

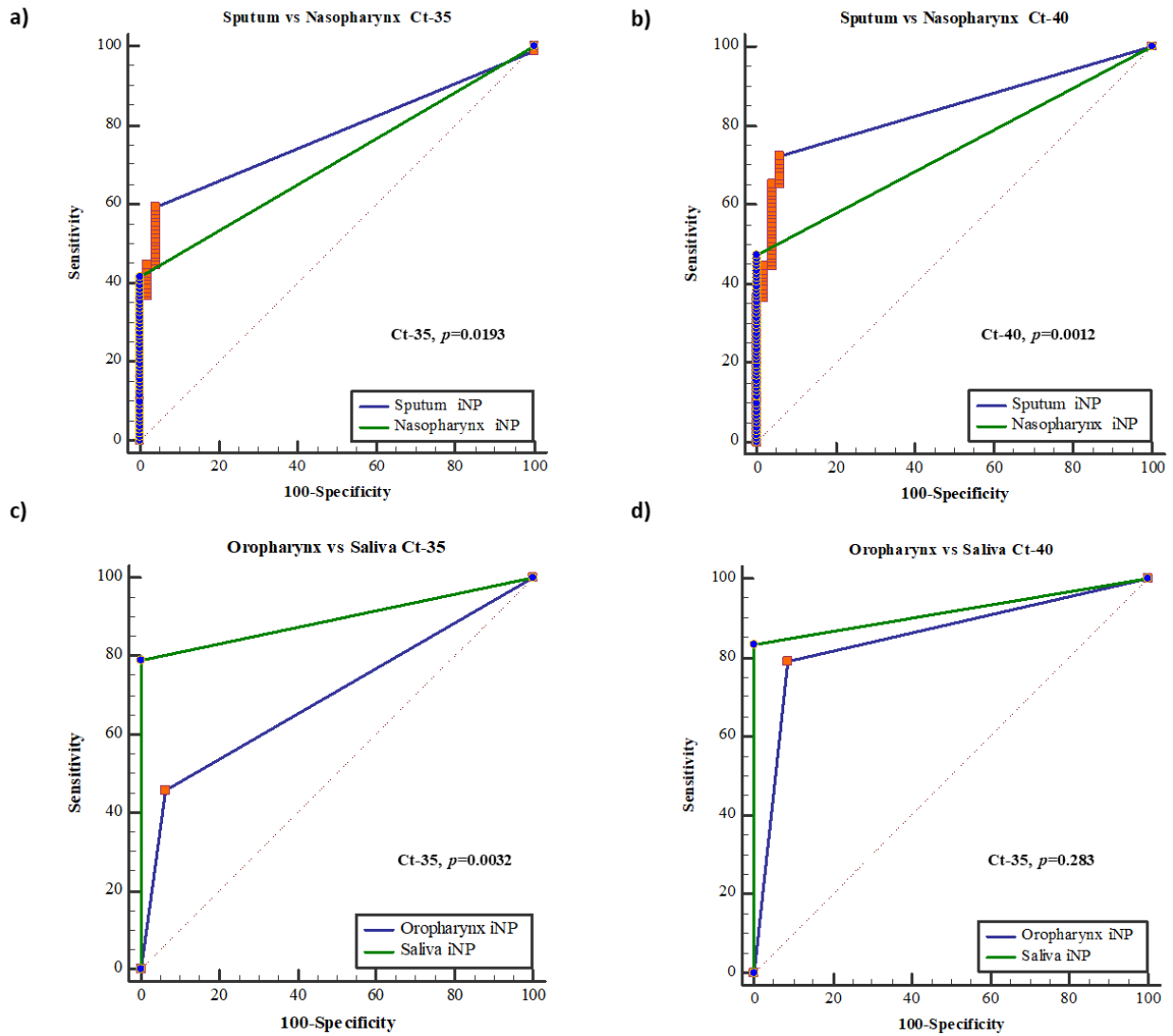
Supplementary Materials

1
2

3 **Supplementary Figures**

4 Figure S1. The specificity and sensitivity of RT-qPCR with the in-house-designed *NP* gene
5 primer set (iNP) in nasopharyngeal vs. sputum samples and oropharyngeal vs. saliva samples.

6
7
8



9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24

25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50

Supplementary Figure 1

The specificity and sensitivity of RT-qPCR with the in-house–designed *NP* gene primer set (iNP) in nasopharyngeal vs. sputum samples and oropharyngeal vs. saliva samples.

- a) Determination of specificity and sensitivity in selected sputum vs. nasopharyngeal samples at a C_t cutoff of 35 (C_t -35).
- b) Determination of specificity and sensitivity in selected sputum vs. nasopharyngeal samples at a C_t cutoff of 40 (C_t -40).
- c) Determination of specificity and sensitivity in selected (first week) oropharyngeal vs. saliva samples at a C_t cutoff of 35 (C_t -35).
- d) Determination of specificity and sensitivity in selected (first week) oropharyngeal vs. saliva samples at a C_t cutoff of 40 (C_t -40).

51

52

53

54

55

56 **Table S1.** All primers, probe, and PCR conditions used in this study.

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

Primer source	Target gene	Primer name	Primer sequence	Target	PCR condition
WHO primer	RdRp gene	RdRP_SARSr-F2	GTGARATGGTCATGTGTGGCGG	Pan Sarbeco	
		RdRP_SARSr-R1	CARATGTTAAASACTATTAGCATA		
		RdRP_SARSr-P1	[TET]CCAGGTGGWACRTCATCMGGTGATGC[BHQ1]		
CDC primer	N gene 1	2019-nCoV_N1-F	GACCCCAAATCAGCGAAAT	SARS-CoV2	50°C for 10 min, 1 cycle for Reverse transcription. 95°C 30sec for 1 cycle for pre incubation, 95°C 5sec for and 57°C for amplification and data detection for 45 cycles
		2019-nCoV_N1-R	TCTGGTTACTGCCAGTTGAATCTG		
		2019-nCoV_N1-P	[FAM]ACCCCGCATTACGTTTGGTGGACC[BHQ1]		
CDC primer	N gene 2	2019-nCoV_N2-F	TTACAAACATTGGCCGCAAA	SARS-CoV2	
		2019-nCoV_N2-R	GCGCGACATTCCGAAGAA		
		2019-nCoV_N2-P	[FAM]ACAATTTGCCCCAGCGCTTCAG[BHQ1]		
CDC primer	N gene 3	2019-nCoV_N3-F	GGGAGCCTTGAATACACCAAAA	SARS-CoV2	
		2019-nCoV_N3-R	TGTAGCACGATTGCAGCATTG		
		2019-nCoV_N3-P	[FAM]AYCACATTGGCACCCGCAATCCTG[BHQ1]		
This study	iNP gene	nCov-NP_572F	GCAACAGTTCAAGAAATTC	SARS-CoV2	59°C for amplification and the rest is same like above
Kogene Kit	E gene	Primer/Probe mix 1 (E-gene)	Manufacturer's trade mark	SARS-CoV2	As per Manufacturer's instructions
	RdRp gene	Primer/Probe mix 2 (RdRp gene)	Manufacturer's trade mark	SARS-CoV2	As per Manufacturer's instructions

77

78 **Table S2.** Diagnostic characteristics of negative control samples.

79

80

81

82

83

84

85

Diagnostic characteristics	Negative samples			
	Nasopharynx	Sputum	Oropharynx	Saliva
No. of samples	55	21	24	28
NP-gene (iNP) RT-PCR	ND	ND	ND	ND
E-gene (Kogene kit) RT-PCR	ND	ND	ND	ND
RdRp-gene (Kogene kit) RT-PCR	ND	ND	ND	ND
WHO RdRp Primers RT-PCR	ND	ND	ND	ND
CDC N1 primers RT-PCR	ND	ND	ND	ND
CDC N2 primers RT-PCR	2 (+)	2 (+)	1 (+)	1 (+)
CDC N3 primers RT-PCR	ND	ND	ND	ND

86

87

88

Fifty-five nasopharyngeal, 21 sputum, 24 oropharyngeal, and 28 saliva samples were obtained from health checkup subjects with no clinical symptoms, no increase in a SARS-CoV-2 antibody titer, or no contact with confirmed COVID-19 patients. Cycle threshold >35 indicates a negative result; ND: not detectable; 2(+): two SARS-CoV-2-positive samples; 1(+): one virus-positive sample

89

90

91

92

93 **Table S3.** Clinical and subclinical characteristics of all COVID-19 patients

Patient	Gender/ Age	Underlying Comorbidity	Symptoms & signs	IFA			ELISA			Cell culture
				IgG titer Initial/ Follow-up	IgM titer Initial/ Follow-up	IgM antibody titer Initial/ Follow-up	IgG antibody titer Initial/ Follow-up	Total antibody titer Initial/ Follow-up		
1	M/46	HTN, dyslipidemia	coughing, chill	<1:32/1:256	<1:32/<1:32	<1:128/1:512	<1:128/1:4096	<1:128/1:4096	(+)	
2	M/30	HTN	coughing, sore throat chill	<1:32/1:128	<1:32/<1:32	<1:128/<1:128	<1:128/1:512	<1:128/1:512	(+)	
3	F/79	HTN,DM, dyslipidemia, Guillain-Barre syndrome	dyspnea	<1:32/1:1,024	<1:32/<1:32	<1:128/1:512	<1:128/1:2048	<1:128/1:2048	(+)	
4	M/30	None	febrile sense	<1:32/1:128	<1:32/<1:32	<1:128/1:128	<1:128/1:1024	<1:128/1:1024	(+)	
5	F/29	None	sore throat, myalgia chill	<1:32/1:64	<1:32/<1:32	<1:128/<1:128	<1:128/1:128	<1:128/<1:128	(+)	
6	M/74	HTN,DM, dyslipidemia	sore throat, rhinorrhea	1:256/1:1,024	<1:32/<1:32	1:128/1:256	1:512/1:8192	1:512/1:4096	(-)	
7	F/75	HTN, dyslipidemia	fever, sore throat	<1:32/>1:1,024	<1:32/1:32	<1:128/1:128	<1:128/1:1024	<1:128/1:1024	(-)	
8	M/79	HTN,DM	fever, headache	<1:32/1:128	<1:32/<1:32	<1:128/1:256	<1:128/1:8192	<1:128/1:4096	(-)	
9	F/61	None	fever, coughing	<1:32/1:256	<1:32/<1:32	<1:128/1:1024	1:256/1:16384	1:128/1:8192	(-)	
10	M/36	None	sore throat, headache chill	<1:32/1:128	<1:32/<1:32	<1:128/<1:128	<1:128/1:128	<1:128/1:128	(-)	
11	F/29	None	No symptoms	1:64/1:64	<1:32/<1:32	<1:128/<1:128	<1:128/1:128	<1:128/<1:128	(-)	
12	M/22	None	No symptoms	1:64/1:64	<1:32/<1:32	<1:128/<1:128	<1:128/1:128	<1:128/1:128	(-)	

94

95 HTN, hypertension; DM, diabetes mellitus. The IFA titer was measured by serial dilution of a patient's serum, and the IgG titer cutoff was $\geq 1:32$. Antibody titer detection was performed
96 (with serial dilution) by an indirect ELISA at OD₄₅₀ against a recombinant SARS-CoV-2 nucleoprotein; the cutoff titer for IgM was 0.5; for IgG, it was 1.1; and for total Ig, it was 0.7

97

98

99

100

101 **Table S4.** RT-PCR results on SARS-CoV-2 in all clinical samples for primer set iNP and primer sets targeting the *E* and
 102 *RdRp* genes (Kogene Kit).

103

104	Specimens and values	Nasopharynx (N=101)	Oropharynx (N=90)	Sputum (N=106)	Pla/Ser/WB (N=77)	Saliva (N=69)	Stool (N=55)	Urine (N=92)
105	NP-gene (iNP)							
	Positive test results, No%	42 (44.68%)	21 (23.59%)	61 (57.54%)	5 (6.32%)	32(46.37%)	6 (10.90%)	1 (1.08%)
106	Cycle threshold, Mean (SD)	26.13 (13.5)	29.39 (4.4)	27.26 (5.1)	30.39 (3.01)	28.47 (4.1)	29.62 (3.5)	NA
107	Ct range	10.04 - 34.94	14.94 - 33.96	14.94 -33.96	27.49 - 33.91	21.60 – 34.69	25.34 – 34.15	NA
108	Sensitivity/Specificity	43.1%/100%	24.3%/100%	58.1%/100%		43.3%/100%		
	95% CI	0.628 - 0.779	14.77 - 32.95	0.709 - 0.841	2.56 - 14.08	0.644 - 0.808	3.64 - 20 .0	NA
109	Specimens and values	Nasopharynx (N=97)		Sputum (N=104)				
110	E-gene (Kogene kit)							
	Positive test results, No%	38 (39.17%)		49 (47.11%)				
111	Cycle threshold, mean (SD)	25.39 (5.2)		27.83 (5.0)				
112	Ct range	12.88 – 34.33		13.76 - 34.93				
113	Sensitivity/Specificity	36.5%/100%		46.1%/100%				
	95% CI	0.593 - 0.750		0.638 - 0.784				
114	Specimens and values	Nasopharynx (N=97)		Sputum (N=104)				
115	RdRp-gene (Kogene kit)							
	Positive test results, No%	37 (38.18%)		47 (45.19%)				
116	Cycle threshold, Mean (SD)	25.62(5.2)		27.96 (5.0)				
117	Ct range	13.40 - 33.68		14.17 – 34.68				
118	Sensitivity/Specificity	36.1%/100%		46.6%/100%				
119	95% CI	0.593 - 0.750		0.652 - 0.796				

120

121 RT-PCR results on SARS-CoV-2 in all clinical samples collected from symptom onset to post-recovery, up to 5 weeks or more. Pla/Ser/WB: plasma/serum/whole blood; NA: not
 122 applicable; N: number; SD: standard deviation; CI: 95% confidence interval

123

124

125

126 **Table S5.** Sensitivity of RT-PCR with the in-house–designed *NP* gene primer set (iNP) in other samples depending on the
 127 time interval after symptom onset.

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

Days Since Symptoms	NP-gene (iNP)									
	Oropharynx (N=90)		Saliva (N=67)		WB/Plasma (N=77)		Urine (N=92)		Stool (N=55)	
	Ct < 35	Ct < 40	Ct < 35	Ct < 40	Ct < 35	Ct < 40	Ct < 35	Ct < 40	Ct < 35	Ct < 40
Admission date Sensitivity	50.0% (N =12)	83.3% (N =12)	70% (N =10)	80.0% (N =10)	12.5% (N =8)	37.5% (N =8)	0% (N =10)	10.0% (N =10)	0% (N =5)	20.0% (N =5)
0 – 3 days Sensitivity	35.7% (N =14)	64.3% (N =14)	69.2% (N =13)	69.2% (N =13)	18.2% (N =11)	27.3% (N =11)	0% (N =8)	12.5% (N =8)	0% (N =6)	16.7% (N =6)
1 week(0 - 7 days) Sensitivity	50.0% (N =26)	84.6% (N =26)	79.1% (N =24)	83.3% (N =24)	10% (N =20)	20.0% (N =20)	0% (N =22)	4.6% (N =22)	6.67% (N =15)	20.0% (N =15)
2 weeks(8 – 14 days) Sensitivity	20% (N =25)	40.0% (N =25)	37.5% (N =24)	62.5% (N =24)	12.5% (N =16)	31.3% (N =16)	3.83% (N =28)	15.4% (N =28)	14.3% (N =28)	31.3% (N =28)
3 weeks(15 – 21 days) Sensitivity	5.88% (N =17)	23.5% (N =17)	27.27% (N =11)	54.6% (N =11)	6.3% (N =16)	31.3 % (N =16)	0% (N =17)	23.5% (N =17)	14.3% (N =14)	31.3% (N =14)
4 weeks(22 – 29 days) Sensitivity	0% (N =8)	37.5% (N =8)	0% (N =4)	25.0% (N =4)	25.0% (N =4)	25.0% (N =4)	0% (N =8)	12.5% (N =8)	0% (N =4)	25.0% (N =4)
5 weeks+(30 + days) Sensitivity	9.1% (N =11)	27.3% (N =11)	0% (N =3)	33.3% (N =3)	0% (N =22)	13.6% (N =22)	0% (N =20)	10.0% (N =20)	0% (N =9)	13.6% (N =9)

Samples were segregated as follows: admission date, 0–3 days after admission, 0–7 days after admission as 1 week, 8–14 days as 2 weeks, 15–21 days as 3 weeks, 22–29 as 4 weeks, and 30+ as 5 weeks+. *N*: number of samples; WB: whole blood