csm3*(D32A) mutations	This study
pWJ191 (gp43)	type III-A CRISPR-Cas with a spacer targeting the *gp43* gene of ϕNM1γ6	[2]
pJTR109 (gp43, dHD)	type III-A CRISPR-Cas with a spacer targeting the *gp43* gene of ϕNM1γ6 and *cas10*(H14A, D15A) mutations	[3]
pWJ242 (gp43, dCsm3/dCsm6)	type III-A CRISPR-Cas with a spacer targeting the *gp43* gene of ϕNM1γ6 and *csm3*(D32A), *csm6*(R364A,H369A) mutations	[2]
pJTR127 (gp43, dHD/dCsm3/dCsm6)	type III-A CRISPR-Cas with a spacer targeting the *gp43* gene of ϕNM1γ6 and *cas10*(H14A, D15A), *csm3*(D32A), *csm6*(R364A,H369A) mutations	This study
pWJ153	pE194-based plasmid vector with an inducible P_{tet} promoter, used as a non-targeting spacer control	[1]
pJTR162	pE194-based plasmid with a target sequence matching the *gp43* spacer under the control of the P_{tet} promoter	[3]
pDVB51 (pSauCas9)	Type II-A CRISPR-Cas system of *S. aureus* with a spacer targeting *gp12* in ϕNM1γ6.	This study
pAV22	SOS reporter plasmid with Green Fluorescent Protein (GFP) under the control of the 4-oxalocrotonate tautomerase promoter of *S. aureus*.	This study

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2 Jiang, W., Samai, P. & Marraffini, L. A. Degradation of phage transcripts by CRISPR-associated RNases enables type III CRISPR-Cas immunity. *Cell* **164**, 710-721.

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