

## Supplementary Materials for

### **Increasing breast milk betaine modulates *Akkermansia* abundance in mammalian neonates and improves long-term metabolic health**

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#### **The PDF file includes:**

Fig. S1. Effect of maternal betaine administration on young mouse offspring.

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Fig. S5. Effect of betaine on bacterial growth in vitro.

Fig. S6. Effect of maternal betaine supplementation on mouse ileum histology and gene expression.

Table S1. No association between milk betaine concentration and change in human infant body length  $z$  score and head circumference.

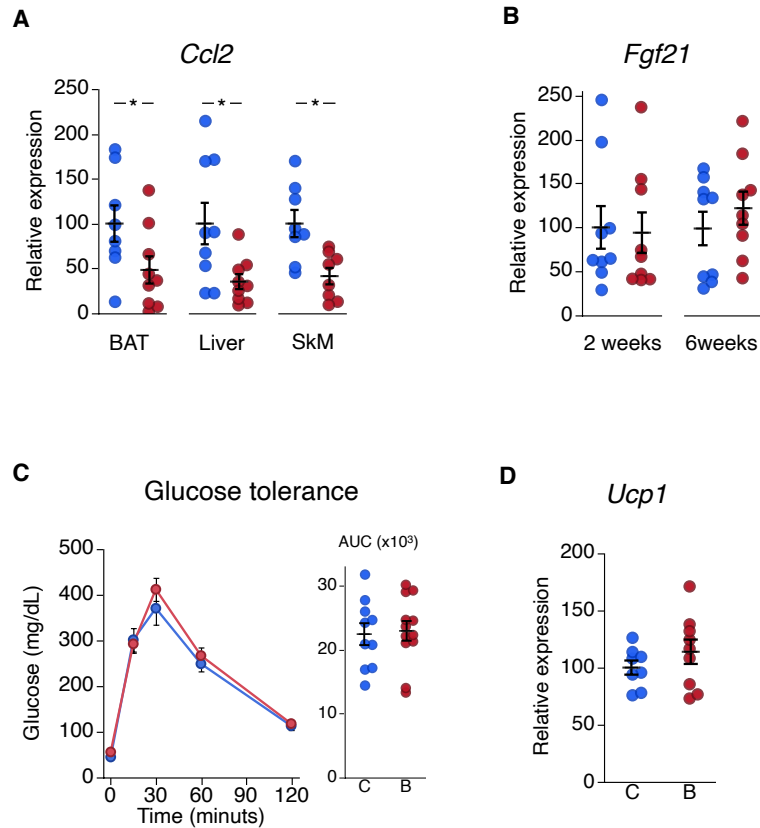
Table S2. Prevalence of *A. muciniphila* in human infants exposed to low and high breast milk betaine content.

Table S3. Primer sequences for qPCR analyses.

#### **Other Supplementary Material for this manuscript includes the following:**

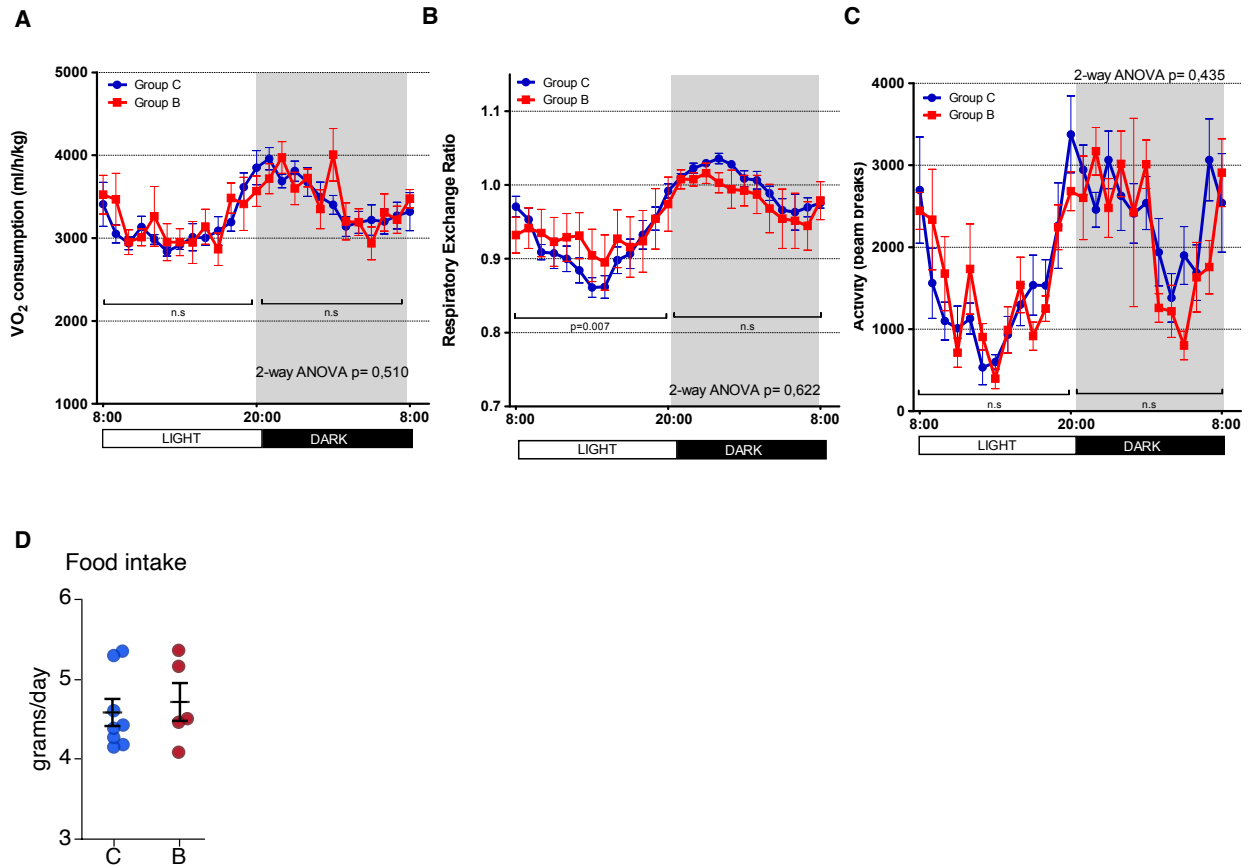
(available at [stm.sciencemag.org/cgi/content/full/13/587/eabb0322/DC1](http://stm.sciencemag.org/cgi/content/full/13/587/eabb0322/DC1))

Data file S1 (Microsoft Excel format). Individual level data for all figures.



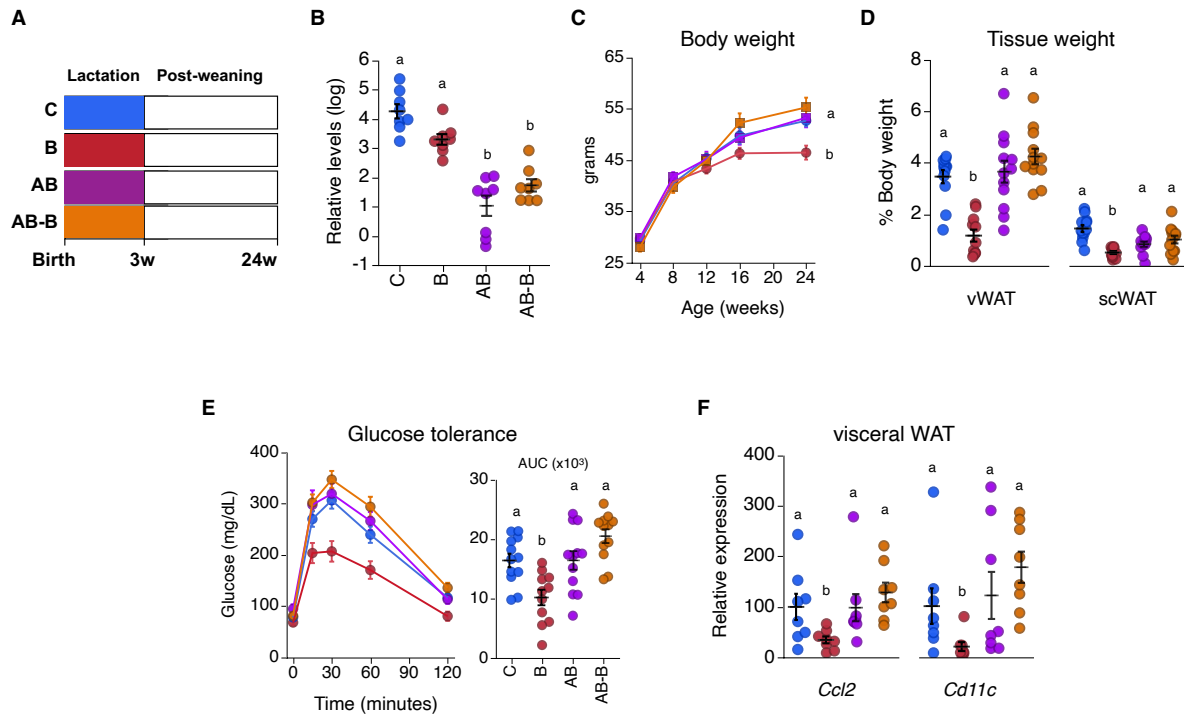
**Figure S1. Effect of maternal betaine administration on young mouse offspring.**

**A**) *Ccl2* mRNA levels in brown adipose (BAT, n=8-9 per group), liver (n=9 per group), and skeletal muscle (SkM) (n=8 per group) in 6-week-old offspring from control (C, blue color) and betaine-treated (B, red color) dams. **B**) *Fgf21* mRNA levels in liver from 2- and 6-week-old mice (n=9 per group). **C**) Glucose tolerance was measured in 8-week-old male offspring (n=10-12 per group). **D**) *Ucp1* mRNA expression in BAT from 6-week-old offspring (n=8-9 per group). Data are mean  $\pm$  SEM. \*, *t*-test  $p < 0.05$ .

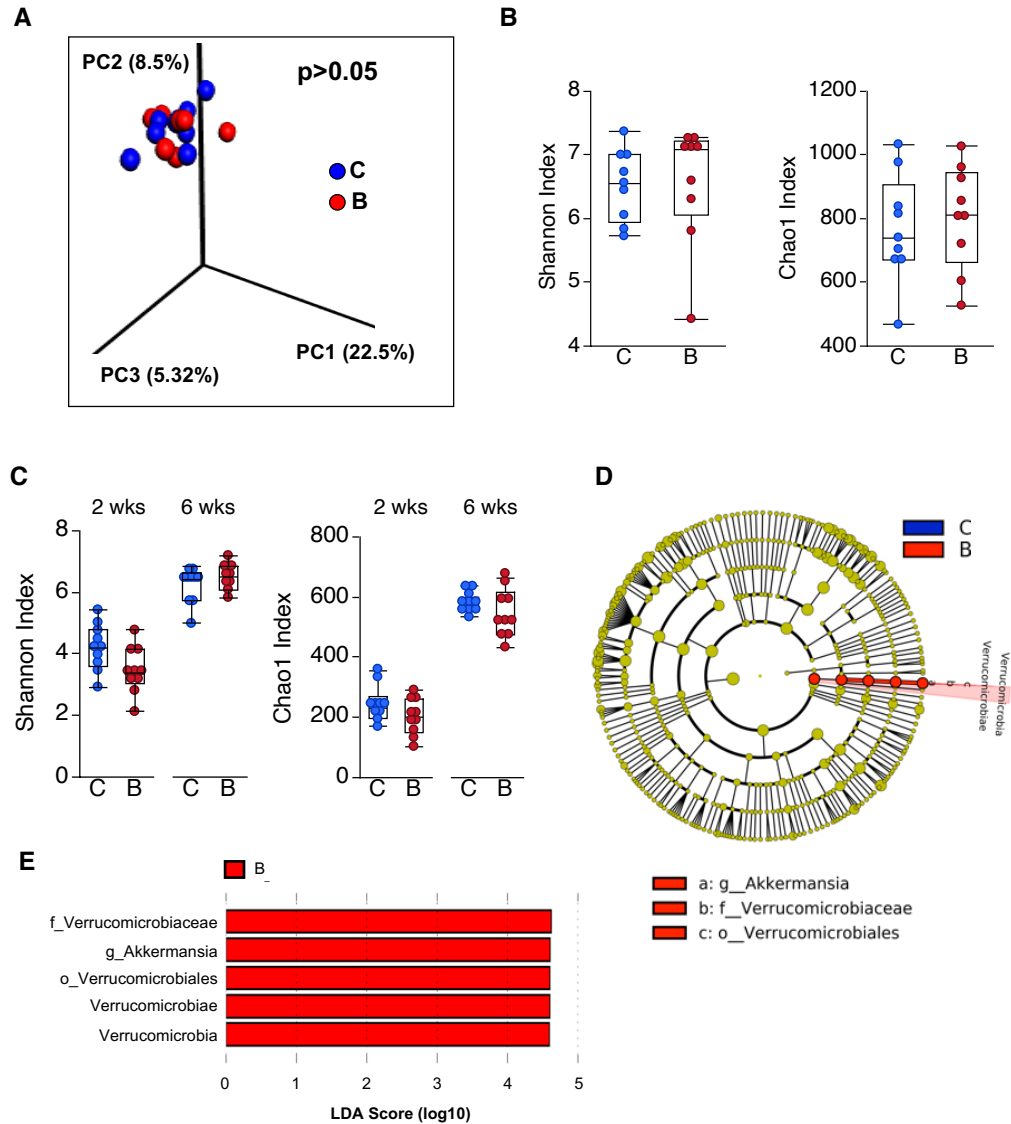


**Figure S2. Effects of maternal betaine administration on mouse offspring energy homeostasis.**

Six-week-old offspring from control (C, blue color, n=8) and betaine-treated (B, red color, n=5) dams were examined in metabolic chambers for 24 hours. **A**) Oxygen consumption, **B**) respiratory exchange ratio, **C**) physical activity, and **D**) food intake were measured. Data are mean  $\pm$  SEM. Differences between groups were assessed by 2-way ANOVA (A-C).

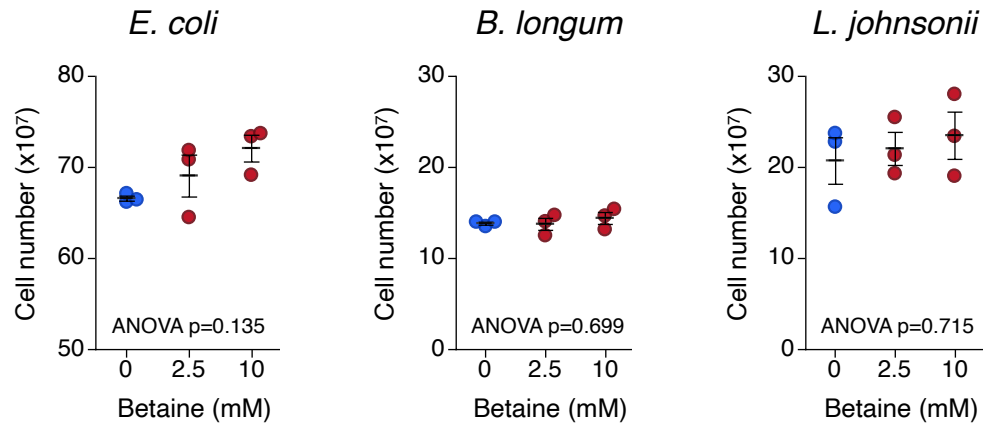


**Figure S3. Effects of maternal antibiotic co-administration on offspring long-term metabolic health in mice.** Dams were treated with betaine (B group, red), ampicillin (1g/L) and neomycin (0.5 g/L) (AB group, purple), antibiotics and betaine (AB-B group, orange) in the drinking water, or with no supplement (C group, blue) during lactation and offspring monitored until adulthood. **A)** Schematic representation of the experimental design. **B)** A subset of mice was sacrificed at 2 weeks of age and bacterial DNA extracted from cecal samples; levels of bacterial 16S gene as a measure of microbial content was determined by qPCR (oligos 5'-ACTCCTACGGGAGGCAGCAG-3' and 5'-ATTACCGCGGCTGCTGG-3') and expressed as log-transformed relative levels per gram of cecal sample (n=8 per group). **C)** Body weight and **D)** WAT weight at sacrifice (n=11-12 per group). **E)** Glucose tolerance at 22 weeks of age (n=11-12 per group). **F)** mRNA expression of vWAT immune markers (n=8 per group). Data are mean  $\pm$  SEM. Different letters indicate statistically significant differences after one-way ANOVA and post-hoc Tukey test ( $p < 0.05$ ).



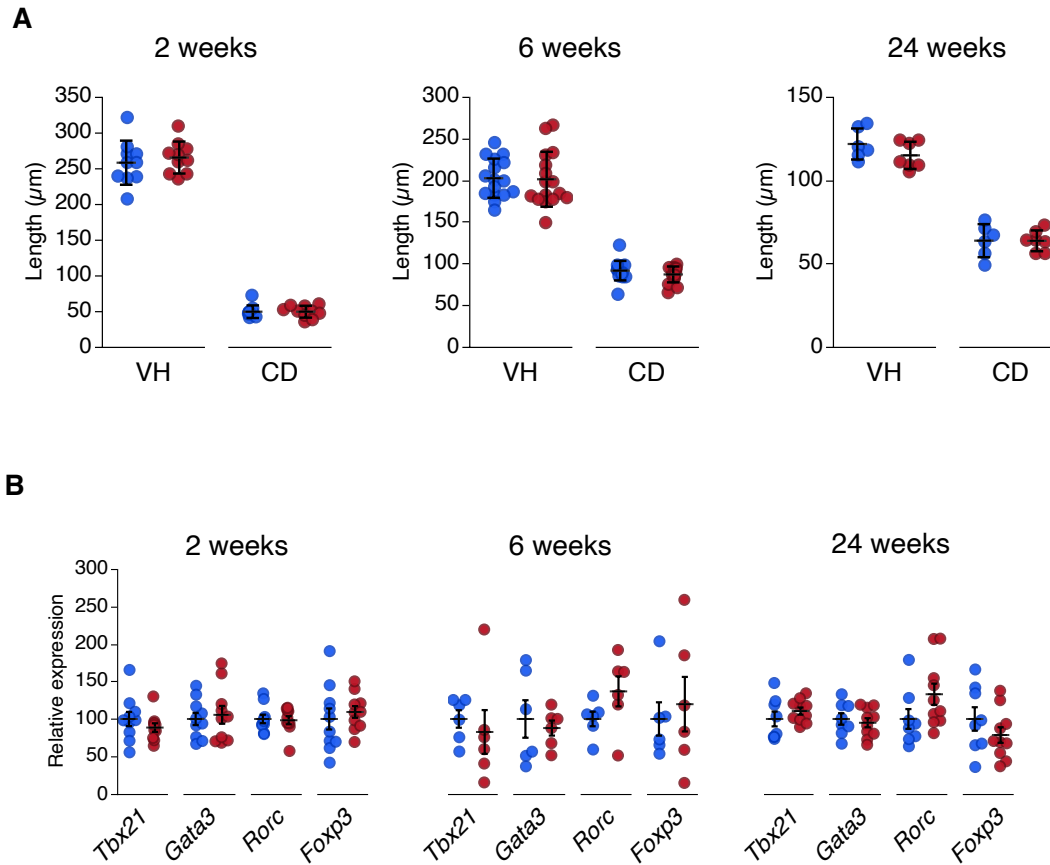
**Figure S4. Effect of betaine administration on the maternal and offspring gut microbiome in mice.**

Betaine-treated (B group, red color, n=8) or control (C group, blue color, n=8) dams and offspring were sacrificed at day 14 after delivery, and at 6 weeks of age (n=10 per group). **A**) Principal coordinate analysis of unweighted UniFrac distances of cecal samples; p value assessed by adonis test. **B**) Diversity Shannon and Chao1 indices from C and B dams. **C**) Diversity Shannon and Chao1 indices from offspring at 2 and 6 weeks of age. **D**) Cladogram and **E**) LDA scores arising from LEfSe analysis in 2-week-old pups from betaine-treated and control groups.



**Figure S5. Effect of betaine on bacterial growth in vitro.**

*Escherichia coli*, *Bifidobacterium longum* and *Lactobacillus johnsonii* were grown in vitro in the absence (blue color) or presence (red color) of 2.5 mM or 10 mM betaine in the media until reaching the stationary phase (n=3 per group). Cell number for the indicated bacterial species was determined based on optical density (600 nm) after 8h, 12h, or 15h of growth, respectively. Data are mean  $\pm$  SEM. One-way ANOVA was applied to assess differences between groups.



**Figure S6. Effect of maternal betaine supplementation on mouse ileum histology and gene expression.**

**A)** Ileal vili height (VH) and crypt depth (CD) in ileum sections from 2- (n=10 per group), 6- (n=16 per group), and 24-week-old (n=6-7 per group) control (blue circles) and betaine-treated mice (red circles).

**B)** Ileal mRNA levels of T cell differentiation markers *Tbx21*, *Gata3*, *Rorc*, and *Foxp3* at 2 weeks (n=10 per group), 6 weeks (n=6 per group), and 24 weeks of age (n=8-10 per group). Data are shown as mean  $\pm$  SEM. Group differences were assessed by Student's *t*-test.

	Cohort I		Cohort II	
	B (CI 95%)	P value	B (CI 95%)	P value
<b>Change in body length z score</b>				
Birth - 1 month	0.01 (-0.57, 0.58)	0.982	0.22 (-0.07, 0.51)	0.138
Birth - 12 months	---		-0.04 (-0.37, 0.30)	0.826
<b>Change in head circumference (cm)</b>				
Birth - 1 month	-0.01 (-0.67, 0.65)	0.973	-0.14 (-0.51, 0.24)	0.478
Birth - 12 months	---		-0.37 (-0.91, 0.15)	0.162

**Table S1. No association between milk betaine concentration and change in human infant body length z score and head circumference.** Least-square regression modelling was applied to assess the correlation between milk betaine level (independent variable) and body length z score change from birth to 1 month or 12 months (dependent variable). Model was adjusted for gestational age, pre-pregnancy BMI, gestational weight gain, birth method, and body length z score at birth (for change in body length) or head circumference at birth (for change in head circumference). B, size effect estimate from the regression model; CI, confidence interval.



		<i>Akkermansia muciniphila</i>		Chi-square p value
		Absent	Present	
<b>1 month (n=83)</b>	Low betaine	32 (38.6 %)	10 (12.1 %)	0.840
	High betaine	32 (38.6 %)	9 (10.8 %)	
<b>12 months (n=91)</b>	Low betaine	28 (30.8 %)	18 (19.8 %)	0.348
	High betaine	23 (25.3 %)	22 (23.5 %)	

**Table S2. Prevalence of *A. muciniphila* in human infants exposed to low and high breast milk betaine content.** Infants were categorized into Low and High betaine groups based on the median value of breast milk betaine concentration (4.1  $\mu$ M). Presence of *A. muciniphila* was determined in fecal samples from 1-month (n=83) and 12-month-old infants (n=91) by qPCR. Data are shown as n (%) of subjects with either absence or presence of *A. muciniphila* in fecal samples. Chi-square test was applied to detect differences in prevalence among low and high betaine groups at 1 month or 12 months of age.

Gene	Forward	Reverse
<i>Hprt</i>	5'-GCCCCAAAATGGTTAAGGTTG-3'	5'-GTCAAGGGCATATCCAACAAC-3'
<i>Muc2</i>	5'-CTGACCAAGAGCGAACACAA-3'	5'-CATGACTGGAAGCAACTGGA-3'
<i>Ocln</i>	5'-ATGTCCGGCCGATGCTCTC-3'	5'-CTTTGGCTGCTCTTGGGTCTGTAT-3'
<i>Zo2</i>	5'-CTAGACCCCCAGAGCCCCAGAAA-3'	5'-TCGCAGGAGTCCACGCATACAAG-3'
<i>Zo1</i>	5'-TTTTGACAGGGGGAGTGG-3'	5'-TGCTGCAGAGCTCAAAGTTCAAG-3'
<i>Ccl2</i>	5'-CAAGATGATCCCAATGAGTAG-3'	5'-TTGGTGACAAAACTACAGC-3'
<i>Tlr4</i>	5'-GCCTCCCTGGCTCCTGGCTA-3'	5'-CAGGGACTTTGCTGAGTTTCTGATCCA-3'
<i>Cd11c</i>	5'-AGTCTGTTGGTTCTGTAAG-3'	5'-ACAGTTCTGTTATGACATGC-3'
<i>Fgf21</i>	5'-AGCTCTCTATGGATCGCCTCACTT-3'	5'-ACACATTGTAACCGTCCTCCAGCA-3'
<i>Ucp1</i>	5'-CAAATCAGCTTTGCCTCACTC-3'	5'-ACACCTCCAGTCATTAAGCC-3'
<i>Tbx21</i>	5'-ACGTCTTTACTTTCCAAGAG-3'	5'-GTACATGGACTCAAAGTTCTC-3'
<i>Gata3</i>	5'-TATTAACAGACCCCTGACTATG-3'	5'-CACCTTTTTCACCTTTTTTCG-3'
<i>Rorc</i>	5'-CTGTGTTTTTCTGAGGATGAG-3'	5'-GCAGAGATGATGATGGAAAG-3'
<i>Foxp3</i>	5'-AATAGTTCCTTCCCAGAGTTC-3'	5'-GGTAGATTTCATTGAGTGTC-3'
<i>A. muciniphila</i>	5'-CAGCACGTGAAGGTGGGGAC-3'	5'-CCTTGCGGTTGGCTTCAGAT-3'

**Table S3. Primer sequences for qPCR analyses.**