Biophysical Journal, Volume 121

Supplemental information

Protein dynamics and lipid affinity of monomeric, zeaxanthin-binding

LHCII in thylakoid membranes

Fatemeh Azadi-Chegeni, Sebastian Thallmair, Meaghan E. Ward, Giorgio Perin, Siewert J. Marrink, Marc Baldus, Tomas Morosinotto, and Anjali Pandit

Supporting Information

Protein dynamics and lipid affinity of monomeric, zeaxanthin-binding LHCII in thylakoid membranes

Fatemeh Azadi-Chegeni¹; Sebastian Thallmair^{2,3}, Meaghan E. Ward⁴; Giorgio Perin⁵; Siewert J. Marrink², Marc Baldus⁴; Tomas Morosinotto⁵; Anjali Pandit¹

¹Leiden Institute of Chemistry, Dept. of Solid-State NMR, Leiden University, Einsteinweg 55, 2333CC, Leiden, The Netherlands

² Groningen Biomolecular Sciences and Biotechnology Institute and Zernike Institute for Advanced Materials, University of Groningen, 9747 AG, Groningen, The Netherlands

³ Frankfurt Institute for Advanced Studies, Ruth-Moufang-Straße 1, 60438 Frankfurt am Main, Germany

⁴NMR Spectroscopy, Bijvoet Center for Biomolecular Research, Utrecht University, 3584 CH, Utrecht, The Netherlands

⁵ Dept. of Biology, University of Padua, Via Ugo Bassi 58B, 35121 Padua, Italy

Table S1

Fluorescence lifetime analysis of U-¹³C-¹⁵N Cr npq2 LHCII in α -DM and in proteoliposomes.

Sample	A1	τ ₁ (ns)	A2	τ₂ (ns)	A3	τ₃ (ns)	τ _{av} (ns)
<i>npq2</i> LHCII in α-DM	62%	3.6	25%	1.7	13%	0.2	3.3
npq2 LHCII in liposomes	50%	1.1	44%	0.4	6%	2.5	1.1



Figure S1 A: LHCII purification from sucrose gradient of thylakoid membranes from npq2 Cr cells. B: HPLC analysis of wt (cw15) and npq2 LHCII fractions. Traces were normalized on the peak of chlorophyll a (Chl a). Identification of lettered peaks is as follows: Loro, loroxanthin; N, neoxanthin; V, violaxanthin; A, antheraxanthin; L, lutein; Z, zeaxanthin; β , β -carotene.



Figure S2. A. Time resolved fluorescence (excitation 440 nm, detection 680 nm) of npq2 LHCII in α -DM detergent (green) and npq2 LHCII in liposomes (black). B. Absorption spectra of npq2 LHCII in α -DM detergent (black) and after inserting into liposomes (red).



Figure S3 ¹³C -¹³C CP-PARIS NMR spectra of wt (blue) and npq2 (red) LHCII. Spectra were collected at -18 °C and with 17 kHz MAS frequency. A. Thr and Ser region. Helix, coil and β strands contribution are presented with black, red and green boxes. B. The Ala region. Arrows indicate resonance signals that are only observed in the spectrum of WT LHCII. C. Linewidth comparison for Ala peak (1D slice ω_1 -¹³C = 55.2 ppm); red, npq2 LHCII, blue, wt LHCII. Data of the wt LHCII proteoliposomes has been presented in previous work. ¹



Figure S4 ¹³C -¹³C CP-PARIS NMR spectra of wt (blue) and npq2 (red) LHCII. Spectra were collected at -18 °C and with 17 kHz MAS, pigment region. Data of the wt LHCII proteoliposomes has been presented in previous work. ¹



Figure S5. ¹³C-¹³C PARIS NMR spectrum of npq2 LHCII, collected at -3 °C and with 14 kHz MAS frequency, galactosyl region with MGDG NMR chemical shift correlation signals.



Figure S6 RMSF of the protein backbone of the wt LHCII trimer and the monomeric npq2 LHCII evaluated for the aggregation simulations containing 4 LHCII trimers and 12 npq2 LHCII monomers, respectively, embedded in thylakoid membrane. The inlay shows a zoom of the Nterminus.

References

 [1] Azadi-Chegeni, F., Ward, M. E., Perin, G., Simionato, D., Morosinotto, T., Baldus, M., and Pandit, A.
(2021) Conformational Dynamics of Light-Harvesting Complex II in a Native Membrane Environment, *Biophys J 120*, 270-283.