Supplemental Materials for

Title: Platelets mediate inflammatory monocyte activation by SARS-CoV-2

Spike protein

Authors:

Tianyang Li¹, Yang Yang¹, Yongqi Li¹, Zhengmin Wang¹, Faxiang Ma¹, Runqi Luo², Xiaoming Xu², Guo Zhou², Jianhua Wang³, Junqi Niu⁴, Guoyue Lv^{4*}, Ian Nicholas Crispe⁵, Zhengkun Tu^{1,4*}.

Affiliations:

¹ Institute of Translational Medicine, The First Hospital of Jilin University, Changchun, Jilin, China.

² Infectious Disease, The Second Affiliated Hospital of Guangzhou Medical University, Guangzhou, Guangdong, China.

³ Guangzhou Institutes of Biomedicine and Health, Chinese Academy of Sciences, Guangzhou, Guangdong, China.

⁴ Institute of Liver Diseases, The First Hospital of Jilin University, Changchun, Jilin, China.

⁵ Department of Laboratory Medicine and Pathology, University of Washington, Seattle, WA, USA

*Correspondence to: Zhengkun Tu, MD. Email: tuzhengkun@jlu.edu.cn; Guoyue Lv, MD, PhD. Email: lvgy@jlu.edu.cn.

This PDF file includes:

Supplemental Figure 1-8

Supplemental Methods

Cell line

Huh7 cells which endogenously express ACE-2 (1) were maintained in Dulbecco's modified Eagle's medium (DMEM) containing 10% heat-inactivated fetal bovine serum (FBS) before flow cytometry, as described in Methods.

Platelet aggregation assay

Platelet aggregations were measured on an optical aggregometer (AG800, Techlink, Beijing) at 37°C as described (2). Briefly, 300 μ l of PRP was stimulated by S protein (1 μ g/mL) or S-pseudovirus (80 TCID₅₀/mL) and light transmission was recorded until trace stabilization.

Biolayer interferometry

Biolayer interferometry analysis was performed on an Octet RED96 (ForteBio) as described (3). Spike RBD protein (20 μ g/mL) were immobilized on AMC sensors to maximum loading levels. Associations and dissociations were performed in PBS supplemented with 0.1% BSA, 0.02% Tween20 at 30°C after sensors were washed to a obtained baseline. K_D value was analyzed using DataAnalysis9 (ForteBio).

Platelet-derived dense granule secretion assay

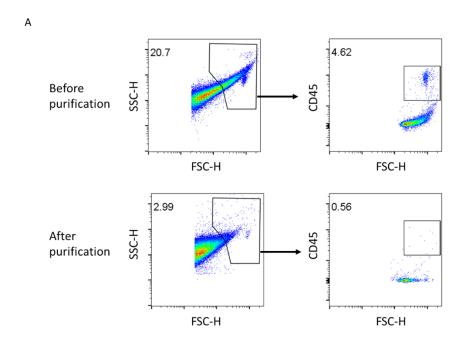
Platelet ATP/ADP secretion from dense granules was detected as described (4). Briefly, 80 µl of PRP was incubated with or without S protein and collagen at indicated concentrations for 5 min at room temperature on an orbital shaker, and 10 µl of Chrono-

lume (Chrono-Log) were injected into each well. Luminescence was detected by Synergy H1 (Biotek).

Enzyme-linked immunosorbent assay (ELISA)

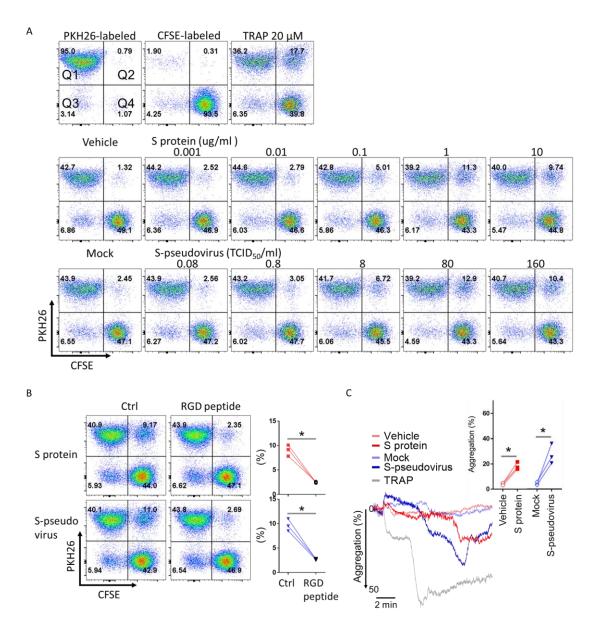
The monocyte culture supernatants were collected, and the concentration of IL-1 β , TNF- α and IL-10 were measured by ELISA Ready-Set-Go® (eBioscience, San Diego, USA) according to the manufacturer's instructions (5).

Supplemental Figures



Supplemental Figure 1: Leukocyte contaminants in purified platelets.

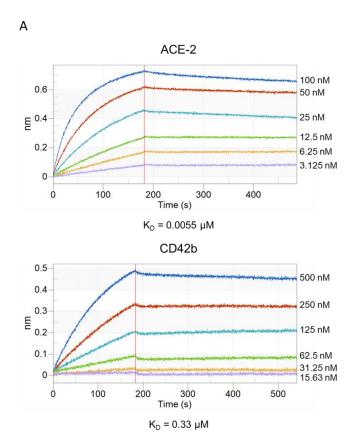
(A) Before and after purification, platelets were stained by anti-CD45 and analyzed by flow cytometry, and leukocyte contaminants were shown.



Supplemental Figure 2: SARS-CoV-2 induced platelet aggregation.

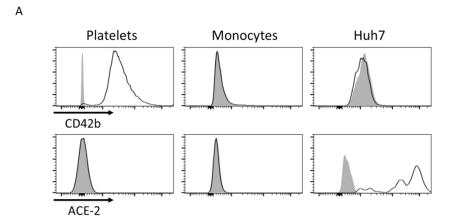
(A) Purified platelets were labeled by PKH26 or CFSE and mixed 1:1 before incubation with S protein or S-pseudovirus at indicated concentration. Platelets stained with PKH26 are enclosed in Quadrant 1. Platelets stained with CFSE are enclosed in Quadrant 4. Double-colored events in Quadrant 2 represent platelet aggregates before and after stimulation. Statistical analysis was displayed in Figure 1A. (B) Purified platelets were labeled by PKH26 or CFSE and mixed 1:1 before incubation with S protein (1.0 μg/mL) or S-pseudovirus (80 TCID50/mL) in the presence or absence of

RGD peptide. (C) Representation and quantification of light transmission curve of platelets aggregation stimulated by S protein or S-pseudovirus were shown. Mean with SD and P value by paired Student's t test are displayed. * Indicated P value < 0.05.



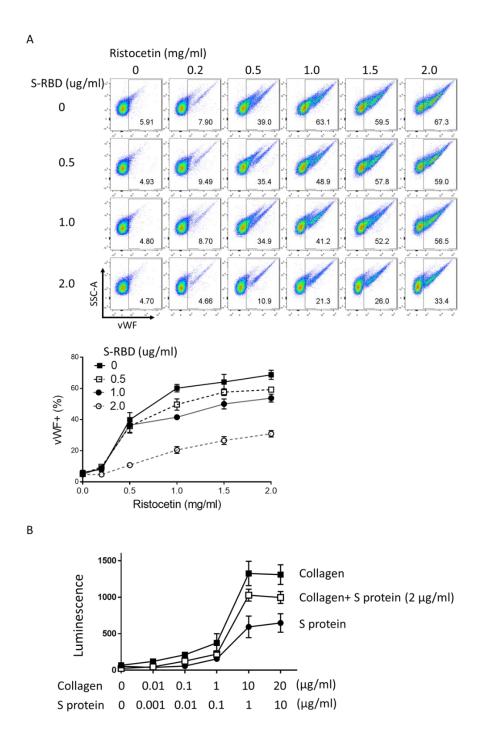
Supplemental Figure 3: The binding affinity of recombinant Spike protein to recombinant CD42b and ACE2 were characterized by kinetics.

(A) Binding curves of immobilized Spike RBD with the ACE-2 (upper) and CD42b (lower) are shown. K_D are analyzed by fitting the data to a 1:1 binding model.



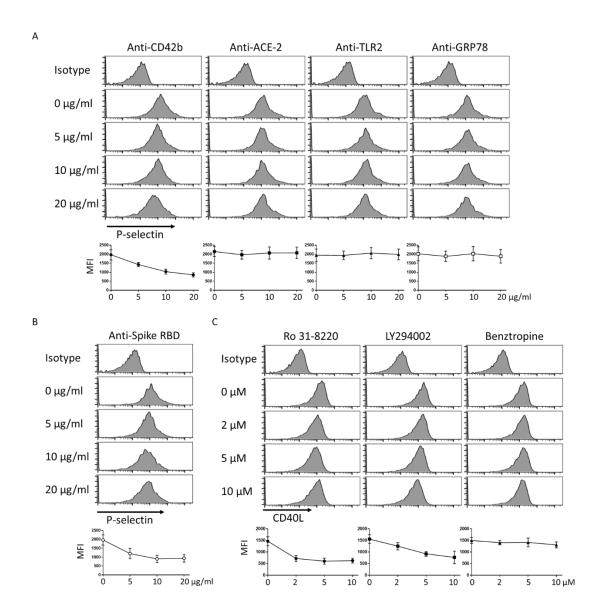
Supplemental Figure 4: ACE-2 was not detected on platelets.

(A) ACE-2 and CD42b expression on purified platelets, monocytes and the Huh7 cell line (which endogenously expresses ACE-2) was tested by flow cytometry.



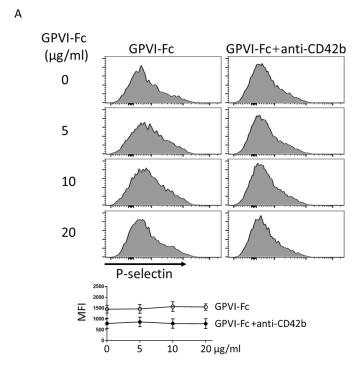
Supplemental Figure 5: Spike protein impeded vWF binding on platelets.

(A) Platelet-vWF complexes were detected by flow cytometry using recombinant vWF and ristocetin. MFI analysis was shown in Figure 2F. (B) Platelet-rich plasma was incubated with S protein and collagen at indicated concentrations, and luminescence derived from ATP/ADP release were measured. Data are reported as the mean with SD.



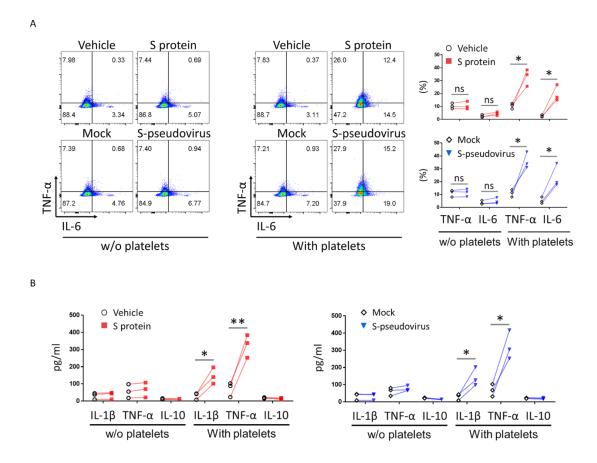
Supplemental Figure 6: Dose response curves for inhibitory antibodies and inhibitors.

Purified platelets were incubated with indicated inhibitory antibodies before stimulation with 1 μ g/ml of S protein (**A**, **C**) or with 80 TCID₅₀/mL of S-pseudovirus (**B**). P-selectin or CD40L expression was detected by flow cytometry. Data are reported as the mean with SD.



Supplemental Figure 7: Blockade of GPVI did not affect Spike protein binding with CD42b-induced P-selectin expression.

(A) Purified platelets were incubated with S protein (1 μ g/ml) and GPVI-Fc chimera protein at indicated concentrations in absence or presence of anti-CD42b (20 μ g/ml). P-selectin expression was detected by flow cytometry. Data are reported as the mean with SD.



Supplemental Figure 8: SARS-COV-2-activated platelets mediated IL-6 and TNF- α expression in monocytes.

(A) Purified monocytes (n=3) were cultured and co-cultured as described in Figure 4B, and IL-6 and TNF- α expression is shown. (B) IL-1 β , TNF- α and IL-10 levels in supernatants from monocyte culture were measured by ELISA. P value by paired Student's t test is displayed. * indicated P value < 0.05, ** P < 0.01, ns indicated not significant.

Supplemental references

- 1. Lai ZW, et al. The identification of a calmodulin-binding domain within the cytoplasmic tail of angiotensin-converting enzyme-2. *Endocrinology*. 2009;150(5):2376-81.
- 2. Dai B, et al. Integrin- α IIb β 3-mediated outside-in signalling activates a negative feedback pathway to suppress platelet activation. *Thromb Haemost*. 2016;116(5):918-30.
- 3. Tortorici MA, et al. Structural basis for human coronavirus attachment to sialic acid receptors. *Nat Struct Mol Biol*. 2019;26(6):481-9.
- 4. Sun B, et al. Luminometric assay of platelet activation in 96-well microplate.

 *Biotechniques. 2001;31(5):1174, 6, 8 passim.
- 5. Li H, et al. Regulatory NK cells mediated between immunosuppressive monocytes and dysfunctional T cells in chronic HBV infection. *Gut*. 2018;67(11):2035-44.