Supplementary Material for:

In vivo CRISPR screens reveal HIF-1α-mTOR-mediated regulation of T follicular helper versus Th1 cells

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Supplementary Figure 1. Additional characterization of CRISPR-mediated gene knockout in primary mouse T cells

a, Schematic of MRCIG and MRIG vectors. **b**, Thy1.2 expression of WT CD4 T cells transduced with MRCIG control- or Thy1-sgRNA after 6 days in culture. c, Same experimental setup as (b), CD45.2 expression with control- or Ptprc-sgRNA. d, Fluorescent PCR *Thy1* amplicon size analysis of subpopulations of the cells from (b) sorted by GFP and Thy1.2 expression. e, Same experimental setup as (d), Ptprc amplicons from cells sorted from (c). f, Thy1.2 expression of Cas9⁺ CD4 T cells transduced with MRIG control- or Thy1-sgRNA after 6 days in culture. g, Matched GFPand GFP⁺ comparisons of Tfh percentages from Cas9⁺ SMARTA cells transduced with control-, Bcl6-, or Prdm1-sgRNA, in each LCMV-infected WT host from the experiment in Fig. 1c. Control-sgRNA n=4 mice, all other sgRNAs n=3. h, Total number of GFP⁺ Tfh cells in (g), replicates pooled per sgRNA target. For *Bcl6* and *Prdm1* sgRNAs, squares, triangles, and diamonds represent sgRNAs #1, 2, and 3, respectively, with similar results for all sgRNAs. Control-sgRNA n=4 mice, Bcl6- and Prdm1-sgRNA n=9 mice. Data presented as mean +SEM. i, TCF-1 expression of Cas9⁺ SMARTA CD4 T cells transduced with MRIG control- or Tcf7-sgRNA after 6 days in culture. j, Representative flow plots of Tfh differentiation of cells from (i) after transfer into WT hosts, on day 6 post-LCMV infection. k, Cas9⁺ SMARTA cells transduced with MRIG Tcf7-sgRNA were spiked into mock-transduced cells at the indicated frequencies, and the mixture was transferred into WT hosts for LCMV infection. Matched mock-transduced and GFP⁺ comparisons of %Tfh among SMARTA cells from each host on day 6 post-LCMV infection. n=2 mice for 5K Tcf7-sqRNA group, n=5 for all other groups. Representative data shown from 1 of 2 independent experiments. *p<0.05; **p<0.01, ***p<0.001, ****p<0.0001 as determined by two-tailed paired (g, k) or unpaired (h) Student's t test. Source data are provided in Source Data file.



Supplementary Figure 2. PID and druggable genes screen details; additional characterization of *Pik3cd-* and *Hif1a-sgRNA SMARTA cells in vivo*

a, Waterfall plot from Fig. 1h with additional genes labeled (all genes \geq 1.2-fold relative enrichment or depletion in Tfh versus Th1). **b**, Gates for sorting Th1, pre-Tfh, and GC Tfh cells in SMARTA cells on day 6-8 p.i. c, Correlations between sgRNAs Z-scores from the druggable target genes screen as performed and analyzed in Fig. 2a. Each symbol represents one gene, averaged over all its sgRNAs. R² values were calculated using Prism least squares regression fitting, without slope or y-intercept constraints. d, Quantification of GFP⁺ SMARTA, Tfh, and Th1 cells from control- versus *Pik3cd*-sgRNA transduced Cas9⁺ SMARTA cells, on day 6 p.i. post-LCMV infection, n=4 mice for controlsgRNA, 6 for *Pik3cd*-sgRNA. e, Total GFP⁺ SMARTA numbers, Tfh percentages and cell numbers, Th1 percentages and cell numbers, and ratios of Tfh:Th1 cells from two independent experiments of control- versus *Hif1a*-sgRNA transduced Cas9⁺ SMARTA cells, on day 6 post-LCMV infection, n=6 mice for control-sgRNA, 8 for Hif1a-sgRNA. For Hif1a sgRNAs, circles, squares, triangles, and diamonds represent sgRNAs #1, 2, 3, and 4, respectively. f, Representative flow plots of Foxp3 staining in control- or Hif1a-sgRNA SMARTA cells, after transfer into WT hosts, compared to host endogenous GFP^{neg} CD4 T cells, on day 7 post-LCMV infection. Data in (d) and (e) are presented as mean values +SEM. Representative data for (d-f) shown from 1 of at least 2 independent experiments. *p<0.05; **p<0.01, ***p<0.001, as evaluated by two-tailed unpaired Student's *t* test (d-e). Source data are provided in Source Data file and Supplementary Data 3 and 5.



Supplementary Figure 3. Additional characterization of and Hif1a-KO polyclonal and SMARTA cells *in vivo*

a, Baseline percentages and numbers of Tfh, GC B, and Tfr cells in mesenteric lymph nodes, spleen, and Peyer's patches of WT or Hif1a^{fl/fl}Cd4-Cre mice, pooled from 2 independent experiments, n=9 mice for WT, 11 for *Hif1a*-KO except n=10 for mesenteric LN Tfh and Tfr plots. Tfh cells were gated as CD4⁺B220⁻CXCR5⁺PD-1⁺Foxp3⁻. GC B cells were gated as B220⁺CD19⁺Fas⁺GL-7⁺, Tfr cells were gated as CD4⁺B220⁻CXCR5⁺PD-1⁺Foxp3⁺. **b**, Quantification of the experiment in Fig. 2d: total SMARTA, Tfh, and Th1 numbers, and the ratio of KO to WT for Tfh and for Th1 populations within each recipient. c, Representative flow plots of TCF-1, BCL-6, T-bet and ICOS staining, in WT versus Hif1a-KO SMARTA cells, on day 8 post-LCMV infection, and guantification of MFI in total SMARTA population, n=6 mice/group for TCF-1, BCL-6 and T-bet stains, n=5 mice/group for ICOS stains. d, Representative flows plots of intracellular IL-21 and surface CD40L staining after in vitro restimulation of WT versus Hif1a-KO SMARTA cells, on day 7 post-LCMV infection, and quantification of percentages in total SMARTA population, n=5 mice/group. e, Representative flow plots, Tfh and Th1 percentages and total numbers, and ratio of total Tfh to Th1 cells, in WT versus *Hif1a*-KO SMARTA, on day 3 post-LCMV infection, n=4 mice for WT, 5 for *Hif1a*-KO. **f**, Frequencies and total numbers of isotype switched (IgD⁻IgM⁻) B cells and IgG2a⁺ B cells, ratio of dark zone to light zone in GC B cells, in the experiment in Fig. 2f. Data in (a), (c)-(f) are presented as mean values +SEM. Representative data for (b-f) shown from 1 of at least 2 independent experiments. *p<0.05; **p<0.01, ***p<0.001, ****p<0.0001 as evaluated by two-tailed unpaired Student's *t* test. Source data are provided as a Source Data file.



Supplementary Figure 4. Characterization of Vhl-sgRNA SMARTA cells in vivo

a, GSEA analysis of the "Hallmark Hypoxia" gene set using publicly available Tfh gene expression data (Moriyama GSE56883 and Yusuf GSE21380). b, Representative flow plots of HIF-1a staining in Cas9⁺ SMARTA cells transduced with control- or Vhl-sgRNA, transferred into WT hosts, on day 6 post-LCMV infection. n=5 mice for control-sgRNA, n=4 for Vhl-sgRNA. c, total number of GFP⁺ SMARTA cells, percent GFP⁺ SMARTA Th1 cells, total GFP⁺ SMARTA Th1 cells, and ratio of #GFP⁺ SM Tfh to #GFP⁺ SM Th1 cells, from the experiment in Fig. 3c. d, Representative flow plots of ICOS and e, BCL-6 and Tbet staining and MFI in Cas9⁺ SMARTA cells transduced with control- or Vhl-sgRNAs, on day 6 post-LCMV infection. For MFIs, populations were gated on total GFP⁺Amet⁺ SMARTA cells or on GFP⁺Amet⁺ SMARTA Th1 cells (CXCR5⁻SLAM⁺). n=5 mice/group. f, total number of GFP⁺ SMARTA cells, percent GFP⁺ SMARTA Th1 cells, total GFP⁺ SMARTA Th1 cells, and ratio of #GFP⁺ SM Tfh to #GFP⁺ SM Th1 cells, from the experiment in Fig. 3d. Data in (b-f) are presented as mean values +SEM. Representative data for (b-f) shown from 1 of at least 2 independent experiments. *p<0.05; **p<0.01, ***p<0.001, ****p<0.0001 as evaluated by two-tailed unpaired (b-f) Student's t test. Source data are provided as a Source Data file.



Supplementary Figure 5. HIF-1a related gene expression changes in vivo

a, Volcano plot of differentially expressed genes and b, schematic of select glycolysis pathway components, from comparison of WT and Hif1a-KO GC Tfh SMARTA cells by RNA-seq, on day 8 post-LCMV infection. In (b), metabolites are in grey, genes are colorcoded in green by RNA-seq expression difference. **c**, gRT-PCR measurement of select glycolytic gene mRNA levels in sorted post-LCMV infection Cas9⁺ SMARTA cells, as in Fig. 3c. d, Comparison of total GLUT1 protein staining MFI in Tfh cells, comparing cotransferred CD45.2/.2 Cas9⁺ SMARTA cells transduced with control-sgRNA and CD45.1/.2 Cas9⁺ SMARTA cells transduced with the indicated test sgRNA vectors, on day 6 post-LCMV infection. n=5 mice for control-sqRNA, 4 for *Hif1a*-sqRNA, 3 for *Pik3cd*sgRNA. e, Representative flow plots of total GLUT1 protein staining and MFI in Th1 and Tfh cells, in WT or *Hif1a*-KO Cas9⁺ SMARTA cells transduced with control- or *Vhl*-sgRNA vector, on day 6 post-LCMV infection, n=4 mice for Vhl-sgRNA, 5 for all other groups. Data in (c) and (e) are presented as mean values +SEM. RNA-seg analyses in (a) used the Wald test to generate p-values, which were adjusted for multiple testing using the Benjamini-Hochberg procedure. Representative data for (d-e) shown from 1 of at least 2 independent experiments. *p<0.05; **p<0.01, ***p<0.001, ****p<0.0001 as evaluated by two-tailed unpaired (c, e) or paired (d) Student's t test. Source data are provided as a Source Data file and in Supplementary Data 6.



Supplementary Figure 6. HIF-1a downstream signaling pathways

a. Tfh percentages in GFP⁺ WT or *Hif1a*-KO SMARTA cells transduced with empty or GLUT1-expressing retroviral vectors, on day 6 post-LCMV infection. n=5 mice/group. b, Representative flow plots, Tfh percentages and total GFP⁺Ametrine⁺ Tfh cells, in Cas9⁺ SMARTA cells transduced with indicated sgRNAs, on day 6 post-LCMV infection. The data in (b) came from the same experiment as the one in Fig. 6d. c, Representative flow plots of BNIP3 staining in Cas9⁺ SMARTA cells transduced with control- or Vhl-sgRNAs, on day 6 post-LCMV infection, n=5 mice/group. For MFIs, populations were gated on total GFP⁺Amet⁺ SMARTA cells or on GFP⁺Amet⁺ SMARTA Th1 cells (CXCR5⁻SLAM⁺). d, Phospho-AKT and phospho-FOXO1 staining in WT or *Hif1a*-KO SMARTA T cells cultured under Tfh-like conditions, with or without PI3Kd inhibitor CAL-101, at day 3. e, LC3b GFP/RFP ratios in GC Tfh cells within Cas9⁺ SMARTA cells co-transduced with a LC3b reporter and control- or Hif1a-sgRNA vector, on day 6 post-LCMV infection. Values are pooled from 3 independent experiments n=17 total control; n=19 total Hif1a-sgRNA. GFP/RFP ratios within each experiment were normalized to the average of control-sgRNA samples. Data in (a)-(c) and (e) are presented as mean values +SEM. Representative data for (a-e) shown from 1 of at least 2 independent experiments. ***p<0.001, ****p<0.0001 as evaluated by two-tailed unpaired Student's t test (b, c, e). Source data are provided as a Source Data file.





Supplementary Figure 7. Expanded screen hits and validation.

a, Hits from comparison of pre-Tfh and Th1 cells, in the expanded CRISPR screen described in Fig. 5a. b, Representative flow plots, Tfh and Th1 cells percentages and numbers, and ratio of GFP⁺ SMARTA Tfh to Th1 cells, from Cas9⁺ SMARTA cells transduced with control- or Ambra1-sgRNA, on day 6 post-LCMV infection, n=4 mice for control-sgRNA, 5 for Ambra1-sgRNA. For Ambra1 sgRNAs, squares and triangles represent sgRNAs #1 and 2, respectively. c, d Gene hits related to metabolism (c) and HIF-1a (d) are highlighted in volcano plots comparing pre-Tfh and Th1 cells. e, Total GFP⁺ SMARTA, percentage GFP⁺ SMARTA Th1 cells, total GFP⁺ SMARTA Th1 cells, and ratio of GFP⁺ SMARTA Tfh to Th1 cells, from WT and Hif1a-KO Cas9⁺ SMARTA cells transduced with the indicated sgRNAs, on day 6 post-LCMV infection, n=5 mice/group, for the experiment in Fig. 5d. f, Gene hits related to mTOR analyzed as in c-d. g, TSC2 staining in endogenous WT B cells and transferred Cas9⁺ SMARTA cells transduced with control-sgRNA, on day 6 post-LCMV infection, n=5 mice/group. GC B cells were gated as B220⁺CD19⁺Fas⁺GL-7⁺ cells, and further sub-gated into dark zone (CXCR4⁺CD86⁻) and light zone (CXCR4⁻CD86⁺). SMARTA Th1, pre-Tfh, and GC Tfh were gated as in Supplementary Fig. 2a. h, TSC2 staining in Cas9⁺ SMARTA cells transduced with controlor Vhl-sgRNA, on day 6 post-LCMV infection, n=5 mice/group. Data in (b), (e), (g) and (h) are presented as mean values +SEM. Representative data for (b, e, g, h) shown from 1 of at least 2 independent experiments. *p<0.05; **p<0.01, ***p<0.001, ****p<0.0001 as evaluated by two-tailed unpaired Student's t test (b, e, h). Source data are provided as a Source Data file and in Supplementary Data 8.



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Supplementary Figure 8. Increased mTORC1 activity rescues Tfh differentiation in *VhI*-deficient T cells

a, Total number of gated SMARTA cells, percentages and total numbers of gated Th1 cells, ratio of gated Tfh to Th1 cells, for the experiment in Fig. 6a. SMARTA cells were gated on GFP⁺Ametrine⁺, except for the *Tsc2*-sgRNA group, which was gated on GFP⁺. b, Total number of GFP⁺ SMARTA cells, percentages and total number of GFP⁺ SMARTA Th1 cells, and ratio of GFP⁺ SMARTA Tfh to Th1 cells, from the experiment in Fig. 6b. c. Representative flow plots and quantification of ICOS staining for the experiment in Fig. 6b. d, Phospho-S6 staining in WT or Vhl-KO SMARTA cells transduced with empty or caRHEB-expressing vector and cultured in vitro for 3d. e, Tfh cells percentages from Cas9⁺ SMARTA cells transduced with the indicated sgRNA vectors, on day 6 post-LCMV infection, n=5 mice for control- and Vhl-sgRNA, 4 for Bnip3-sgRNA, 6 for Vhl-sgRNA + Bnip3-sgRNA. f, Tfh cell percentages from WT or Hif1a-KO SMARTA cells transduced with empty or BNIP3-expressing vector, on day 6 post-LCMV infection, n=5 mice/group. **q**. Total numbers of GFP⁺Ametrine⁺ cells and GFP⁺Ametrine⁺ Tfh and Th1 cells from Cas9⁺ SMARTA cells transduced with the indicated sgRNA vectors, on day 6 post-LCMV infection, n=5 mice/group, from the experiment in Fig. 6d. Data in (a)-(c), (e)-(g) are presented as mean values +SEM. Representative data for (a-g) shown from 1 of at least 2 independent experiments. *p<0.05; **p<0.01, ***p<0.001, ****p<0.0001 as evaluated by two-tailed unpaired Student's t test (a, b, c, g). Source data are provided as a Source Data file.



Supplementary Figure 9. Vhl is required for optimal Myc and cell proliferation.

a-b, WT or Vhl-KO SMARTA cells were cultured under Tfh-like conditions and evaluated for size by FSC (a) and GLUT1 expression (b). c, Representative CellTrace Violet dilution in Tfh and Th1 cells from WT or Hif1a-KO SMARTA cells transduced with control- or Vhl-sgRNA and transferred into WT hosts, day 3 post-LCMV infection. d, Representative viability dye staining in total SMARTA, Th1, or Tfh cells from WT or Hif1a-KO SMARTA cells transduced with controlor Vhl-sgRNA, day 3 post-LCMV infection, as in (c), n=4 mice for WT + control-sgRNA and WT + Vhl-sgRNA groups, 5 for Hif1a-KO + control-sgRNA and Hif1a-KO + Vhl-sgRNA /groups. Ratio of %dead among Tfh divided by %dead among Th1, for each group. e, WT or Hif1a-KO SMARTA cells were cultured under Tfh-like conditions, and stained as indicated on day 3. f, gRT-PCR measurement of Myc mRNA levels in sorted p.i. SMARTA cells, as described in Fig. 2c. g, WT or Vhl-KO SMARTA cells were stained with CellTrace Violet, transduced with empty or caRHEBexpressing vector at 24h post activation, and cultured in vitro, day 3. h, Phospho-AKT and phospho-FOXO1 staining in WT or Vhl-KO SMARTA cells cultured under Tfh-like conditions, day 3. Data in (d) and (f) are presented as mean values +SEM. Representative data for (a-e, q, h) shown from 1 of at least 2 independent experiments. *p<0.05 as evaluated by two-tailed unpaired Student's t test (d, f). Source data are provided as a Source Data file.







Supplementary Figure 10. Expanded screen reveals feedback regulation of mTORC1 signaling through HIF-1a and Myc

Network map of signaling components in mTOR/HIF-1a pathways, including canonical genes and genes found to be statically significant hits (thick grey border) from the expanded CRISPR screen as described in Fig. 5a. Genes from the expanded screen are in ovals and Z-scores of GC Tfh versus Th1 enrichment is color-coded by shading as shown in the legend. Genes identified by the druggable screen or individual sgRNAs are indicated in blue or red boxes. Genes known from the literature are outlined in blue or red ovals.

FIG S11



Supplementary Figure 11. Gating strategies for in vivo experiments

a, Gating strategy from total cells to live cells. The initial gating strategy included: FSC-A/SSC-A, exclusion of doublets (through FSC-H/FSC-W then SSC-H/SSC-W), and live cells (negative for Aqua or Blue fixable viability stain). These were followed by specific gating strategies. b, Gating strategy from live cells to CD4 T cells. c, Gating strategy from CD4 T cells to GFP⁺ SMARTA cells, then Th1 and Tfh cells (by SLAM vs CXCR5, d6-8 post-infection), or Th1, pre-Tfh, and GC Tfh cells (by PD-1 vs CXCR5, d6-8 postinfection), or Tfh and Th1 cells (by TIM-3 vs CXCR5, d3 post-infection). Used for analysis in Fig 1c, 2b, 3c-d, 5d, 6b, Supplementary Fig. S1g-h, S1j-k, S2d-f, S4b, S4c, S4f, S5de, S6a, S7b, S7e, S8b, S8-c, S8f, S9c, S9d. Used for sort for Fig 1d-h (PID genes screen, SLAM vs CXCR5), Fig 2a, Supplementary Fig S2a-c (druggable gene targets screen, PD-1 vs CXCR5), Fig 2c, 4d-e, Supplementary Fig S5a-c, S9f (RT-PCR samples, PD-1 vs CXCR5), 5a-c, 5e, Supplementary Fig S7a, S7c, S7d, S7f, S10 (expanded screen, PD-1 vs CXCR5). Post-sort fractions had higher than 95% purify, as verified by flow cytometry analysis on the same FACSAria machine used to sort the cells. d. Gating strategy from CD4 T cells to GFP⁺Ametrine⁺ SMARTA cells and GFP⁺ SMARTA cells, then Tfh and Th1 cells. GFP⁺Ametrine⁺ SMARTA cells gating used for Fig. 3e, 6a (control + control sgRNAs, Vhl + Vhl sgRNAs, and Tsc2 + Vhl sgRNAs conditions), 6d, Supplementary Fig. S4d, S4e, S6b, S6c, S7g (right), S7h, S8a (control + control sgRNAs, Vhl + Vhl sgRNAs, and Tsc2 + Vhl sgRNAs conditions), S8e, S8g. GFP⁺ SMARTA cells gating used for Fig. 6a (Tsc2-sgRNA only condition) and Fig. S8a (Tsc2-sgRNA only condition). e, Gating strategy from live CD4 T cells to co-transferred WT and Hif1a-KO SMARTA cells, then Tfh and Th1 cells. Used for Fig. 2d-e. f, Gating strategy from live CD4 T cells to SMARTA cells, then Tfh and Th1 cells. Used for analysis in Fig. 3b, Supplementary Fig. S3b-e, S4a. Used for sort for Fig 4a-c, Supplementary Fig S5a, S5b (RNA-seq samples). g, Gating strategy from live cells to plasma cells. Used for Fig 2f. h, Gating strategy from live cells to B cells, to GC B cells, to DZ and LZ GC B cells, used for Fig. 2f, 3b, Supplementary Fig. S3f, S7g (left). i, Gating strategy from B cells to isotype-switched B cells, to IgG2a+ switched B cells. Used for Supplementary Fig. S3f.