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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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FOr	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on statistics for biologists contains articles on many of the points above

Software and code

Policy information about <u>availability of computer code</u>

Data collection

Data used for this analysis comes from the AD Knowledge Portal (adknowledgeportal.synapse.org) and the PsychENCODE Knowledge Portal (synapse.org/pec) and is hosted on the Sage Bionetworks' Synapse platform for access by qualified investigators. Data was generated from post-mortem tissue and has be de-identified according to the Synapse terms of use, and is available through the submission of an AD Knowledge Portal Data Use Certificate (https://adknowledgeportal.synapse.org/DataAccess/DataUseCertificates) or by application to the NIMH Repository and Genomics Resources (http://nimhgenetics.org), respectively. We also used publicly available data from GTEX (gtexportal.org).

Data analysis

eQTL discovery was performed with a linear mixed model using the mmQTL software developed in this work (https://github.com/jxzb1988/mmQTL and https://doi.org/10.5281/zenodo.5560014). All downstream analysis was performed with R v4.0.3.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Analysis results are available at http://icahn.mssm.edu/brema

Field and				
Fleid-Spe	ecific reporting			
Please select the o	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.			
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Life scier	nces study design			
All studies must dis	close on these points even when the disclosure is negative.			
Sample size	We included samples with genotype and brain gene expression data from GTEx, PsychENCODE, and ROSMAP datasets. The sample size was based on the availability of the existing data. No statistical method was used to determine what sample size to use			
Data exclusions	Samples with < 8 million well imputed SNPs (info < 0.3) were excluded from PsychENCODE. All other samples with both genotype and brain gene expression were retained.			
Replication	We evaluated the replication of our eQTL analyses in independent datasets of 1) bulk brain tissue (Wang, et al), 2) granule cell layer of the dentate gyrus enriched for excitatory neurons (Jaffe, et al, 2020) and 3) purified microglia (Kosoy, et al. in prep). The genome-wide replication rate is consistent with the rest of the field.			
Randomization	We performed analysis on existing data, so randomization of the experimental design does not apply			
Blinding	We performed analysis on existing data, so blinding of the experimental design does not apply			
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Reportin	g for specific materials, systems and methods			
'	on from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, sed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.			
Materials & ex	perimental systems Methods			
n/a Involved in th	n/a Involved in the study			
Antibodies	ChIP-seq			
Eukaryotic	cell lines Flow cytometry			
Palaeontol	ogy and archaeology MRI-based neuroimaging			
Animals and other organisms				
Human res	earch participants			
Clinical dat	a			
Dual use re	esearch of concern			

Antibodies

Antibodies used

Data generated by Bendl, et al (in revision, https://doi.org/10.1101/2021.01.11.426303) was show in Figure 7 involves use of NeuN antibody. However no data was generated as part of the current study. We refer the reader to that work for experimental details.

Validation

Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.