

**Design, Synthesis, and Biological Evaluation of 4-Quinoline Carboxylic Acids as Inhibitors of Dihydroorotate Dehydrogenase**

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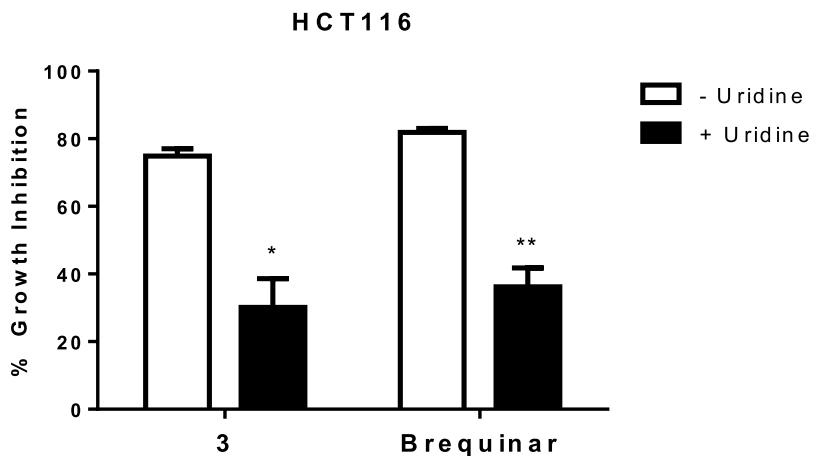
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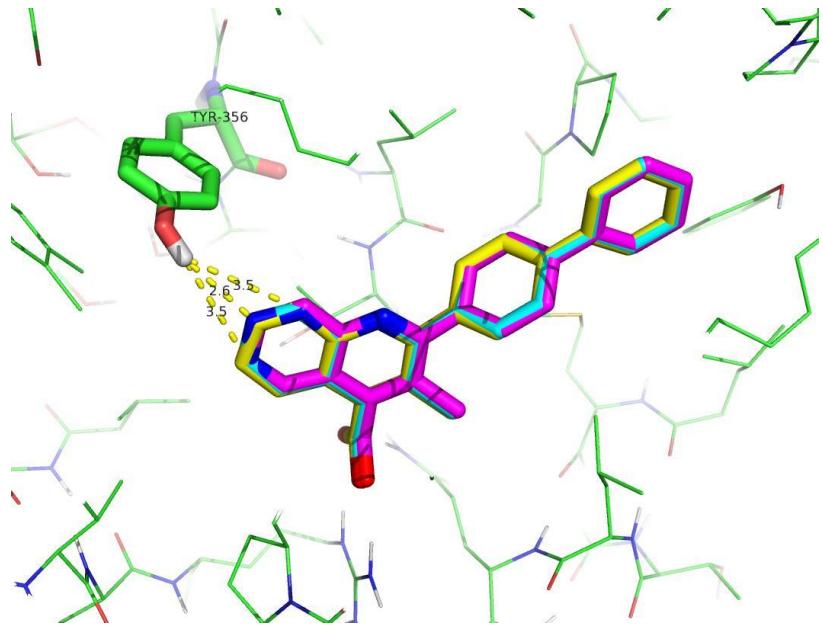
## Supplemental Figures

Analysis Type by Cancer	Cancer vs. Normal	Cancer vs. Cancer		Cancer Subtype Analysis							Cancer vs. Baseline (DNA only)	Pathway and Drug		Outlier	
		Cancer Histology	Mutli-cancer	Clinical Outcome	Metastasis vs. Primary	Molecular Subtype: Biomarker	Molecular Subtype: Mutation	Pathology Subtype: Grade	Pathology Subtype: Stage	Patient Treatment Response		Drug Sensitivity	Perturbation		
Bladder Cancer														2 2	
Brain and CNS Cancer														2 7	
Breast Cancer														6 8	
Cervical Cancer														5 6	
Colorectal Cancer	1													1	
Esophageal Cancer														4 2	
Gastric Cancer														2 4	
Head and Neck Cancer														3 4	
Kidney Cancer	1	1 1												5 14	
Leukemia														4 6	
Liver Cancer	6 1 1													2 4	
Lung Cancer	1													4 7	
Lymphoma														4 5	
Melanoma														9	
Myeloma														4 10	
Other Cancer														3 1	
Ovarian Cancer														2 1	
Pancreatic Cancer														1 14	
Prostate Cancer														5 8	
Sarcoma														51 101	
Significant Unique Analyses	3	6	2 2									1		51 964	
Total Unique Analyses	464	743	255	622	50	254	634	211	462	100	27	118	243	942	291

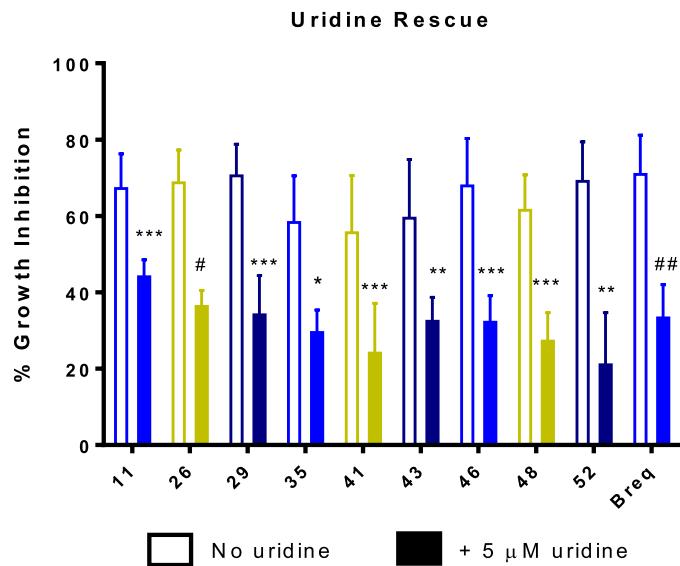
**Supplemental Figure 1: Overexpression of DHODH in various cancers.** This data was generated using the Oncomine database.<sup>1-3</sup>



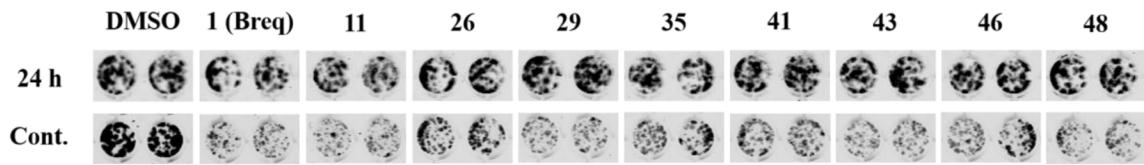
**Supplemental Figure 2: Uridine supplementation rescues cell growth inhibition induced by lead compound 3.** Compound 3 and brequinar were evaluated at twice their IC<sub>50</sub> values. Antiproliferative effects were determined in media ± 5 µM uridine. Data are represented as the mean and standard deviation from three independent experiments. \* p < 0.001, \*\* p < 0.0005



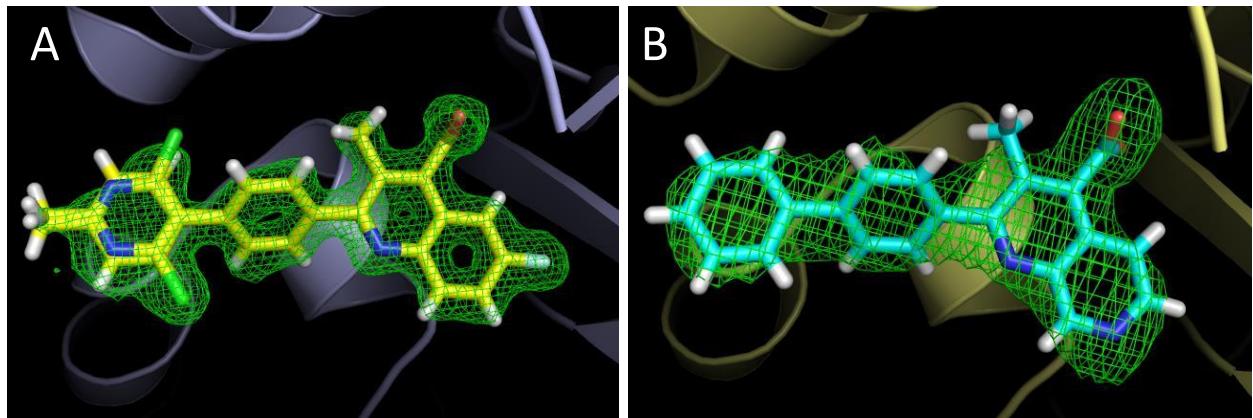
**Supplemental Figure 3: 1,7-naphthyridines show optimal positioning to form a hydrogen bond with Y356.** DHODH (1D3G, green), **45** (yellow), **46** (cyan), and **47** (magenta).



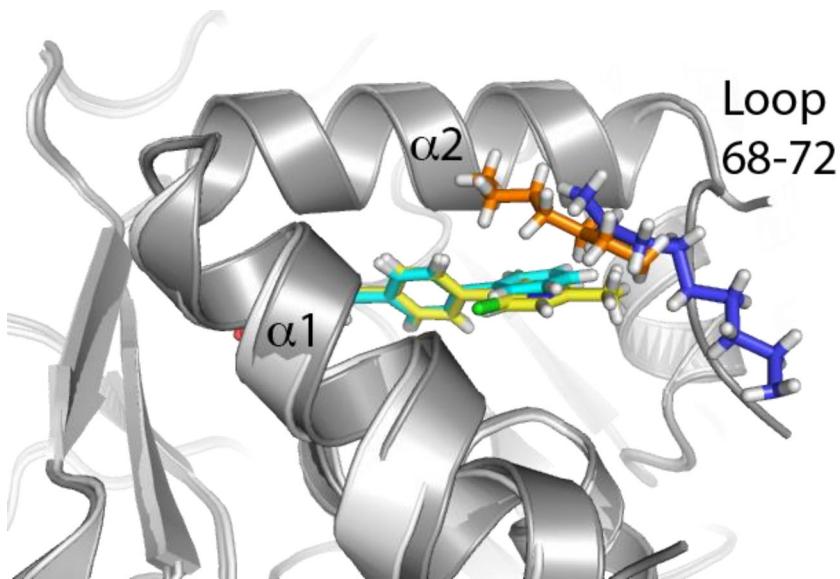
**Supplemental Figure 4: Uridine supplementation rescues cell growth inhibition induced by select optimized analogs.** Compounds were evaluated at twice their IC<sub>50</sub> values. Antiproliferative effects were determined in media ± 5 μM uridine. Data are represented as the mean and standard deviation from at least three independent experiments. \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.005, # p = 0.0005, ## p ≤ 0.0001.



**Supplemental Figure 5: DHODH inhibitors are cytostatic.** HCT-116 cells were treated with 10  $\mu$ M of the indicated analogs for either 24 hr or continuously for a period of seven days.



**Supplemental Figure 6:** Difference electron density (green mesh) corresponding to **43** (yellow, left) and **46** (cyan, right) contoured to  $3\sigma$ .



**Supplemental Figure 7:** Overlay of DHODH structure with inhibitors **43** (polypeptide in light gray, inhibitor in yellow, Zwittergent aliphatic chain shown in orange) compared DHODH bound to **46** (polypeptide in dark gray, inhibitor in cyan, Zwittergent aliphatic chain shown in blue) showing differences in the loop near the binding site corresponding to residues 68-72 where peptide was able to be modeled. Also shown are modeled aliphatic chains associated with detergent packed against the 68-72 loop and inhibitors.

**Supplemental Table 1: Molecular docking binding energies**

Compound #	Affinity (kcal/mol) <sup>a</sup>	Predicted K <sub>i</sub>
1D3G brequinar analog	-13.0	3.91E-11
13	-12.1	2.05E-10
14	-12.2	1.71E-10
16	-12.2	1.71E-10
22	-12.9	4.7E-11
26	-12.6	8.17E-11
28	-12.6	8.17E-11
32	-12.6	8.17E-11
45	-12.4	1.18E-10
46	-12.3	1.42E-10
47	-12.3	1.42E-10

<sup>a</sup>All binding energies shown are for the highest-ranking pose.

**Supplemental Table 2: Crystallography Data Collection and Refinement Statistics**

Data Collection	DHODH:43	DHODH:46
PDBID	6CJF	6CJG
Space Group	P1	P3 <sub>2</sub> 21
Unit Cell (Å)	a = 50.753 b = 51.528 c = 65.088 α = 89.15° β = 81.61° γ = 76.44°	a = b = 90.098 c = 123.176 α = β = 90° γ = 120°
Wavelength (Å)	0.9786	0.9786
Resolution (Å) <sup>1</sup>	1.63 (1.63-1.66)	2.85 (2.85-2.90)
Rsym <sup>2</sup>	0.056 (0.429)	0.223 (1.075)
<I/Ī> <sup>3</sup>	10 (2)	10 (2)
Completeness (%) <sup>4</sup>	97.2 (95.2)	100 (100)
Redundancy	4.0 (4.0)	16.2 (15.2)
<b>Refinement</b>		
Resolution (Å)	1.63	2.85
R-Factor <sup>5</sup>	0.163	0.170
Rfree <sup>6</sup>	0.188	0.206
Protein atoms	5303	2654
Compounds	2	1
Water Molecules	365	67
Unique Reflections	75537	13950
R.m.s.d. <sup>7</sup>		
Bonds	0.006	0.008
Angles	0.865	0.991
MolProbity Score <sup>8</sup>	1.44	1.54
Clash Score <sup>8</sup>	3.93	4.22
RSR <sup>9</sup>	0.10/0.09	0.16
RSCC <sup>9</sup>	0.93/0.93	0.97

<sup>1</sup>Statistics for highest resolution bin of reflections in parentheses.

<sup>2</sup>R<sub>sym</sub> =  $\sum_h \sum_j |I_{hj} - \langle I_h \rangle| / \sum_h \sum_j I_{hj}$ , where I<sub>hj</sub> is the intensity of observation j of reflection h and  $\langle I_h \rangle$  is the mean intensity for multiply recorded reflections.

<sup>3</sup>Intensity signal-to-noise ratio.

<sup>4</sup>Completeness of the unique diffraction data.

<sup>5</sup>R-factor =  $\sum_h |IF_o - IF_c| / \sum_h |F_o|$ , where F<sub>o</sub> and F<sub>c</sub> are the observed and calculated structure factor amplitudes for reflection h.

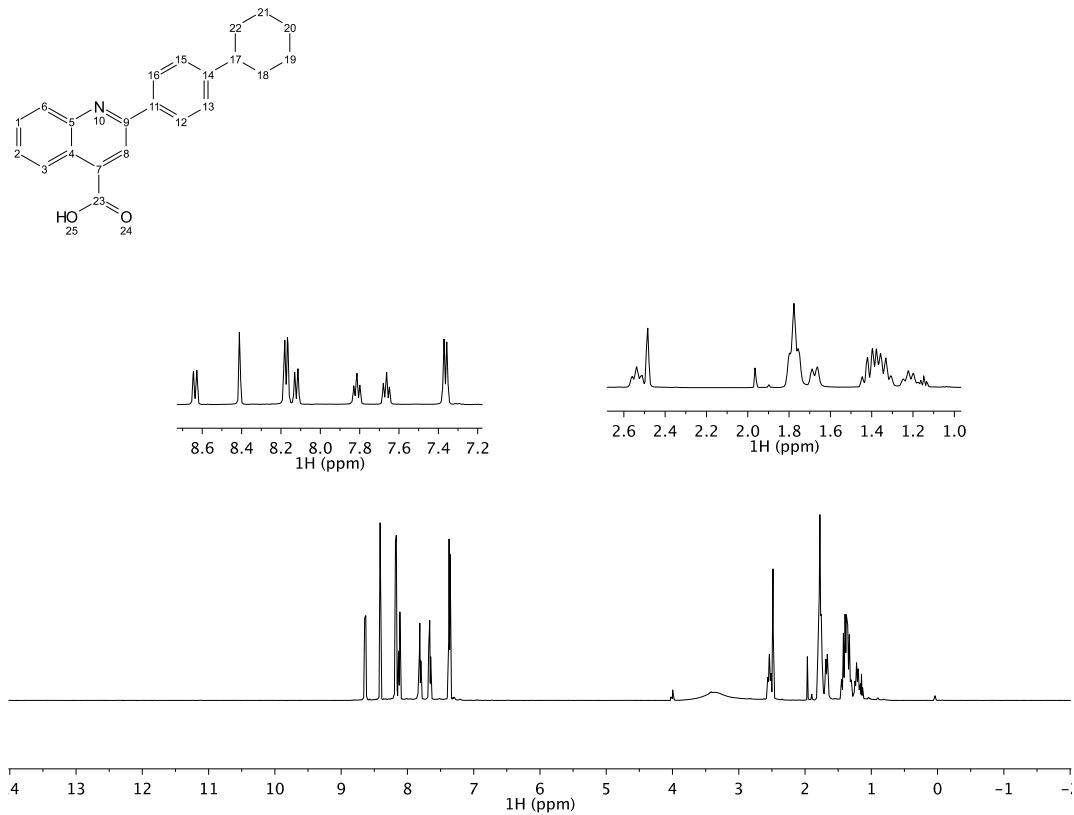
<sup>6</sup>R<sub>free</sub> is calculated against a 5% random sampling of the reflections that were removed before structure refinement.

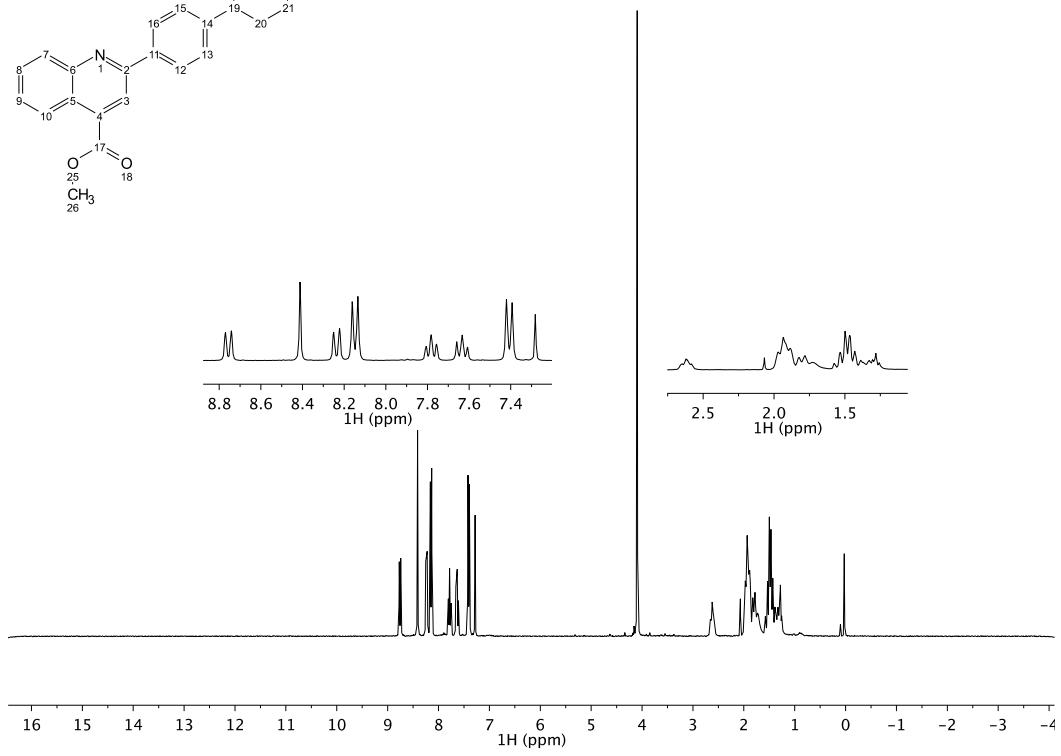
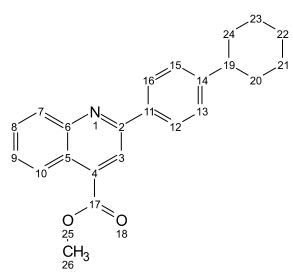
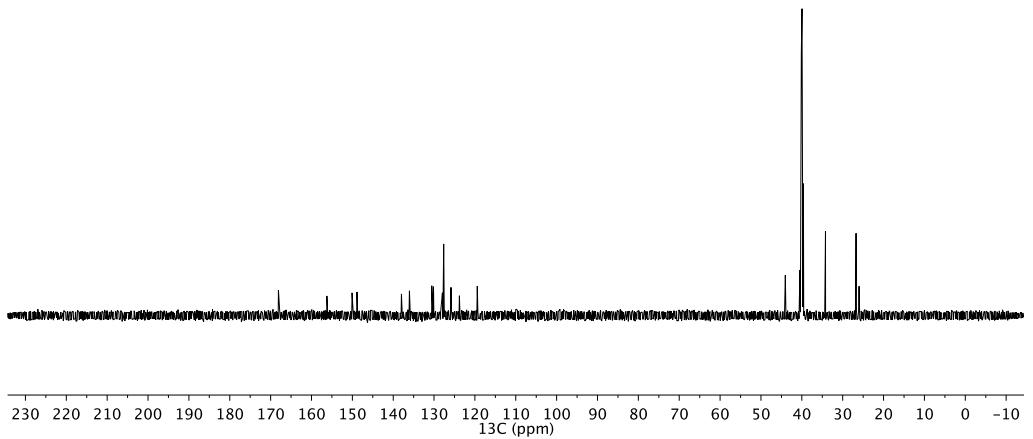
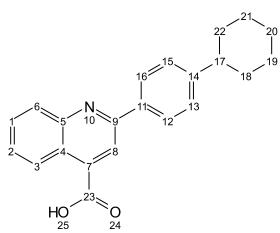
<sup>7</sup>Root mean square deviation of bond lengths and bond angles.

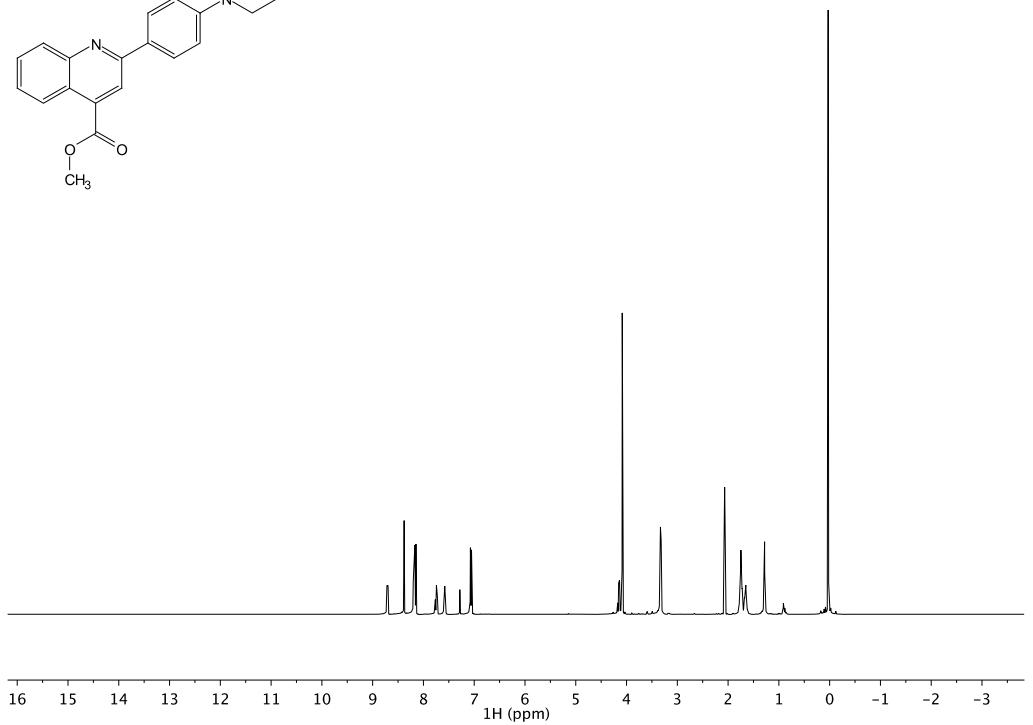
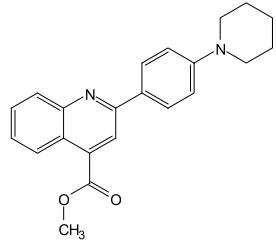
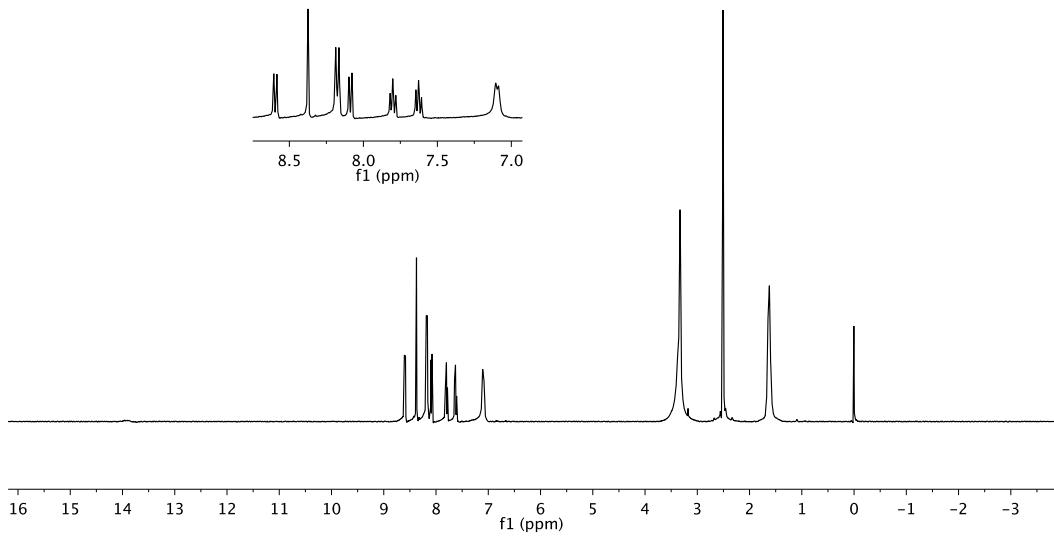
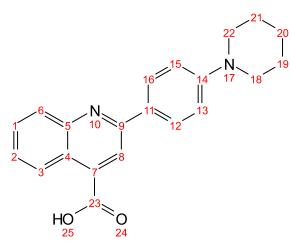
<sup>8</sup>Chen et al. (2010) MolProbity: all-atom structure validation for macromolecular crystallography. Acta Crystallographica D66:12-21.

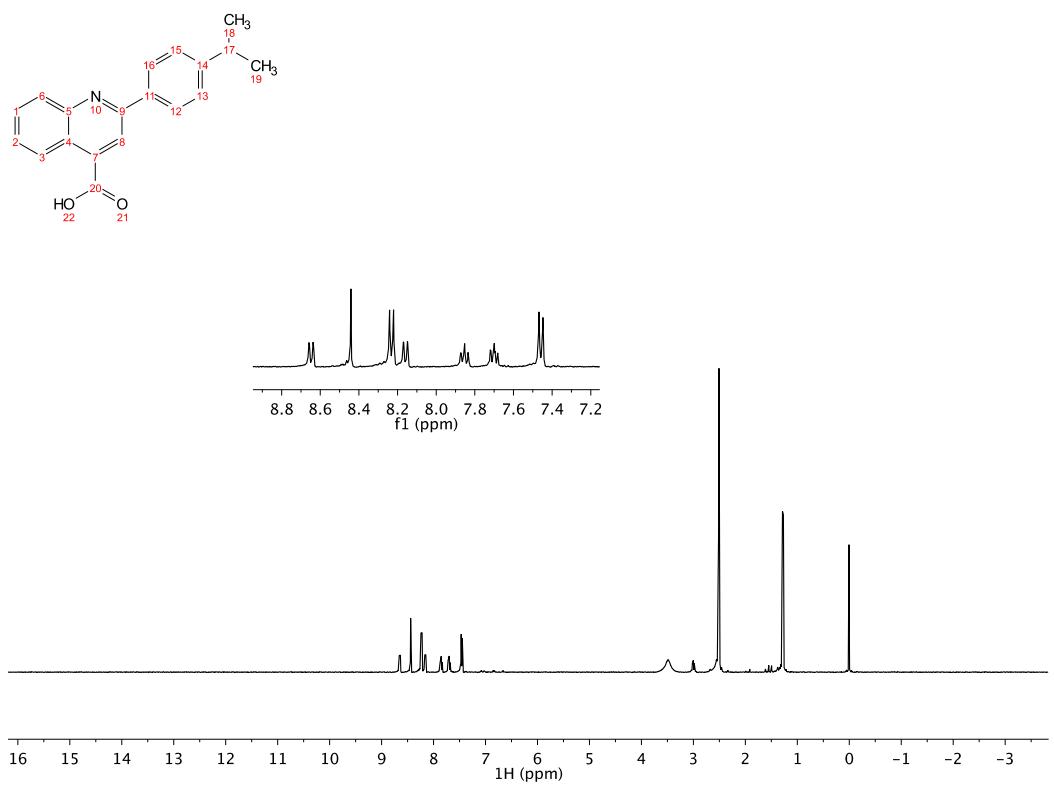
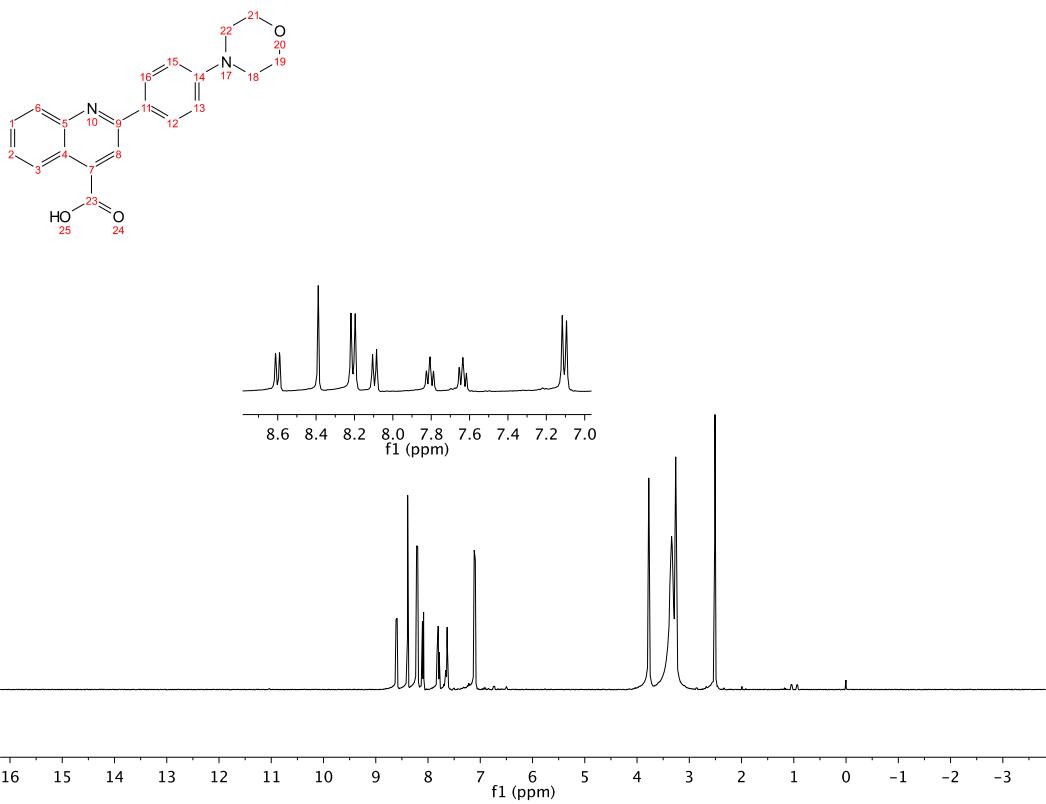
<sup>9</sup>wwPDB Validation Server.

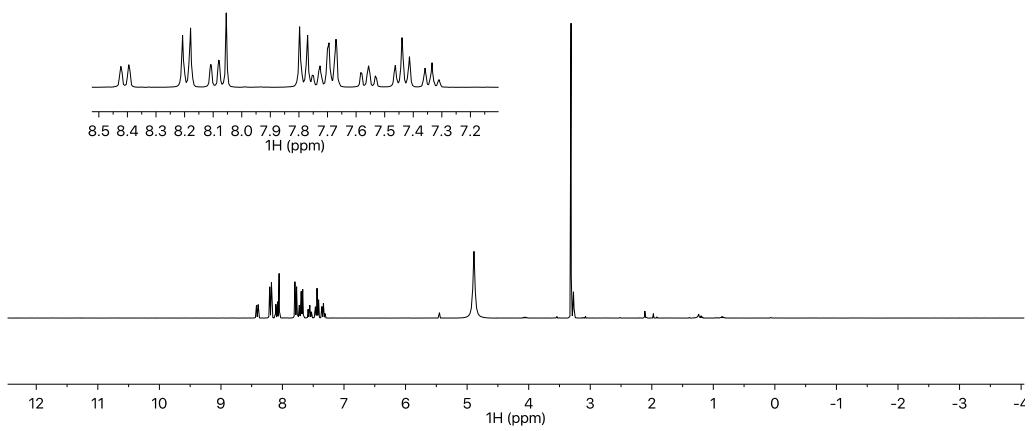
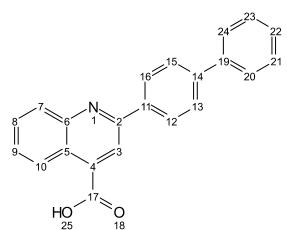
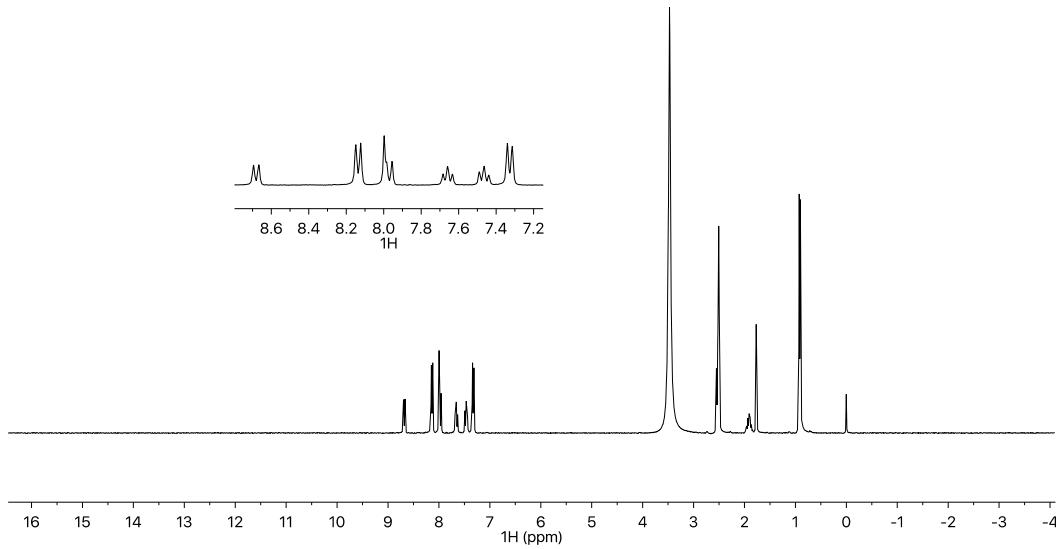
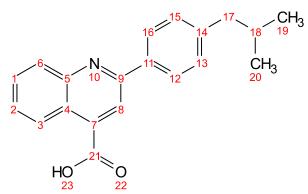
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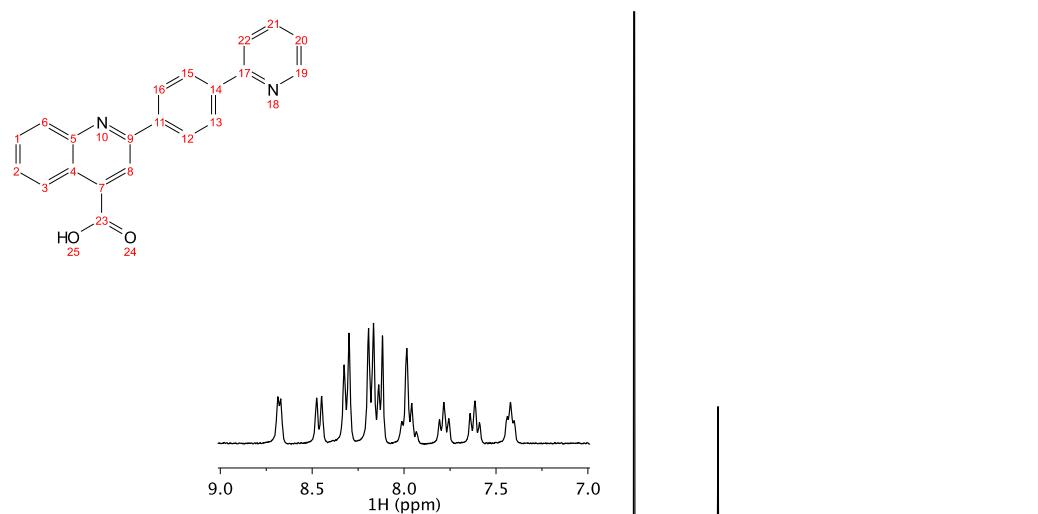
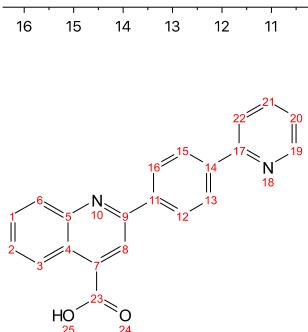
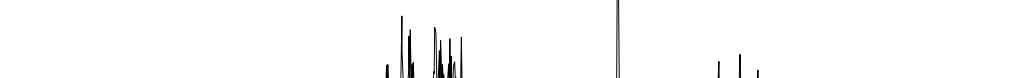
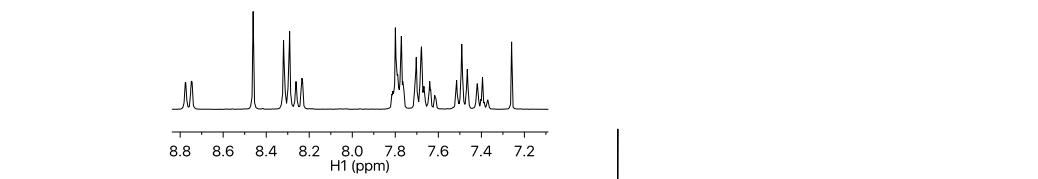
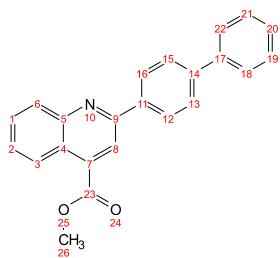


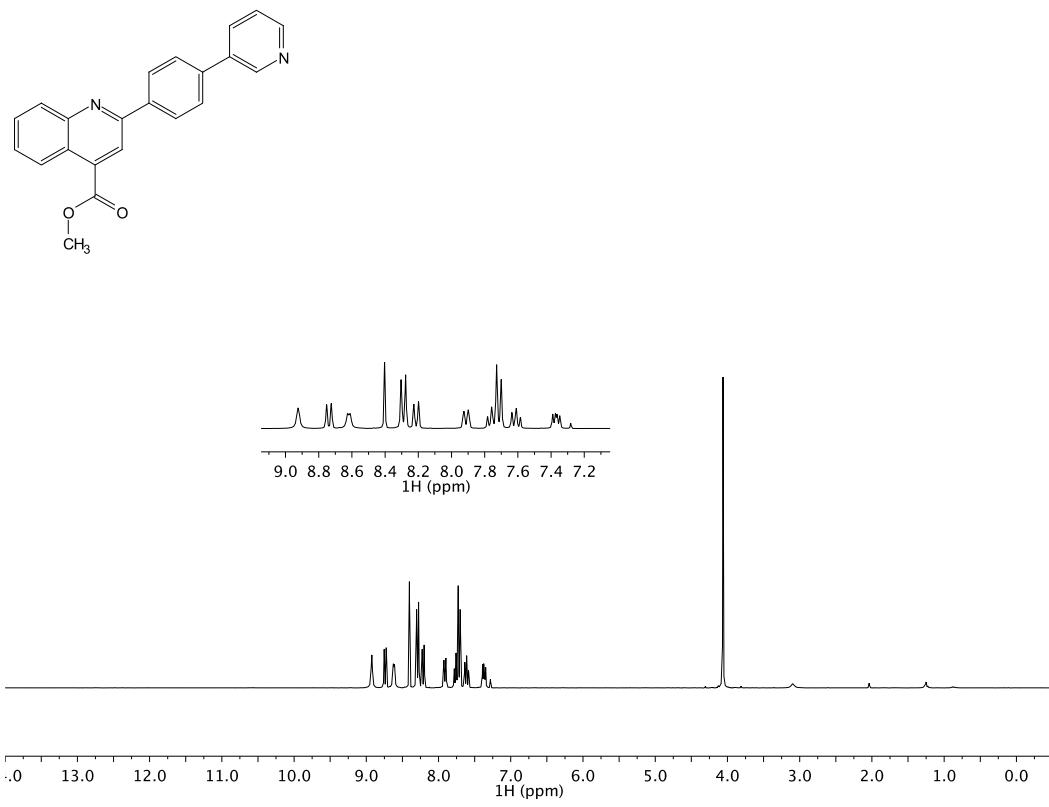
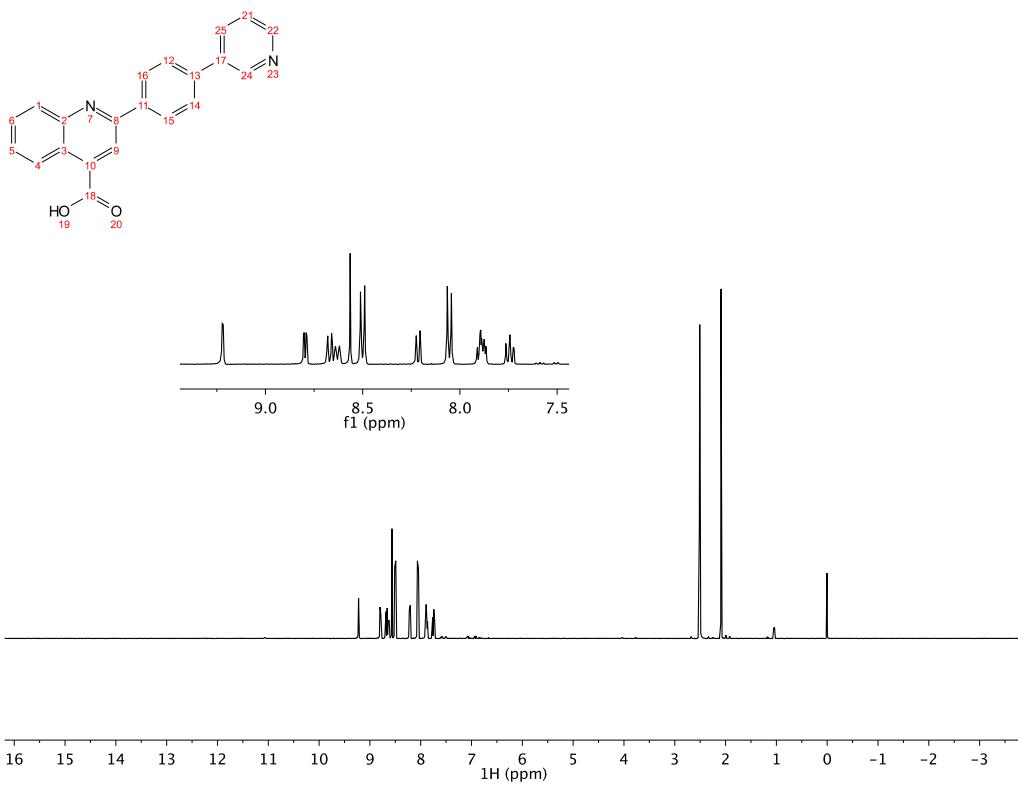


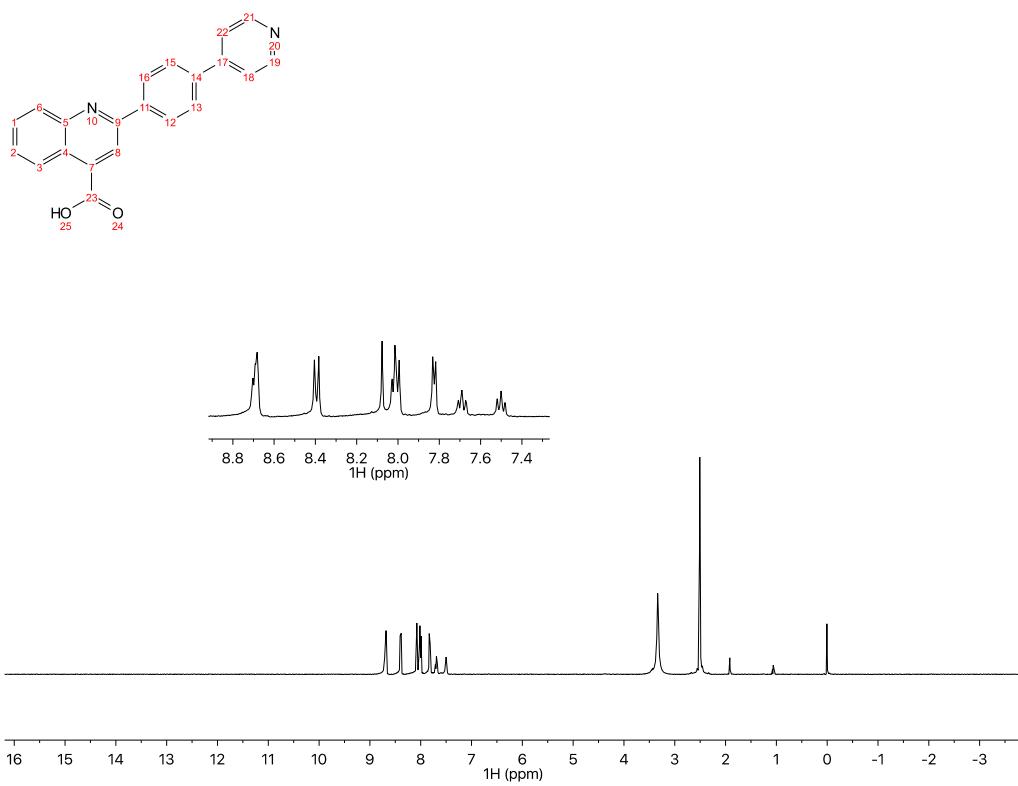


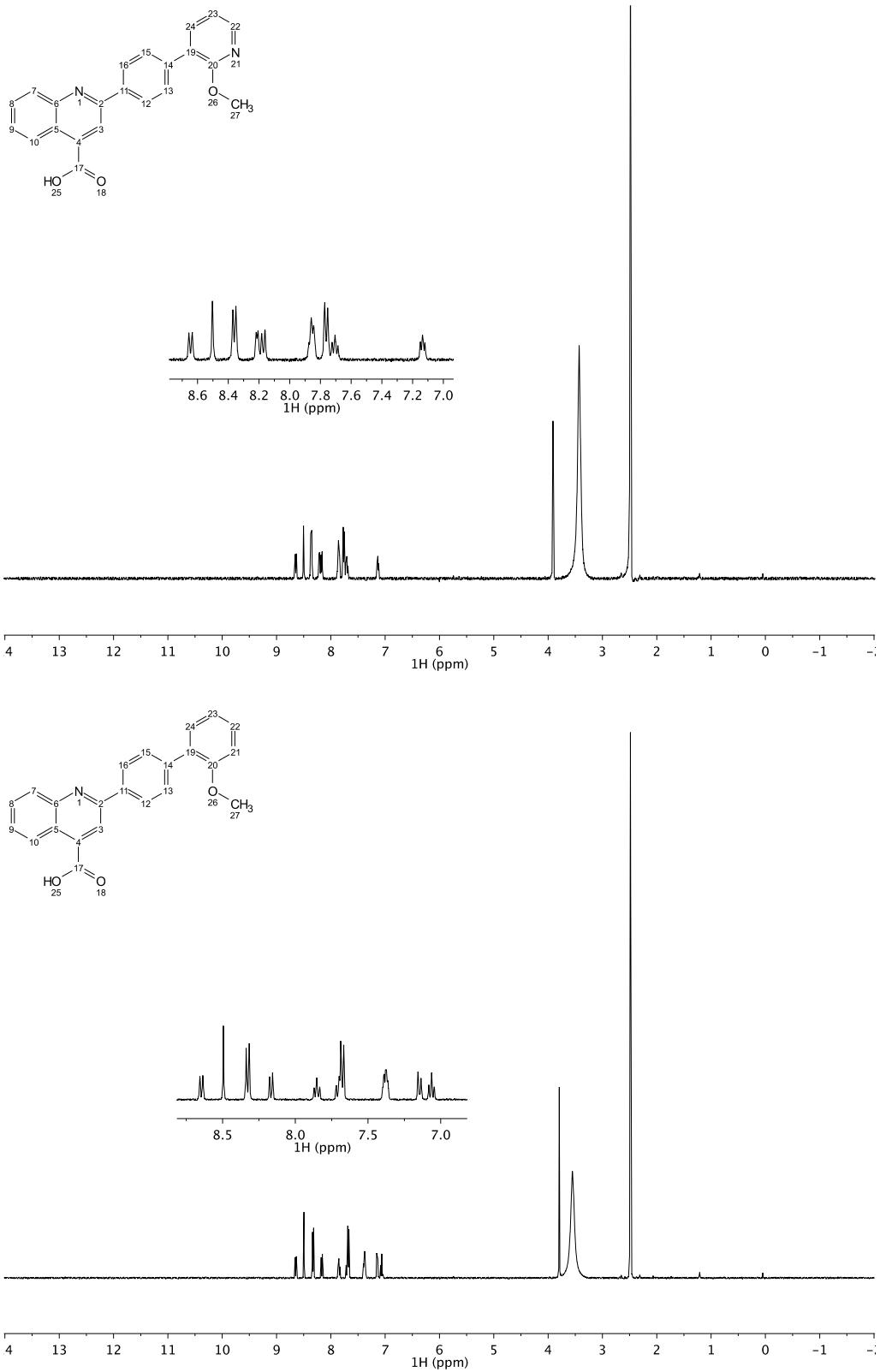


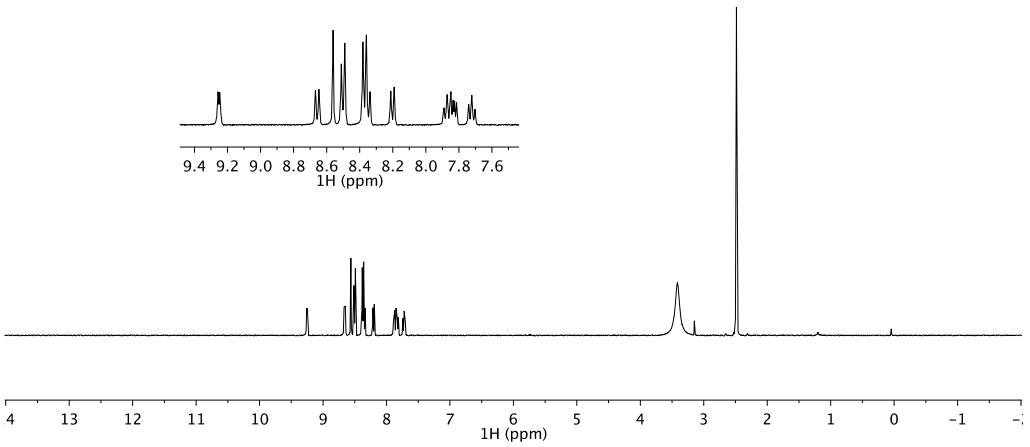
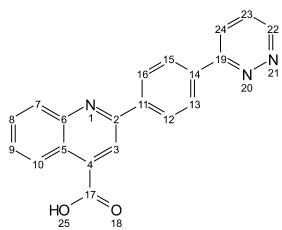
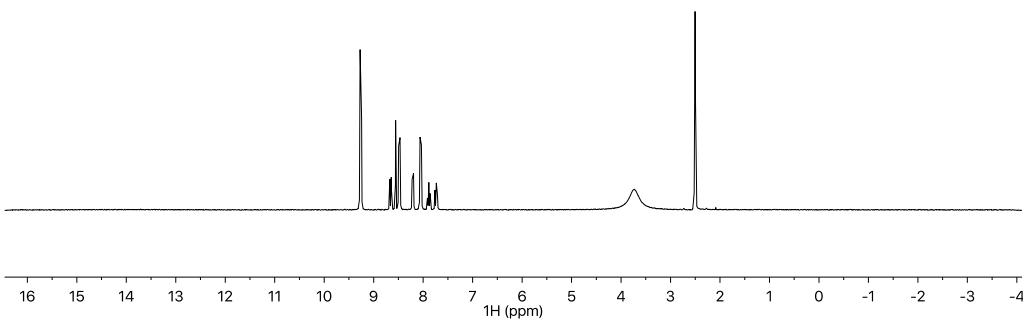
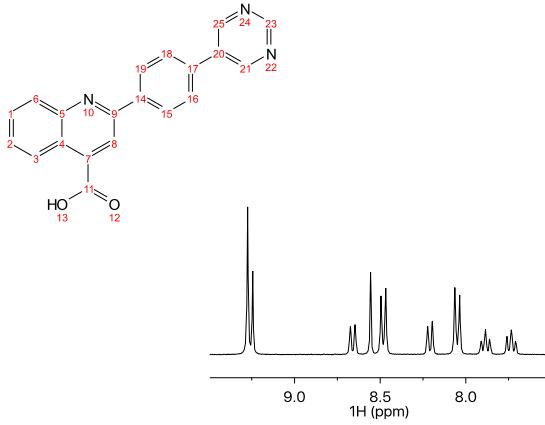


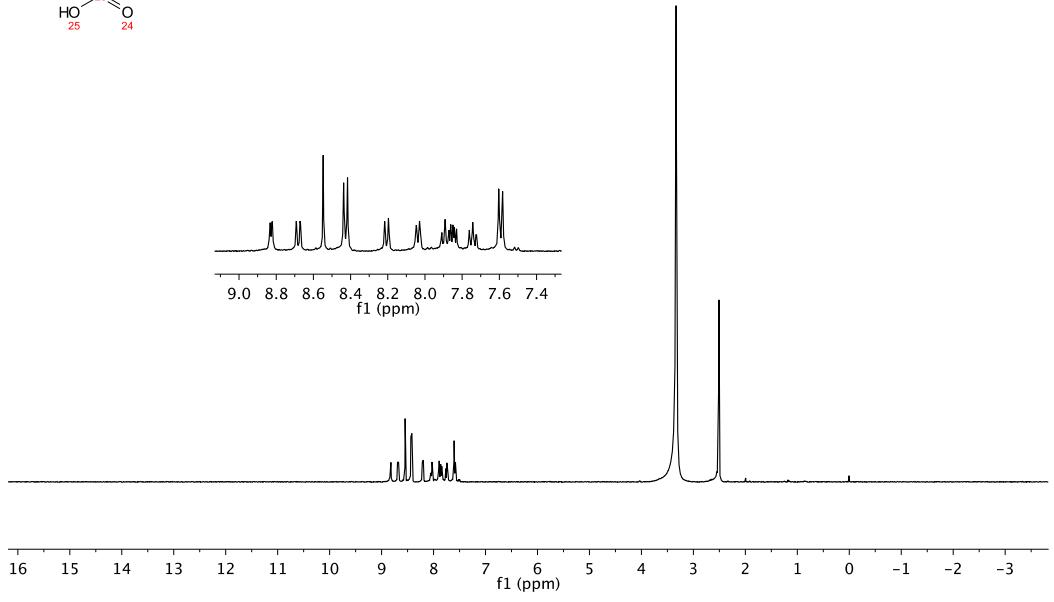
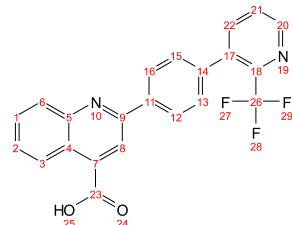
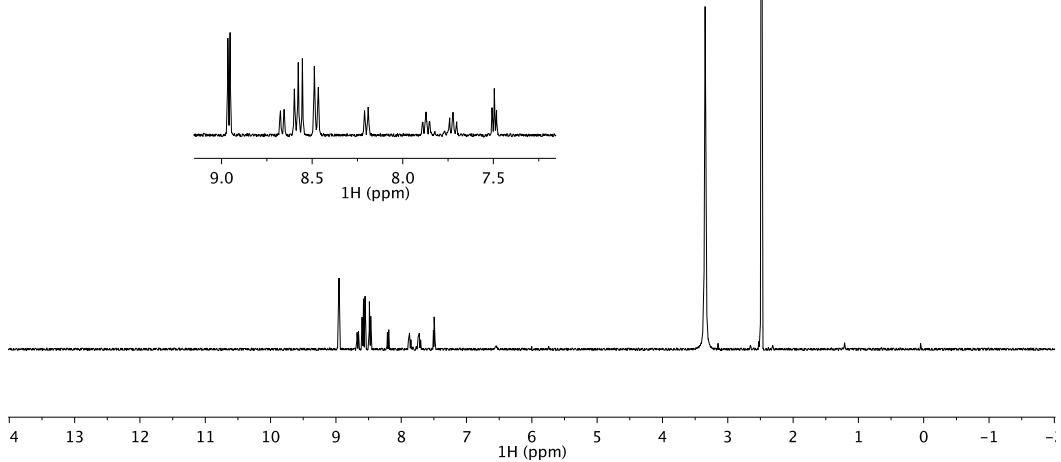
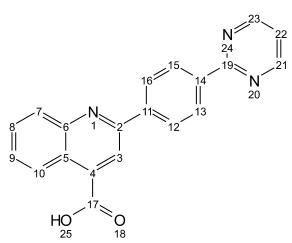


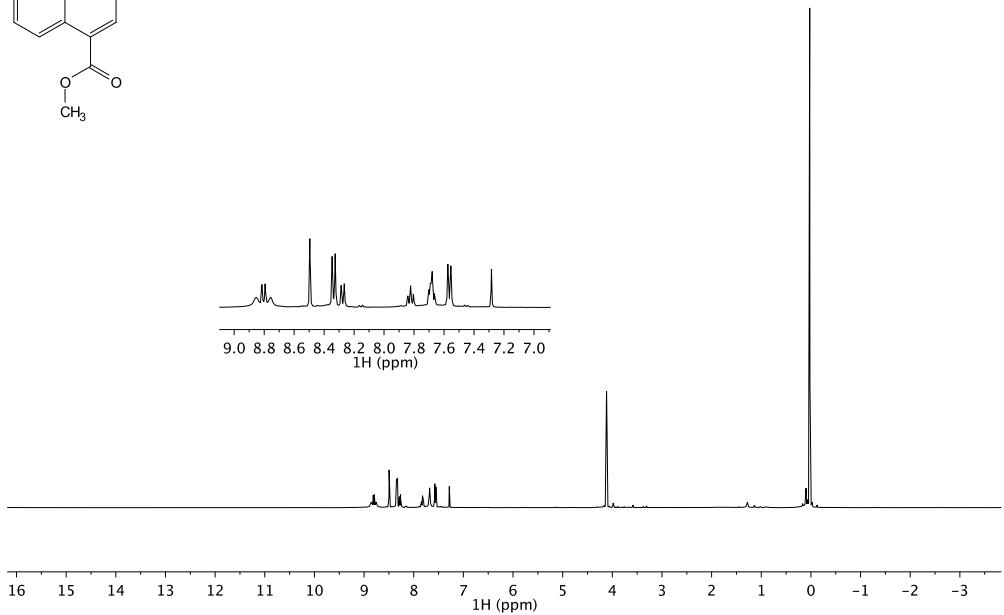
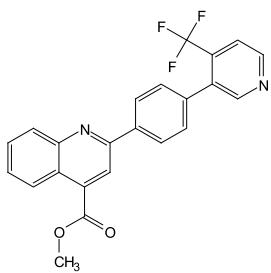
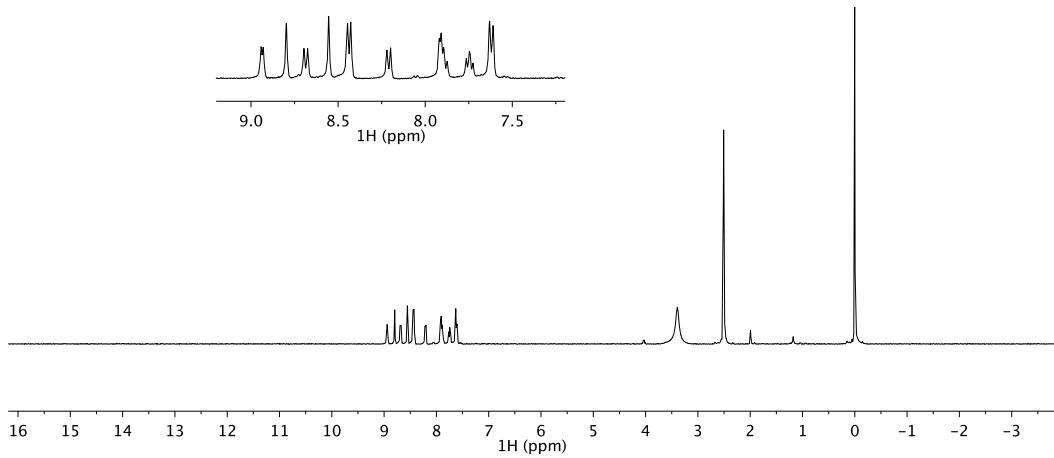
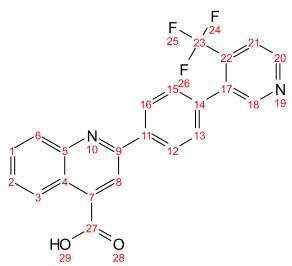


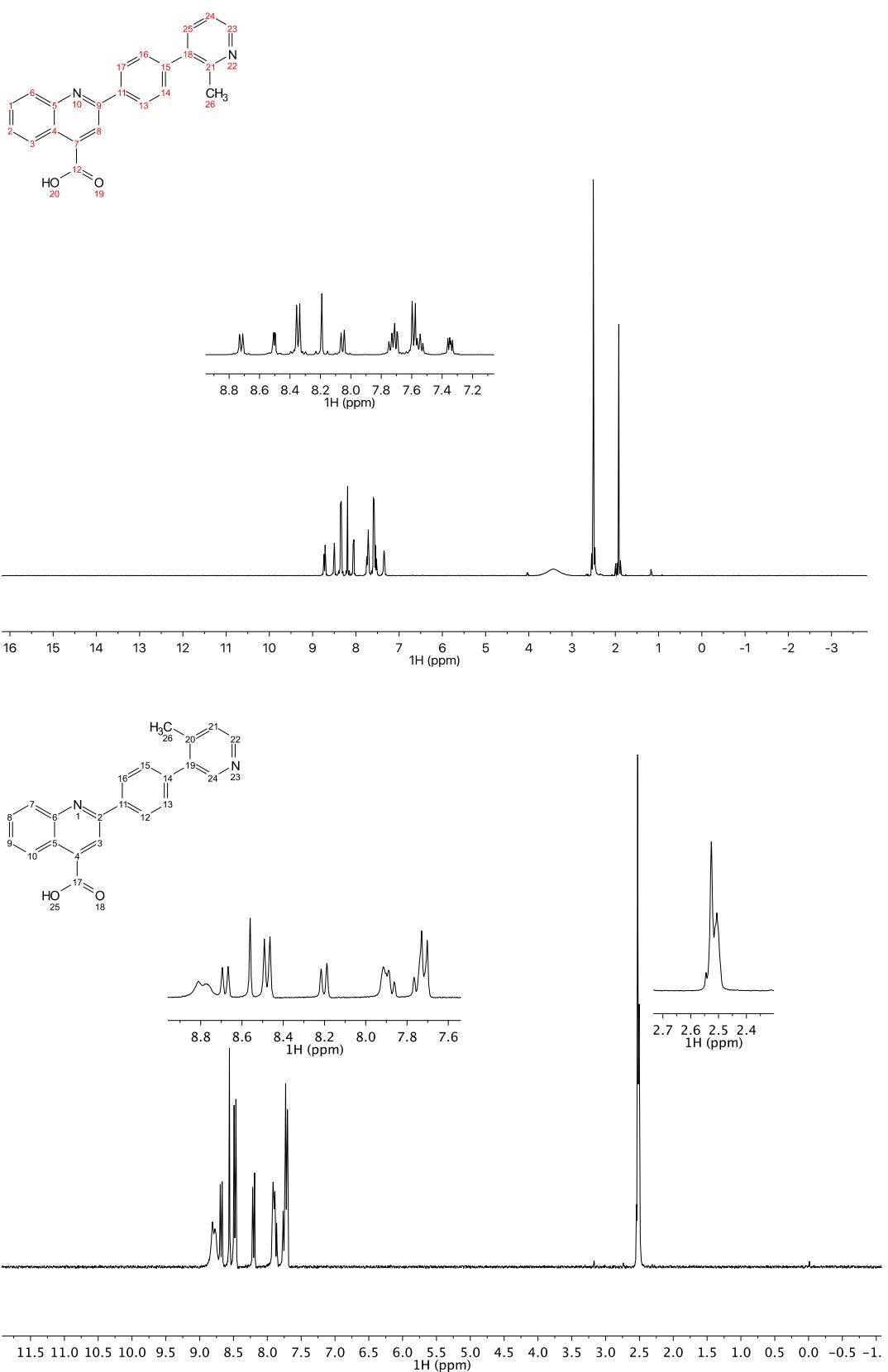


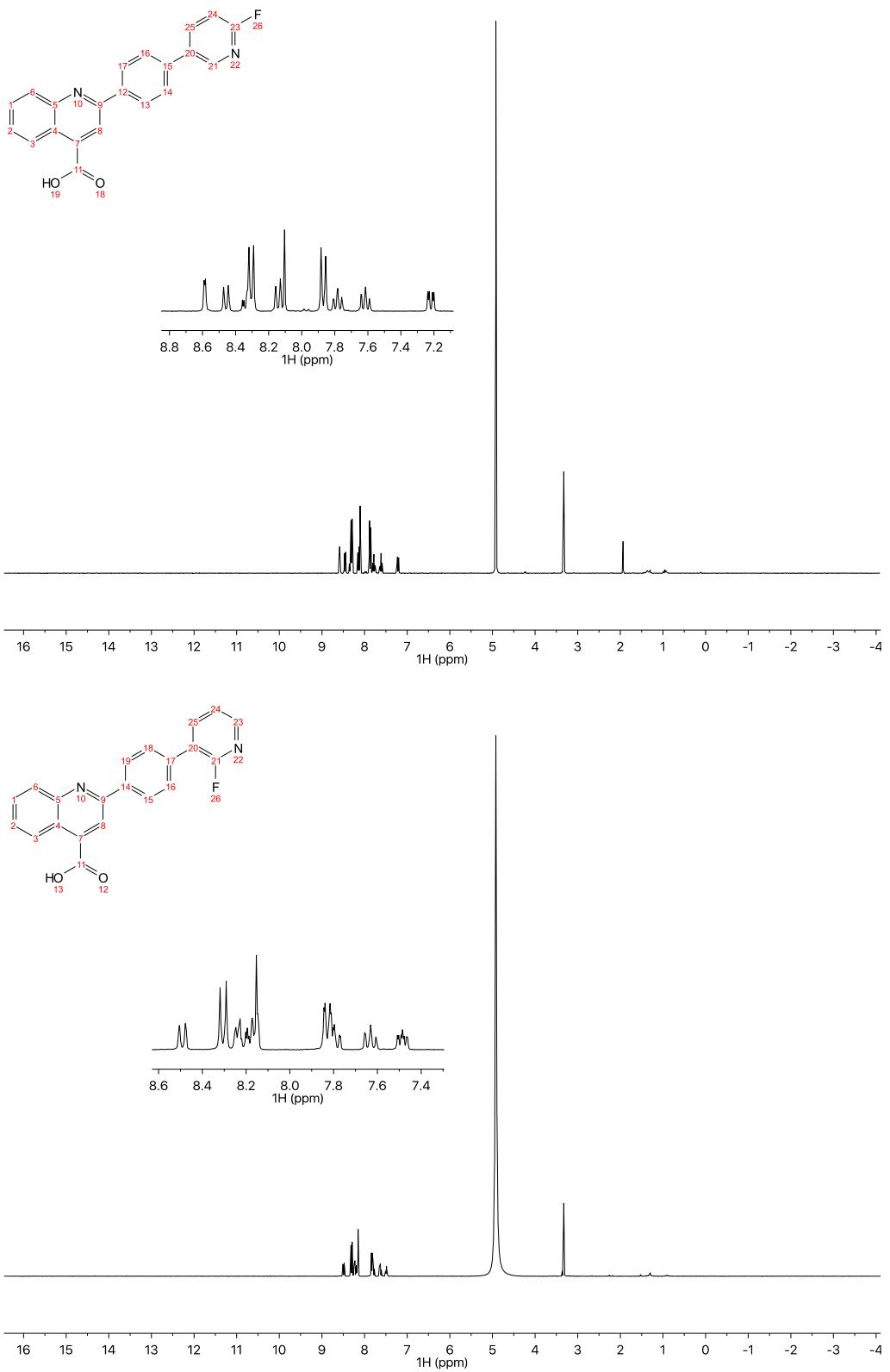


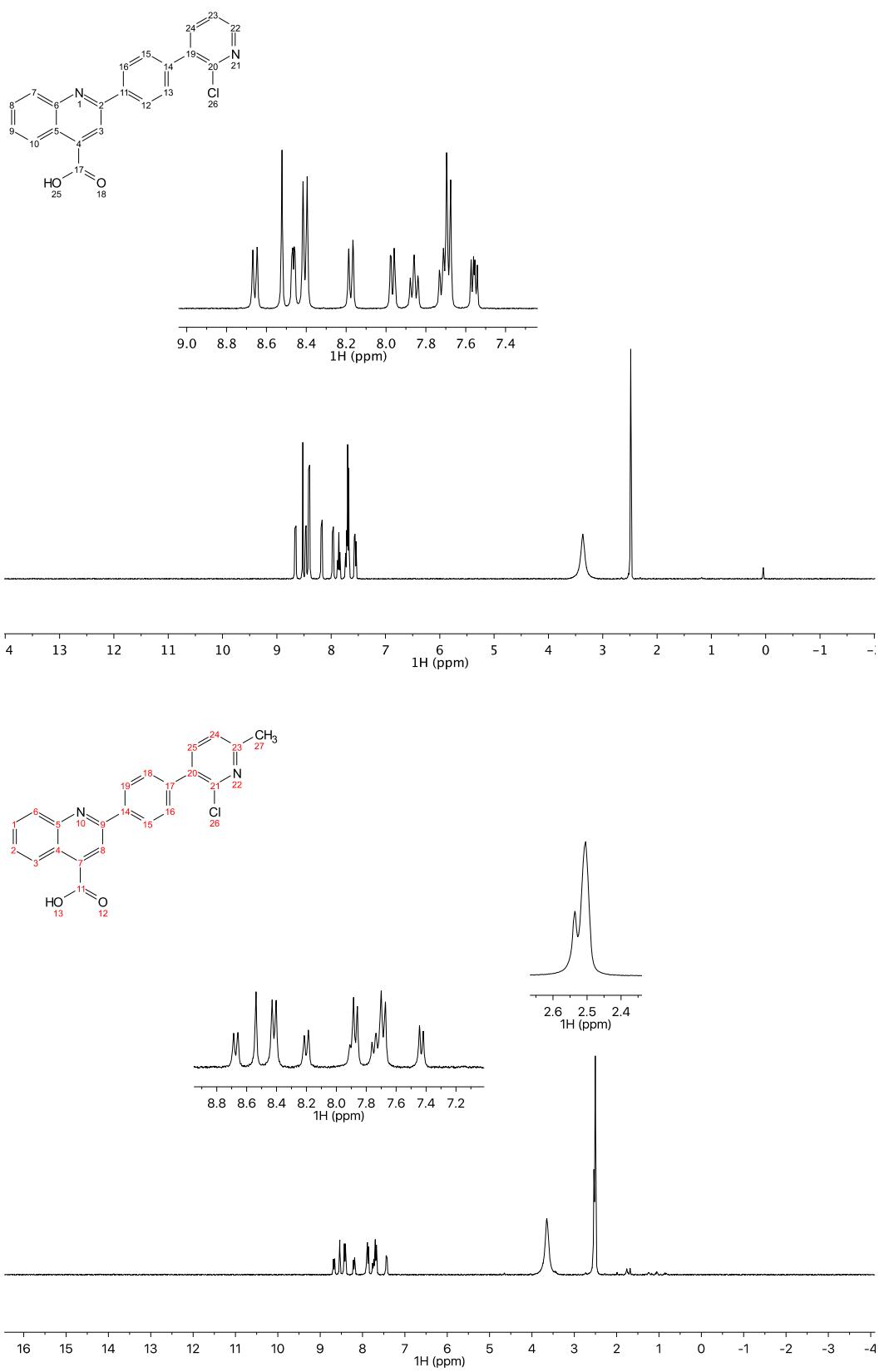


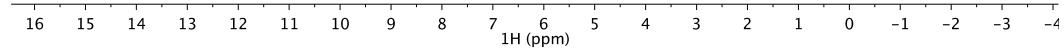
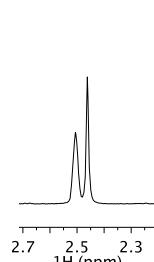
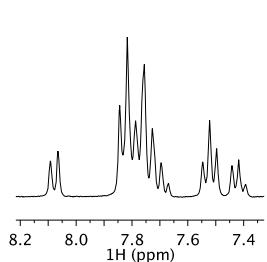
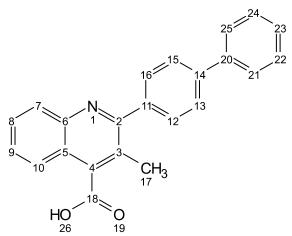
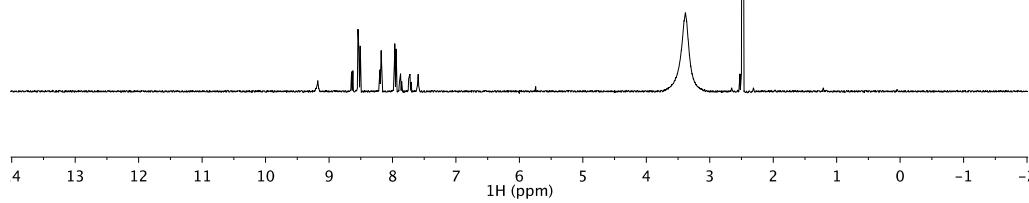
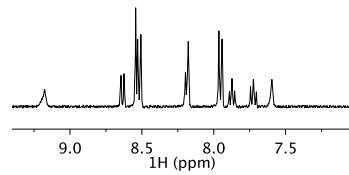
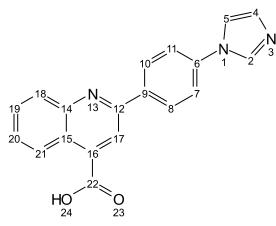


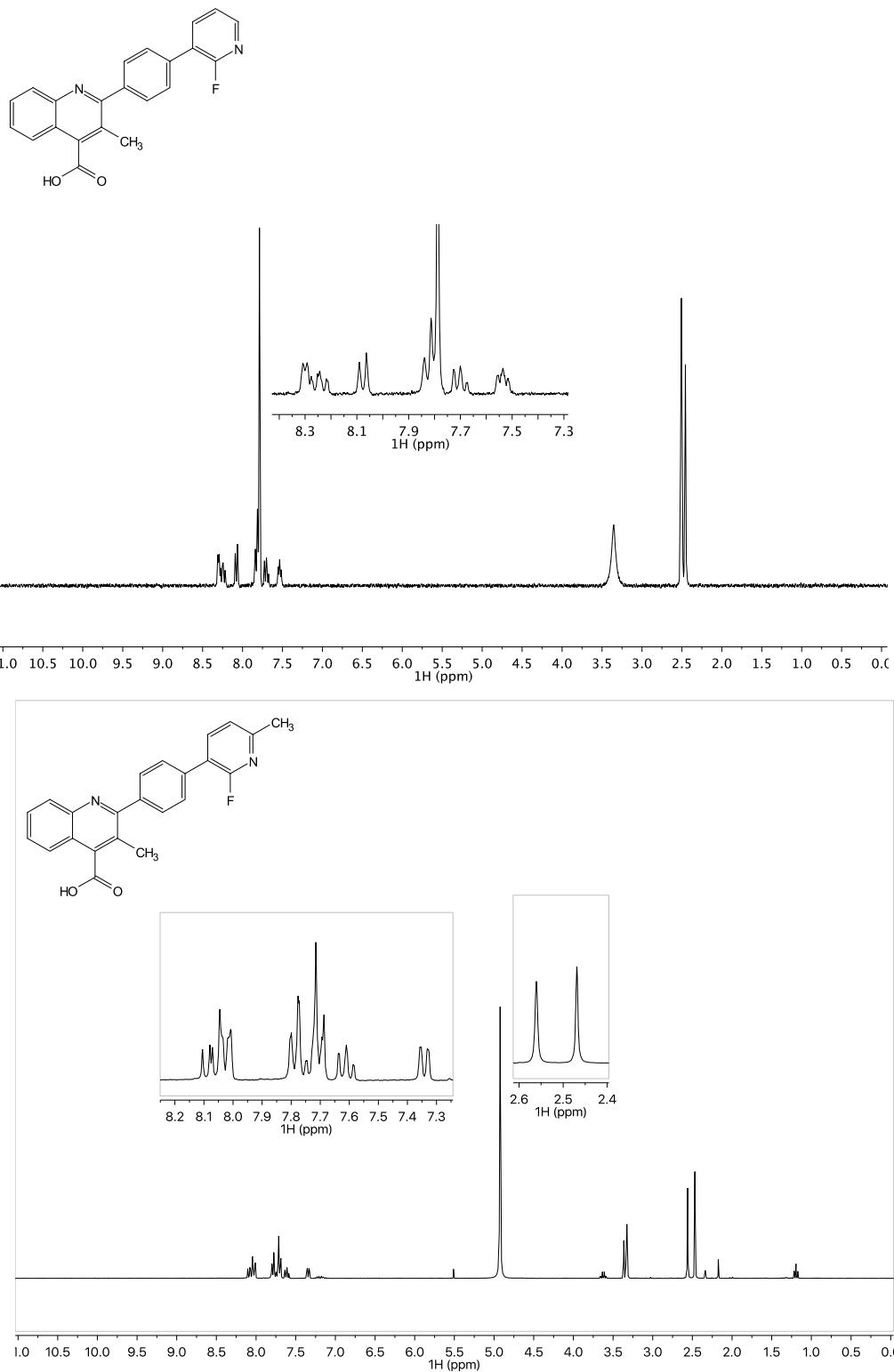


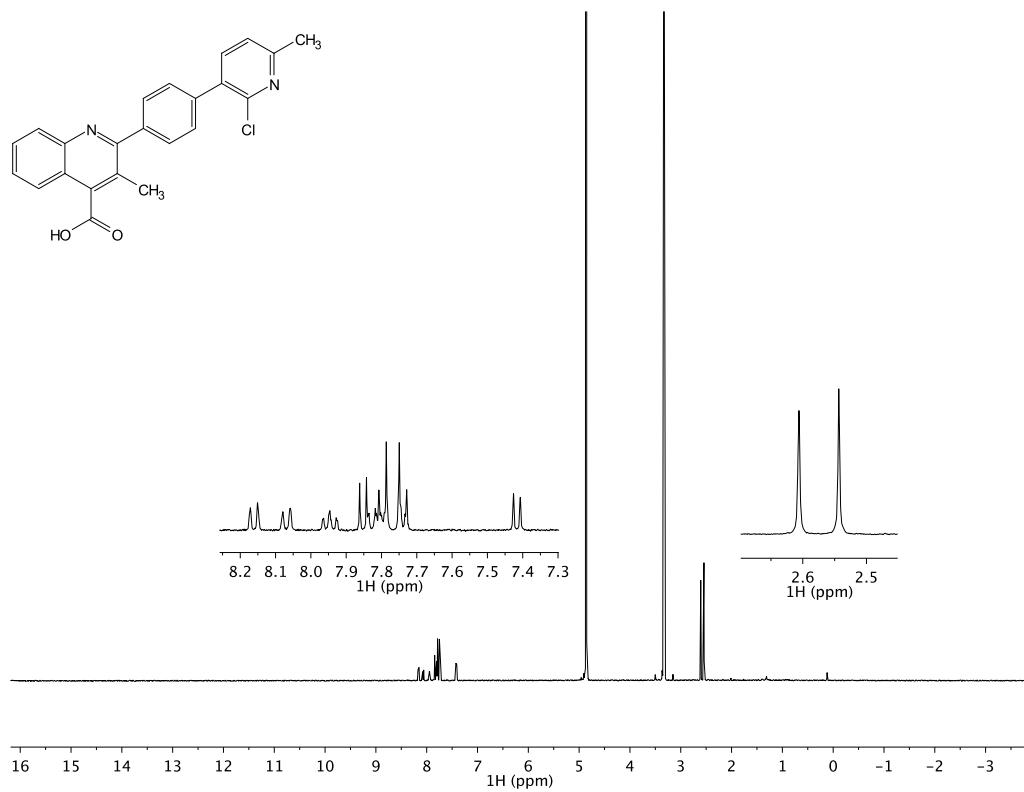
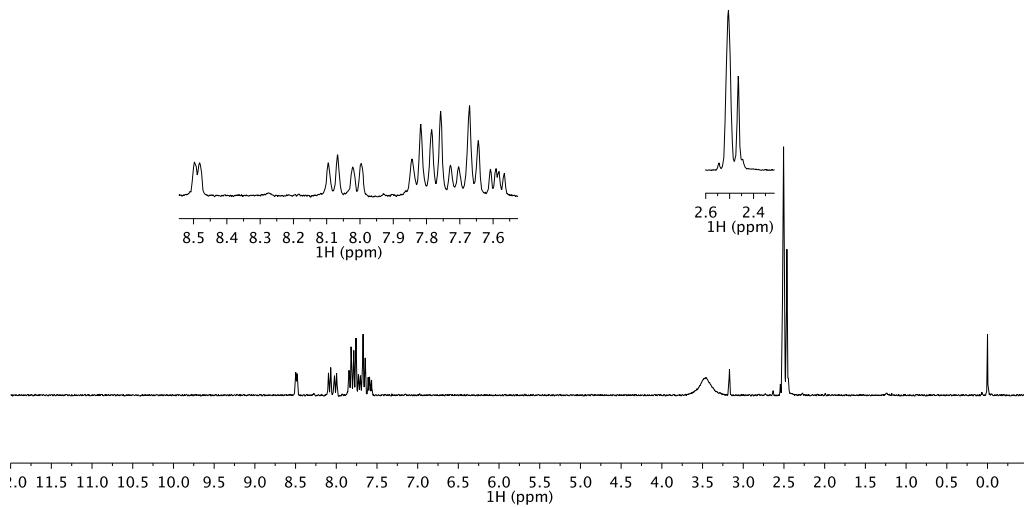
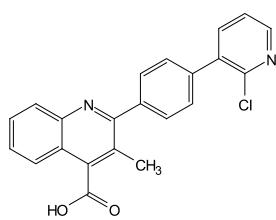


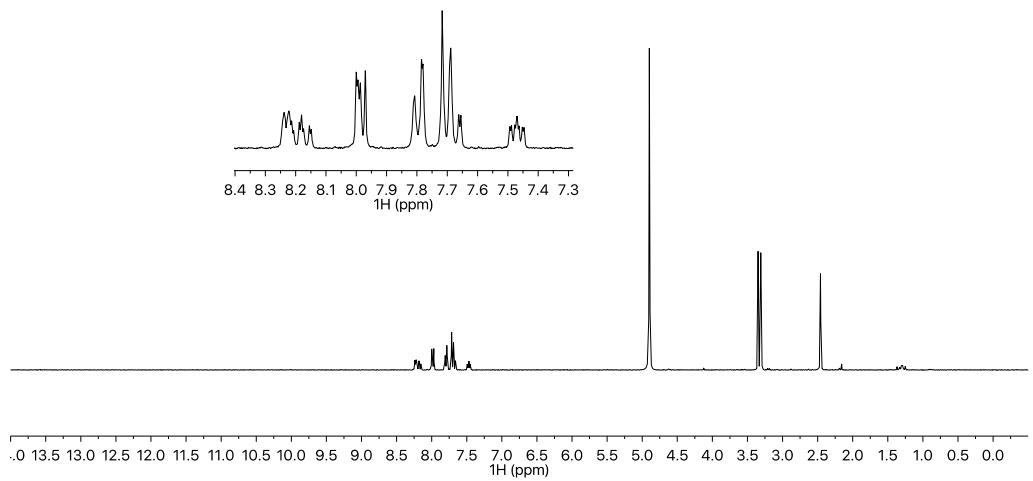
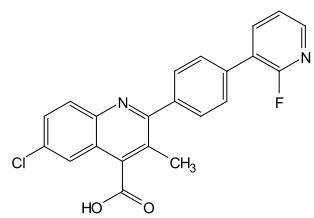
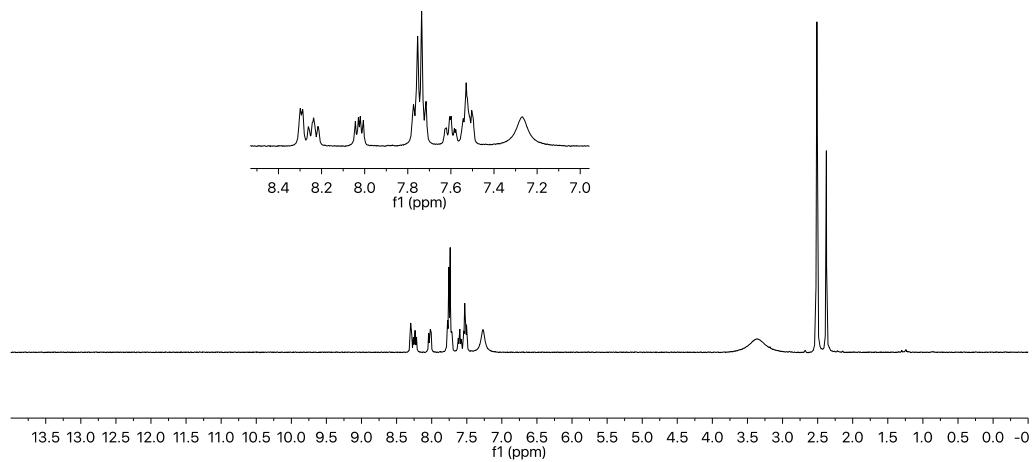
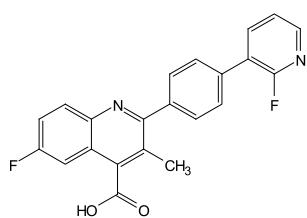


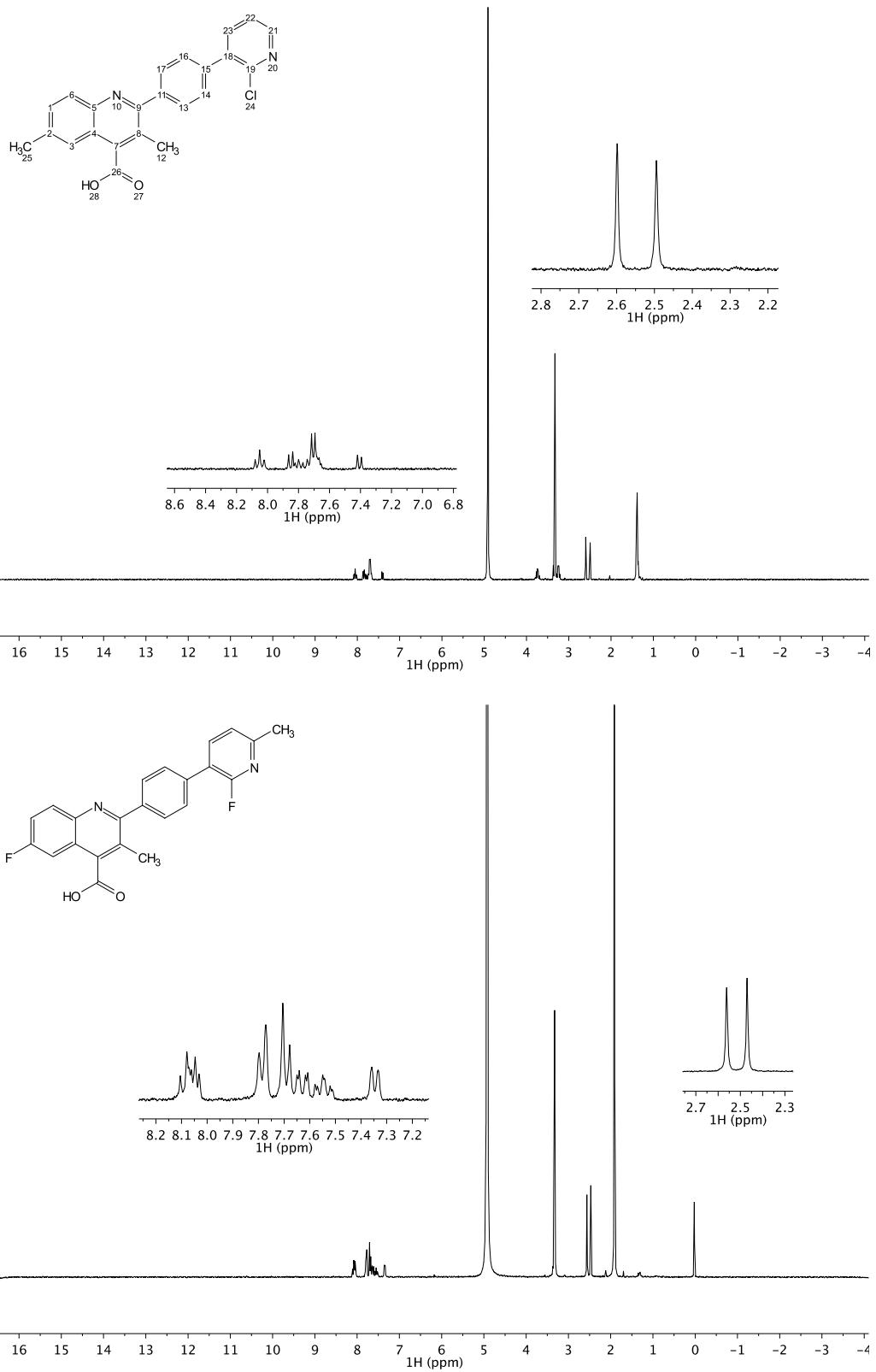


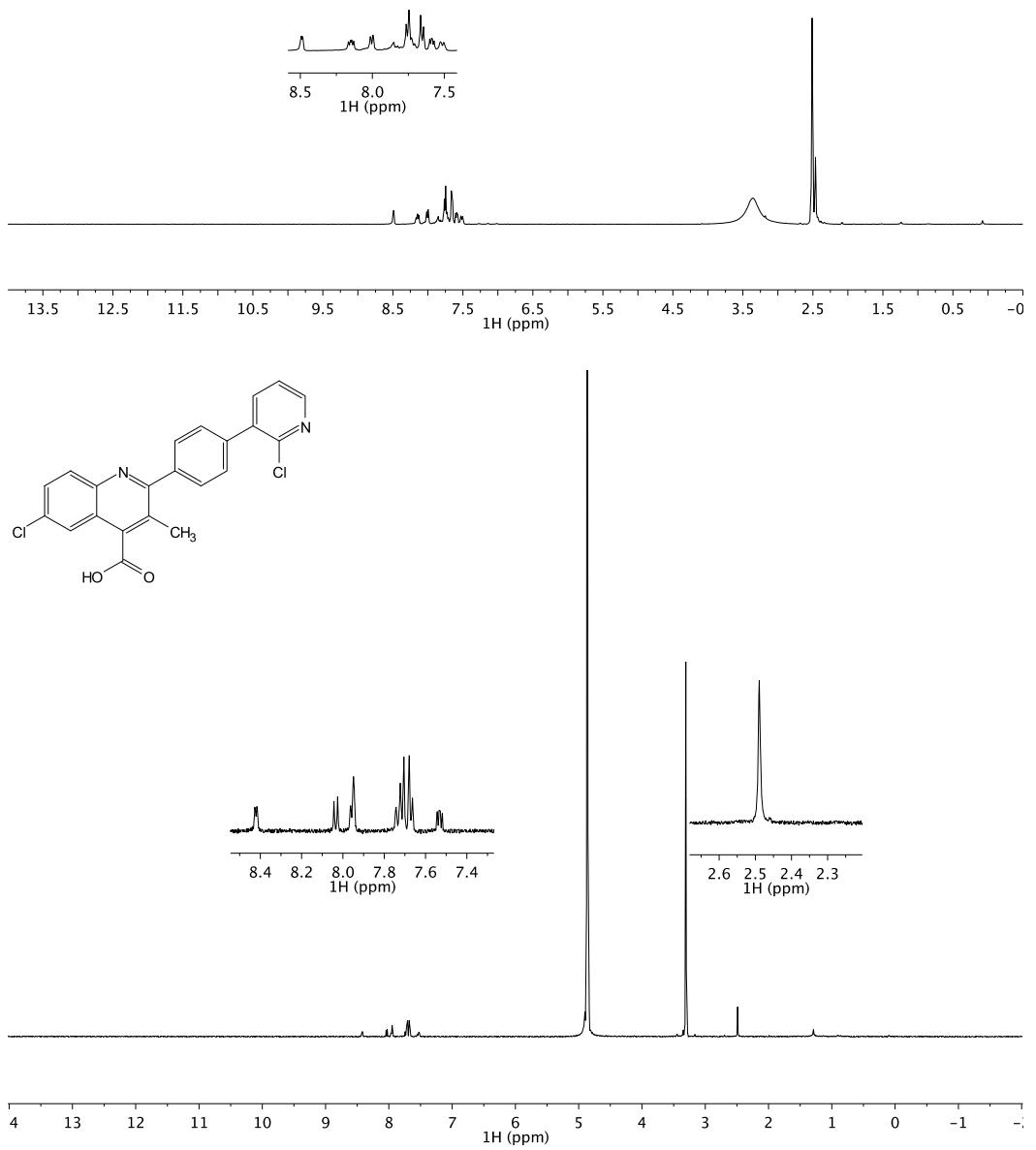
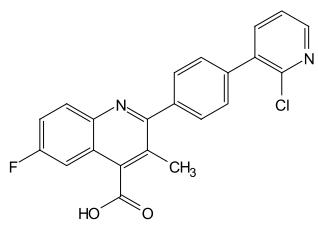


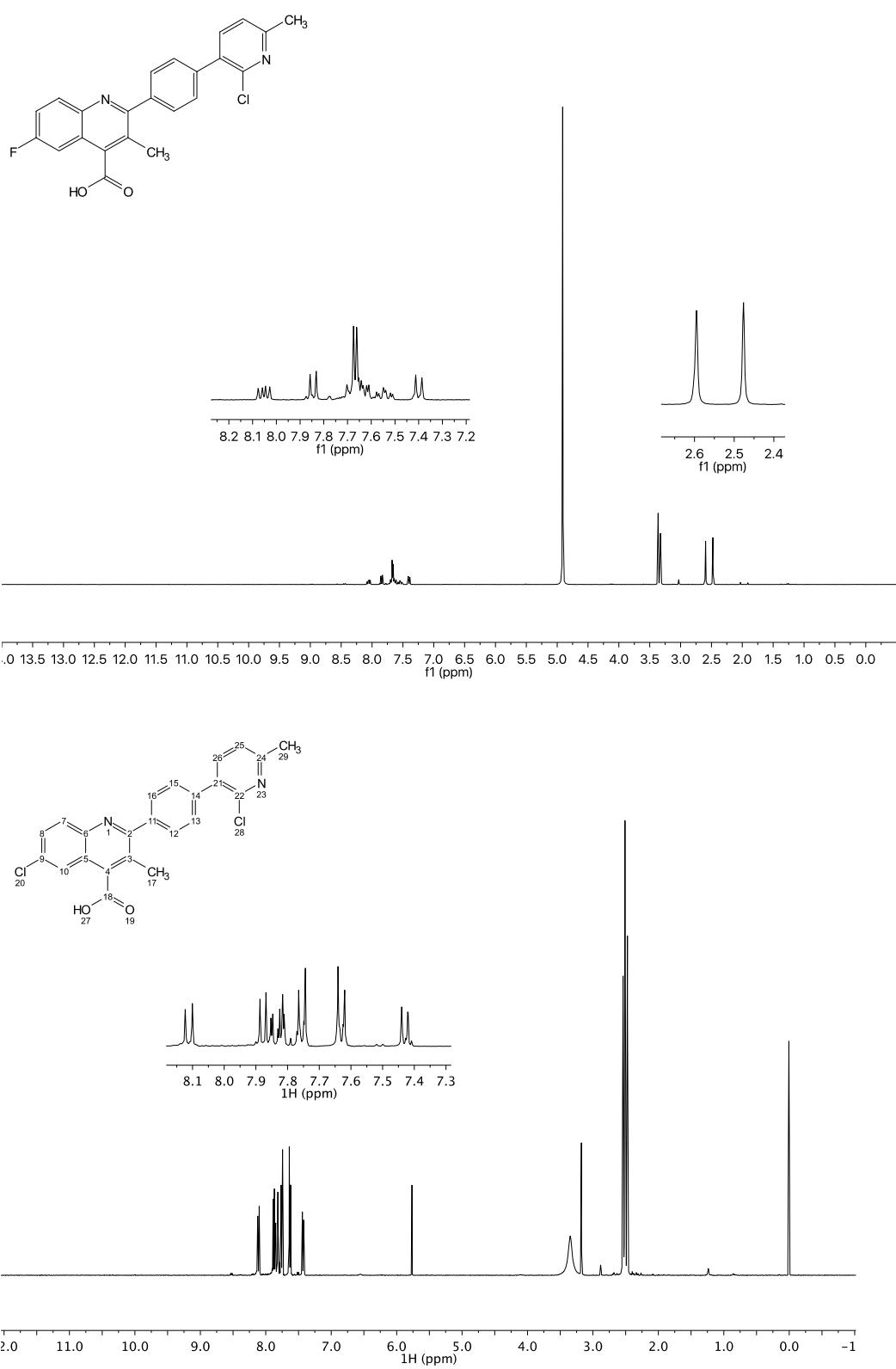


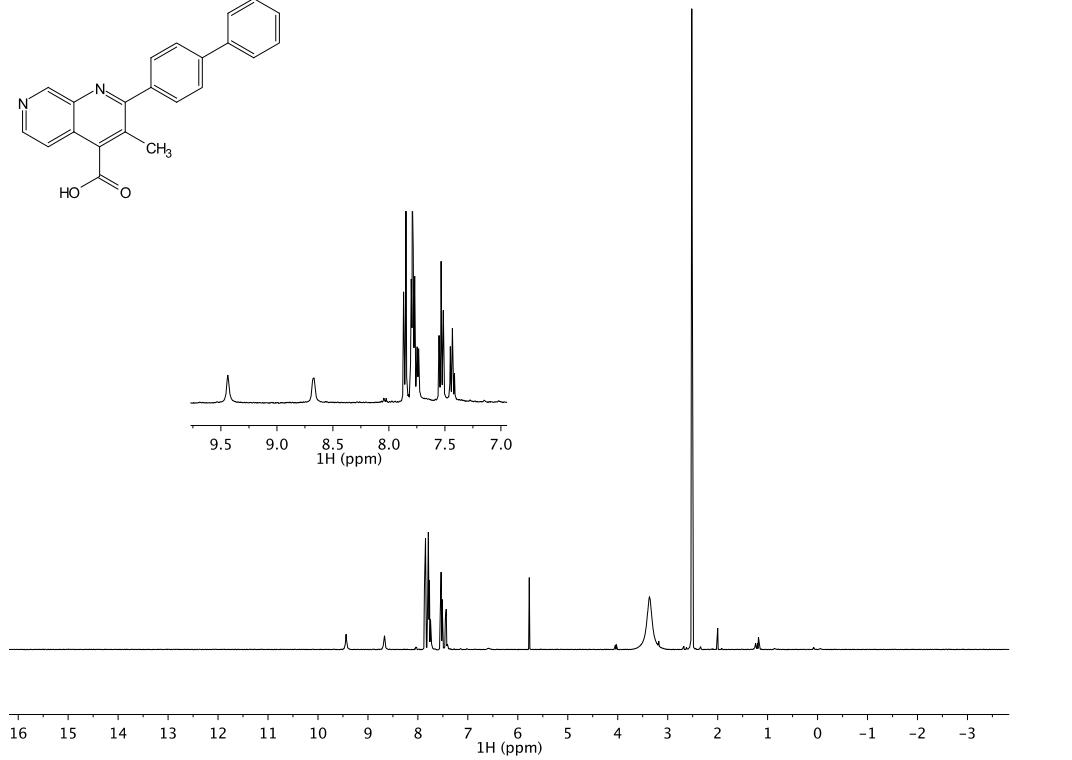
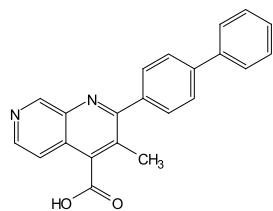
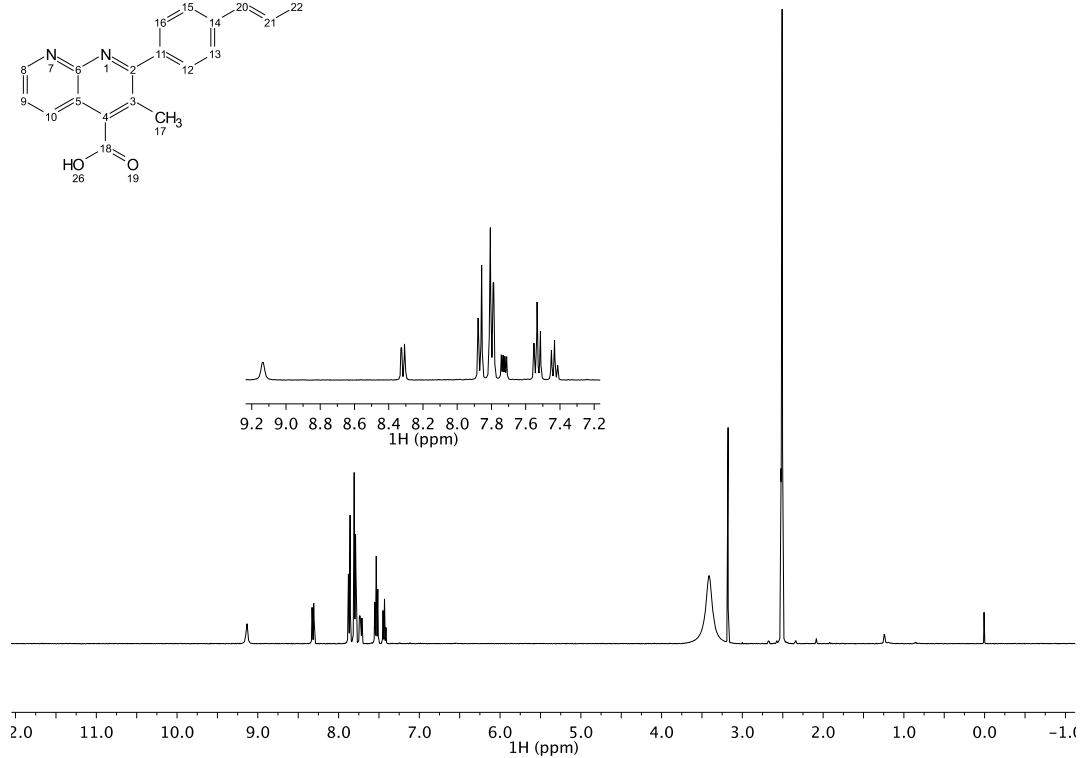
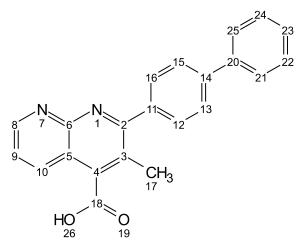


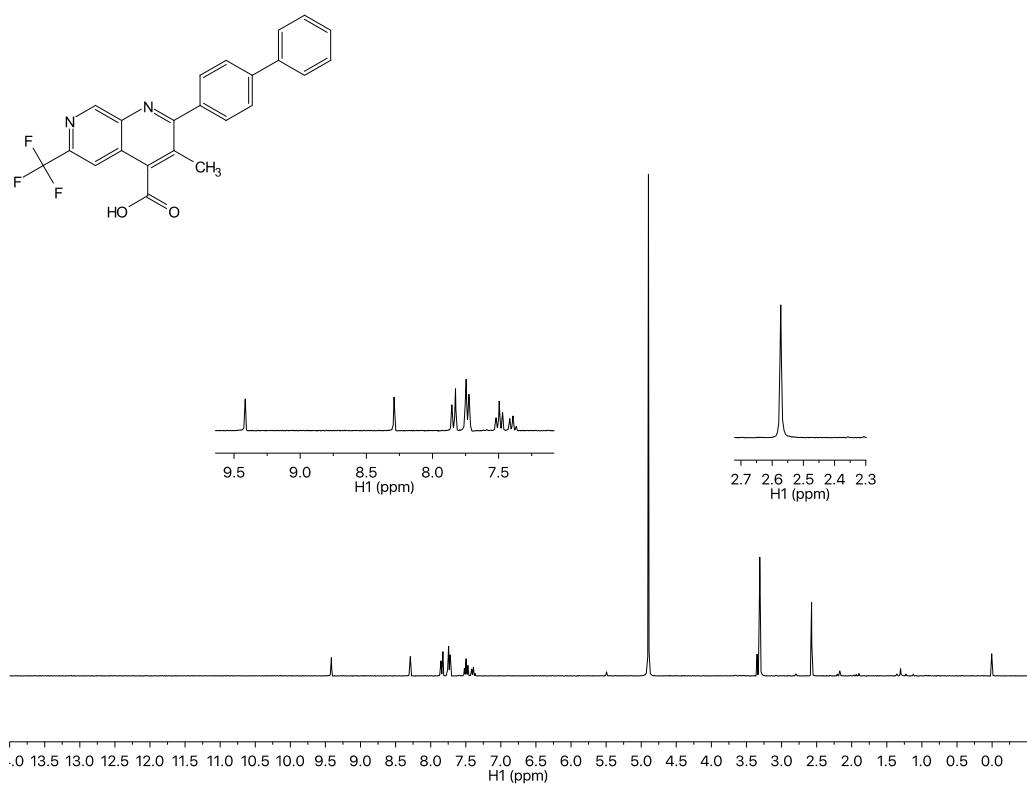
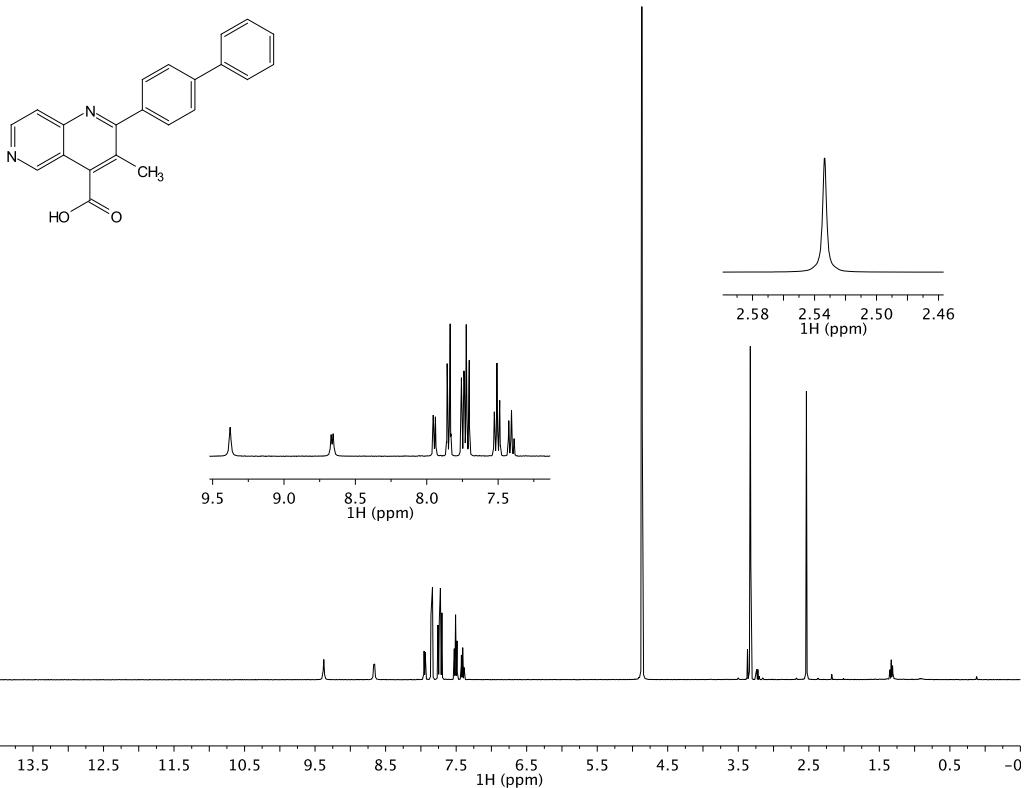


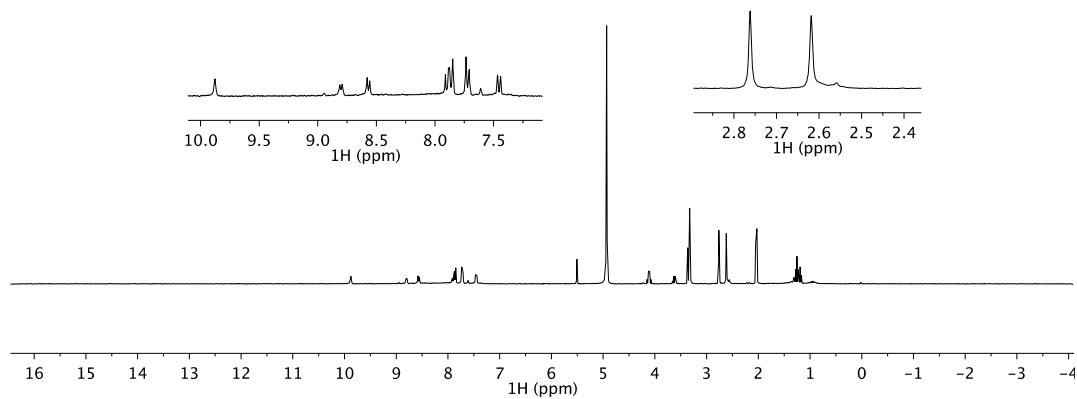
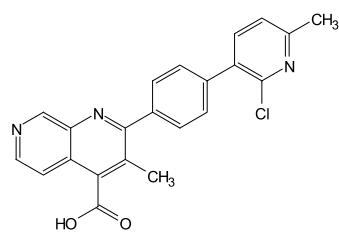
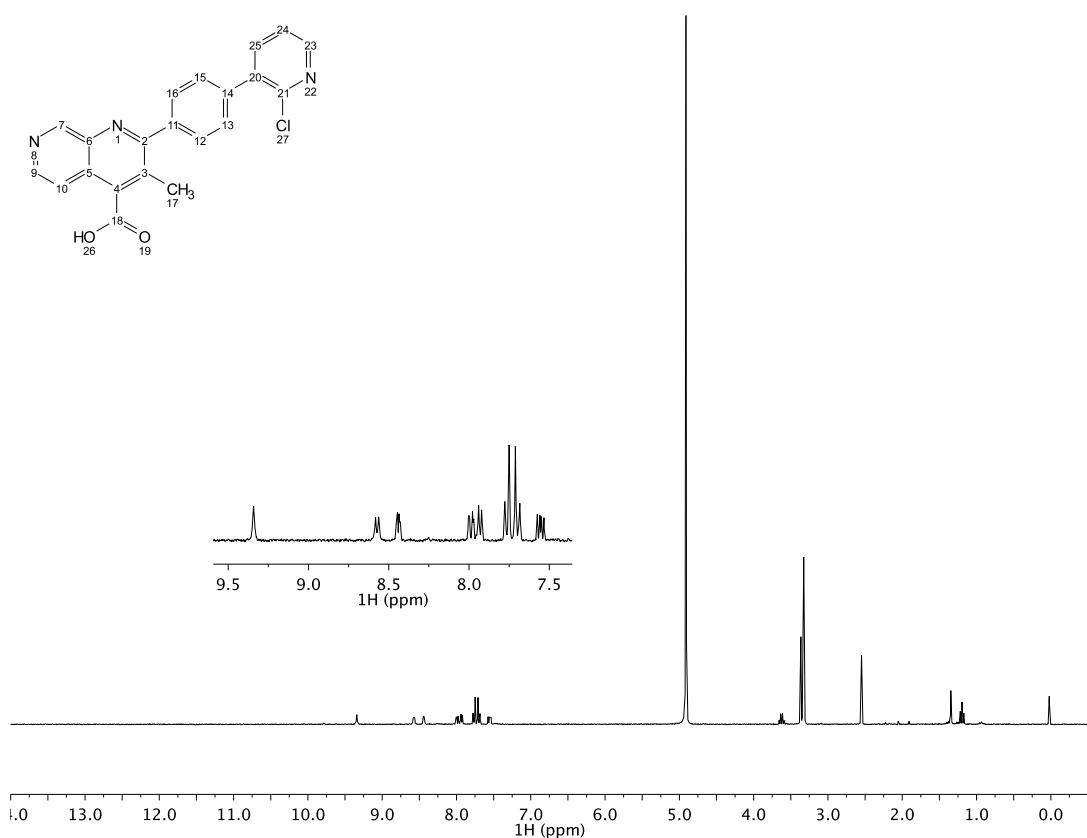
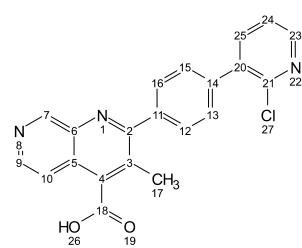


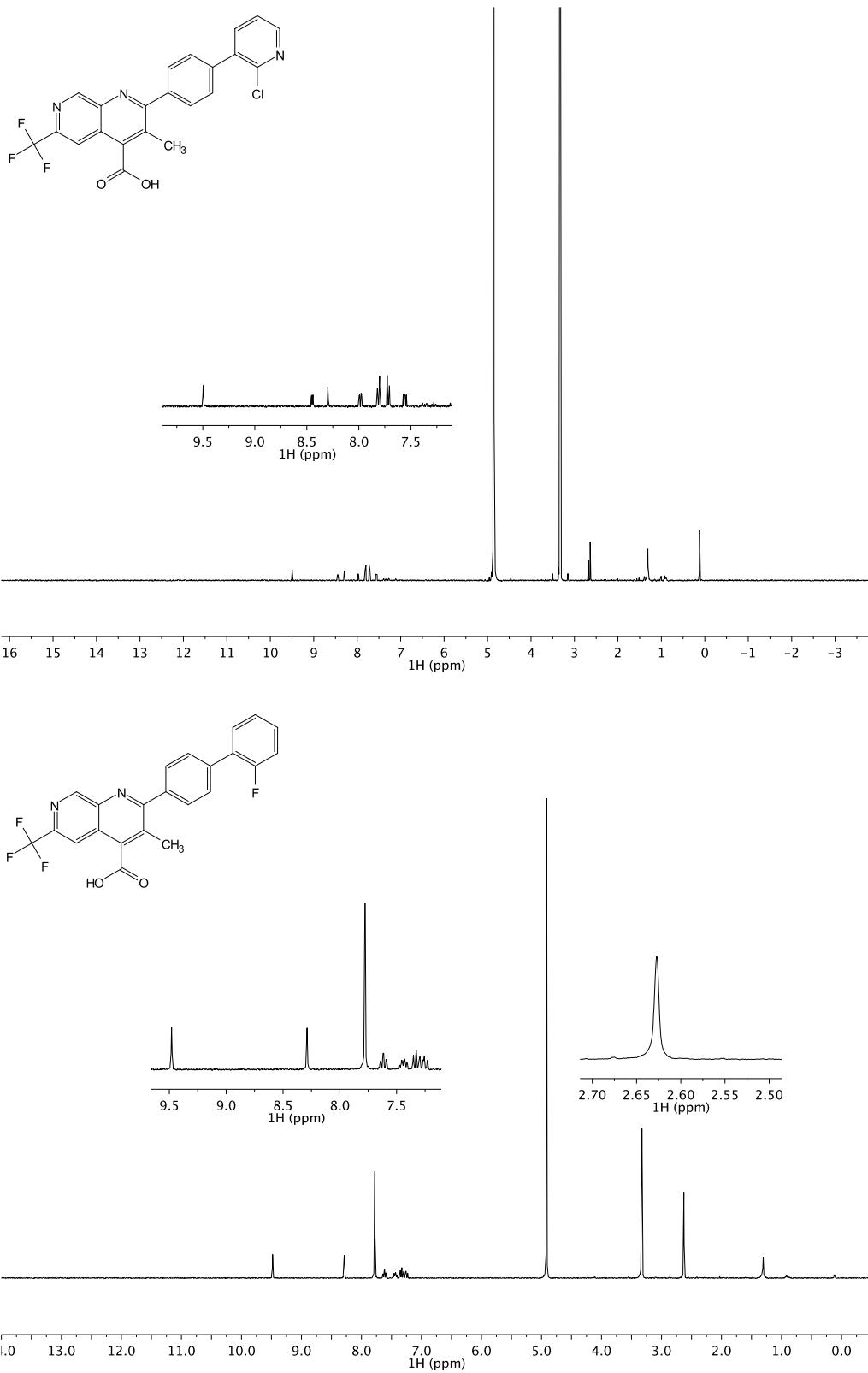












## **Supplemental References:**

1. The Oncomine Platform (Thermo Fisher, Ann Arbor, MI)  
[https://www.oncomine.com/resource/main.html - a:1780;cso:sizeSmallLarge;cv:detail;d:1197;dso:geneOverex;dt:predefinedClass;ec:\[2\];et:over;f:2507079;g:1723;gt:boxplot;p:200001263;pg:1;pvf:10222,35244,35245,35246,35247,35248,35249,35250;scr:summary;ss:analysis;v:18](https://www.oncomine.com/resource/main.html - a:1780;cso:sizeSmallLarge;cv:detail;d:1197;dso:geneOverex;dt:predefinedClass;ec:[2];et:over;f:2507079;g:1723;gt:boxplot;p:200001263;pg:1;pvf:10222,35244,35245,35246,35247,35248,35249,35250;scr:summary;ss:analysis;v:18)
2. Rhodes, D. R.; Yu, J.; Shanker, K.; Deshpande, N.; Varambally, R.; Ghosh, D.; Barrette, T.; Pandey, A.; Chinnaiyan, A. M. ONCOMINE: a cancer microarray database and integrated data-mining platform. *Neoplasia* **2004**, 6, 1-6.
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