Supplemental Figure Legends

SI Appendix Figure S1: Transcriptional changes in LXR-deficient liver.

A. Differential expression analysis of the LXRDKO liver RNA-Seq results showed 246 upregulated and 321 downregulated genes. B. Volcano plot showing differential gene expression between WT and LXRDKO samples. Genes are color-coded based on their fold change and adjusted p value. C. Top 10 downregulated (left) and upregulated (right) genes from LXRDKO liver RNA-Seq based on adjusted p value.

SI Appendix Figure S2: Expression profiles of LXRDKO livers.

A. Pathway enrichment results for downregulated (top) and upregulated (bottom) genes in LXRDKO liver. B. Transcription Factor ChIP-Seq enrichment analysis (ChEA) for downregulated genes in LXRDKO liver. Publicly available ChIP-Seq data were used to estimate enrichment of transcription factor binding in the promoters of genes downregulated in LXRDKO liver.

SI Appendix Figure S3: Chromatin accessibility profiles of LXRDKO livers

A. Correlation heatmap of ATAC-Seq samples using all peaks. B. t-SNE plot of ATAC-Seq samples. C. Distribution of top 1000 peaks with largest loss and gain in LXRDKO liver based on relative distance to closest TSS. D. Average change in accessibility of peaks associated with differentially expressed genes in LXRDKO liver. E. Average change in accessibility in intronic (left), intergenic (right), and promoter (bottom) regions for all genes (All) and those whose expression was reduced (Down) or increased (Up) in LXRDKO livers. * comparison to all genes; # comparison between downregulated and upregulated genes.

SI Appendix Figure S4: Pathway enrichment.

Pathway enrichment analysis of the top 1000 peaks that lost (top) and gained (bottom) accessibility.

SI Appendix Figure S5: Changes to LXR binding sites in LXRDKO livers.

A. Proportion of LXR basal or agonist induced binding genes among genes differentially expressed in LXRDKO livers. B. Average number of (basal) LXR binding sites per gene for all LXR target genes and genes that are differentially expressed in LXRDKO liver. C. Changes in accessibility across LXR binding sites in LXRDKO liver compared to WT. D. Proportion of LXR binding sites within intergenic, intronic and promoter regions plotted as a function of the change in accessibility in LXRDKO liver.

SI Appendix Figure S6: Loss of LXRs reduces motif accessibility of other transcription factors.

A. Heatmaps of motif enrichment of selected transcription factors from Figure 4A across binned intergenic, intronic and promoter ATAC-Seq peaks based on the change in accessibility. B. ATAC-Seq signal intensity heatmap and profile across peaks with PPARE motif, among the top 1000 peaks losing accessibility in LXRDKO livers. C. (Left) Hierarchical clustering heatmap for

transcription factor motif co-occurrence among the least accessible 10 bins from Figure 4A. Eachcolumn represents a peak and red indicates presence of the corresponding motif in that peak. (Right) Hierarchical clustering heatmap for transcription factor motif co-occurrence among LXR binding genes that are associated with the least accessible 10 bins in LXRDKO livers. Each column represents a gene and red indicates presence of the corresponding motif in the peaks associated with the same gene. D. Normalized counts for the expression of transcription factors identified in Figure 4A. (* is used to indicate the FDR < 0.05)

SI Appendix Figure S7: Impact of the loss of LXR on other transcription factor motif activity.

Footprint profile of HNF6 (A) and HNF1B (B) using the ATAC-Seq samples for LXRDKO and WT with merged replicates. C. Pathway enrichment analysis of genes associated with increased accessibility of the NFY motif (top 10 bin from Figure 5A). D. Volcano plot of the expression profile of genes with NFY motif containing peaks within the top 10 bins that gain accessibility (from Figure 5A).

SI Appendix Figure S8: Change in accessibility and genomic features at the LXR binding sites in relation to different modes of LXR action.

A. Proportions of genomic annotations for LXR binding peaks that are associated with genes that are differentially upregulated or downregulated in LXRDKO and either downregulated, upregulated or not changing in response to either GW3965 and T0901317 treatment are plotted. B. Pathway enrichment analysis of genes that were downregulated in LXRDKO and have an LXR ChIP-Seq peak but did not change in expression in response to agonist treatment. C. LXR ChIP-Seq (basal) sites are segregated based on their genomic feature and ranked and binned based on their change in accessibility in LXRDKO livers. Motif analysis was performed for each of the bins across the accessibility and genomic features. Similar motifs were combined (>.90 Similarity) and motif with low enrichment were not displayed. Transcription factor motifs enriched in the LXR binding intergenic (left), intron (middle) and promoter (right) regions are displayed. D. Signal intensity heatmap and profiles of ATAC peaks at LXR binding sites proximal to ligand-dependent repressed genes.



В 125 Not sig
Log2FC
Adj. p-value
Adj. p-value &Log2FC Cd5I 100 75 -Log₁₀ P Gm42047 Gpnmb 50 PhIda3 Cd163 Nr1h3 GotCyp2b10 Acach 25 Eda2r Srebf1 Nr h2 DIF 0 Ccr3 -5 Ō 5 Log₂ fold change

WT1 WT2 KO1 KO2

2.66E-27

3.87E-26

5.26E-26

3.91E-24

9.02E-24

1.84E-22

C Top 10 Downregulated Genes					
Name	Adj. p value	Fold Change	Туре		
Cd5l	2.05E-115	0.0040	protein coding		
Cd163	4.39E-35	0.0335	protein coding		
Clec4f	3.45E-28	0.1402	protein coding		
Nr1h3	3.45E-28	0.3239	protein coding		

0.3733

0.3219

0.2306

0.3331

0.0939

0.1875

protein coding

protein coding

protein coding

protein coding

protein coding

lincRNA

Top 10 Upregulated Genes

Name	Adj. p value	Fold Change	Туре	
Gm42047	4.36E-66	9.6148	lincRNA	
Gpnmb	1.44E-45	32.1408	protein coding	
PhIda3	3.49E-34	21.7534	protein coding	
Sult2a7	1.15E-31	14.4144	protein coding	
Eda2r	2.66E-22	18.0955	protein coding	
Cyp2b10	2.70E-21	4.4866	protein coding	
Got1	1.20E-18	2.8311	protein coding	
Cplane1	4.55E-18	5.3950	protein coding	
Gm48199	9.53E-18	8.6393	lincRNA	
Fkbp5	9.54E-18	2.6513	protein coding	

С

Ces1e

Nr1h2

Acacb

Hamp2

Srebf1

1810008I18Rik

Downregulated Pathwaysz	Adjusted	Genes
PPAR signaling pathway	3.92E-08	Acsl5, Lpl, Nr1h3, Dbi, Sorbs1, Fabp1, Fads2, Fabp2, Fabp5, Acox1, Me1, Cd36, Ppara
Lipid and lipoprotein metabolism	2.11E-06	Slc44a1, Mttp, Lpl, Sgms2, Acacb, Acaca, Mtm1, Helz2, Fads2, Me1, Cd36, Arsa, Srebf1, Stard4, Hsd3b2, Elovi5, Srd5a1, Slc10a2, Elovi2, Acsi5, Fabp1, Gpam, Acox1, Fasn, Lpcat3, Ppara, Mgll, Abcg1
Fatty acid, triacylglycerol, and ketone body metabolism	1.70E-04	Srebf1, Elovi5, Elovi2, Acsi5, Acacb, Acaca, Helz2, Fabp1, Gpam, Acox1, Fasn, Me1, Cd36, Ppara
	Adjusted	
Upregulated Pathways	P-value	Genes
Cysteine and methionine metabolism	0.001861	Sds, Got1, Tat, Cth, Mat1a, Cdo1
p53 activity regulation	0.004402	Cdkn1a, Zmat3, Igfbp3, Sesn2, Ccng1, Mdm2, Tnfrsf10b, Bax, Gtse1
MicroRNA regulation of DNA damage response	0.004582	Cdkn1a, Myc, Ccng1, Mdm2, Tnfrsf10b, Bax, Brca1



Α







Top 1000 Peaks with Largest Gain of Accessibility













total = 1925 variables





В

Α

Mechanism of gene regulation by peroxisome proliferators via PPAR-alpha Alpha-linolenic (omega3) and linoleic (omega6) acid metabolism Lipid and lipoprotein metabolism Biosynthesis of unsaturated fatty acids

Fatty acid, triacylglycerol, and ketone body metabolism

1.00E-13	1.00E-11	1.00E-09	1.00E-07	1.00E-05	1.00E-03	1.00E-01
			p value	e		



PPAR signaling pathway

Supplemental Table 1. PCR Primers used

Primer name (m f Sequence

mLpin1_F	CATGCTTCGGAAAGTCCTTCA
mLpin1_R	GGTTATTCTTTGGCGTCAACCT
mCxcl1_F	CTGGGATTCACCTCAAGAACATC
mCxcl1_R	CAGGGTCAAGGCAAGCCTC
mSaa4_F	CTCTGTTCTTTGTTCCTGGGAG
mSaa4_R	CTAGGTTGTCCCGATAGGCTC
mSlc25a15_F	GCTGCCTCAAGACCTACTCC
mSlc25a15_R	CCGTAACACATGAACAGCACC
mSorbs3_F	TTCAGCTTCGTCTTTGAACAACA
mSorbs3_R	CTTGGGTCAAGGTTGGAGGA
mNnmt_F	TGTGCAGAAAACGAGATCCTC
mNnmt_R	AGTTCTCCTTTTACAGCACCCA
mCdkn1a_F	CCTGGTGATGTCCGACCTG
mCdkn1a_R	CCATGAGCGCATCGCAATC
mTbc1d8_F	AGCCTAGCCAGATCACAAAGA
mTbc1d8_R	CGTCCAGAGGGAACAGTCT
mTymp_F	CGCGGTGATAGATGGAAGAGC
mTymp_R	CACACCTCCTGTGGAGTGTT
mGot1_F	GCGCCTCCATCAGTCTTTG
mGot1_R	ATTCATCTGTGCGGTACGCTC
mEtnppl_F	GCTCTCCGTTTGCTACTTCAC
mEtnppl_R	CCCTCTTGACATCTTTGCCCTT
mll1r1_F	CGAACCGTGAACAACACAAA
mll1r1_R	CAGAGGCACCATGAGACAAA
mLurap1l_F	TCTCTTGGGTCTCTCGGTATAA
mLurap1l_R	TCCACAGCCAGCAAGATTAG
mRetreg1_F	GCCATCAAAGACCAGCTAGAA
mRetreg1_R	GTCCCAGCTCACTCTCAATTT