1	SUPPLEMENTARY INFORMATION for
2	TLR4 is a regulator of trained immunity in a murine model of
3	Duchenne muscular dystrophy
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Necrotic phase Ctrl WT mdx kDa p-STAT6 100-STAT6 Actin 37 Fibrotic phase WT mdx **k**Da p-STAT6 100 100 STAT6 Actin 37 21 Supplementary Figure 1: Lack of STAT6 phosphorylation in mdx BMDM. Levels of 22 p-STAT6 in total cell lysate of WT and mdx BMDM were evaluated by western blot at 23 24 necrotic (top panel) and fibrotic (bottom panel) phases of the disease. WT BMDM treated with 20 ng/ml IL4 for 2 hours, served as a positive control (Ctrl, in the top panel). 25 Total STAT6 was detected in all samples, and actin served as an additional loading 26 control. Each lane represents an independent biological sample obtained from a 27

different animal, and all experimental replicates are shown in the figure.



30 Supplementary Figure 2: mdx BMDM at the fibrotic phase of disease show non-

31 specific amplified responses to heterologous stimuli. BMDM generated from WT

- 32 and mdx mice at the fibrotic phase of disease were exposed to (a) LPS (100 ng/ml) +
- IFN γ (20 ng/ml) and IL4 (20 ng/ml) (n=4/group for WT, n=5/group for mdx), (**b**)
- fibrinogen (1 mg/ml) (n=4/group for WT, n=5/group for mdx), or (**c**) β-glucan (100 μ g/ml)
- 35 (n=4/group for WT, n=5/group for mdx except IL6 at 24h n=4). Bar graphs show mRNA
- transcript levels of prototypical pro-inflammatory ("M1") and anti-inflammatory ("M2")
- marker genes at 4 and 24 hours after exposure. All values are expressed relative to the
- mean basal WT (unstimulated) level determined on the same PCR plate. Data
- represent means \pm SEM of biologically independent samples from different mice.
- 40 **P*<0.05 vs unstimulated WT BMDM and †P<0.05 vs stimulated WT BMDM at a given

- 41 time point (one-way ANOVA followed by Tukey post-hoc test, two-tailed). See Source
- 42 Data file for the exact P values.



48 IFNγ or IL4 (n=4/group) (**b**) fibrinogen (n=4/group), or (**c**) β-glucan (n=4/group except

49 TGF β in mdx 24h group n=3). Bar graphs show mRNA transcript levels of prototypical pro-inflammatory ("M1") and anti-inflammatory ("M2") marker genes at 4 and 24 hours 50 after exposure. (d-e) BMDM generated from uninjured and cardiotoxin-injured WT were 51 exposed to heterologous stimuli as in (a-c) (n=4/group for Uninjured, n=5/group for 52 Injured). All values are expressed relative to the mean basal WT level determined on 53 the same PCR plate. Data represent means \pm SEM of biologically independent samples 54 from different mice. *P<0.05 vs unstimulated WT/Uninjured BMDM and †P<0.05 vs 55 stimulated WT/Uninjured BMDM (one-way ANOVA followed by Tukey post-hoc test, 56 two-tailed). See Source Data file for the exact P values. 57



Supplementary Figure 4: M1 and M2 marker gene expression after "training" by
muscle extract or serum in vitro. As indicated in Fig. 4a, WT BMDM were exposed for
24 hours to muscle extract (ME) derived from either (a) WT (n=5 per group) or (b) mdx
muscles (n=5/group except for TGFβ at 24h n=4). Gene expression was measured

immediately after ME exposure (24 hours) as well as after removal of the ME stimulus 64 and subsequent "resting" of the cells (5 days). The dashed line indicates the mean 65 basal expression level for PBS-exposed WT BMDM at each time point. (c-d) Expression 66 of inflammatory genes 8 hours after fibrinogen exposure in WT BMDM previously 67 "trained" with mdx-ME at (c) different concentrations (0.1, 0.5 and 1 mg/ml) for 24 hours 68 69 (n=4 per group), or (d) different exposure durations (2, 8, 24 hours) using the same concentration (1 mg/ml) (n=4 per group). Data are expressed relative to the PBS-trained 70 group identically stimulated with fibrinogen. (e) Identically to the ME protocol (Fig. 4a), 71 72 WT BMDM (n=4 per group) were "trained" with WT or mdx (4-6 weeks old) serum (5%) followed by resting for 5 days and secondary stimulation with fibringen for 8 hours. The 73 graph shows the M1 and M2 markers gene expression levels relative to the PBS-trained 74 group stimulated with fibrinogen. (f) Donor macrophage (defined as CD45.2+CD11c-75 CD11b+F4/80+) number (left panel) and percentage (right panel) in the tibialis anterior 76 (TA) muscle of mice (n=5 per group) transplanted with WT or mdx bone marrow as 77 shown in Fig. 4d. There were no differences between groups. All data represent means 78 ± SEM of independent biological or mice (**a-b**, **f**: *P<0.05 unpaired t-test, two-tailed; **c-e**: 79 80 *P<0.05 vs. PBS- trained WT BMDM one-way ANOVA followed by Tukey post-hoc test, two-tailed). See Source Data file for the exact P values. 81



84 Supplementary Figure 5: Pathway analysis (Reactome database) for the Gene-

85 based Patterns (GP) designated GP2, GP3 and GP4 for H3K27me3. Pathway

- enrichment analysis is shown for genes showing the (a) GP2, (b) GP3, and (c) GP4
- 87 configurations. The vertical line indicates the cut-off P value = 0.05. The extended gene
- lists are found in the Supplementary Data 2 table.



- 90 Supplementary Figure 6: Transcription factor enrichment analysis for H3K27me3
- 91 in WT, mdx and mdxTLR4^{-/-} BMDM. (a) Stacked heatmaps of the Peak-based Pattern
- 92 (PP) analysis showing the dynamically regulated patterns in mdx versus WT
- 93 (designated PP1 to PP4); the normalized H3K27me3 read intensity is plotted ±12 kb
- 94 over the center of peaks. The values in parentheses indicate the percentages for each
- 95 pattern (I=Increased and U=Unchanged relative to Input) as follows: PP1=IUI,
- 96 PP2=IUU, PP3=UII and PP4=UIU. (b-c) Transcription factor enrichment Reactome
- pathway analysis for (b) PP1, and (c) PP2 patterns. Vertical lines indicate the cut-off P
- value = 0.05. The extended lists for these data can be found in Supplementary Data 4
- 99 and 5 tables.



Supplementary Figure 7: H3K4me3 promoter occupancy in BMDM. ChIP-qPCR
was performed to determine promoter region occupancy of H3K4me3 on both M1 and
M2 marker genes in (a) WT versus mdx BMDM (n=5 per group) and (b) mdx versus
mdxTLR4^{-/-} BMDM (n=5 per group). IgG antibody was used as a control for non-specific
binding of antibody. Data represent means ± SEM of independent biological samples
from different mice. *P<0.05 (unpaired t-test, two tailed). See Source Data file for the
exact P values.



109 Supplementary Figure 8: Gene-based Pattern (GP) analysis of H3K27ac ChIP

sequencing for WT, mdx and mdxTLR4^{-/-} BMDM at the necrotic phase of disease.

(a) Violin/box plots representing normalized intensity fold change (H3K27ac vs. Input) in 111 the region 5kb upstream to the transcription start site (TSS) of all genes across the 112 whole genome; *P<0.0001 compared to WT BMDM, and + P<0.0001 compared to the 113 mdx group (two-sided Mann-Whitney U-test). Maxima and minima are shown at the 114 extreme limits of the plot, box boundaries indicate the 25th and 75th percentiles, 115 whiskers represent the 10th and 90th percentiles, and the horizontal line within the box 116 denotes the median value. (b) Heatmaps showing different Gene-based Patterns 117 (I=Increased and U=Unchanged relative to Input) in order of frequency: GP1 (IUU), GP2 118 119 (UIU), GP3 (IUI) and GP4 (UII) of the normalized H3K27ac read intensity (proportional to red color intensity) in the 5 kb promoter region of the genes. The values in 120 parentheses indicate the percentages for each pattern. (c-f) Pathway enrichment 121 analysis (Reactome database) for genes showing the GP1-4 configuration (see 122 Supplementary Data 1 and 2 tables for the extended list of all Gene-based Patterns and 123 enriched pathways, respectively). The vertical line indicates the cut-off P value = 0.05. 124







- enrichment Reactome pathway analysis for (**b**) PP1 and (**c**) PP2 patterns are shown.
- 134 Vertical lines indicate the cut-off P value = 0.05. The extended lists for these data can
- be found in Supplementary Data 4 and 5 tables.



137 Supplementary Figure 10: Phenotype of macrophages in WT and mdx limb

muscles. Tibialis anterior (TA) muscles from WT and mdx mice were processed and139the percentages of pro-inflammatory macrophages (iNOS+ CD206- population, in upper140panel and iNOS+ TGF β - population, in lower panel) were determined by flow cytometry.141Data are mean values ± SEM of independent biological samples from different mice142(n=3/group). *P<0.05 (unpaired t-test, two-tailed). See Source Data file for the exact P</td>143values.



Supplementary Figure 11: Phenotype of host macrophages in injured muscles 146 after adoptive transfer with WT or mdx BMDM. No significant differences in host (a) 147 pro-inflammatory macrophages (iNOS+ CD206- TGFβ- population) (n=10/group) and 148 (b) anti-inflammatory macrophages (iNOS- CD206+ TGF β + population) (n=9/group), 149 150 were found in comparisons between mice adoptively transferred with either WT or mdx 151 BM. Similarly, no significant differences in the host (c) pro-inflammatory macrophages (n=5/group) and (d) anti-inflammatory macrophages (n=5/group) were observed 152 153 between groups adoptively transferred with either mdx or mdxTLR4^{-/-} BM. All data are means ± SEM of independent biological samples from different mice. There are no 154 significant differences between groups (unpaired t-test, two-tailed). See Source Data file 155 for the exact P values. 156





- shown for: (a) Hematopoietic progenitor cells, (b) Innate myeloid cells, and (c)
- 161 Lymphocytes, in bone marrow. (d) Gating strategy for the analysis of pro- and anti-
- 162 inflammatory macrophages in the injured host (CD45.1) limb muscles of mice after
- adoptive transfer with donor (CD45.2) bone marrow cells.