

Cell Reports, Volume 37

Supplemental information

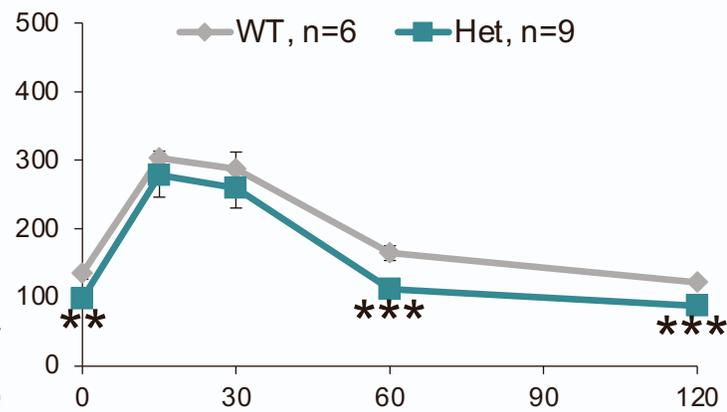
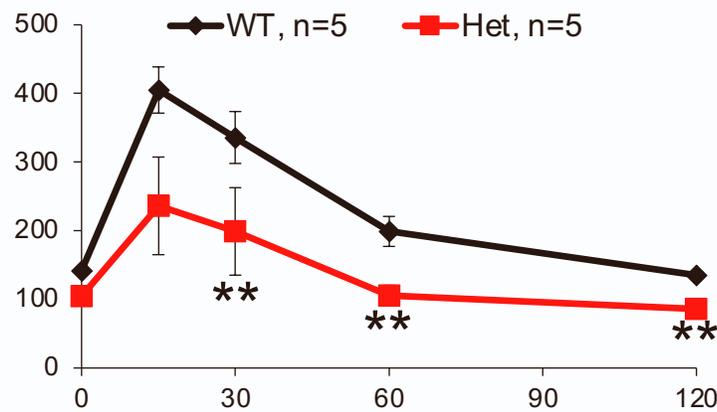
**Sex-biased islet β cell dysfunction is caused
by the MODY MAFA S64F variant by inducing
premature aging and senescence in males**

Emily M. Walker, Jeeyeon Cha, Xin Tong, Min Guo, Jin-Hua Liu, Sophia Yu, Donato Iacovazzo, Franck Mauvais-Jarvis, Sarah E. Flanagan, Márta Korbonits, John Stafford, David A. Jacobson, and Roland Stein

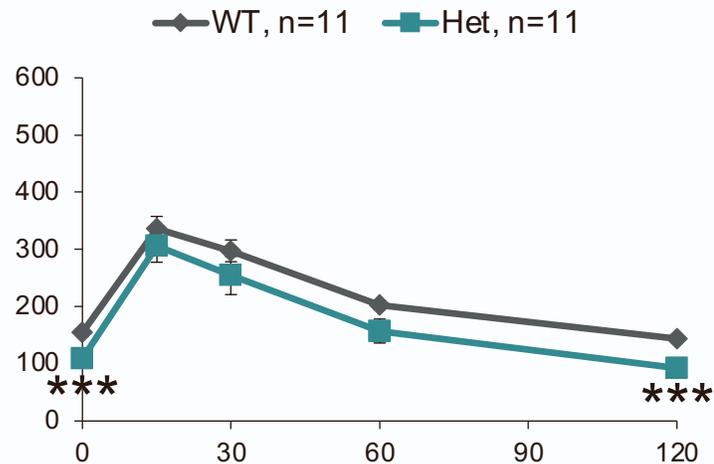
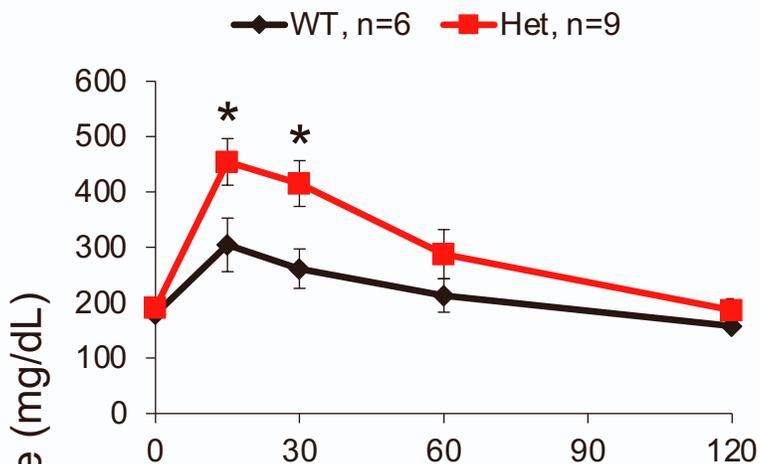
Male

Female

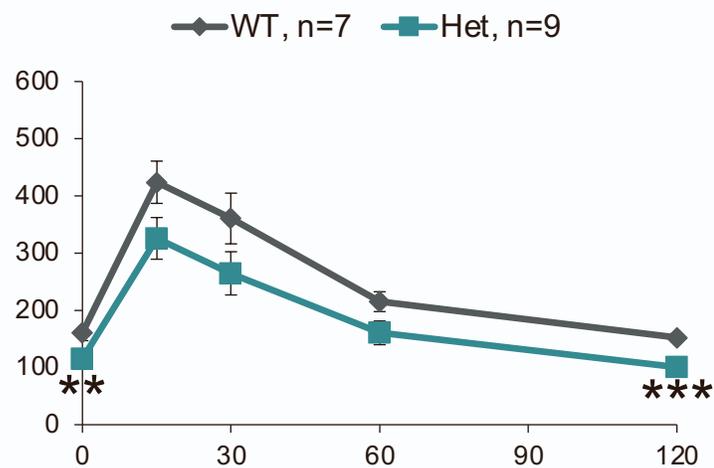
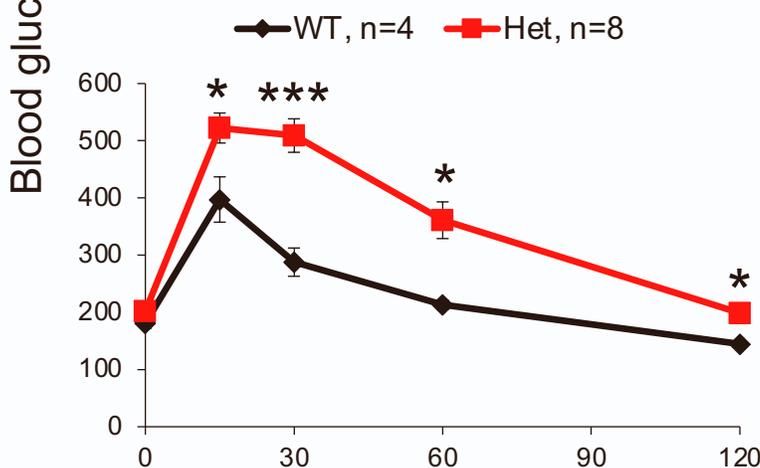
3 weeks



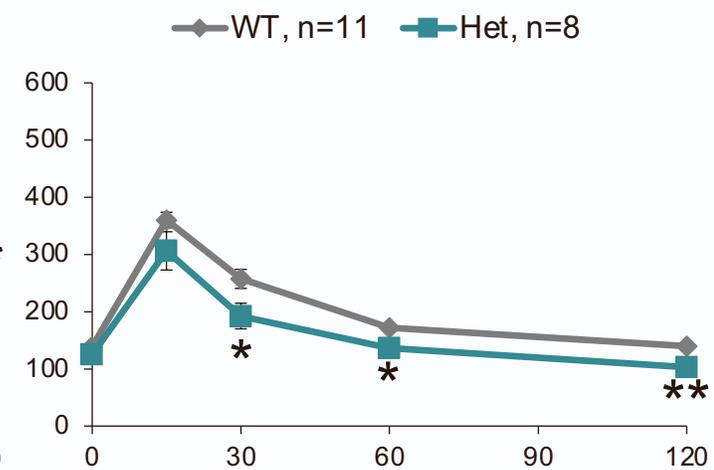
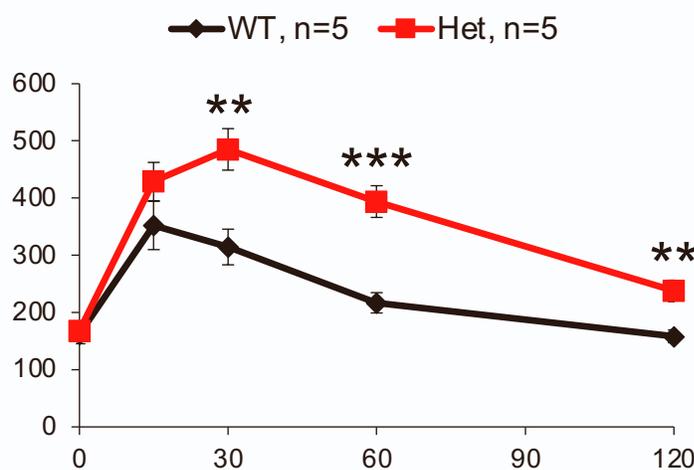
6 weeks



7 weeks

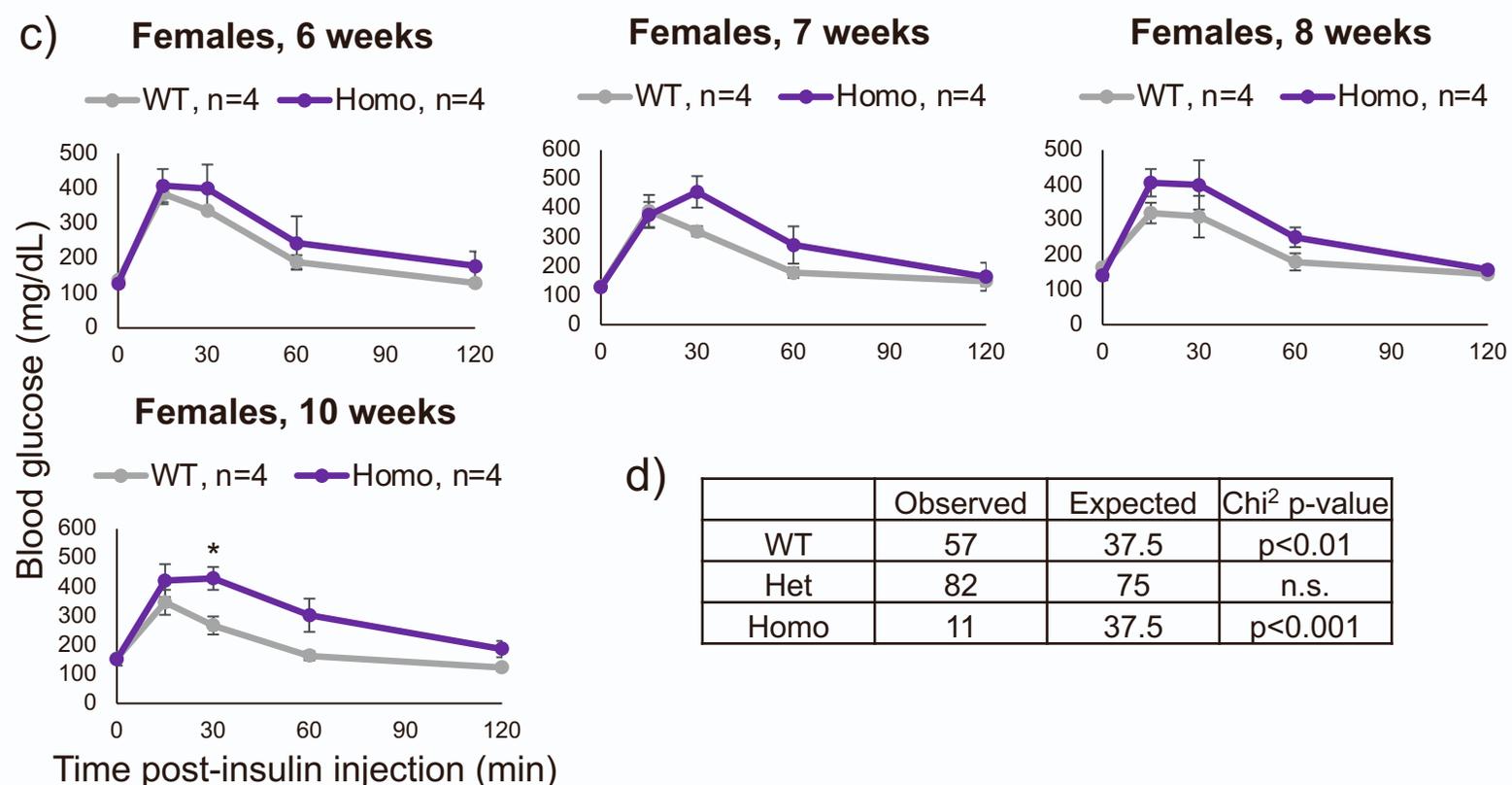
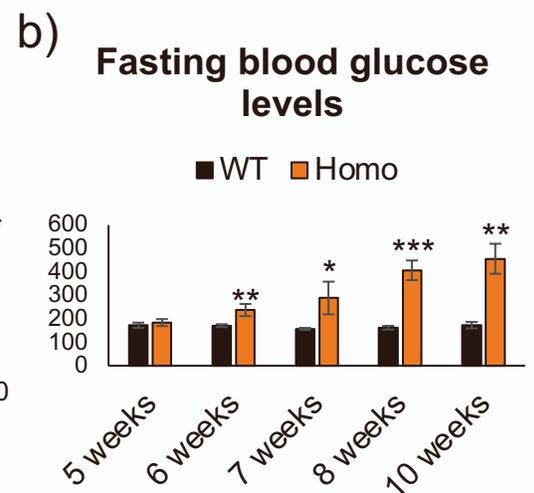
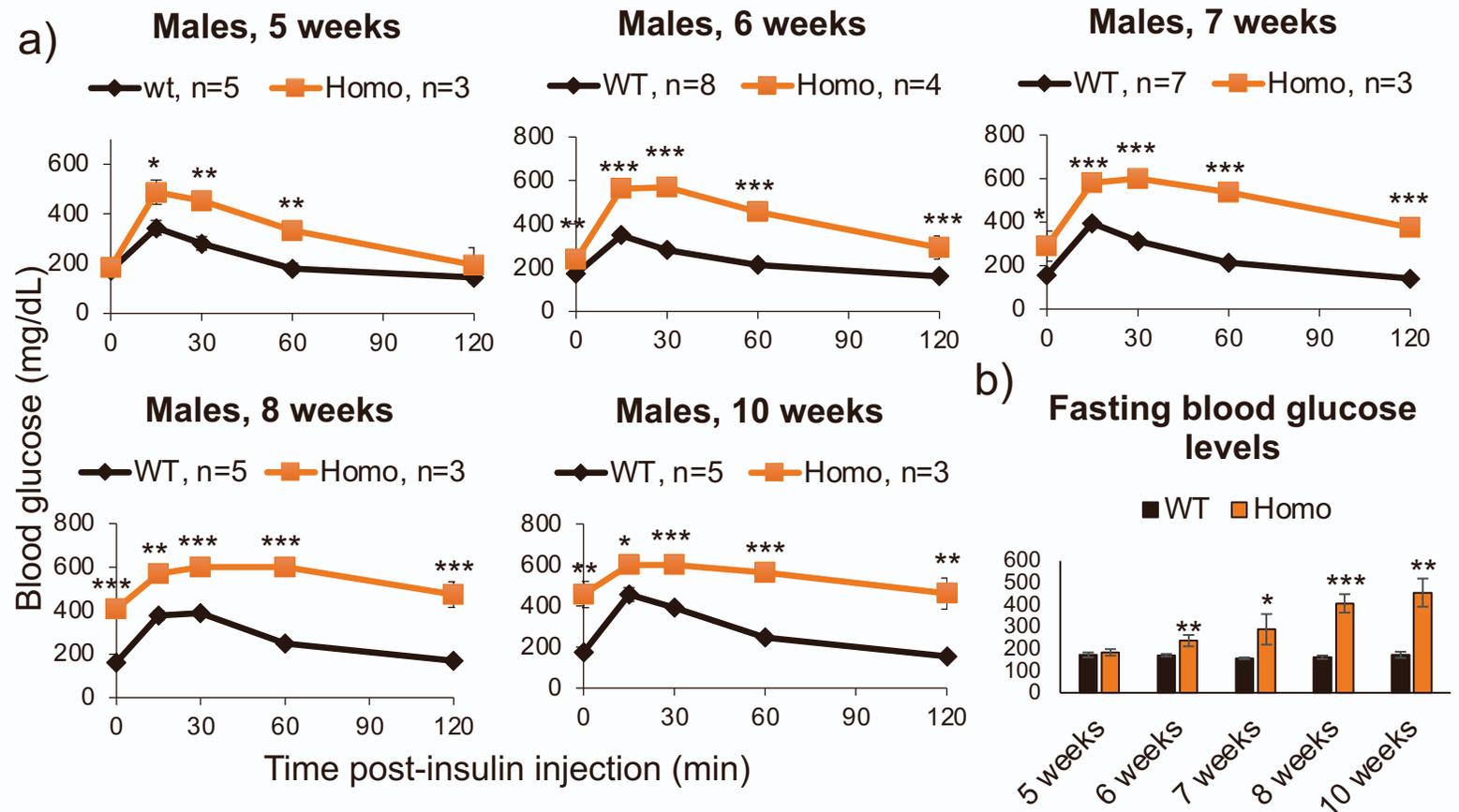


16 weeks



Time post-glucose injection (min)

Figure S1: Glucose tolerance phenotypes were stably maintained in male and female *MafA*^{S64F/+} mice. Related to Figure 1. GTT was performed in 3-, 6-, 7-, and 16-week-old mice. Glucose (2mg/kg) was injected following a 6 hour fast and blood glucose was measured at the indicated time points. Two-tail Student *t* test; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.



d)

	Observed	Expected	Chi ² p-value
WT	57	37.5	p<0.01
Het	82	75	n.s.
Homo	11	37.5	p<0.001

Figure S2: Homozygous male *MafA*^{S64F/S64F} mice were hyperglycemic, with glucose tolerance and blood glucose levels worsening with age. Related to Figure 1. a) *MafA*^{S64F/S64F} mice showed progressively worsening glucose tolerance and fasting blood glucose levels between 5 to 10 weeks of age. b) Fasting blood glucose levels in homozygous male mutant mice increased significantly over time. c) Female S64F *MafA* homozygous mice were only mildly glucose intolerant at 10 weeks of age although their temporal responses to glucose was variable between animals. d) Chi-square analysis revealed that *MafA*^{S64F/S64F} male and female animals were observed with significantly less frequency at weaning than WT animals. In contrast, *MafA*^{S64F/+} numbers were as expected. Two-tail Student *t* test; **p*<0.05; ***p*<0.01; ****p*<0.001.

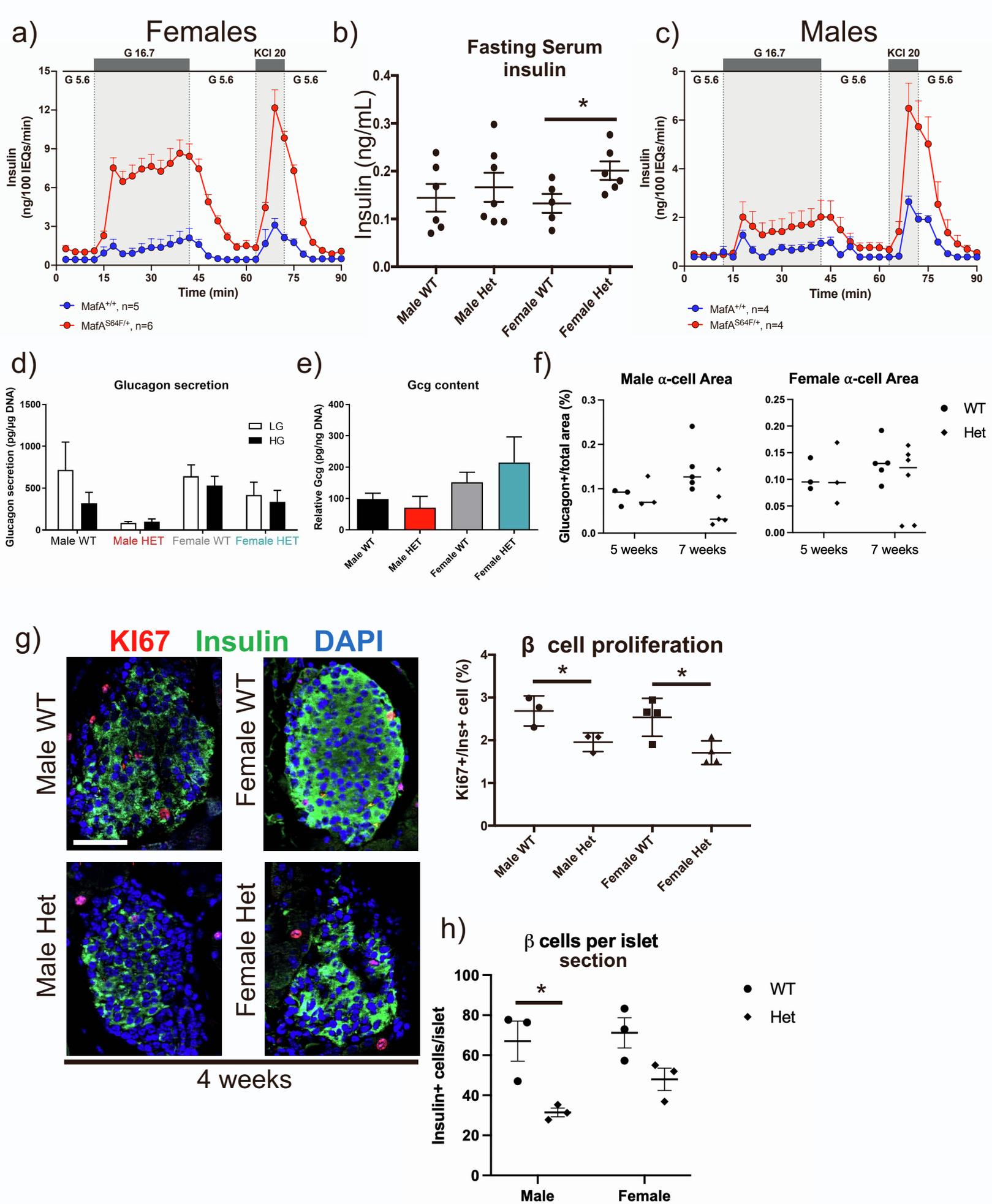


Figure S3: Female and male *MafA*^{S64F/+} islets secreted more insulin in response to high glucose with decreased proliferation and no change in glucagon secretion. Related to Figure 1. a) Islet perfusion demonstrated islets from *MafA*^{S64F/+} female mice secreted higher levels of insulin at low (G 5.6) and high (G 16.7) glucose and in response to KCl. b) Serum insulin levels (ng/mL) were higher in 6-hour fasted female *MafA*^{S64F/+} animals while male *MafA*^{S64F/+} levels were unchanged. c) Islet perfusion results from *MafA*^{S64F/+} male mice at high (G 16.7) glucose and in response to KCl. Islets were isolated from 8-10 week-old animals in both genders. Glucagon secretion (d) and content (e) were unchanged in *MafA*^{S64F/+} islets, which were incubated in 4.6 mM (LG) and 16.7 mM (HG) glucose for 1 hour prior to collection. Secretion and content were normalized to DNA. f) Islet α cell area was not reduced in male or female *MafA*^{S64F/+} mice, though measurements were highly variable between samples. Islet α cell area was calculated by dividing the total glucagon⁺ area by the total pancreas area (eosin staining) multiplied by 100 to obtain percent (%). Two-way ANOVA. g) Representative islets stained for insulin, Ki67 (proliferation marker), and DAPI (nuclei). Islet β cell proliferation was calculated by dividing the number of Ki67⁺ cells by total insulin⁺ β cells. Greater than 1000 β cells were counted per animal. h) β cell number per islet section was significantly reduced in *MafA*^{S64F/+} male mice calculated by dividing the total number of Insulin⁺ cells by the number of islet sections (n \geq 20 islets per sample) quantitated. Two-tail Student *t* test; *p<0.05.

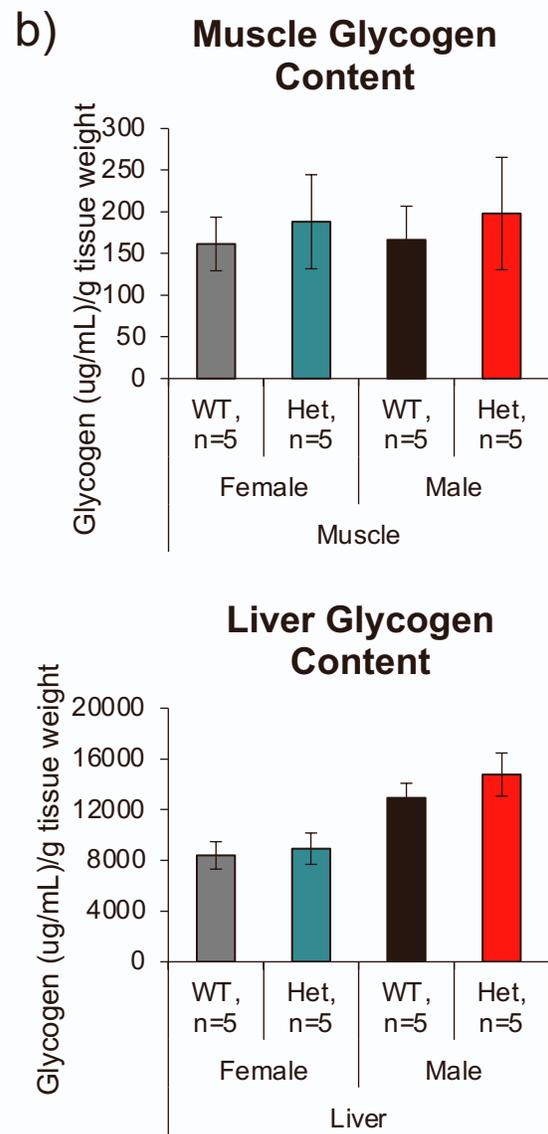
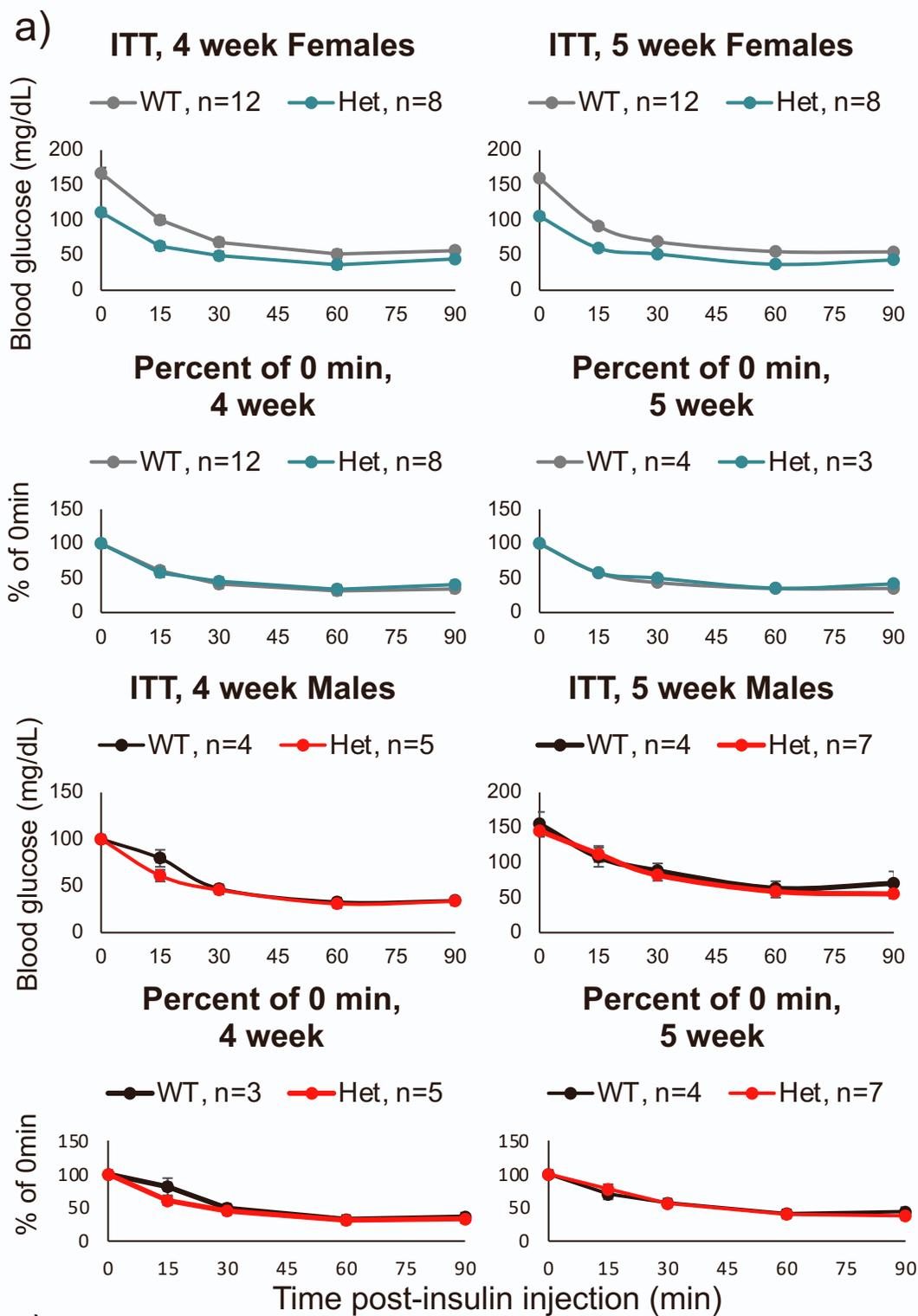


Figure S4: Insulin tolerance and glycogen content are unaltered in *MafA*^{S64F/+} animals. Related to Figure 1. a) Insulin tolerance was unchanged at 4 and 5 weeks in *MafA*^{S64F/+} animals once corrected for initial fasted blood glucose levels (Percent of 0 min). Insulin (0.5U/kg body weight) was injected following a 6-hour fast. b) Glycogen content in muscle (soleus) and liver, normalized to wet tissue weight, was also unaffected in 7-week-old *MafA*^{S64F/+} mice. c) There was no change in average body weight between 3-6 weeks except a small decrease (-1.07-fold) in female *MafA*^{S64F/+} mice at 5 weeks of age. Two-tail Student *t* test; *p<0.05.

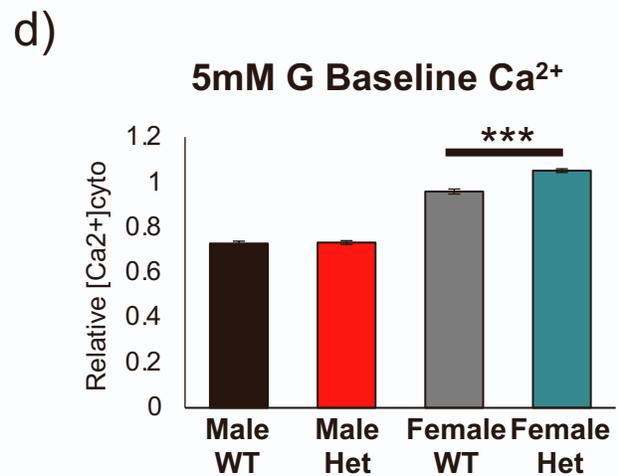
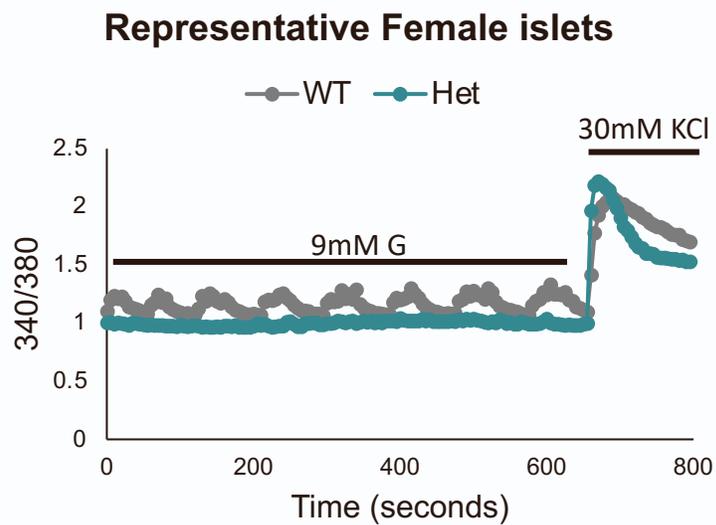
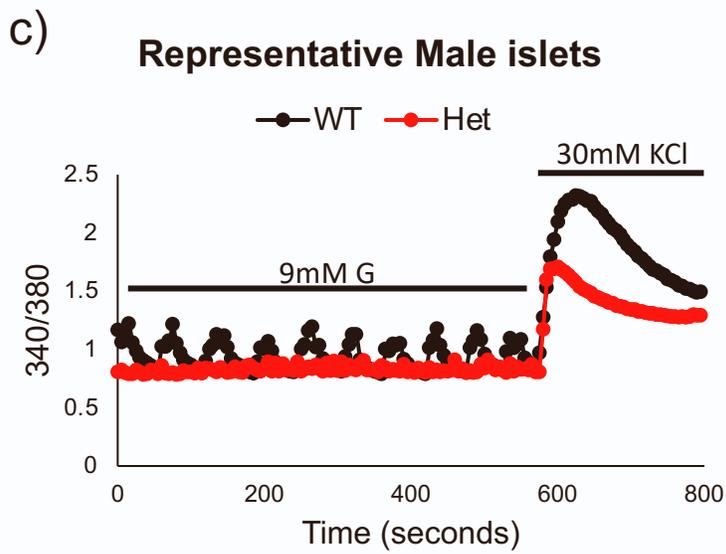
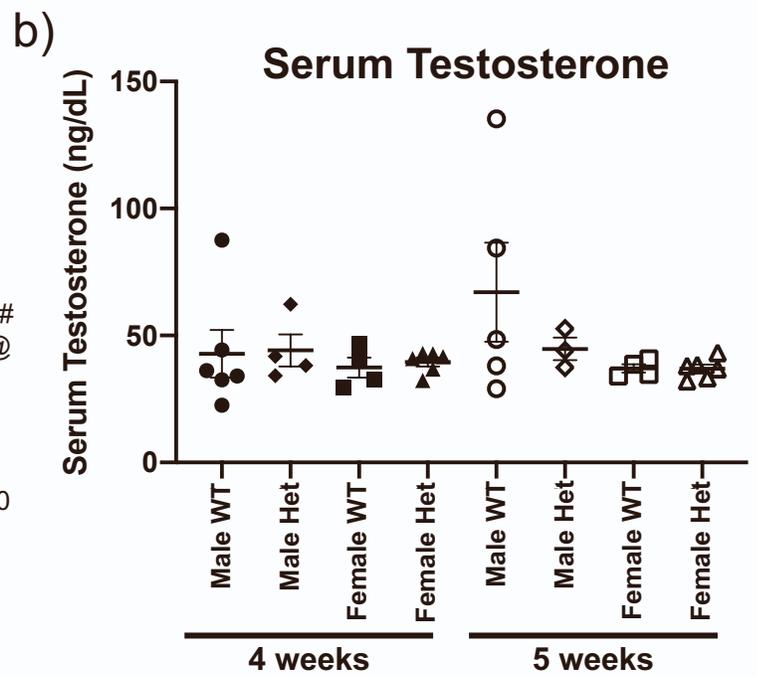
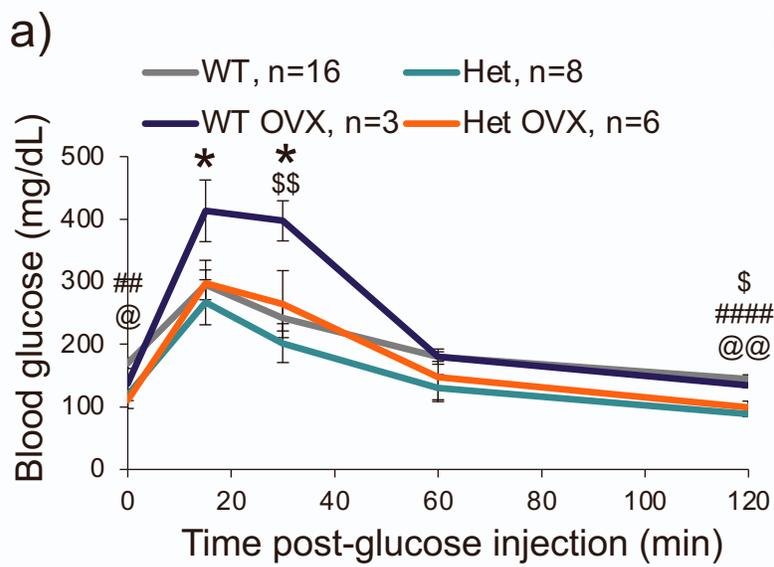


Figure S5: Ovariectomy (OVX) did not alter glucose tolerance in female *MafA*^{S64F/+} mice, nor are testosterone levels changed while male *MafA*^{S64F/+} islets had a reduced response to KCl stimulation. Related to Figures 1 and 4. a) Compared to 4-week-old control mice (i.e., non OVX: WT, grey line; Het, teal line), the GTT at 1-week post ovariectomy showed glucose intolerance in WT female mice (WT OVX, purple line) and no change in *MafA*^{S64F/+} mice (Het OVX, orange line). One-way ANOVA; WT vs. WT OVX: *p<0.05; Het vs. WT OVX: \$p<0.05; \$\$p<0.01; WT vs. Het: ##p<0.01; ####p<0.0001; WT vs. Het OVX: @p<0.05; @@p<0.01. b) Serum testosterone levels were unchanged in male or female *MafA*^{S64F/+} mice. Male and female testosterone levels are similar at 4 and 5 weeks of age. Two-way ANOVA. c) Representative Ca²⁺ traces quantitated in Figure 4c in response to both high glucose (9 mM) and KCl (30 mM). d) Female *MafA*^{S64F/+} islets have increased baseline Ca²⁺ at 5mM compared with WT islets (right) while male *MafA*^{S64F/+} islets do not (left).

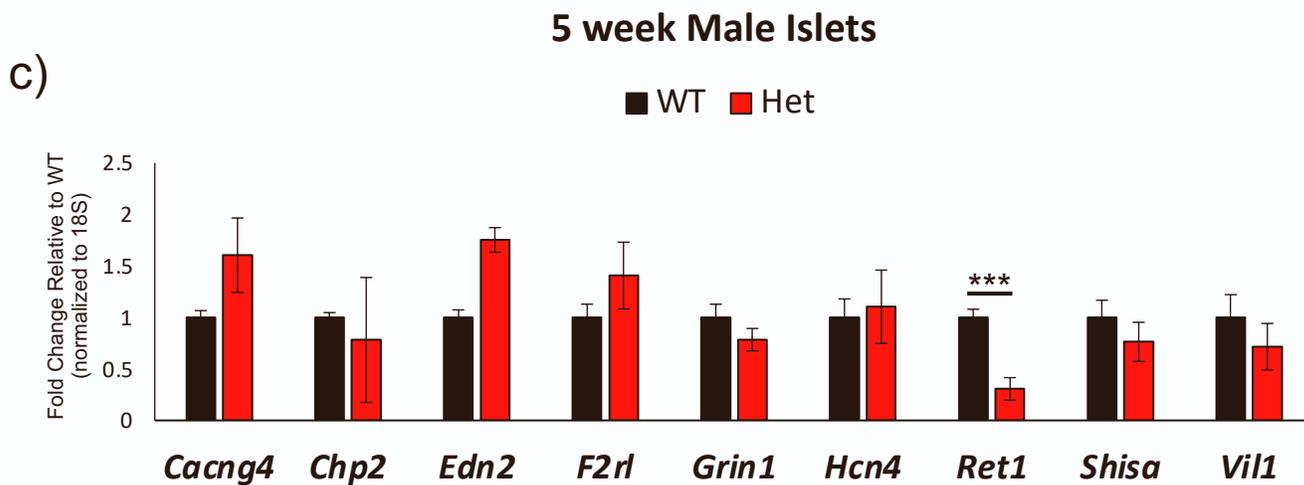
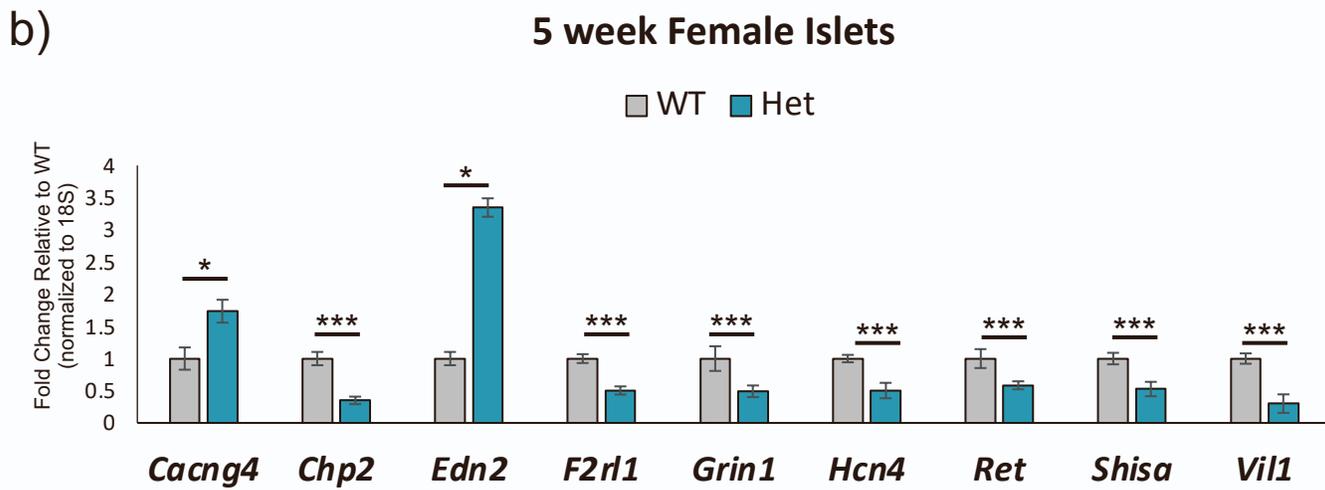
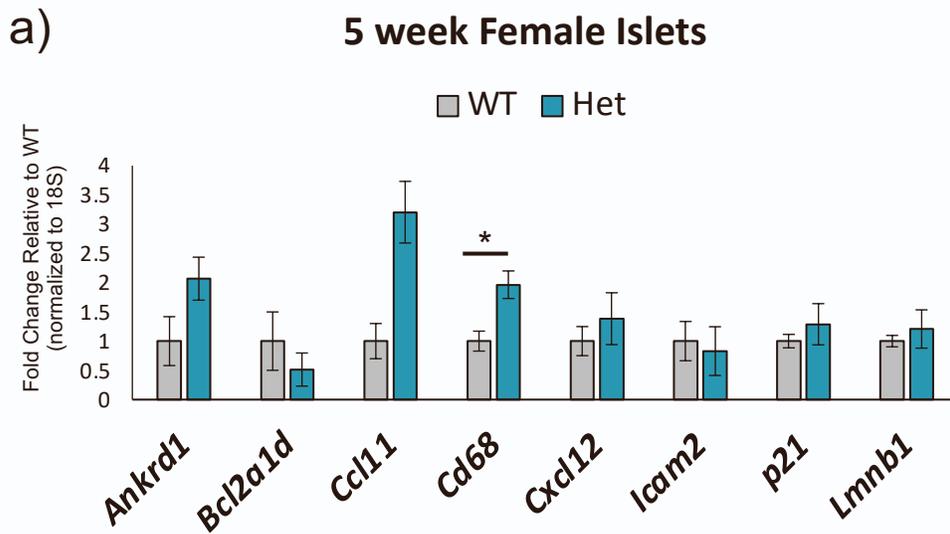


Figure S6: qPCR confirmation of changes in islet gene expression identified by RNA-seq. Related to Figure 5. a) Senescence markers were primarily unchanged in female *MafA*^{S64F/+} islets. b-c) Genes altered specifically in female *MafA*^{S64F/+} islets (b) identified by RNA-seq were mostly unchanged in male *MafA*^{S64F/+} islets (c). Two-tail Student *t* test; **p*<0.05; ****p*<0.001.

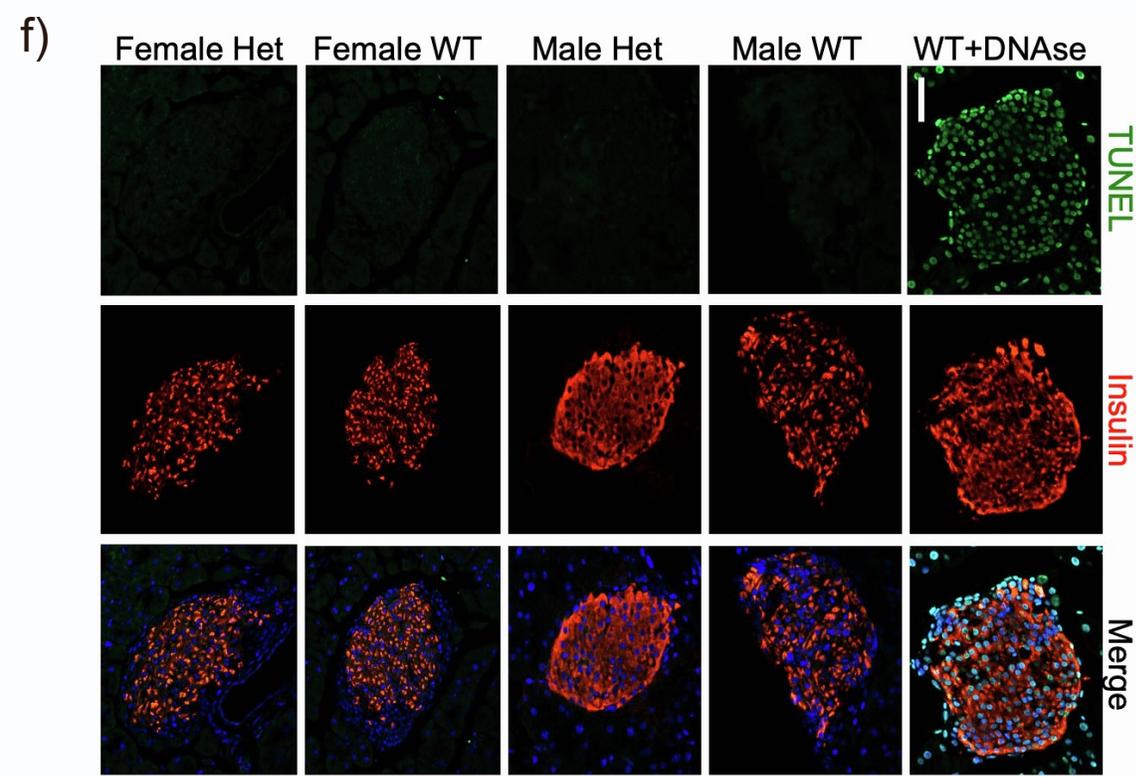
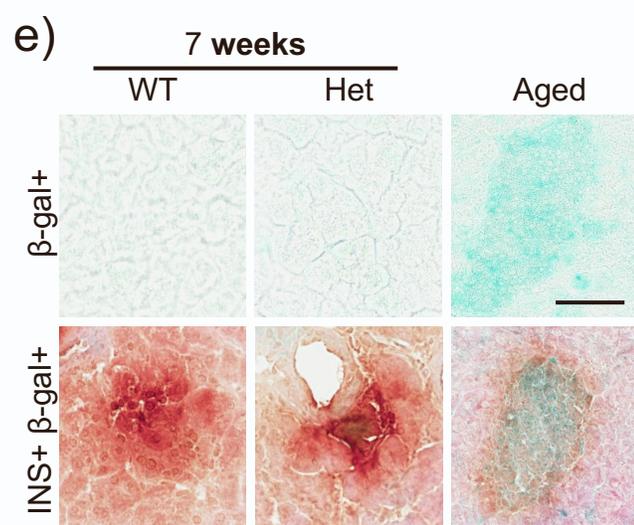
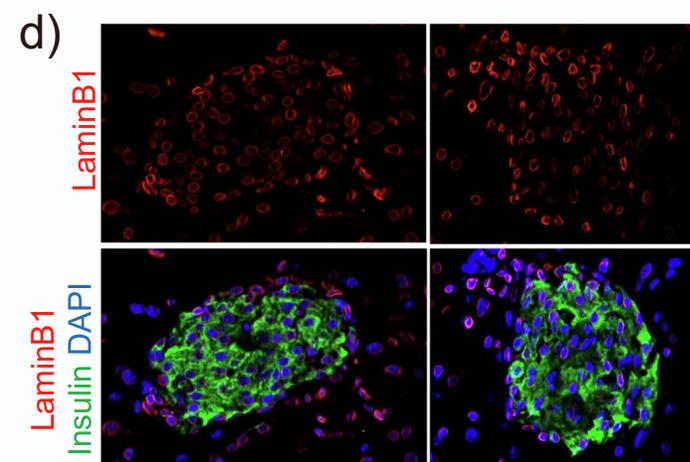
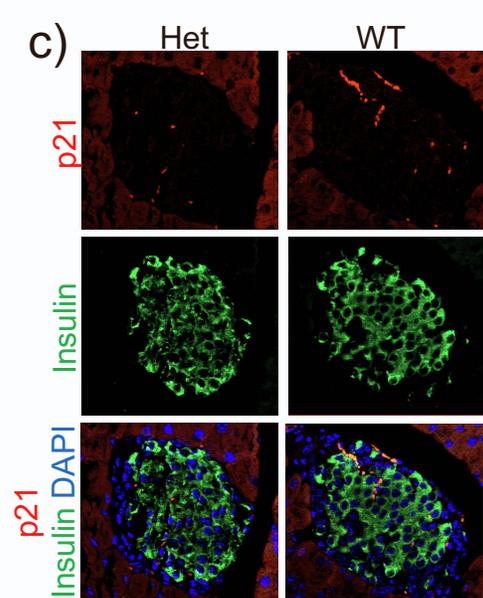
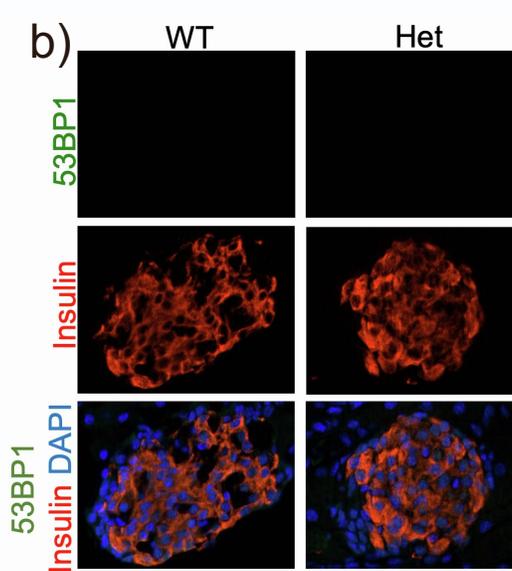
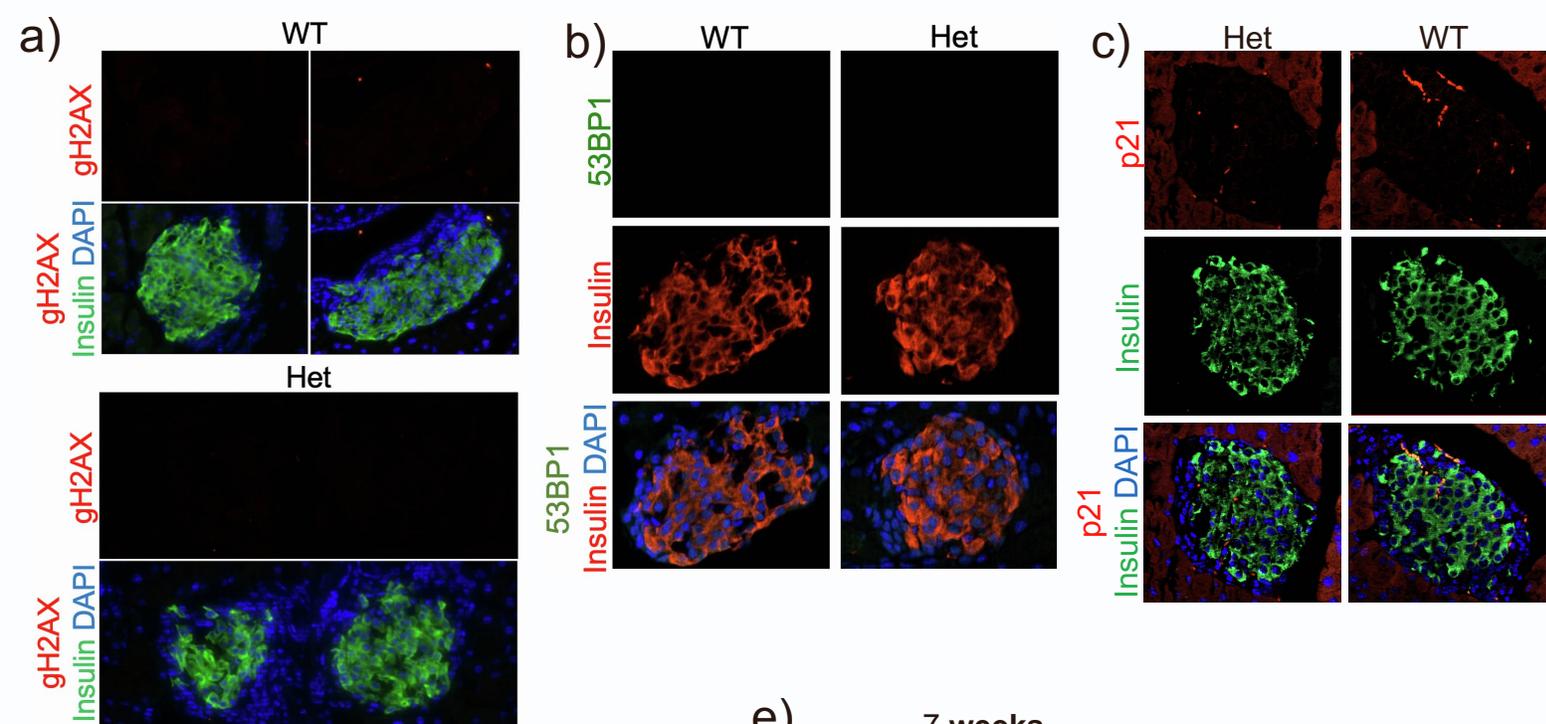


Figure S7: DNA damage and cell cycle inhibition markers were not detected in 5-week-old female *MafA*^{S64F/+} islets, nor were significant levels of apoptosis found in *MafA*^{S64F/+} islet cells. Related to Figure 6. a-e) Changes in immunostaining for γ H2AX (DNA double strand break; a), 53BP1 (DNA double strand break; b), p21 (cell cycle inhibitor; c), LaminB1 protein (nuclear integrity marker; d) and endogenous staining for SA- β -gal (senescence marker; e) were not altered in female *MafA*^{S64F/+} islets. f) TUNEL⁺ nuclei were barely detected in *MafA*^{S64F/+} islets at 6 weeks of age but easily visible in DNase treated controls (top panels).

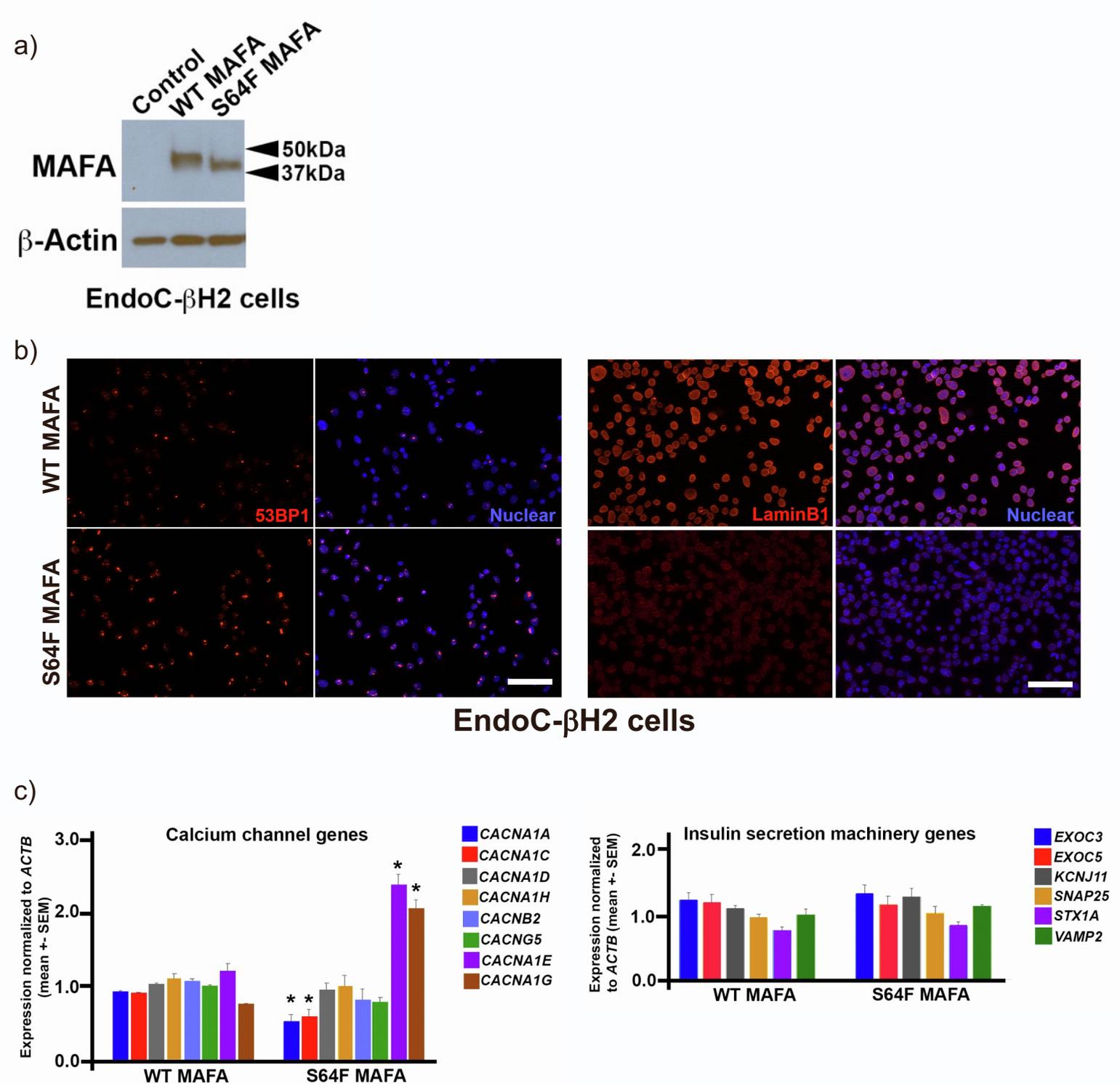


Figure S8: MAFA^{S64F} expressing human cells have changes in DNA damage, senescence, and Ca²⁺ channel gene expression. Related to Figure 7. a) Western blotting for EndoC-βH2 cells transduced to express WT MAFA or MAFA^{S64F} shows faster migration in the MAFA^{S64F} lane consistent with under-phosphorylated MAFA. Higher MAFA^{S64F} protein level is likely due to increased MAFA protein stability seen in (Iacovazzo et al., 2018). b) 53BP1 staining (left) intensity was higher and LaminB1 staining (right) intensity was lower in MAFA^{S64F} expressing EndoC-βH2 cells compared with MAFA^{WT}. c) qRT-PCR for Ca²⁺ channel genes (left) showed some alterations in MAFA^{S64F} expressing EndoC-βH2 cells while no changes in insulin secretion machinery genes were detected, consistent with findings in *MafA*^{S64F/+} mouse islets.

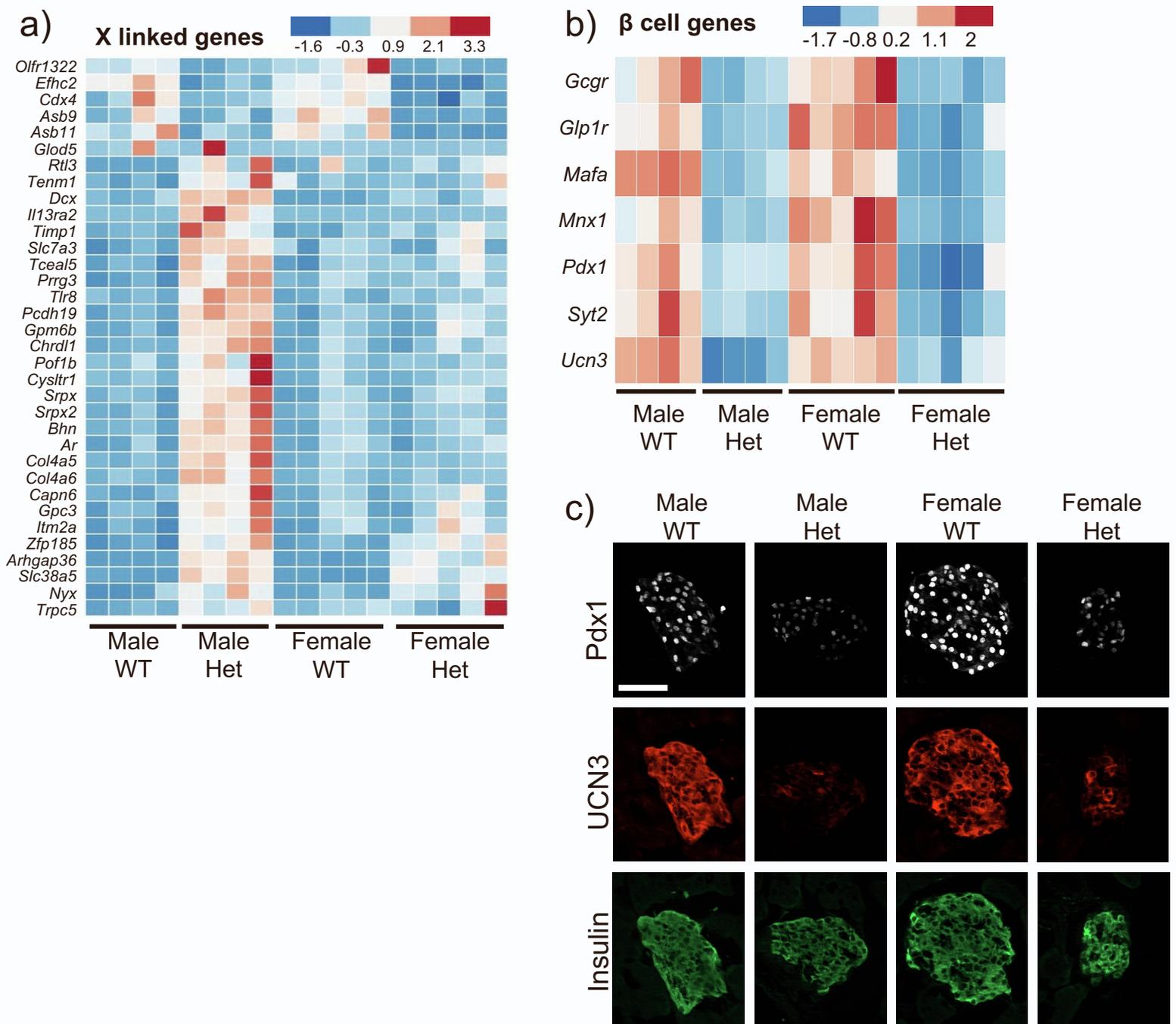


Figure S9: Many X chromosome linked genes were altered in male *MafA*^{S64F/+} islets and production of some β cell identity gene products are downregulated in both male and female *MafA*^{S64F/+} islets. Related to Figure 3. a) Heat map developed from the 5-week-old islet RNA-Seq data. FDR<0.05. b) Heat map showing β identity genes decreased in both male and female Het islets. FDR<0.05. c) Immunostaining illustrating decreased Pdx1 and Ucn3 protein levels in 7-week-old Het islets.

Table S1: Primer information

Mouse primers	Forward	Reverse
<i>MafA</i>	CCTGTAGAGGAAGCCGAGGAA	CCTCCCCAGTCGAGTATAGC
<i>Ins1</i>	CACTTCCT ACCCTGCTGG	ACCACAAAGATGCTGTTTGACA
<i>pre-Ins2</i>	GGGGAGCGTGGCTTCTTCTA	GGGGACAGAATTCAGTGGCA
<i>Ins2</i>	CCACCCAGGCTTTTGTCAA	CCCAGCTCCAGTTGTTCCAC
<i>Pdx1</i>	CGGCTGAGCAAGCTAAGGTT	TGGAAGAAGCGCTCTCTTTGA
<i>Ucn3</i>	AGCACCCGGTACAGATACCAA	GGCCTTGTGATGTTGAAGAG
<i>Ankrd1</i>	AAACGGACGGCACTCCACCG	CGCTGTGCTGAGAAGCTTGCTCT
<i>Bcl2a1d</i>	GTATATCCACTCCCTGGCTGAG	TAGTCACAATCCTTCCCCAGTT
<i>Ccl11</i>	TCCACAGCGCTTCTATTCTG	GGAGCCTGGGTGAGCCA
<i>Cd68</i>	ACTTCGGGCCATGTTTCTCT	GCTGGTAGGTTGATTGTCGT
<i>Cxcl12</i>	CAGTGACGGTAAACCAGTCAGC	TGGCGATGTGGCTCTCG
<i>Icam1</i>	CAATTTCTCATGCCGCACAG	AGCTGGAAGATCGAAAGTCCG
<i>p21</i>	GCCTTAGCCCTCACTCT TG	AGCTGGCCTTAGAGGTGACA
<i>Cacng4</i>	TCCGGAAGACGGGACTAC	ATGATGTTGTGGCGTGTCTTG
<i>Chp2</i>	CGCCTAGACCTCCAGCAGATC	GCCTGCGAAATACAGTCTCTGAC
<i>Edn2</i>	CTGGCAAGATGTGGACTGCTGA	GCCTTTCTTGTACCTCTGGCT
<i>F2r1</i>	CGGACCCGAGAACCTTGACCCG	GTGAGGATGGACGCAGAGAACT
<i>Grin1</i>	CCTTTTCAGAGCACACTGTGGCT	CCAGGAAAACCATGGCAGAG
<i>Hcn4</i>	CGTGCTCACTAAGGGCAACAAAG	GCACCTCATTGAAGTTGTCCACG
<i>Ret</i>	TTCCAGCATCAACTGCACTG	GTCAGTGGCTACCACCGTGT
<i>Shisa2</i>	TGGCACAACGACCGCCAGCAG	TGAAGGCAACGAACACTGAGCC
<i>Vil1</i>	TTCTACGGTGGTGACTGCTACC	TGGTCCAACAGGACGGCTTGAT
18S rRNA	GTAACCCGTTGAACCCATT	CCATCCAATCGGTAGTAGCG
<i>Lmb1</i>	CAGGAATTGGAGGACATGCT	GAAGGCTTGGAGAGAGCTT
<i>β-Actin</i>	AGGTCATCACTATTGGCAACGA	CACTTCATGATGGAATTGAATGATGT

Human primers	Forward	Reverse
<i>MAFA</i>	GAGAGCGAGAAGTGCCAACCT	TTCTCCTTGACAGGTCCCG
<i>MAFB</i>	CATAGAGACGTGGCAGCAA	ATGCCCGGAACCTTTTCTTT
<i>INS</i>	AGAGGCCATCAAGCAGATCACTGT	ACAGGTGTTGGTTCACAAGGCTG
<i>ANKRD1</i>	AGACTCCTTCAGCCAACATGATG	CTCTCCATCTCTGAAATCCTCAGG
<i>BCL2A1</i>	GGATAAGGCAAAAACGGAGGCTG	CAGTATTGCTTCAGGAGAGATAGC
<i>CCL11</i>	GCTACAGGAGAATCACCAGTGG	GGAATCCTGCACCCACTTCTTC
<i>CD68</i>	CGAGCATATTCTTTCACCAGCT	ATGAGAGGCAGCAAGATGGACC
<i>CXCL12</i>	CTCAACACTCCAAACTGTGCC	CTCCAGGTAATCCTGAATCCAC
<i>ICAM1</i>	AGCGGCTGACGTGTGAGTAAT	TCTGAGACCTCTGGCTTCGTCA
<i>P21</i>	CCTGTCACTGTCTTGTACCCT	GCGTTTGGAGTGGTAGAAATCT
<i>BCL2</i>	ATCGCCCTGTGGATGACTGAGT	GCCAGGAGAAATCAAACAGAGGC
<i>CXCL2</i>	GGCAGAAAGCTTGTCTCAACCC	CTCCTTCAGGAACAGCCACCAA
<i>ICAM3</i>	AGATCGTCTGCAACGTGACCCT	TGGTGTGAGGTTTACAATGGGTC
<i>IGFBP2</i>	CGAGGGCACTTGTGAGAAGCG	TGTTTATGGTGTGCTCCACGTG
<i>IGFBP4</i>	GAGCTGGGTGACACTGCTTG	CCCACGAGGACCTCTACATCA
<i>IGF1R</i>	CTCCTGTTTCTCTCCGCCG	ATAGTCGTTGCGGATGTCGAT
<i>LMNB1</i>	TGGAGTGG TTGTTGAGGAAG	GAGAAGGCTCTGCACTGTATAC
<i>CACNA1A</i>	CTGGTAGCCTTTGCCTTCACTG	ACACAGCCTTGAGCTTTGGCAG
<i>CACNA1C</i>	GCAGGAGTACAAGAACTGTGAGC	CGAAGTAGGTGGAGTTGACCAC
<i>CACNA1E</i>	AGCGTGAGACAGGCAAGCCAT	GGATGCACATCTCAAAGTAGCGC
<i>CACNA1G</i>	TTCACCGCAGTCTTCTGGCTG	TGACGGAGATGAGCACCAACAG
<i>EXOC3</i>	GAGCCATTGCTTTCTCCACAG	TGGCTCTGTCTTTGACCCAG
<i>EXOC5</i>	CTTCAGTAATCCAGAAACAGTCTT	TGCTCTGCATCGGACTTCTTAC
<i>SNAP25</i>	CGTCGATGCTGCAACTGGTTG	GGTTCATGCCTTCTTGCACAG
<i>VAMP2</i>	CTCCAAACCTACCAGTAACAGG	AGTCCGACAGTCTGGTCTC
<i>STX1A</i>	GGAACACGCGGTAGACTATGT	CTGGAGTGGAGTGGCAGTTT
<i>KCNJ11</i>	TGTGTACCAGCATCCACTCCT	GTTCTGCACGATGAGGATCAGG
<i>GAPDH</i>	CTCACCGGATGCACCAATGTT	CGCGTTGCTCACAATGTTTCT