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Dynamic clinical and biomarker data for mortality risk prediction in COVID-19

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Dynamic clinical and biomarker data for mortality risk prediction in COVID-19

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39 40 41 42 **Contributor and Guarantor Information** 43

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45 All authors were involved in the concept and design of the study which was led by HP and AV. HP,
46
47 CH, MW acquired the data, and CB, LB, AV, HP, CH analysed and interpreted the data. CB, DB, SA,
48
49 OP, JG, AV, CH, HP drafted the manuscript. All authors were involved in revising the manuscript and
50
51 contributed to the final draft. The corresponding author attests that all listed authors meet authorship
52
53 criteria and that no others meeting the criteria have been omitted. HP acts as guarantor for the study.
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Competing Interests Declaration

Swedish Orphan Biovitrum have provided investigational medicinal product for public-funded, peer-reviewed trials on which AK, AV, JG, HP, and SH are coinvestigators. The other authors declare no competing interests.

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Abstract

Objectives

Being able to predict which COVID-19 patients are going to deteriorate is important to help identify patients for clinical and research practice. Clinical prediction models play a critical role in this process, but current models are of limited value because they are typically restricted to baseline predictors and don't always use state of the art methods. We sought to explore the benefits of incorporating dynamic changes in routinely measured biomarkers, non-linear effects and applying 'state of the art' statistical methods in the development of a prognostic model to predict death in hospitalised COVID-19 patients.

Design

Data were analysed from COVID-19 admissions to three hospital sites. Exploratory data analysis included a graphical approach to partial correlations. Dynamic biomarkers were considered up to five days following admission rather than depending solely on baseline or single time-point data. Marked departures from linear effects of covariates were identified by employing smoothing splines within a generalised additive modelling framework.

Setting

3 secondary and tertiary level centres in Greater Manchester

Participants

392 hospitalised patients with a diagnosis of COVID-19

Results

392 patients with a COVID-19 diagnosis were identified. Area under the receiver operating characteristic (ROC) curve increased from 0.73 using admission data alone to 0.75 when also considering results of baseline blood samples and to 0.83 when considering dynamic values of routinely collected markers. There was clear non-linearity in the association of age with patient outcome.

Conclusions

This study shows that clinical prediction models to predict death in hospitalised COVID-19 patients can be improved by taking into account both non-linear effects in covariates such as age and dynamic changes in values of biomarkers.

Strengths and Limitations.

- Using contemporary statistical methods, and by incorporating routinely available blood tests performed over the first 5 days of hospital admission we have shown the importance of using dynamic blood biomarker data to enhance patient-level prediction of COVID-19 progression.
- This approach should inform how future clinical prediction models are generated.
- We did not have sufficient data to construct definitive prediction models.
- More sophisticated exploitation of biomarker trajectories through, for example, approaches based on random effects models of biomarker evolution or ‘conditional on outcome’ models of biomarker evolution, could make clinical predictions models better still.

Introduction

Most patients with severe acute respiratory syndrome coronavirus 2 (SARS-Cov-2) experience mild symptoms. Some patients however experience significant symptoms requiring hospitalisation. The pandemic nature of the covid-19 outbreak has meant that hospital services and capacity can be overwhelmed.¹ A tool to predict which patients are likely to deteriorate or need intensive care would help clinicians, hospital managers and researchers make better decisions.

Several such models are reported for COVID-19 patients but have been criticised for risk of bias using the PROBAST (prediction model risk of bias assessment tool) criteria.² We have further concerns regarding the statistical tools used to develop models. First, current models typically only consider patient characteristics available at baseline and do not consider that COVID-19 patients' presentation and in hospital course is variable. Secondly, models routinely seek only linear effects of potential predictors on the outcome of interest although these are not always clinically plausible.

We sought here to explore the benefits of incorporating dynamic changes in routinely measured biomarkers, non-linear effects and applying 'state of the art' statistical methods in the development of a prognostic model to predict death in hospitalised COVID-19 patients.

Methods

Study Population

Admissions with confirmed COVID-19 (according to World Health Organisation guidance) at three hospitals in the Northern Care Alliance (Greater Manchester, UK) between 11th March and 17th April 2020 with a minimum of a three week follow-up were studied.³

Data Collection

Necessary approvals were obtained from the local Research and Innovation department.

Research nurses abstracted data from the electronic patient records based on the International Severe Acute Respiratory and emerging Infection Consortium (ISARIC) data collection tool but modified for use with this study.⁴ The ISARIC study data were supplemented from electronic patient records with results of blood analyses performed as part of routine clinical care. The date of diagnosis was considered Day 1.

Data analysis

All data were subjected to range checks and validated for internal consistency and missing items then anonymised prior to transfer.

Selection of biomarkers

The initial list of potential markers was determined through review of the literature and availability within routinely collected data. Candidate variables were further screened using a graphical representation of the partial correlation structure stratified by survival status.⁵

Routine bloods were typically analysed on alternate days. The assumption that the unrecorded values were missing at random was corroborated by inspection of joint bivariate plots of complete and incomplete observations made on each particular marker on consecutive days.⁶ Then, each missing value of a marker was imputed by iterative sampling from its conditional predictive distribution given its past values, using R package MICE.^{6 7}

Modelling

A binary logistic model for the all-cause mortality outcome using only clinical features at presentation was fitted initially. For each of the first five days following admission, additional potential predictors from the routinely measured biomarkers available by that day were then selected resulting in a sequence of day specific mortality prediction models. We subsequently fitted each model within a generalised additive modelling (GAM) framework involving smoothing splines to detect marked departures from linearity for continuous predictors and undertook data transformations (e.g. log transformation of concentrations) as indicated.⁸ A standard logistic version of the model was then fitted. We used the Akaike Information Criterion (AIC) to choose between logistic models and assessed predictive performance using the area under a ten-fold, cross-validated Receiver Operating Characteristic (ROC) curve.

Results

A total of 392 patients with a COVID-19 diagnosis were admitted during the study period. Table 1 provides a summary of their demographic and clinical features including medical history. Blood samples were typically requested every other day following admission (supplementary Table).

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3 For an informal analysis of biomarker relationships we analysed partial correlations between
4 seven potential inflammatory markers in each of the two survival groups (Fig 1). Amongst
5 survivors, the anticipated correlations between lymphocytes (Lym) and neutrophils (Neu, via
6 white cell count) and between creatinine (Cre) and urea (Ure) were present, but these
7 correlations were found to be significantly lower among the decedents, suggesting the
8 possible presence of differences in the neutrophil/lymphocyte and urea/creatinine ratios
9 between the two outcome groups.
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16 Inclusion of admission biomarker data did not improve the predictive value of the model over
17 clinical data alone. Incorporation of post admission dynamic biomarker data did however
18 increase the discriminative ability of the model (Fig 2). Estimates from the best fitting model
19 at Day 5 (Table 2) show strongly statistically significant term(s) reflecting post-baseline
20 biomarker changes that that can be readily visualised (Fig 3). In addition to age and disease
21 severity, the most recent neutrophil:lymphocyte ratio and the two most recent (and therefore
22 recent change in) urea:creatinine ratios were generally predictive. There was a marked non-
23 linearity in the effect of age (Fig 4).
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30 **Discussion**

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32 These results suggest that using dynamic data is better than using baseline initial presentation
33 data to predict death in COVID-19 patients. Even with a local dataset of just 392 COVID-19
34 admissions we were able to identify clear benefit from exploiting dynamic biomarker data and
35 marked non-linearity in the effects of commonly used factors to predict outcomes. Our findings
36 should be taken as indicative of the benefit of applying more recent developments in statistical
37 methodology than are commonly found in the clinical literature. Identification and validation
38 of anything approaching a definitive predictive model would require substantially larger
39 sample sizes.⁹
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47 Neither the non-linear effect of age after allowance for other factors nor the particular
48 biomarkers identified within this dataset are surprising. Others have also observed associations
49 of mortality with age, and clinical and biochemical markers of disease severity (e.g.
50 neutrophil/lymphocyte ratio).¹⁰⁻¹² Similarly, renal injury has also been shown to be common
51 in COVID-19 patients and is associated with a worse outcome.¹³ The reason for this is not
52 clear. There is emerging evidence that SARS-CoV-2 infection can directly harm the kidneys.
53 The worsening urea/creatinine ratio observed in our data set may also reflect either the
54 therapeutic effects of fluid restriction to treat severe ARDS or evidence of multiorgan
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dysfunction.^{11 14} Regardless of the cause, the impact of the urea/creatinine level on death was not evident at presentation but became a significant predictor of death in our model over time. This illustrates the benefit of taking into account improvements and deterioration in daily blood test results as well as initial presentation factors when calculating the probability of death. Improving the accuracy of prediction models using this approach is likely to be successful in informing clinical decision making, resource planning and communication with patients and relatives.

We acknowledge that our study has some limitations. We would have liked to consider other outcomes including ARDS and ICU admission, but a consistent diagnosis and dates were not routinely available. Although we have undertaken internal cross-validation to ensure unbiased comparison of ROC curves we have not considered calibration. We do not wish to make any claim for the value of our current models at each day based on the small sample size available to us locally. With three hospital sites contributing during the first wave of the pandemic we do not have sufficient data to construct definitive prediction models. More sophisticated exploitation of biomarker trajectories through, for example, approaches based on random effects models of biomarker evolution or ‘conditional on outcome’ models of biomarker evolution, would also require more data and be expected to add further insights.¹⁵⁻¹⁷

Clinical prediction models are important and can help in clinical decision making, resource allocation and optimal selection of trial participants for investigational treatments. In the setting of an infectious disease pandemic- affecting all geographic and socioeconomic groups – using routinely available and performed blood tests to inform prediction models has obvious advantages over less widely available, but perhaps more specific, biomarkers of disease severity. Until investigators incorporate such data in participant selection it is unlikely that future trials will be able to accurately target those patients most likely to benefit from therapies such as immunomodulation. Overall benefits will be ‘diluted’ and potentially reversed by inclusion of participants who have nothing to gain and, in theory, may be harmed by restriction of a healthy inflammatory response.¹⁸ The consequence of poorly considered eligibility criteria may therefore be to erroneously dismiss therapies that could benefit those at highest risk from COVID-19.

Data sharing statement.

Our data would be made available to reasonable requests.

Patient and Public involvement.

There was no involvement of patients or public involvement in the design or delivery of this study. This was because of the acute nature and fast moving pace of the disease studied and because access to patients and the public was limited at this time.

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Figure Legends:

Figure 1. Partial correlations between biomarkers. Nodes represent average marker levels day 2 to 5 and edges represent partial correlations, as calculated from the survivors (left) and from the decedents (right). Broader lines indicate stronger relationships. Blr-Bilirubin, CRP-C-Reactive Protein, Crt-Creatinine, Lym-Lymphocytes, Ntr-Neutrophils, WCC- White Cell Count, Ure-Urea.

Figure 2. ROC curves for three models: solid line indicates model considering only clinical factors at baseline (Area under ROC curve = 0.73); finely dotted line indicates model extended to consider also biomarker data from baseline sample (Area under ROC curve = 0.75); top line indicates model at 5 days extended to consider dynamic changes in biomarker data (Area under ROC curve = 0.83). Note that models are not nested

Figure 3. Violin plots showing distribution at each day of admission, stratified by survival status, for biomarkers identified by statistical modelling. Panel A: log transformed neutrophil ($\times 10^9/L$):lymphocyte ($\times 10^9/L$) ratio. Panel B: log transformed urea (mmol/L):creatinine ($\mu\text{mol/L}$) ratio. Survivors (white) on left and decedents (shaded) on right.

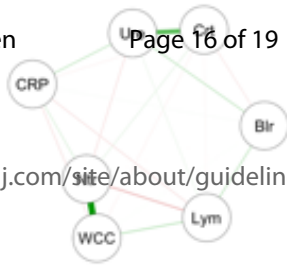
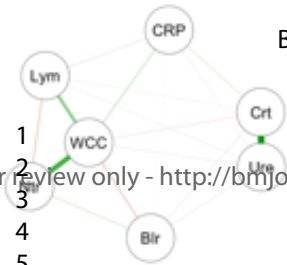
Figure 4. Spline plot demonstrating marked non-linearity in relationship between age and outcome after adjustment for other factors included in the final model.

Table 1. Demographic, clinical and medical history factors considered at baseline.

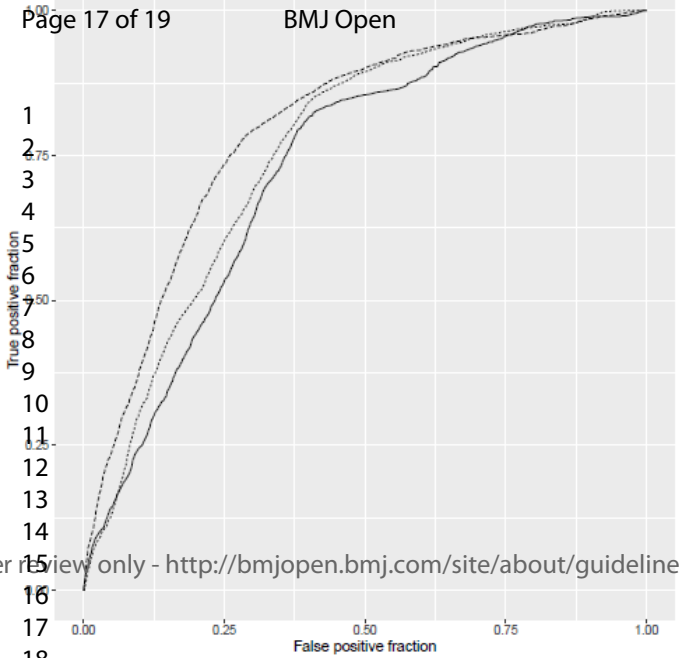
| | Overall dataset |
|---|-----------------|
| Number of patients | 392 |
| Age, median (IQR) | 71(22) |
| Gender Male: Female ratio | 65:35 |
| Median time to hospitalisation following disease onset (IQR) days | 5(8) |
| Initial symptoms (%) | |
| Fever | 223 (57%) |
| Cough | 240(61%) |
| Dyspnoea | 245 (65%) |
| Fatigue | 127 (37%) |
| Muscle ache | 53 (16%) |
| Co-morbidities | |
| Cardiovascular disease | 108 (28%) |
| Chronic Respiratory disease (inc asthma) | 110 (28%) |
| Chronic Renal disease | 45 (12%) |
| Chronic Liver disease | 14 (2%) |
| Obesity | 34 (10%) |
| Diabetes | 95 (24%) |
| Dementia | 49 (13%) |
| Current smoker | 24 (7%) |
| Presenting clinical features | |
| Requirement for supplemental O ₂ | 125(37%) |
| Oxygen Saturation < 90 | 59 (17%) |
| Respiratory rate>24 | 109 (30%) |
| Temperature ≥38°C | 168 (45%) |
| MAP≤70mmHg | 30 (8%) |
| Outcomes | |
| Acute Respiratory Distress Syndrome | 47 (17%) |
| Non-Invasive Ventilation | 25 (9%) |
| Need for ICU care | 31 (12%) |
| Invasive Ventilation | 14 (5%) |
| Death | 110 (27%) |

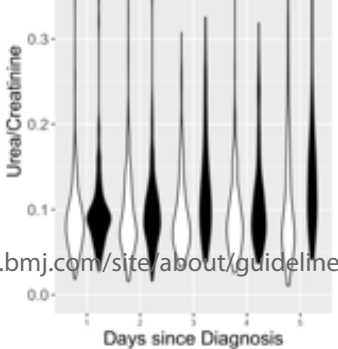
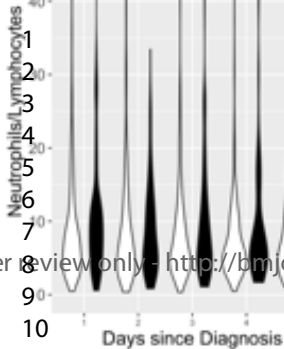
Table 2. Estimated coefficients (Est) with their standard error (se) and p-value. Note that different variables are selected at different days so that models are not nested. Neut/Lymp- Neutrophil/Lymphocyte, Creat- Creatinine, D- Day, O₂- Oxygen. * The addition of biomarker data on day 2 did not contribute any additional predictive power of that obtained at Day 1.

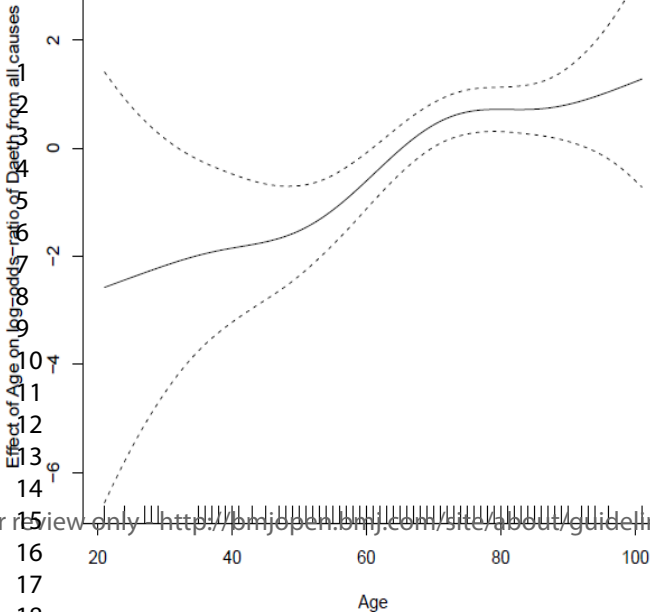
| Predictor | Clinical data alone Day 1 | | | Clinical data + Day 1* biomarker data | | | Day 3 | | | Day 4 | | | Day 5 | | |
|---------------------------|---------------------------|-------|---------|---------------------------------------|-------|---------|-------|-------|---------|-------|-------|---------|-------|-------|---------|
| | Est | se | p | Est | se | p | Est | se | p | Est | se | p | Est | se | p |
| Intercept | 0.31 | 1.36 | 0.8 | -4.36 | 1.35 | 0.001 | 0.67 | 1.97 | 0.73 | 0.005 | 1.86 | 0.99 | -0.20 | 1.68 | 0.9 |
| log Neut/Lymp D1 | | | | 0.28 | 0.16 | 0.08 | | | | | | | | | |
| log Neut/Lymp D3 | | | | | | | 0.41 | 0.19 | 0.03 | | | | | | |
| log Neut/Lymp D4 | | | | | | | | | | 0.48 | 0.2 | 0.02 | | | |
| log Neut/Lymp D5 | | | | | | | | | | | | | 0.52 | 0.21 | 0.01 |
| log Urea/Creat D2 | | | | | | | -4.22 | 1.24 | 0.0007 | | | | | | |
| log Urea/Creat D3 | | | | | | | 5.13 | 1.30 | 0.0001 | | | | | | |
| log Urea/Creat D4 | | | | | | | | | | 1.08 | 0.35 | 0.002 | -4.97 | 1.72 | 0.0003 |
| log Urea/Creat D5 | | | | | | | | | | | | | 6.32 | 1.77 | 0.0004 |
| Age (Years) | 0.13 | 0.026 | <0.0001 | 0.073 | 0.012 | <0.0001 | 0.069 | 0.013 | <0.0001 | 0.071 | 0.012 | <0.0001 | 0.066 | 0.012 | <0.0001 |
| O ₂ Saturation | -0.03 | 0.013 | 0.05 | -0.03 | 0.012 | 0.01 | -0.03 | 0.013 | 0.03 | -0.03 | 0.012 | 0.02 | | | |
| Respiratory Rate | 0.05 | 0.022 | 0.02 | 0.085 | 0.022 | 0.0001 | 0.08 | 0.023 | 0.0003 | 0.087 | 0.022 | 0.0001 | 0.09 | 0.022 | 0.0001 |
| Smoking | 0.44 | 0.29 | 0.1 | 0.7 | 0.267 | 0.01 | 0.8 | 0.27 | 0.004 | 0.71 | 0.27 | 0.008 | 0.76 | 0.28 | 0.006 |



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| | Day 1 | Day 2 | Day 3 | Day 4 | Day 5 |
|----------------------------------|-----------|-----------|-----------|-----------|-----------|
| White Cell count $\times 10^9/L$ | 7.5 (4.2) | 7.1 (4.3) | 7.5 (4.3) | 7.1 (4.7) | 7.6 (4.6) |
| Lymphocytes $\times 10^9/L$ | 0.8 (0.6) | 0.8 (0.5) | 0.8 (0.5) | 0.8 (0.6) | 0.8 (0.5) |
| Neutrophils $\times 10^9/L$ | 6.1 (4.6) | 5.7 (4.2) | 5.6 (4.2) | 5.6 (4.4) | 6.4 (4.2) |
| Platelets $\times 10^9/L$ | 202 (121) | 209 (125) | 235 (131) | 262 (161) | 265 (164) |
| Bilirubin mg/dL | 10 (8) | 9 (7) | 11 (7) | 10(5) | 11 (6) |
| Urea mmol/L | 8 (6) | 7(6) | 7(6) | 7(6) | 8 (6) |
| Creatinine $\mu\text{mol/L}$ | 90 (56) | 80(51) | 74 (45) | 75(40) | 74 (52) |
| CRP mg/ml | 98 (123) | 115 (98) | 122 (121) | 121 (125) | 117 (146) |

Supplementary Table: Values of biomarkers at each day. Data is presented as median (interquartile range).

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The value of dynamic clinical and biomarker data for mortality risk prediction in COVID-19: A multi-centre retrospective cohort study

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The value of dynamic clinical and biomarker data for mortality risk prediction in COVID-19: A multi-centre retrospective cohort study

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45 All authors (CB, CH, AK, AV, CO, DB, JG, KO, MW, OP, SH, SA, LB, HP) were involved in the
46
47 concept and design of the study which was led by HP and AV. HP, CH, MW acquired the data, and CB,
48
49 LB, AV, HP, CH analysed and interpreted the data. CB, DB, SA, OP, JG, AV, CH, HP drafted the
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53 in revising the manuscript and contributed to the final draft. The corresponding author attests that all
54
55 listed authors meet authorship criteria and that no others meeting the criteria have been omitted. HP
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57 acts as guarantor for the study.
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Competing Interests Declaration

Swedish Orphan Biovitrum have provided investigational medicinal product for public-funded, peer-reviewed trials on which AK, AV, JG, HP, and SH are coinvestigators. The other authors declare no competing interests.

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Abstract

Objectives

Being able to predict which COVID-19 patients are going to deteriorate is important to help identify patients for clinical and research practice. Clinical prediction models play a critical role in this process, but current models are of limited value because they are typically restricted to baseline predictors and don't always use contemporary statistical methods. We sought to explore the benefits of incorporating dynamic changes in routinely measured biomarkers, non-linear effects and applying 'state of the art' statistical methods in the development of a prognostic model to predict death in hospitalised COVID-19 patients.

Design

Data were analysed from COVID-19 admissions to three hospital sites. Exploratory data analysis included a graphical approach to partial correlations. Dynamic biomarkers were considered up to five days following admission rather than depending solely on baseline or single time-point data. Marked departures from linear effects of covariates were identified by employing smoothing splines within a generalised additive modelling framework.

Setting

3 secondary and tertiary level centres in Greater Manchester, UK.

Participants

392 hospitalised patients with a diagnosis of COVID-19

Results

392 patients with a COVID-19 diagnosis were identified. Area under the receiver operating characteristic (ROC) curve increased from 0.73 using admission data alone to 0.75 when also considering results of baseline blood samples and to 0.83 when considering dynamic values of routinely collected markers. There was clear non-linearity in the association of age with patient outcome.

Conclusions

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3 This study shows that clinical prediction models to predict death in hospitalised COVID-19
4 patients can be improved by taking into account both non-linear effects in covariates such as
5 age and dynamic changes in values of biomarkers.
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8 9 **Strengths and Limitations.**

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12 • Incorporating routinely available blood tests performed over the first 5 days of
13 hospital admission with clinical presentation data can enhance patient-level prediction
14 of COVID-19 progression.
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17 • A larger dataset is needed to construct definitive prediction models.
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20 • More sophisticated statistical exploitation of biomarker trajectories e. g using random
21 effects models of biomarker evolution or ‘conditional on outcome’ models of
22 biomarker evolution, could make clinical predictions models better still.
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Introduction

Most patients with severe acute respiratory syndrome coronavirus 2 (SARS-Cov-2) experience mild symptoms. Some patients however experience significant symptoms requiring hospitalisation. The pandemic nature of the covid-19 outbreak has meant that hospital services and capacity can be overwhelmed.¹ A tool to predict which patients are likely to deteriorate or need intensive care would help clinicians, hospital managers and researchers make better decisions.

Several such models are reported for COVID-19 patients but have been criticised for risk of bias using the PROBAST (prediction model risk of bias assessment tool) criteria.² We have further concerns regarding the statistical tools used to develop models. First, current models typically only consider patient characteristics available at baseline and do not consider that COVID-19 patients' presentation and in hospital course is variable. Secondly, models routinely seek only linear effects of potential predictors on the outcome of interest although these are not always clinically plausible.

We sought here to explore the benefits of incorporating dynamic changes in routinely measured biomarkers, non-linear effects and applying 'state of the art' statistical methods in the development of a prognostic model to predict death in hospitalised COVID-19 patients.

Methods

Study Population

Admissions with confirmed COVID-19 (according to World Health Organisation guidance) at three hospitals in the Northern Care Alliance (Greater Manchester, UK) between 11th March and 17th April 2020 with a minimum of a three week follow-up were studied.³

Data Collection

Necessary approvals were obtained from the local Research and Innovation department.

Research nurses abstracted data from the electronic patient records based on the International Severe Acute Respiratory and emerging Infection Consortium (ISARIC) data collection tool

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3 but modified for use with this study.⁴ The ISARIC study data were supplemented from
4 electronic patient records with results of blood analyses performed as part of routine clinical
5 care. The date of diagnosis was considered Day 1.
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9 Data analysis

10 All data were subjected to range checks and validated for internal consistency and missing
11 items then anonymised prior to transfer.
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15 Selection of biomarkers

16 The initial list of potential markers was determined through review of the literature and
17 availability within routinely collected data. Candidate variables were further screened using
18 a graphical representation of the partial correlation structure stratified by survival status.⁵
19 Routine bloods were typically analysed on alternate days. The assumption that the
20 unrecorded values were missing at random was corroborated by inspection of joint bivariate
21 plots of complete and incomplete observations made on each particular marker on
22 consecutive days.⁶ Then, each missing value of a marker was imputed by iterative sampling
23 from its conditional predictive distribution given its past values, using R package MICE.^{6 7}
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33 Modelling

34 In this study we used the information contained in the clinical presentation data and available
35 biomarkers (creatinine, lymphocyte count, etc.) to update, on a day-by-day basis, the patient's
36 probability of death within 21 days
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42 Initially, a binary logistic model for the all-cause mortality outcome using only clinical
43 features at presentation was fitted initially. We then fitting separate logistic models for death
44 for each day, using predictive variables identified from the partial correlation analysis
45 described above. For each of the five days following hospital admission, we fitted a model
46 based exclusively on data from subjects still alive at that day, with candidate predictors
47 chosen out of the set of clinical variables and biomarker values collected until that day. This
48 approach meant that for each of the first five days following admission, a sequence of day
49 specific mortality prediction models were available. We subsequently fitted each model
50 within a generalised additive modelling (GAM) framework involving smoothing splines to
51 detect marked departures from linearity for continuous predictors and undertook data
52 transformations (e.g. log transformation of concentrations) as indicated.⁸ A standard logistic
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3 version of the model was then fitted. We used the Akaike Information Criterion (AIC) to
4 choose between logistic models and assessed predictive performance using the area under a
5 ten-fold, cross-validated Receiver Operating Characteristic (ROC) curve.
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8 9 Patient and Public involvement.

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12 There was no involvement of patients or the general public in the design or delivery of this
13 study. This was because of the acute nature and fast moving pace of the disease studies and
14 because access to patients and the public was limited at this time.
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17 18 **Results**

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20 A total of 392 patients with a COVID-19 diagnosis were admitted during the study period.
21 Table 1 provides a summary of their demographic and clinical features including medical
22 history. Blood samples were typically requested every other day following admission
23 (supplementary Table).
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27 For an informal analysis of biomarker relationships we analysed partial correlations between
28 seven potential inflammatory markers in each of the two survival groups (Fig 1). Amongst
29 survivors, the anticipated correlations between lymphocytes (Lym) and neutrophils (Neu, via
30 white cell count) and between creatinine (Cre) and urea (Ure) were present, but these
31 correlations were found to be significantly lower among the decedents, suggesting the
32 possible presence of differences in the neutrophil/lymphocyte and urea/creatinine ratios
33 between the two outcome groups.
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37 Inclusion of admission biomarker data did not improve the predictive value of the model over
38 clinical data alone. Incorporation of post admission dynamic biomarker data did however
39 increase the discriminative ability of the model (Fig 2). Estimates from the best fitting model
40 at Day 5 (Table 2) show strongly statistically significant term(s) reflecting post-baseline
41 biomarker changes that that can be readily visualised (Fig 3). In addition to age and disease
42 severity, the most recent neutrophil:lymphocyte ratio and the two most recent (and therefore
43 recent change in) urea:creatinine ratios were generally predictive. There was a marked non-
44 linearity in the effect of age (Fig 4).
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47 48 **Discussion**

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50 These results suggest that using dynamic data is better than using baseline initial presentation
51 data to predict death in COVID-19 patients. Even with a local dataset of just 392 COVID-19
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3 admissions we were able to identify clear benefit from exploiting dynamic biomarker data and
4 marked non-linearity in the effects of commonly used factors to predict outcomes. Our findings
5 should be taken as indicative of the benefits of 'state of the art' statistical methodology but also
6 the necessary collaboration between statisticians and clinicians as this statistical methodology
7 is not readily accessible to most researchers. Identification and validation of anything
8 approaching a definitive predictive model would require substantially larger sample sizes.⁹
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14 Neither the non-linear effect of age after allowance for other factors nor the particular
15 biomarkers identified within this dataset are surprising. Others have also observed
16 associations of mortality with age, and clinical and biochemical markers of disease severity
17 (e.g. neutrophil/lymphocyte ratio).¹⁰⁻¹² There have been few studies investigating dynamic
18 changes in patient biomarkers for mortality prediction in COVID-19; one such study of 548
19 patients in China also demonstrated that the neutrophil:lymphocyte ratio in survivors and
20 non-survivors became increasingly divergent throughout their hospital admission.¹³ Chen et
21 al derived their prognostic score from an analysis based on a Cox's regression model with
22 their candidate predictive variables taken at baseline. They incorporated in their analysis the
23 slope of a line fitted to the first and last measurements of each particular marker to model
24 changes over time. Chen et al approach has advantages and disadvantages. Their model
25 captures duration information but does not involve choice of time horizon for prediction.
26 Their predictions are arguably limited because they are not updated daily and depend on the
27 assumption that marker evolution is linear and summarised by a straight line between initial
28 and final values.
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41 A smaller study limited to patients with severe COVID-19 also revealed a progressive
42 increase in neutrophil count and plasma interleukin-6 concentration in the decedents when
43 compared to the survivors, but the authors did not perform any assessment of the predictive
44 value associated with dynamic changes in these laboratory parameters.¹⁴
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50 Similarly, renal injury has also been shown to be common in COVID-19 patients and is
51 associated with a worse outcome.¹⁵ The reason for this is not clear. There is emerging evidence
52 that SARS-CoV-2 infection can directly harm the kidneys. The worsening urea/creatinine ratio
53 observed in our data set may also reflect either the therapeutic effects of fluid restriction to
54 treat severe ARDS or evidence of multiorgan dysfunction.^{11 16} Regardless of the cause, the
55 impact of the urea/creatinine level on death was not evident at presentation but became a
56 significant predictor of death in our model over time. This observation illustrates the benefit of
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3 taking into account improvements and deterioration in daily blood test results as well as initial
4 presentation factors when calculating the probability of death. Improving the accuracy of
5 prediction models using this approach is likely to be successful in informing clinical decision
6 making, resource planning and communication with patients and relatives.
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10 We acknowledge that our study has some limitations. We would have liked to consider other
11 outcomes including dynamic changes in clinical variables, as well as disease end points such
12 as the incidence of ARDS and ICU admission. Dynamic clinical data were not included
13 because it was less reliable to obtain compared to blood biomarker data, and a consistent
14 diagnosis of ARDS, and dates of onset or admission to ICU were also not routinely available.
15 Although we have undertaken internal cross-validation to ensure unbiased comparison of
16 ROC curves we have not considered calibration. We do not wish to make any claim for the
17 value of our current models at each day based on the small sample size available to us locally.
18 With only three hospital sites contributing during the first wave, and because of significant
19 time/resource pressures during the pandemic we did not have sufficient data to construct
20 definitive prediction models or to follow-up patients beyond 3 weeks. More sophisticated
21 exploitation of biomarker trajectories through, for example, approaches based on random
22 effects models of biomarker evolution or ‘conditional on outcome’ models of biomarker
23 evolution, would also require more data and be expected to add further insights.¹⁷⁻¹⁹
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35 Clinical prediction models are important and can help in clinical decision making, resource
36 allocation and optimal selection of trial participants for investigational treatments. In the setting
37 of an infectious disease pandemic- affecting all geographic and socioeconomic groups – using
38 routinely available blood tests to inform prediction models has obvious advantages over less
39 widely available, but perhaps more specific, biomarkers of disease severity. Until investigators
40 incorporate such data in participant selection it is unlikely that future trials will be able to
41 accurately target those patients most likely to benefit from therapies such as
42 immunomodulation. Overall benefits will be ‘diluted’ and potentially reversed by inclusion of
43 participants who have nothing to gain and, in theory, may be harmed by restriction of a healthy
44 inflammatory response.²⁰ The consequence of poorly considered eligibility criteria may
45 therefore be to erroneously dismiss therapies that could benefit those at highest risk from
46 COVID-19.
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56 **Data sharing statement.**

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59 Our data would be made available to reasonable requests.
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Figure Legends:

Figure 1. Partial correlations between biomarkers. Nodes represent average marker levels day 2 to 5 and edges represent partial correlations, as calculated from the survivors (left) and from the decedents (right). Broader lines indicate stronger relationships. Blr-Bilirubin, CRP-C-Reactive Protein, Crt-Creatinine, Lym-Lymphocytes, Ntr-Neutrophils, WCC- White Cell Count, Ure-Urea.

Figure 2. ROC curves for three models: solid line indicates model considering only clinical factors at baseline (Area under ROC curve = 0.73); finely dotted line indicates model extended to consider also biomarker data from baseline sample (Area under ROC curve = 0.75); top line indicates model at 5 days extended to consider dynamic changes in biomarker data (Area under ROC curve = 0.83). Note that models are not nested

Figure 3. Violin plots showing distribution at each day of admission, stratified by survival status, for biomarkers identified by statistical modelling. Panel A: log transformed neutrophil ($\times 10^9/L$):lymphocyte ($\times 10^9/L$) ratio. Panel B: log transformed urea (mmol/L):creatinine ($\mu\text{mol/L}$) ratio. Survivors (white) on left and decedents (shaded) on right.

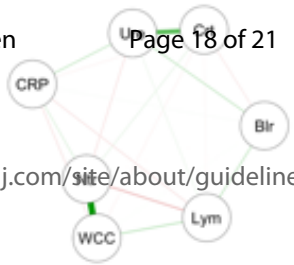
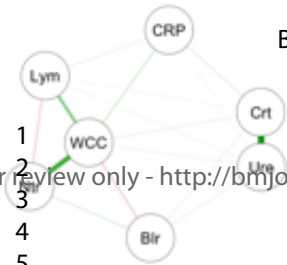
Figure 4. Spline plot demonstrating marked non-linearity in relationship between age and outcome after adjustment for other factors included in the final model.

Table 1. Demographic, clinical and medical history factors considered at baseline.

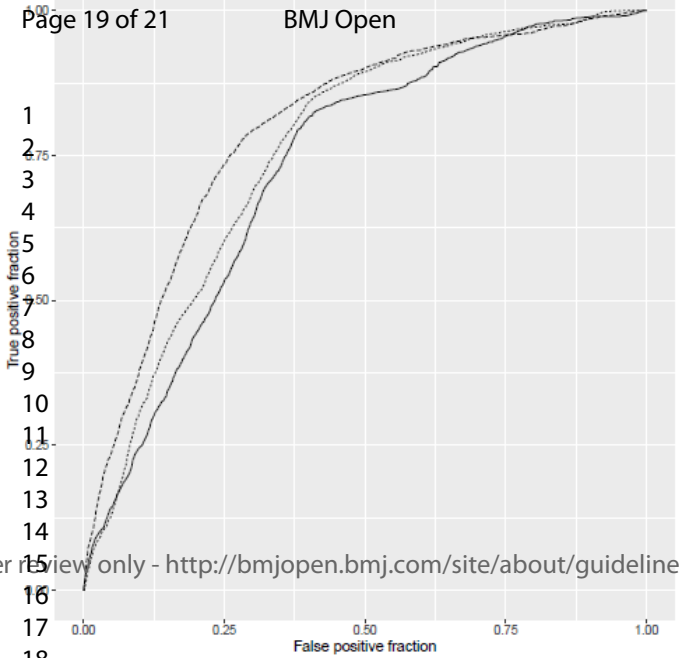
| | Overall dataset |
|---|-----------------|
| Number of patients | 392 |
| Age, median (IQR) | 71(22) |
| Gender Male: Female ratio | 65:35 |
| Median time to hospitalisation following disease onset (IQR) days | 5(8) |
| Initial symptoms (%) | |
| Fever | 223 (57%) |
| Cough | 240(61%) |
| Dyspnoea | 245 (65%) |
| Fatigue | 127 (37%) |
| Muscle ache | 53 (16%) |
| Co-morbidities | |
| Cardiovascular disease | 108 (28%) |
| Chronic Respiratory disease (inc asthma) | 110 (28%) |
| Chronic Renal disease | 45 (12%) |
| Chronic Liver disease | 14 (2%) |
| Obesity | 34 (10%) |
| Diabetes | 95 (24%) |
| Dementia | 49 (13%) |
| Current smoker | 24 (7%) |
| Presenting clinical features | |
| Requirement for supplemental O ₂ | 125(37%) |
| Oxygen Saturation < 90 | 59 (17%) |
| Respiratory rate>24 | 109 (30%) |
| Temperature ≥38°C | 168 (45%) |
| MAP≤70mmHg | 30 (8%) |
| Outcomes | |
| Acute Respiratory Distress Syndrome | 47 (17%) |
| Non-Invasive Ventilation | 25 (9%) |
| Need for ICU care | 31 (12%) |
| Invasive Ventilation | 14 (5%) |
| Death | 110 (27%) |

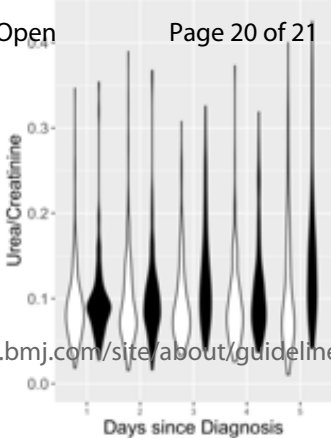
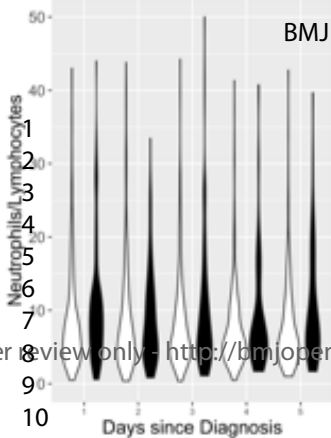
Table 2. Estimated coefficients (Est) with their standard error (se) and p-value. Note that different variables are selected at different days so that models are not nested. Neut/Lymp- Neutrophil/Lymphocyte, Creat- Creatinine, D- Day, O₂- Oxygen. * The addition of biomarker data on day 2 did not contribute any additional predictive power of that obtained at Day 1.

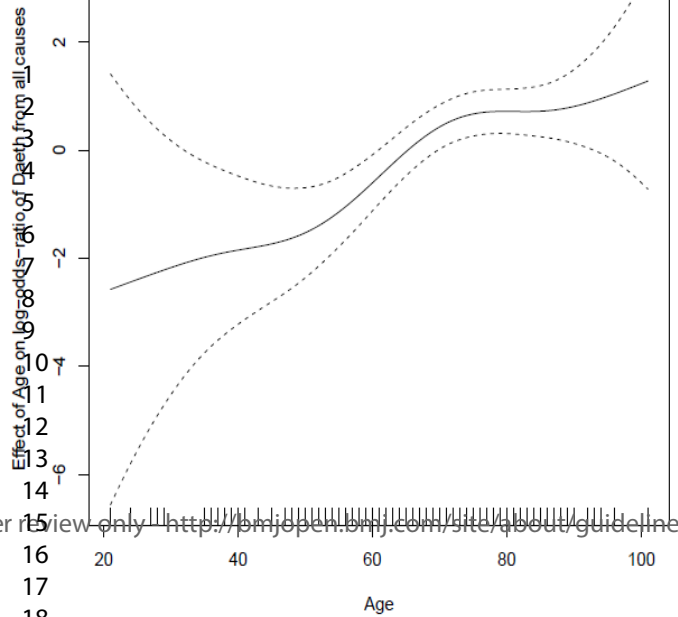
| Predictor | Clinical data alone Day 1 | | | Clinical data + Day 1* biomarker data | | | Day 3 | | | Day 4 | | | Day 5 | | |
|---------------------------|---------------------------|-------|---------|---------------------------------------|-------|---------|-------|-------|---------|-------|-------|---------|-------|-------|---------|
| | Est | se | p | Est | se | p | Est | se | p | Est | se | p | Est | se | p |
| Intercept | 0.31 | 1.36 | 0.8 | -4.36 | 1.35 | 0.001 | 0.67 | 1.97 | 0.73 | 0.005 | 1.86 | 0.99 | -0.20 | 1.68 | 0.9 |
| log Neut/Lymp D1 | | | | 0.28 | 0.16 | 0.08 | | | | | | | | | |
| log Neut/Lymp D3 | | | | | | | 0.41 | 0.19 | 0.03 | | | | | | |
| log Neut/Lymp D4 | | | | | | | | | | 0.48 | 0.2 | 0.02 | | | |
| log Neut/Lymp D5 | | | | | | | | | | | | | 0.52 | 0.21 | 0.01 |
| log Urea/Creat D2 | | | | | | | -4.22 | 1.24 | 0.0007 | | | | | | |
| log Urea/Creat D3 | | | | | | | 5.13 | 1.30 | 0.0001 | | | | | | |
| log Urea/Creat D4 | | | | | | | | | | 1.08 | 0.35 | 0.002 | -4.97 | 1.72 | 0.0003 |
| log Urea/Creat D5 | | | | | | | | | | | | | 6.32 | 1.77 | 0.0004 |
| Age (Years) | 0.13 | 0.026 | <0.0001 | 0.073 | 0.012 | <0.0001 | 0.069 | 0.013 | <0.0001 | 0.071 | 0.012 | <0.0001 | 0.066 | 0.012 | <0.0001 |
| O ₂ Saturation | -0.03 | 0.013 | 0.05 | -0.03 | 0.012 | 0.01 | -0.03 | 0.013 | 0.03 | -0.03 | 0.012 | 0.02 | | | |
| Respiratory Rate | 0.05 | 0.022 | 0.02 | 0.085 | 0.022 | 0.0001 | 0.08 | 0.023 | 0.0003 | 0.087 | 0.022 | 0.0001 | 0.09 | 0.022 | 0.0001 |
| Smoking | 0.44 | 0.29 | 0.1 | 0.7 | 0.267 | 0.01 | 0.8 | 0.27 | 0.004 | 0.71 | 0.27 | 0.008 | 0.76 | 0.28 | 0.006 |



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| | Day 1 | Day 2 | Day 3 | Day 4 | Day 5 |
|----------------------------------|-----------|-----------|-----------|-----------|-----------|
| White Cell count $\times 10^9/L$ | 7.5 (4.2) | 7.1 (4.3) | 7.5 (4.3) | 7.1 (4.7) | 7.6 (4.6) |
| Lymphocytes $\times 10^9/L$ | 0.8 (0.6) | 0.8 (0.5) | 0.8 (0.5) | 0.8 (0.6) | 0.8 (0.5) |
| Neutrophils $\times 10^9/L$ | 6.1 (4.6) | 5.7 (4.2) | 5.6 (4.2) | 5.6 (4.4) | 6.4 (4.2) |
| Platelets $\times 10^9/L$ | 202 (121) | 209 (125) | 235 (131) | 262 (161) | 265 (164) |
| Bilirubin mg/dL | 10 (8) | 9 (7) | 11 (7) | 10(5) | 11 (6) |
| Urea mmol/L | 8 (6) | 7(6) | 7(6) | 7(6) | 8 (6) |
| Creatinine $\mu\text{mol/L}$ | 90 (56) | 80(51) | 74 (45) | 75(40) | 74 (52) |
| CRP mg/ml | 98 (123) | 115 (98) | 122 (121) | 121 (125) | 117 (146) |

Supplementary Table: Values of biomarkers at each day. Data is presented as median (interquartile range).

TRIPOD Checklist: Prediction Model Development

| Section/Topic | Item | Checklist Item | Page |
|------------------------------|------|---|----------|
| Title and abstract | | | |
| Title | 1 | Identify the study as developing and/or validating a multivariable prediction model, the target population, and the outcome to be predicted. | 1 |
| Abstract | 2 | Provide a summary of objectives, study design, setting, participants, sample size, predictors, outcome, statistical analysis, results, and conclusions. | 4 |
| Introduction | | | |
| Background and objectives | 3a | Explain the medical context (including whether diagnostic or prognostic) and rationale for developing or validating the multivariable prediction model, including references to existing models. | 6 |
| | 3b | Specify the objectives, including whether the study describes the development or validation of the model or both. | 7 |
| Methods | | | |
| Source of data | 4a | Describe the study design or source of data (e.g., randomized trial, cohort, or registry data), separately for the development and validation data sets, if applicable. | 7 |
| | 4b | Specify the key study dates, including start of accrual; end of accrual; and, if applicable, end of follow-up. | 7 |
| Participants | 5a | Specify key elements of the study setting (e.g., primary care, secondary care, general population) including number and location of centres. | 6 |
| | 5b | Describe eligibility criteria for participants. | 6 |
| | 5c | Give details of treatments received, if relevant. | n/a |
| Outcome | 6a | Clearly define the outcome that is predicted by the prediction model, including how and when assessed. | 6,7 |
| | 6b | Report any actions to blind assessment of the outcome to be predicted. | n/a |
| Predictors | 7a | Clearly define all predictors used in developing or validating the multivariable prediction model, including how and when they were measured. | 7 |
| | 7b | Report any actions to blind assessment of predictors for the outcome and other predictors. | n/a |
| Sample size | 8 | Explain how the study size was arrived at. | n/a |
| Missing data | 9 | Describe how missing data were handled (e.g., complete-case analysis, single imputation, multiple imputation) with details of any imputation method. | 7 |
| Statistical analysis methods | 10a | Describe how predictors were handled in the analyses. | 7 |
| | 10b | Specify type of model, all model-building procedures (including any predictor selection), and method for internal validation. | 7 |
| | 10d | Specify all measures used to assess model performance and, if relevant, to compare multiple models. | 7 |
| Risk groups | 11 | Provide details on how risk groups were created, if done. | n/a |
| Results | | | |
| Participants | 13a | Describe the flow of participants through the study, including the number of participants with and without the outcome and, if applicable, a summary of the follow-up time. A diagram may be helpful. | 13 |
| | 13b | Describe the characteristics of the participants (basic demographics, clinical features, available predictors), including the number of participants with missing data for predictors and outcome. | 13 |
| Model development | 14a | Specify the number of participants and outcome events in each analysis. | 13 |
| | 14b | If done, report the unadjusted association between each candidate predictor and outcome. | n/a |
| Model specification | 15a | Present the full prediction model to allow predictions for individuals (i.e., all regression coefficients, and model intercept or baseline survival at a given time point). | 14 |
| | 15b | Explain how to use the prediction model. | 8 |
| Model performance | 16 | Report performance measures (with CIs) for the prediction model. | Figure 3 |
| Discussion | | | |
| Limitations | 18 | Discuss any limitations of the study (such as nonrepresentative sample, few events per predictor, missing data). | 9 |
| Interpretation | 19b | Give an overall interpretation of the results, considering objectives, limitations, and results from similar studies, and other relevant evidence. | 8 |
| Implications | 20 | Discuss the potential clinical use of the model and implications for future research. | 9 |
| Other information | | | |
| Supplementary information | 21 | Provide information about the availability of supplementary resources, such as study protocol, Web calculator, and data sets. | 10 |
| Funding | 22 | Give the source of funding and the role of the funders for the present study. | 11 |

We recommend using the TRIPOD Checklist in conjunction with the TRIPOD Explanation and Elaboration document.