



TRE

(a) Schematic diagram of the ChIP-Seq procedure. Wild type (WT) and homozygous TR α knockout (TR $\alpha^{-/-}$) tadpoles at stage 54 were treated without or with T3 for 18 hours. Chromatin was isolated from the intestine and sonicated with a Bioruptor. The chromatin fragments were cross-linked and immunoprecipitated by using a rabbit polyclonal TR antibody. After DNA purification, the quality of the ChIP DNA was checked by qPCR analysis for the well-known TRE region of TR β gene (see B) before 100 bp paired-end sequencing with Illumina HiSeq2500 was performed. ChIP-Seq reads were aligned to *X. tropicalis* genome assembly (v9.1). Finally, the data were normalized by the base coverage profile of the pile-up and the peak sites were used for identification of the TR binding sites.

Exon5

(b) Validation of the ChIP samples from the WT and $TR\alpha^{-/-}$ intestine. The ChIP samples in (a) was analyzed by PCR for the presence of the region containing the TR β TRE or TR β exon 5, the negative control region lacking a TRE.

Tissue	Genotype	Т3	No	Total peaks	Common peaks	Pileup			Mean height by base
						max	min	mean	
Intestine	WT	-	1	46608	11469	738	2	4.31	4.24
			2	60134		1694	3	7.68	7.55
			3	50207		2508	4	12.06	11.26
		+	1	47106	17930	866	3	7.87	7.64
			2	57418		1540	4	12.44	11.73
			3	64193		1977	4	12.70	12.13
	TRα ^{-/-}	-	1	59774	9770	315	2	3.81	3.79
			2	50729		498	3	6.73	6.63
			3	54233		511	3	6.61	6.53
		+	1	22150	8160	481	3	7.11	7.77
			2	33834		665	3	7.51	3.91
			3	42420		717	3	7.80	7.42

Fig. S2. Summary of the peak and pileup values for different ChIP-Seq samples. ChIP-Seq were performed for 3 technical replicates of the intestine samples from wild-type and TR $\alpha^{-/-}$ tadpoles with or without 18 hour treatment with 10 nM T3. The ChIP-Seq FASTQ sequences were aligned to the *Xenbase* genome sequence to identify the peaks and determine the peak heights (Pileup, in base pairs). Note that the common peaks refers to peaks present in all three replicates of the samples.

Motif	Name	Sample	P-value	Target Sequences with Motif	% of Targets Sequences with Motif
	THRa(NR)	WT T3-	1.00E-97	6285	7.63%
		WT T3+	1.00E-153	7904	8.71%
	THRb(NR)	WT T3-	1.00E-60	7847	9.53%
<u>GGTCANNNNAGGACA</u>		WT T3+	1.00E-87	9692	10.69%
	LXRE(NR)	WT T3-	1.00E-12	1188	1.44%
AGGTLANNNNAGGLCA	DR ² 4	WT T3+	1.00E-35	1476	1.63%

Fig. S3. The DR4 TRE sequences are highly enriched in the TR-binding peaks detected by ChIP-Seq in the wild type intestine. Homer de novo motif search of all peak regions in the wild type intestine showed that THRa, THRb and LXRE (DR4) motifs were highly enriched. Note that the software used for the analysis did not show the first residue of the first repeat for THRa and THRb motifs. In any case, the result indicate clearly that DR4 TREs bound by TR have the consensus sequence of GGTCA and (A/G)GGTCA separated by 4 bases, basically identical to the idealized DR4 TRE made of AGGTCAnnnAGGTCA.



Figure S4. Representative views of ChIP-Seq and RNA-Seq data: (a) TR β and (b) TH/bZip. Blue peaks in each wiggle plot represent the normalized ChIP-seq and RNA-seq reads, respectively. There were three technical replicates for each sample (+T3 or -T3). The red-boxed region in the ChIP-Seq plot was enlarged to show the presence of previously characterized TREs in the TR β and TH/bZip genes, with the known TREs are shown in bold letters.



Fig. S5. TR α knockout reduces TR binding to the TRE region in the tadpole intestine. Five stage-matched wild type and TR α knockout tadpoles at stage 54 were treated with or without 10 nM T3 for 18 hr. The intestine was isolated for ChIP assay by using an anti-TR antibody. The immunoprecipitated DNA was analyzed by real-time PCR for the presence of the TRE region in Mucin3A, ATP6V1B2 and TH/bZip. Note that TR binding to the TRE region was significantly reduced in the intestine of TR α knockout tadpoles. Error bars indicate SEM. ** p<0.01.