

Supplementary Material

1 Supplementary Data

1.1 DNA sequences used in EMSA.

Sticky ends (single stranded DNA) are annotated in red.

5500 bp DNA sequence - pCX sfGFP(150TAA) linearised with HindIII digestion:



ATTTATGCAGAGGCCGAGGCCGCCTCGGCCTCTGAGCTATTCCAGAAGTAGTGAGGAGG CTTTTTTGGAGGCCTAGGCTTTTGCAAAAAGCTAACTTGTTTATTGCAGCTTATAATGGT TACAAATAAAGCAATAGCATCACAAATTTCACAAATAAAGCATTTTTTTCACTGCATTCT AGTTGTGGTTTGTCCAAACTCATCAATGTATCTTATCATGTCTGGATCCGCTGCATTAAT GAATCGGCCAACGCGCGGGGGGGGGGGGGGGGGGTTTGCGTATTGGGCGCTCTTCCGCTTCCTCG GGCGGTAATACGGTTATCCACAGAATCAGGGGGATAACGCAGGAAAGAACATGTGAGCA AAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAG GCTCCGCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACC CGACAGGACTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTG TTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTGGCGC TTTCTCAATGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAGCTGG GCTGTGTGCACGAACCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAACTATCGTC TTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCCACTGGTAACAGG ATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAACTA CGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCTTCGG AAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCGCTGGTAGCGGTGGTTTTT TTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATC TTTTCTACGGGGTCTGACGCTCAGTGGAACGAAAACTCACGTTAAGGGATTTTGGTCATG AGATTATCAAAAAGGATCTTCACCTAGATCCTTTTAAATTAAAAATGAAGTTTTAAATCA ATCTAAAGTATATATGAGTAAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGC ACCTATCTCAGCGATCTGTCTATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAG ATAACTACGATACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGA GCAGAAGTGGTCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAG CTAGAGTAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAGGCA TCGTGGTGTCACGCTCGTCGTTTGGTATGGCTTCATTCAGCTCCGGTTCCCAACGATCAA GGCGAGTTACATGATCCCCCATGTTGTGCAAAAAGCGGTTAGCTCCTTCGGTCCTCCGA TCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCAGCACTGCATA ATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCA AGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTTGCCCGGCGTCAATACGG GATAATACCGCGCCACATAGCAGAACTTTAAAAGTGCTCATCATTGGAAAACGTTCTTC GGGGCGAAAACTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGTAACCCACTC GTGCACCCAACTGATCTTCAGCATCTTTTACTTTCACCAGCGTTTCTGGGTGAGCAAAAA CAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGGGCGACACGGAAATGTTGAATACT CATACTCTTCCTTTTTCAATATTATTGAAGCATTTATCAGGGTTATTGTCTCATGAGCGGA TACATATTTGAATGTATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTCCCCG AAAAGTGCCACCTGGTCGACATTGATTATTGACTAGTTATTAATAGTAATCAATTACGGG GTCATTAGTTCATAGCCCATATATGGAGTTCCGCGTTACATAACTTACGGTAAATGGCCC GCCTGGCTGACCGCCCAACGACCCCCGCCCATTGACGTCAATAATGACGTATGTTCCCAT AGTAACGCCAATAGGGACTTTCCATTGACGTCAATGGGTGGACTATTTACGGTAAACTG CCCACTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATG ACGGTAAATGGCCCGCCTGGCATTATGCCCAGTACATGACCTTATGGGACTTTCCTACTT GGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGGTCGAGGTGAGCCCCACGTTC

CGGCGCGCTCCGAAAGTTTCCTTTTATGGCGAGGCGGCGGCGGCGGCGGCGGCCCTATAAAA GCCGCCTCGCGCCGCCCGGCCCCGGCTCTGACTGACCGCGTTACTCCCACAGGTGAGCGG GCGGGACGGCCCTTCTCCTCCGGGCTGTAATTAGCGCTTGGTTTAATGACGGCTCGTTTC TTTTCTGTGGCTGCGTGAAAGCCTTAAAGGGCTCCGGGAGGGCCCTTTGTGCGGGGGGG CTGCCCGGCGGCTGTGAGCGCTGCGGGGCGCGCGGGGGCTTTGTGCGCTCCGCGTGTG GTCGGGCTGTAACCCCCCCTGCACCCCCCCCCGAGTTGCTGAGCACGGCCCGGCTTC AGGGGCGCGGCGGCCCCGGAGCGCCGGCGGCGGCGAGCCGAGCCGAGCCA TTGCCTTTTATGGTAATCGTGCGAGAGGGGCGCAGGGACTTCCTTTGTCCCAAATCTGTGC TGCGGCGCCGGCAGGAAGGAAATGGGCGGGGGGGGGCCTTCGTGCGTCGCCGCGCCGCC GTCCCCTTCTCCCACCTCGGGGGCTGTCCGCGGGGGGGACGGCTGCCTTCGGGGGG GACGGGGCAGGGCGGGGTTCGGCTTCTGGCGTGTGACCGGCGGCTCTAGAGCCTCTGCT AACCATGTTCATGCCTTCTTTTTTCCTACAGCTCCTGGGCAACGTGCTGGTTGTTGTGC TGTCTCATCATTTTGGCAAAGAATTCGCCACCATGGTGAGCAAGGGCGAGGAGCTGTTC ACCGGCGTGGTGCCCATCCTGGTGGAGCTGGACGGCGACGTGAACGGCCACAAGTTCAG CGTGCGCGGCGAGGGCGAGGGCGACGCCACCAACGGCAAGCTGACCCTGAAGTTCATC TGCACCACCGGCAAGCTGCCCGTGCCCTGGCCCACCCTGGTGACCACCCTGACCTACGG CGTGCAGTGCTTCAGCCGCTACCCCGACCACATGAAGCGCCACGACTTCTTCAAGAGCG CCATGCCCGAGGGCTACGTGCAGGAGCGCACCATCAGCTTCAAGGACGACGGCACCTAC AAGACCCGCGCCGAGGTGAAGTTCGAGGGCGACACCCTGGTGAACCGCATCGAGCTGA AGGGCATCGACTTCAAGGAGGACGGCAACATCCTGGGCCACAAGCTGGAGTACAACTTC AACAGCCACTAAGTGTACATCACCGCCGACAAGCAGAAGAACGGCATCAAGGCCAACT TCAAGATCCGCCACAACGTGGAGGACGGCAGCGTGCAGCTGGCCGACCACTACCAGCA GAACACCCCCATCGGCGACGGCCCCGTGCTGCTGCCCGACAACCACTACCTGAGCACCC AGAGCGTGCTGAGCAAGGACCCCAACGAGAAGCGCGACCACATGGTGCTGCTGGAGTT CGTGACCGCCGCCGGCATCACCCACGGCATGGACGAGCTGTACAAGTGAGAATTCACTC CTCAGGTGCAGGCTGCCTATCAGAAGGTGGTGGCTGGTGGGCCAATGCCCTGGCTCAC AAATACCACTGAGATCTTTTTCCCTCTGCCAAAAATTATGGGGACATCATGAAGCCCCTT GAGCATCTGACTTCTGGCTAATAAAGGAAATTTATTTTCATTGCAATAGTGTGTTGGAAT TTTTTGTGTCTCTCACTCGGAAGGACATATGGGAGGGCAAATCATTTAAAACATCAGAA GGCTATAAAGAGGTCATCAGTATATGAAACAGCCCCCTGCTGTCCATTCCTTATTCCATA ACATCCCTAAAATTTTCCTTACATGTTTTACTAGCCAGATTTTTCCTCCTCTCCTGACTAC **TCCCAGTCATAGCTGTCCCTCTTCTCTTATGAAGATCCCTCGACCTGCAGCCCAAGCT**



2700 bp DNA sequence - pUC18 linearised with HindIII digestion:

AGCTTGCATGCCTGCAGGTCGACTCTAGAGGATCCCCGGGTACCGAGCTCGAATTCGTA ATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTATCCGCTCACAAATTCCACACAACAT TAATTGCGTTGCGCTCACTGCCCGCTTTCCAGTCGGGAAACCTGTCGTGCCAGCTGCATT AAGGCGGTAATACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACATGTGAG CAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCAT AGGCTCCGCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCGAA ACCCGACAGGACTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCT CCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGGAAGCGTG GCGCTTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGTAGGTCGTTCGCTCCAAG CTGGGCTGTGTGCACGAACCCCCGTTCAGCCCGACCGCTGCGCCTTATCCGGTAACTAT CGTCTTGAGTCCAACCCGGTAAGACACGACTTATCGCCACTGGCAGCAGCACTGGTAA CAGGATTAGCAGAGCGAGGTATGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTA ACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCTGAAGCCAGTTACCT TTTTTTGTTTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTT GATCTTTTCTACGGGGTCTGACGCTCAGTGGAACGAAAACTCACGTTAAGGGATTTTGGT CATGAGATTATCAAAAAGGATCTTCACCTAGATCCTTTTAAATTAAAAATGAAGTTTTAA ATCAATCTAAAGTATATATGAGTAAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGA GGCACCTATCTCAGCGATCTGTCTATTTCGTTCATCCATAGTTGCCTGACTCCCCGTCGTG TAGATAACTACGATACGGGAGGGCTTACCATCTGGCCCCAGTGCTGCAATGATACCGCG AAGCTAGAGTAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTGCTACAG CAAGGCGAGTTACATGATCCCCCATGTTGTGCAAAAAGCGGTTAGCTCCTTCGGTCCTC CGATCGTTGTCAGAAGTAAGTTGGCCGCAGTGTTATCACTCATGGTTATGGCAGCACTGC ATAATTCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAA CCAAGTCATTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTTGCCCGGCGTCAATA CGGGATAATACCGCGCCACATAGCAGAACTTTAAAAGTGCTCATCATTGGAAAACGTTC TTCGGGGCGAAAACTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGTAACCCA CTCGTGCACCCAACTGATCTTCAGCATCTTTTACTTTCACCAGCGTTTCTGGGTGAGCAA AAACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGGGCGACACGGAAATGTTGAA TACTCATACTCTTTCCATTTCAATATTATTGAAGCATTTATCAGGGTTATTGTCTCATGAG CGGATACATATTTGAATGTATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTC CCCGAAAAGTGCCACCTGACGTCTAAGAAACCATTATTATCATGACATTAACCTATAAA AATAGGCGTATCACGAGGCCCTTTCGTCTCGCGCGTTTCGGTGATGACGGTGAAAACCT CTGACACATGCAGCTCCCGGAGACGGTCACAGCTTGTCTGTAAGCGGATGCCGGGAGCA GCGGCATCAGAGCAGATTGTACTGAGAGTGCACCATATGCGGTGTGAAATACCGCACAG ATGCGTAAGGAGAAAATACCGCATCAGGCGCCATTCGCCATTCAGGCTGCGCAACTGTT GGGAAGGGCGATCGGTGCGGGCCTCTTCGCTATTACGCCAGCTGGCGAAAGGGGGGATGT GCTGCAAGGCGATTAAGTTGGGTAACGCCAGGGTTTTCCCAGTCACGACGTTGTAAAAC GACGGCCAGTGCCAAGCT

1300 bp PCR fragment - PCR of gene encoding maltose binding protein:

RBS	MBP	Factor Xa site
signal) —

TTTTGTTTAACTTTAAGAAGGAGATATACATATGAAAATAAAAACAGGTGCACGCATCC TCGCATTATCCGCATTAACGACGATGATGTTTTCCGCCTCGGCTCTCGCCAAAATCGAAG AAGGTAAACTGGTAATCTGGATTAACGGCGATAAAGGCTATAACGGTCTCGCTGAAGTC GGTAAGAAATTCGAGAAAGATACCGGAATTAAAGTCACCGTTGAGCATCCGGATAAACT GGAAGAGAAATTCCCACAGGTTGCGGCAACTGGCCGATGGCCCTGACATTATCTTCTGGG CACACGACCGCTTTGGTGGCTACGCTCAATCTGGCCGGATGGCCGTACGTTACAACGGAC AAAGCGTTCCAGGACAAGCTGTATCCGTTTACCTGGGATGCCGTACGTTACAACGGCAA GCTGATTGCTTACCCGATCGCTGTTGAAGCGTTATCGCTGATTTATAACAAAGATCTGCT GCCGAACCCGCCAAAAACCTGGGAAAGAAGATCCCGGCGCTGGATAAAGAACTGAAAGCG

2 Supplementary Figures and Tables

2.1 Supplementary Figures



Supplementary Figure 1. Mass spectrometry analysis of purified proteins from **Figure 2** of the main manuscript. Mass spectrometry analysis of purified (**A**) HU α , (**B**) HU β , (**C**) HU β (K67ac) and (**D**) HU β (K86ac).



Supplementary Figure 2. Effect of acetylation of K67ac and K86ac on interaction with 30-bp DNA containing 2-nucleotide gap. EMSA of *E. coli* HU β_2 (**A**) or HU $\alpha\beta$ (**B**) with 30-bp DNA containing a 2-nt gap (shown in **Figure 3A**). Free DNA remaining in HU-containing conditions was calculated relative to the band intensity of the DNA-only lane (= 100%). Average free DNA remaining (Rel %) and standard deviation (SD) are presented below three independent repeats (i, ii, and iii).

A 2-nt gap repeats



B Nicked backbone repeats



Supplementary Figure 3. Repeats of EMSAs from **Figure 4** of the main manuscript. (A) EMSA comparing the effect of acetylation on the HU β_2 homodimer vs the HU $\alpha\beta$ heterodimer when interacting with a 30 bp DNA fragment containing a 2-nucleotide gap. Repeats of the representative

figure in panel **A** of **Figure 4** of the main manuscript. (**B**) EMSA comparing the effect of acetylation on the HU β_2 homodimer vs the HU $\alpha\beta$ heterodimer when interacting with a 30 bp with a nicked phosphate backbone. Repeats of the representative figure in panel **B** of **Figure 4** of the main manuscript. (**C**) EMSA comparing the effect of acetylation on the HU β_2 homodimer vs the HU $\alpha\beta$ heterodimer when interacting with a 30 bp DNA fragment with a complete double-stranded duplex. Repeats of the representative figure in panel **C** of **Figure 4** of the main manuscript.



Supplementary Figure 4. EMSA with alternative 30 bp fully complementary DNA fragment and 420 bp DNA fragment. (**A**) Sequence of alternative fully complementary 30 bp DNA fragment. The phosphate backbone is indicated by a solid line and hydrogen bonds between base pairs are indicated by dashed lines. (**B**) EMSA comparing acetylation of Lys86 of HU β_2 homodimer vs the HU $\alpha\beta$

heterodimer when interacting with an alternative 30 bp DNA fragment. Three repeats (i, ii and iii) are shown. (C) EMSA comparing acetylation of HU β_2 when interacting with a 420 bp DNA fragment.



Supplementary Figure 5. Repeats of EMSAs from Figure 4 of the main manuscript. (A) EMSA investigating the effect of acetylation of the HU β_2 homodimer on interaction with a 5500 bp linear DNA fragment. Repeats of the representative figure in Figure 4D i) of the main manuscript. (B) EMSA investigating the effect of acetylation of the HU β_2 homodimer on interaction with a 2700 bp linear DNA fragment. Repeats of the representative figure in Figure 4D ii) of the main manuscript.

(C) EMSA investigating the effect of acetylation of the $HU\beta_2$ homodimer on interaction with a 1300 bp linear DNA fragment. Repeats of the representative figure in **Figure 4D iii**) of the main manuscript.



Supplementary Figure 6. Effect of acetylation at Lys86 on DNA supercoiling ability of HU β_2 in the presence of topoisomerase I. Two repeats, **A** and **B**, are shown. A miniprepped 5500 bp plasmid (non-relaxed) was relaxed with *E. coli* topoisomerase I then purified (relaxed). The relaxed DNA was then incubated with HU β_2 , or HU β (K86ac)₂ in the presence of calf thymus topoisomerase I. The samples were treated with proteinase K before electrophoresis.



Supplementary Figure 7. Analytical size exclusion high-performance liquid chromatography. (**A**) Analytical size exclusion chromatography of Bio-Rad Gel Filtration standards. The absorbance at 214 nm is plotted against the retention time of each of the five protein standards. The known molecular weights of the standards are 670, 158, 44, 17 and 1.35 kDa. (**B**) Analytical size exclusion chromatography of HU proteins. The absorbance at 214 nm is plotted against the retention time. (**C**) A standard curve generated from the retention times of the 5 protein standards in A and the log of their molecular weights. (**D**) A standard curve generated from the retention times of the 44, 17 and 1.35 kDa standards and the log of their molecular weights. For estimated molecular weight calculations, please see **Supplementary Tables 7 - 9**.

2.2 Supplementary Tables

Supplementary Table 1. Calculations of free DNA remaining for EMSAs of $HU\beta_2$ homodimers incubated with a 30 bp DNA fragment containing a 2-nucleotide gap. Average and standard deviation were calculated for three independent repeats. A single outlier was omitted from the calculations and is annotated in red.

		Н	$J\beta_2$			Ηυβ(κ	K67ac) ₂		HUβ(K86ac) ₂			
μM HU	0.5	1	2	4	0.5	1	2	4	0.5	1	2	4
Repeat 1	84	37	21	12	77	59	32	15	86	87	74	57
Repeat 2	93	82	29	11	83	52	28	14	97	93	69	41
Repeat 3	87	77	24	12	78	65	32	15	89	81	70	49
Average	88	80	25	12	79	59	31	15	91	87	71	49
Standard Deviation	4.6	3.5	4.0	0.6	3.2	6.5	2.3	0.6	5.7	6.0	2.7	8.0

Supplementary Table 2. Calculations of free DNA remaining for EMSAs of HU $\alpha\beta$ heterodimers incubated with a 30 bp DNA fragment containing a 2-nucleotide gap. Average and standard deviation were calculated for three independent repeats.

	ΗUαβ					HUαβ(K67ac)				HUαβ(K86ac)			
μM HU	0.5	1	2	4	0.5	1	2	4	0.5	1	2	4	
Repeat 1	69	27	5	4	68	35	5	4	70	45	10	8	
Repeat 2	78	39	7	2	68	35	5	4	68	41	5	5	
Repeat 3	82	61	9	2	77	48	23	9	79	58	12	2	
Average	76	42	7	3	71	39	11	6	72	48	9	5	
Standard Deviation	6.7	17.2	2.0	1.2	5.2	7.5	10.4	2.9	5.9	8.9	3.6	3.0	

Supplementary Table 3. Calculations of free DNA remaining for EMSAs comparing the effects of acetylation on $HU\beta_2$ homodimers vs $HU\alpha\beta$ heterodimers incubated with a 30 bp DNA fragment containing a 2-nucleotide gap. Average and standard deviation were calculated for three independent repeats.

		ΗUαβ			αβ(K86	K86ac) HUβ ₂				HUβ(K86ac) ₂		
μM HU	0.5	1	2	0.5	1	2	0.5	1	2	0.5	1	2
Repeat 1	72	36	9	77	52	9	74	52	27	89	85	60
Repeat 2	64	26	4	71	42	3	66	48	20	75	76	67
Repeat 3	83	51	8	80	56	15	92	75	42	94	84	72
Average	73	38	7	76	50	9	77	58	30	86	82	66
Standard Deviation	9.5	12.6	2.7	4.6	7.2	6.0	13.3	14.6	11.2	9.6	4.9	6.0

Supplementary Table 4. Calculations of free DNA remaining for EMSAs comparing the effects of acetylation on HU β_2 homodimers vs HU $\alpha\beta$ heterodimers incubated with a 30 bp DNA fragment with a nicked phosphate backbone. Average and standard deviation were calculated for three independent repeats.

		Ηυαβ		HU	αβ(K86	bac)	$HU\beta_2$			HUβ(K86ac) ₂		
μM HU	1	2	4	1	2	4	1	2	4	1	2	4
Repeat 1	45	11	6	16	4	3	60	57	27	69	62	46
Repeat 2	52	13	9	54	9	5	98	65	35	102	91	71
Repeat 3	42	13	8	44	15	6	79	58	37	81	68	48
Average	46	12	8	38	9	5	79	60	36	84	74	47
Standard Deviation	5.1	1.2	1.5	19.7	5.5	1.5	19.0	4.4	1.4	16.7	15.3	1.4

		Ηυαβ		HU	Ηυαβ(Κ86ac)			$HU\beta_2$			HUβ(K86ac) ₂		
μM HU	1.5	3	6	1.5	3	6	1.5	3	6	1.5	3	6	
Repeat 1	84	37	14	76	36	11	90	68	51	82	86	70	
Repeat 2	83	50	7	79	50	10	94	77	45	89	86	73	
Repeat 3	89	76	21	81	42	14	100	89	80	78	73	67	
Average	85	54	14	79	43	12	95	78	59	83	82	70	
Standard Deviation	3.2	19.9	7.0	2.5	7.0	2.1	5.0	10.5	18.7	5.6	7.5	3.0	

Supplementary Table 5. Calculations of free DNA remaining for EMSAs comparing the effects of acetylation on $HU\beta_2$ homodimers vs $HU\alpha\beta$ heterodimers incubated with a 30 bp fully complementary DNA fragment. Average and standard deviation were calculated for three independent repeats.

Supplementary Table 6. Calculations of free DNA remaining for EMSAs comparing the effects of acetylation on $HU\beta_2$ homodimers vs $HU\alpha\beta$ heterodimers incubated with an alternative 30 bp fully complementary DNA fragment. Average and standard deviation were calculated for three independent repeats.

	ΗUαβ			HU	Uαβ(K86ac) HU			$HU\beta_2$	B ₂ HUβ(K			ac)2
μM HU	1	2	4	1	2	4	1	2	4	1	2	4
Repeat 1	88	59	17	85	61	15	85	75	48	82	88	75
Repeat 2	82	43	20	94	55	19	100	99	81	100	96	79
Repeat 3	92	64	15	93	60	22	100	98	74	100	98	91
Average	87	55	17	91	59	19	95	91	68	94	94	82
Standard Deviation	5	11	3	5	3	4	9	14	17	10	5	8

Supplementary Table 7. Retention times of Bio-Rad Gel Filtration Standards. The retention times of three repeats are provided alongside the known molecular weights of the proteins. The log of the molecular weights were plotted against the average retention times to generate a calibration curve (**Supplementary Figure 7A** and **B**).

		Retention	Time (min)			
	1	2	3	Average	MW (Da)	Log(WW)
Thyroglobulin	5.636	5.626	5.624	5.629	670000	5.826
y-globulin	6.242	6.232	6.224	6.233	158000	5.199
Ovalbumin	7.135	7.125	7.122	7.127	44000	4.643
Myoglobin	8.215	8.209	8.205	8.210	17000	4.230
Vitamin B12	12.595	12.573	12.563	12.577	1350	3.130

Supplementary Table 8. Verifying standard curve molecular weight estimations. The line of best fits generated in the calibration curves (**Supplementary Figure 7A** and **B**) were used to calculate the molecular weights of the protein standards based on their retention times. This calculated value was compared to the known molecular weight of the standards. Equation 2 provides more accurate estimations of molecular weight at lower molecular weight ranges (**1350 – 44000 Da**) and was used for further analyses as the estimated molecular weights of HU protein dimers are within this range (*ca.* 24000 Da).

	MW		Average	Equa	tion 1	Equation 2		
	(Da)	Log(MW)	RT (min)	Log(MW)	MW (Da)	Log(MW)	MW (Da)	
Thyroglobulin	670000	5.826	5.629	5.430	269340	4.995	98758	
y-globulin	158000	5.199	6.233	5.216	164529	4.831	67813	
Ovalbumin	44000	4.643	7.127	4.899	79282	4.589	38858	
Myoglobin	17000	4.230	8.210	4.516	32779	4.297	19812	
Vitamin B12	1350	3.130	12.577	2.968	929	3.116	1307	

Supplementary Table 9. Molecular weight estimations of HU proteins based on their retention times in analytical size exclusion chromatography. A standard curve based on the retention times of protein standards of known molecular weights (**Supplementary Figure 7D**, **Supplementary Tables 7-8**) was used to estimate the molecular weight of HU proteins. The estimations suggest that the HU proteins exist in their dimeric forms.

	RT (min)	Log (MW)	Calculated MW (Da)	Expected dimer MW	Expected monomer MW
HUa ₂	7.908	4.378	23904	20716	10358
ΗUαβ	7.952	4.367	23258	20649	10358+10291
$HU\alpha\beta^{(K67ac)}$	7.94	4.370	23432	20691	10358+10333
$HU\alpha\beta^{(K86ac)}$	7.902	4.380	23993	20691	10358+10333
HUβ ₂	7.915	4.377	23800	20582	10291
$HU\beta^{(K67ac)}_{2}$	7.916	4.376	23785	20666	10333
$HU\beta^{(K86ac)}_{2}$	7.901	4.380	24008	20666	10333