

## Supporting information

### Dynamic contrast enhanced CEST MRI using a low molecular weight dextran

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## Supplementary Methods

### S1. Estimation of exchange rate $k_{ex}$ using $T_2$ relaxation times

The exchange rate was estimated using the classic Swift-Connick equation (Eq S1) according to our previously published protocol (1-3). In brief, the Swift-Connick equation describes the relationship between the chemical shift difference of the exchangeable proton with respect to water resonance ( $\Delta\omega = 1.2$  ppm for Dex1 in the present study), exchange rate ( $k_{ex}$ , to be determined), and  $R_{2ex}$  is the measured  $T_2$  relaxation rate in the presence of exchange ( $R_{2ex} = 1/T_{2ex}$ ), which were measured using the CPMG sequences as described below.

Of note, the unit of  $\Delta\omega$  is rad/s (4,5).

$$R_{2ex} = k_{ex} P_B \frac{\Delta\omega^2}{k_{ex}^2 + \Delta\omega^2} \quad \text{Eq. S1}$$

$P_B$  is the fraction of exchangeable protons. Using an average of 3 OH groups per glucose unit (6) and 6 glucose units in each Dex1 and 110 M protons in water,  $P_B$  equals  $1.63 \times 10^{-5}$  per mM glucose unit.

$$r_{2ex} = (1.63 \times 10^{-5}) \frac{k_{ex} \Delta\omega^2}{k_{ex}^2 + \Delta\omega^2} \quad \text{Eq. S2}$$

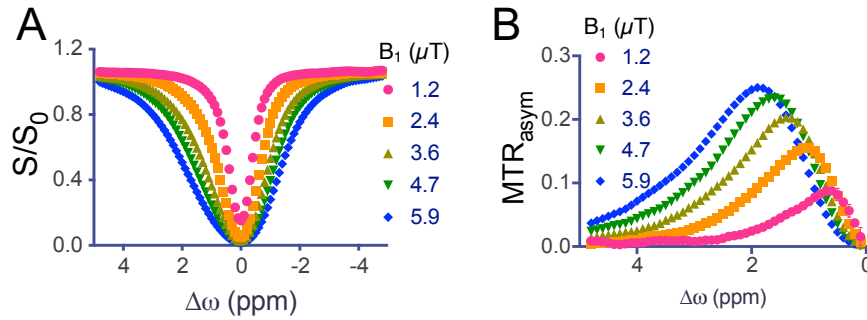
$R_2$  relaxation rates were acquired using a Carr-Purcell-Meiboom-Gill (CPMG) method.(3) Briefly, a  $T_2$  preparation module was added in the front of a fast spin-echo imaging readout, i.e., Rapid Acquisition with Relaxation Enhancement (RARE) pulse sequence. The  $T_2$  preparation period consisted of different numbers of CPMG segments with the inter-echo time  $t_{CPMG}$  fixed at 10 ms. A total of 16 CPMG-weighted images were acquired with the number of segments varied from 2 to 1024, corresponding to echo times of 20 ms - 10.24 sec. The imaging parameters were: TR/TE = 25000/4.3 ms, RARE factor = 16, a 64x64 acquisition matrix with a spatial resolution of c.a.  $250 \times 250 \mu\text{m}^2$ , and slice thickness of 1 mm. The acquisition time for each  $T_2$ -weighted image was 1 min 40 s. To obtain the exchange-based relaxivity rate,  $r_{2ex}$ , of each compound, the  $R_2 = 1/T_2$  water proton relaxation rates of the compound solutions at different concentrations, i.e., 1, 2, 5 and 10 mM, were measured and fitted to Equation S3.

$$R_2 = R_2^0 + r_{2ex} \times [C] \quad \text{Eq. S3}$$

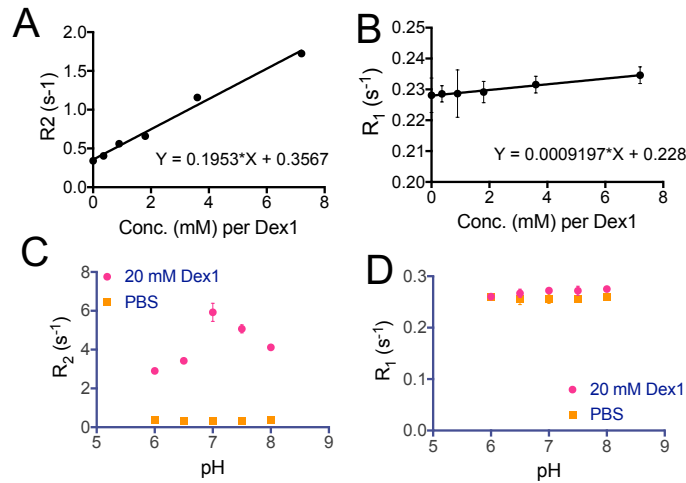
Where  $R_2^0$  is the inherent water proton  $R_2$  transverse relaxation rate of the solvent and  $[C]$  is the concentration of the agent.

It should be noted that, with a single measured  $r_{2ex}$ , two  $k_{ex}$  values are obtained when solving the Swift-Connick equation. Because it is well known that hydroxyl protons are fast-exchanging at physiological pH and temperature, the fast  $k_{ex}$  was chosen.

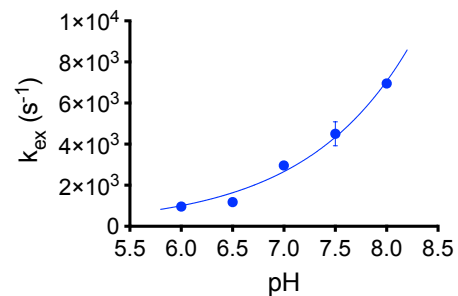
### Supplementary Results



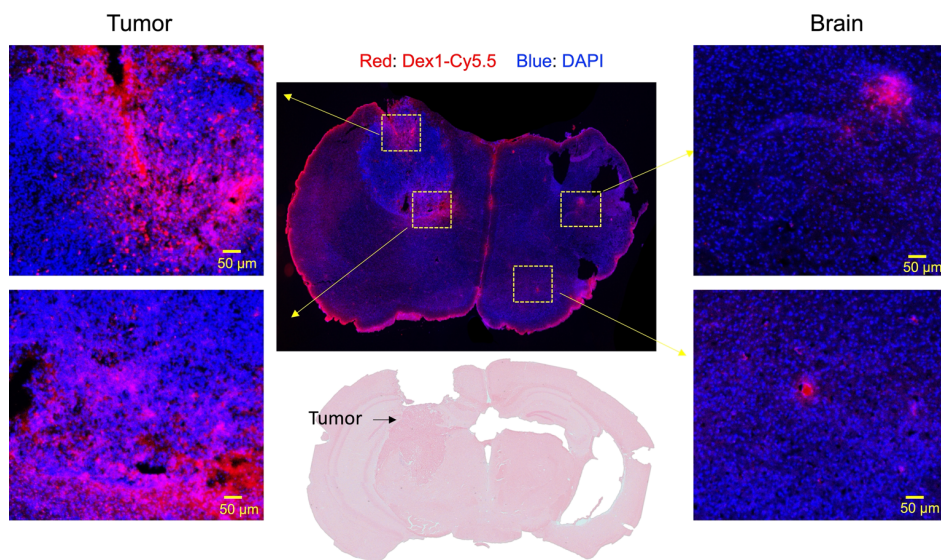
**Figure S1.  $B_1$  dependence of the CEST MRI signal of 20 mM Dex1 at 11.7T. A)** Z-spectra; **B)**  $MTR_{asym}$  plots.  $B_1$  is in  $\mu$ T. Note that the peak values shift with respect to  $B_1$  due to the effect of direct water saturation.



**Figure S2. The  $T_1$  and  $T_2$  contrast-enhancing ability of Dex1 at 11.7T. A)**  $R_2 = 1/T_2$  and **B)**  $R_1 = 1/T_1$  of Dex1 at different concentrations. **C)**  $R_2$  and **D)**  $R_1$  of 20 mM Dex1 and PBS (control) as a function of pH.



**Figure S3. Exchange rate of OH protons in Dex1 as the function of pH at 37 °C in PBS.** The exchange rate of hydroxyl proton at each pH was estimated using the measured  $R_2$  relaxation time as described above.



**Figure S4. Immunofluorescence images showing the distribution of Dex1 in a representative tumor-bearing mouse brain.** On the images, blue indicates nuclei (stained with DAPI), whereas red indicates Dex1 (labeled with Cy5.5). Four zoomed views of different tumor and brain regions are shown on the left and right panels, respectively. Shown below is a H&E-stained image as the reference.

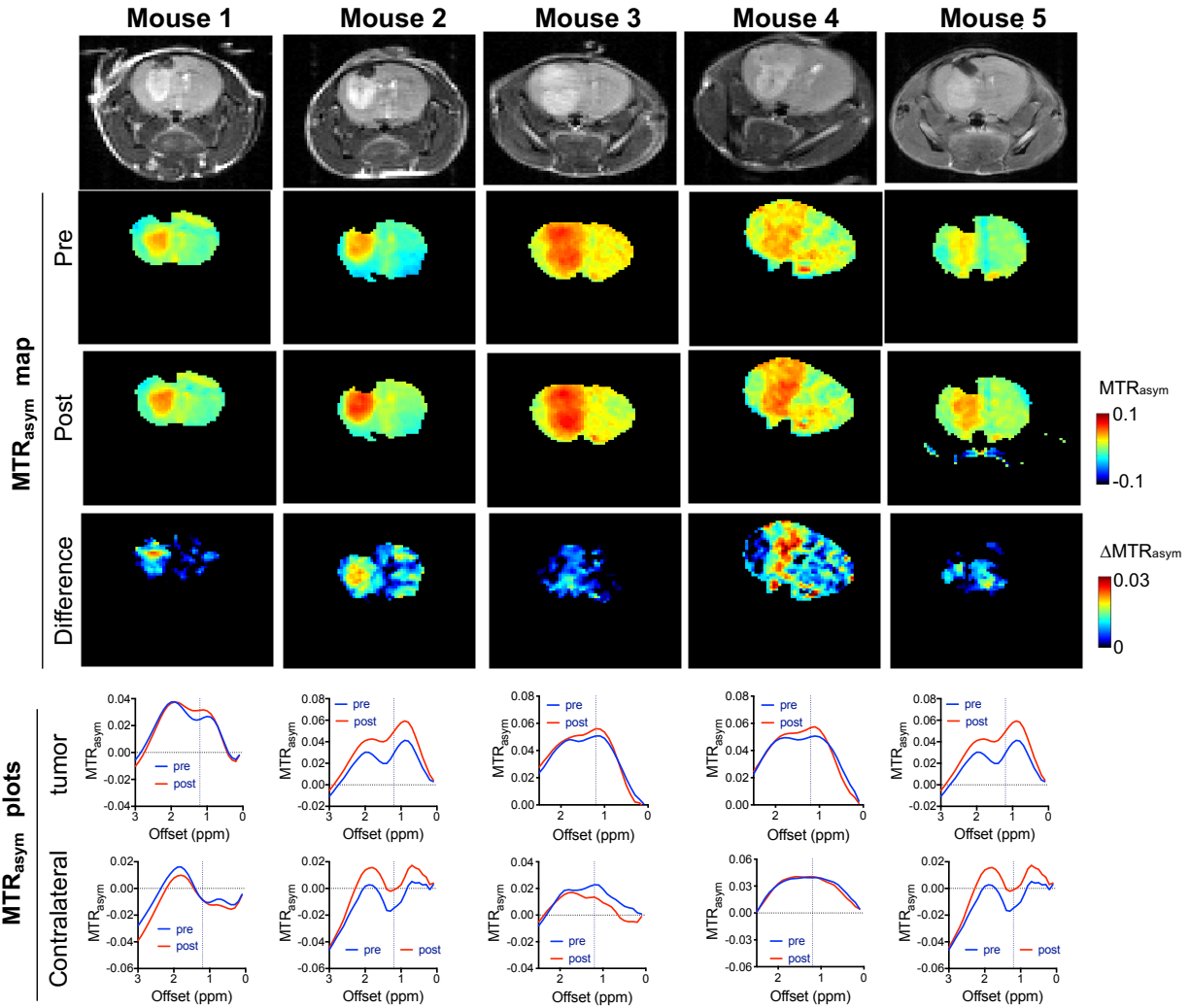


Figure S5. Dex1-enhanced CEST MRI at 11.7T in all five mice.

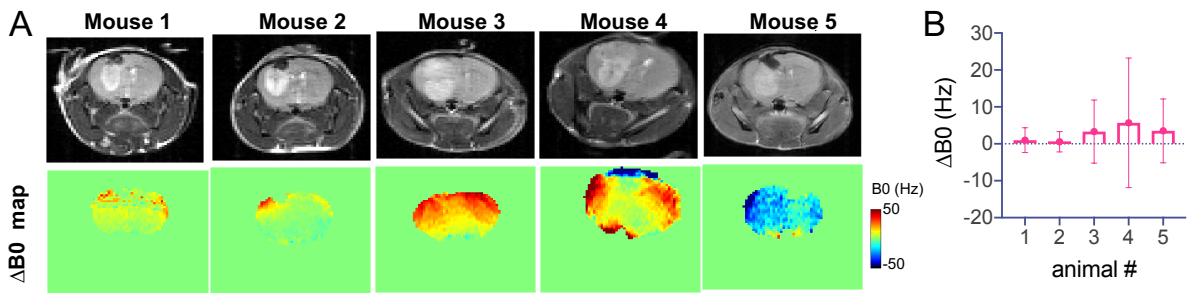
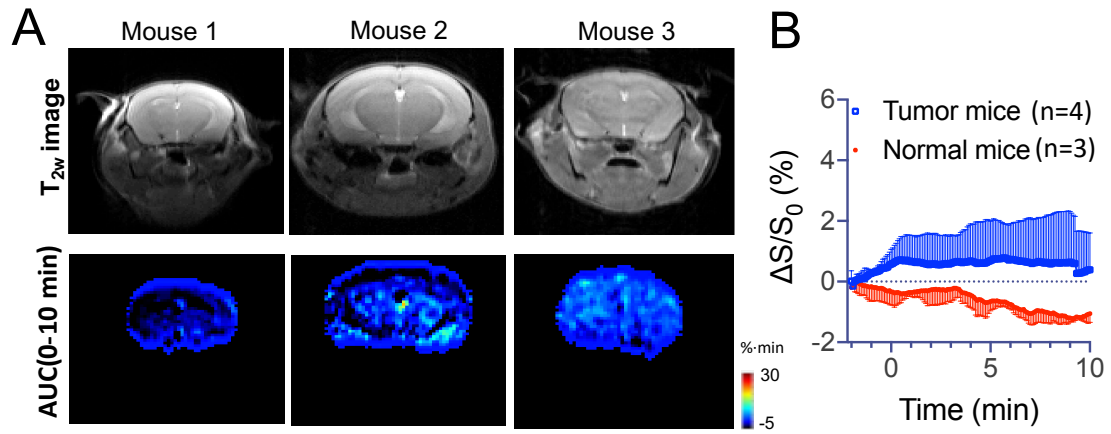
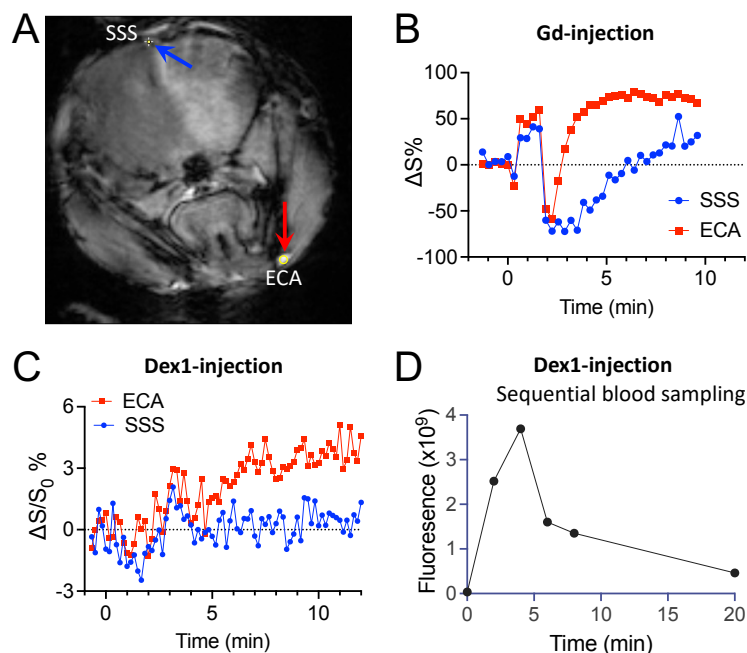


Figure S6. Assessment of the changes in  $B_0$  inhomogeneity after Dex1 infusion at 11.7T.

A)  $\Delta B_0$  maps of all five animals. B) Bar plots showing the means and standard deviations of the whole brain region of each animal.



**Figure S7. DDE CEST MRI in the normal mouse brain at 11.7T. A)** T2w and corresponding AUC(0-10 min) maps of all three non-tumor-bearing normal mice studied. **B)** Mean dynamic CEST signal change in the brain ROIs in either non-tumor-bearing mice (red, n=3) or tumor-bearing mice (blue, n=4).



**Figure S8.** A) T2w image showing the locations of ROIs in the superior sagittal sinus (SSS, blue arrow) and an external carotid artery (ECA, red arrow) used to assess AIF and VOF. B) Dynamic MRI signal changes in the SSS and ECA ROIs in the Gd-CE MRI study. C) Dynamic MRI signal changes in the same SSS and ECA ROIs in the Dex1-CEST CE MRI study. D) Fluorescence intensities of blood samples drawn at different time points in a mouse receiving 10 nmol fluorescent Dex1-Cy5.5. Method: Under anesthesia, a Balb/c mouse was inserted with a 10-cm catheter (flushed with 600 unit/mL heparin solution) into the tail vein. Prior to Dex1-Cy5.5 injection, ten  $\mu\text{L}$  blood sample was drawn to measure the background signal of the blood. Following the injection of 10 nmol Dex1-Cy5.5 (200  $\mu\text{L}$  in PBS), ten  $\mu\text{L}$  blood samples were drawn via the catheter at 2, 4, 6, 8, and 20 min after Dex1-Cy5.5 injection and immediately mixed with 50  $\mu\text{L}$  600 unit/mL heparin solutions. The fluorescence intensities of blood samples were measured using a Caliper IVIS Spectrum (Caliper Life Sciences, Hopkinton, MA, USA) and quantified the Live Image™ software (Caliper Life Sciences).

#### Reference:

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