

**Supplemental Information: Investigation and alteration of organic acid synthesis
pathways in the mammalian gut symbiont *Bacteroides thetaiotaomicron***

Nathan T. Porter, and Johan Larsbrink

2 Supplemental Figures

8 Supplemental Tables

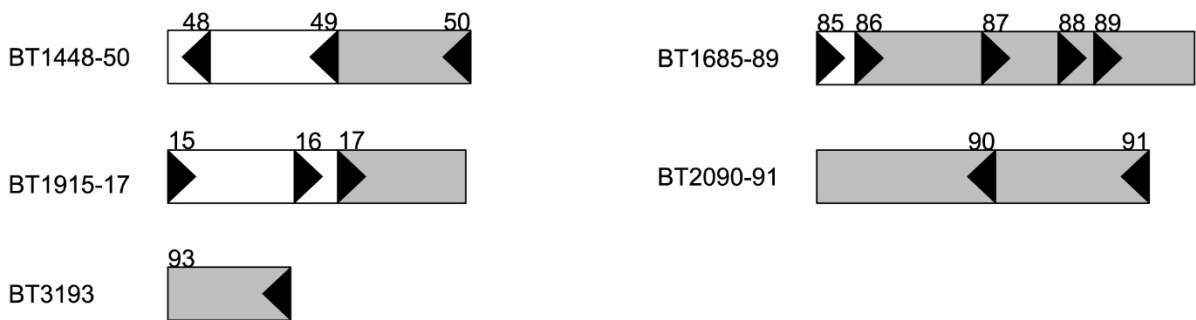
Potential genes involved in formate production



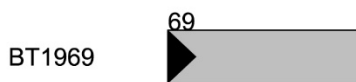
Potential genes involved in lactate production



Potential genes involved in propionate production



Potential genes connecting reductive and oxidative metabolism



Potential genes involved in acetate production



Figure S1. Maps of genetic loci investigated in this study, drawn to scale. Loci are organized by metabolic pathway in which they are putatively involved, with corresponding locus tags shown. Gray boxes indicate genes which were deleted in this study. The blue box indicates the BT3692-93 promoter replaced in this study.

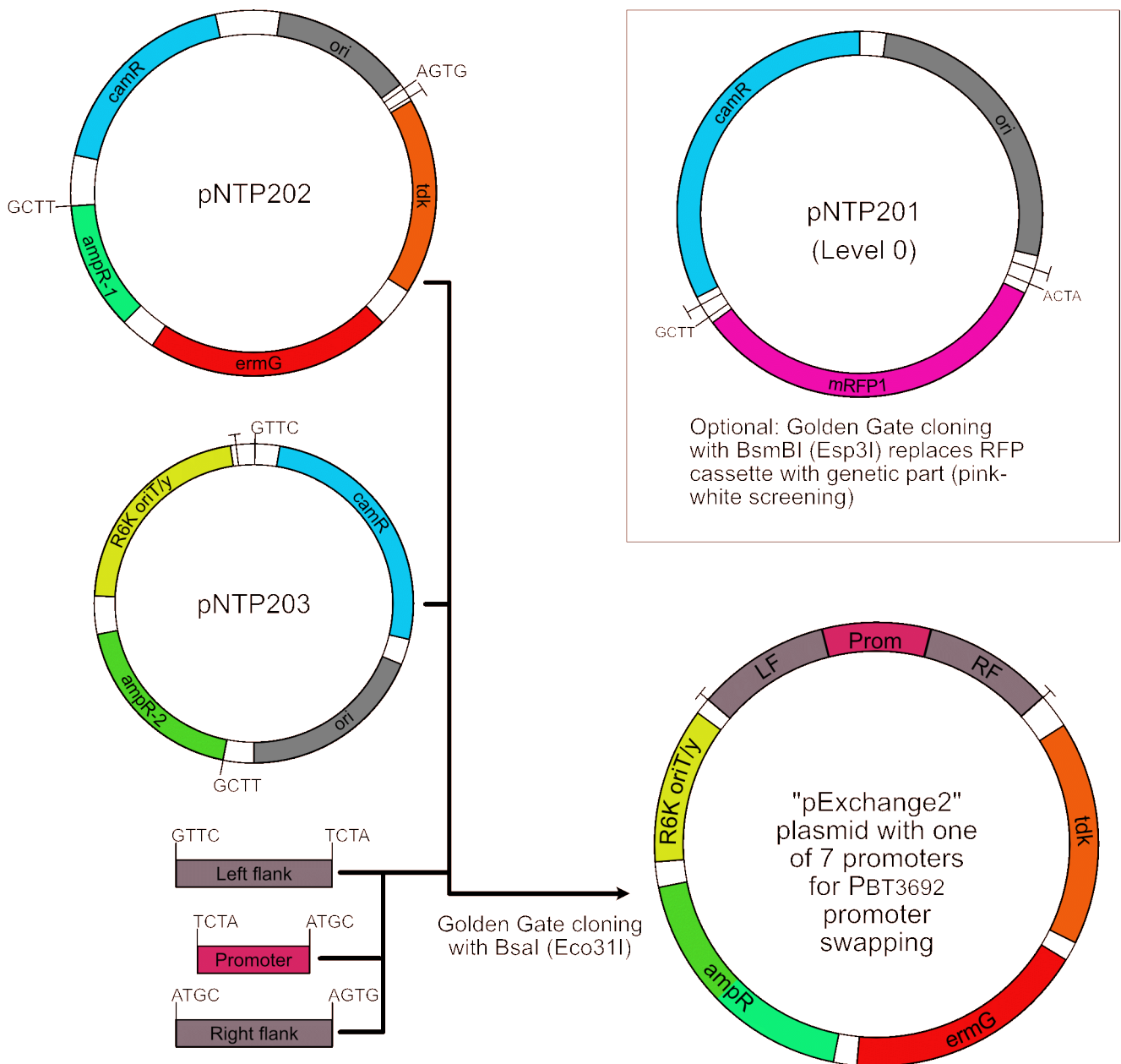


Figure S2. New Golden Gate cloning plasmid set for *Bacteroides* non-replicating plasmids based on pExchange-tdk. Optionally, genetic parts can first be incorporated into pNTP201 with Golden Gate cloning and BsmBI. Then the final "pExchange2" vector is assembled from pNTP202, pNTP203, and inserts (PCR amplicons, oligonucleotides, or plasmids) with BsaI-based Golden Gate cloning. 4-nucleotide overhang sequences are shown for each part used here. Flanking termination sequences are also shown with a "T" on pNTP201-3 and the final plasmid.

Table S1. Organic acid concentrations (mM \pm standard deviation) and glucose remaining over time for wild-type *B. thetaiotaomicron* (n = 3)

Time (hr)	Acetate	α -keto glutarate	Formate	Lactate	Malate	Propionate	Pyruvate	Succinate	Glucose (%) ^a
0	ND ^b	ND	ND	ND	ND	ND	ND	ND	96.4 \pm 4.4
6	1.81 \pm 0.32	0.02 \pm 0.00	1.18 \pm 0.18	ND	0.68 \pm 0.01	1.01 \pm 0.07	ND	0.90 \pm 0.16	90.5 \pm 3.2
7	2.69 \pm 0.48	0.03 \pm 0.00	1.84 \pm 0.19	ND	0.73 \pm 0.03	1.09 \pm 0.05	ND	1.51 \pm 0.26	81.1 \pm 4.1
8	4.26 \pm 0.66	0.03 \pm 0.00	2.46 \pm 0.38	ND	0.83 \pm 0.06	1.23 \pm 0.07	ND	2.33 \pm 0.40	68.9 \pm 4.7
12	9.77 \pm 0.50	ND	6.48 \pm 0.17	2.69 \pm 0.56	1.90 \pm 0.12	2.70 \pm 0.10	0.20 \pm 0.00	5.29 \pm 0.30	25.2 \pm 4.5
24	14.07 \pm 0.43	0.05 \pm 0.00	7.73 \pm 0.07	7.87 \pm 0.17	2.39 \pm 0.02	5.07 \pm 0.07	0.11 \pm 0.01	6.93 \pm 0.07	ND
30	14.20 \pm 0.42	0.04 \pm 0.00	7.64 \pm 0.09	7.76 \pm 0.10	2.23 \pm 0.02	5.34 \pm 0.03	0.08 \pm 0.00	7.00 \pm 0.08	ND
36	14.27 \pm 0.12	0.04 \pm 0.00	7.61 \pm 0.03	7.59 \pm 0.10	2.12 \pm 0.01	5.82 \pm 0.05	0.07 \pm 0.00	7.22 \pm 0.13	ND
48	14.85 \pm 0.62	0.03 \pm 0.03	7.17 \pm 0.44	6.79 \pm 0.51	2.03 \pm 0.20	6.21 \pm 0.16	0.23 \pm 0.30	7.56 \pm 0.46	ND
72	15.25 \pm 0.54	0.04 \pm 0.00	7.75 \pm 0.18	7.34 \pm 0.14	1.57 \pm 0.02	6.16 \pm 0.25	0.02 \pm 0.00	7.99 \pm 0.20	ND

^a Glucose calculated as percent remaining glucose relative to minimal medium with glucose

^b ND: Not detected; butyrate and fumarate were not detected in any samples

Table S2. Genes putatively involved in the metabolic pathways outlined in Figure 2.

Gene ID	Derived predicted annotation	GenBank Accession	Uniprot Entry	Size (aa)	TH ^a	Locus	TSS ^b	Putative source node	Putative destination node	RPKM EEP ^c	RPKM MEP ^d	RPKM Stat ^e	Reference
BT0309	Dihydrolipoamide dehydrogenase (E3 component, 2-oxoglutarate dehydrogenase complex)	NP 809222	Q8AB03	447		BT0309-BT0313	Yes	2-oxo glutarate	Succinyl-CoA	1.92	3.92	59.63	
BT0310	Lipoate-protein ligase A	NP 809223	Q8AB02	239		BT0309-BT0313		2-oxo glutarate	Succinyl-CoA	1.25	2.69	23.65	
BT0311	Dihydrolipoamide succinyltransferase (E2 component, 2-oxoglutarate dehydrogenase complex)	NP 809224	Q8AB01	456		BT0309-BT0313	Yes	2-oxo glutarate	Succinyl-CoA	2.19	8.57	95.24	
BT0312	2-oxoglutarate decarboxylase (E1 component, 2-oxoglutarate dehydrogenase complex)	NP 809225	Q8AB00	678		BT0309-BT0313		2-oxo glutarate	Succinyl-CoA	2.74	6.21	81.91	
BT0313	Flavodoxin associated with 2-oxoglutarate dehydrogenase complex	NP 809226	Q8AAZ9	171		BT0309-BT0313				2.74	10.65	155.37	
BT0329	2-oxoglutarate/2-oxoacid ferredoxin oxidoreductase subunit gamma	NP 809242	Q8AAY3	180		BT0329-BT0333		2-oxo glutarate	Succinyl-CoA	53.59	68.99	37.83	
BT0330	2-oxoglutarate/2-oxoacid ferredoxin oxidoreductase subunit beta	NP 809243	Q8AAY2	235		BT0329-BT0334	Yes	2-oxo glutarate	Succinyl-CoA	40.46	46.53	26.05	
BT0331	2-oxoglutarate/2-oxoacid ferredoxin oxidoreductase subunit alpha	NP 809244	Q8AAY1	295		BT0329-BT0334		2-oxo glutarate	Succinyl-CoA	43.77	46.77	24.72	
BT0332	2-oxoglutarate/2-oxoacid ferredoxin oxidoreductase subunit alpha	NP 809245		59		BT0329-BT0334		2-oxo glutarate	Succinyl-CoA	51.56	57.49	29.21	
BT0333	2-oxoglutarate/2-oxoacid ferredoxin oxidoreductase subunit delta	NP 809246	Q8AAX9	75		BT0329-BT0334	BT0334	2-oxo glutarate	Succinyl-CoA	65.06	79.51	37.66	
BT0644	Pyruvate phosphate dikinase	NP 809557	Q8AA21	906			Yes	Pyruvate	PEP	11.98	7.82	15.96	
BT0787	Succinyl-CoA synthetase alpha chain	NP 809700	Q8A9M8	286		BT0787-BT0788		Succinate	Succinyl-CoA	0.20	0.59	4.07	
BT0788	Succinyl-CoA synthetase beta chain	NP 809701	Q8A9M7	376		BT0787-BT0788	Yes	Succinate	Succinyl-CoA	0.49	0.62	5.11	
BT1196	Pyruvate carboxylase	NP 810109	Q8A8H1	591			Yes	Pyruvate	Oxaloacetate	1.28	3.59	46.60	
BT1448	Biotin-dependent carboxylase, biotin carboxyl carrier protein	NP 810361	Q8A7S6	174		BT1448-BT1450				0.57	0.83	13.84	

BT1449	Biotin-dependent carboxylase, biotin carboxylase subunit	NP 810362	Q8A7S5	503			BT1448-BT1450				0.59	0.63	13.67	
BT1450	Biotin-dependent carboxylase, carboxytransferase subunit	NP 810363	Q8A7S4	514			BT1448-BT1450	Yes			0.52	0.59	13.28	This work
BT1575	D-lactate dehydrogenase	NP 810488	Q8A7E9	333					Pyruvate	Lactate	2.87	4.99	14.15	This work
BT1685	Methylmalonyl-CoA epimerase	NP 810598	Q8A740	134			BT1685-BT1689	Yes	Methyl malonyl-CoA	Methylmalonyl-CoA	89.81	72.00	27.45	
BT1686	Methylmalonyl-CoA decarboxylase (alpha subunit)	NP 810599	Q8A739	517			BT1685-BT1689		Methyl malonyl-CoA	Propionyl-CoA	96.91	65.63	23.72	This work, (1)
BT1687	Methylmalonyl-CoA decarboxylase (membrane associated subunit)	NP 810600	Q8A738	306	1		BT1685-BT1689		Methyl malonyl-CoA	Propionyl-CoA	87.26	53.66	14.83	This work, (1)
BT1688	Methylmalonyl-CoA decarboxylase (biotin carboxylase component)	NP 810601	Q8A737	144			BT1685-BT1689		Methyl malonyl-CoA	Propionyl-CoA	98.86	65.01	18.77	This work, (1)
BT1689	Methylmalonyl-CoA decarboxylase (beta subunit)	NP 810602	Q8A736	386	10		BT1685-BT1689		Methyl malonyl-CoA	Propionyl-CoA	77.89	62.45	18.84	This work, (1)
BT1696	Oxaloacetate decarboxylase, Na ⁺ transporting (beta subunit)	NP 810609	Q8A729	411	9		BT1696-BT1698		Oxalo acetate	Pyruvate	8.78	7.22	5.11	
BT1697	Oxaloacetate decarboxylase, Na ⁺ transporting (alpha subunit)	NP 810610	Q8A728	609			BT1696-BT1698		Oxalo acetate	Pyruvate	7.89	8.79	6.35	
BT1698	Oxaloacetate decarboxylase, Na ⁺ transporting (gamma subunit)	NP 810611	Q8A727	86	2		BT1696-BT1698	Yes	Oxalo acetate	Pyruvate	13.32	11.87	14.88	
BT1747	Pyruvate ferredoxin oxidoreductase (PFOR)	NP 810660	Q8A6X8	1185					Pyruvate	Acetyl-CoA	74.55	99.46	49.95	(2)
BT1820	Pyruvate oxidase (split by transposable element BT1821)	NP 810733	Q8A6Q6	362			BT1820-BT1822	Yes	Pyruvate	Acetate	0.89	4.10	47.66	This work
BT1822	Pyruvate oxidase (split by transposable element BT1821)	NP 810735	Q8A6Q5	228			BT1820-BT1822		Pyruvate	Acetate	0.24	1.01	10.60	This work
BT1915	Biotin-dependent carboxylase, biotin carboxylase subunit	NP 810828	Q8A6G9	503			BT1915-BT1917	Yes	Pyruvate	Oxalo acetate	73.37	57.03	40.94	
BT1916	Biotin-dependent carboxylase, biotin carboxyl carrier protein	NP 810829	Q8A6G8	169			BT1915-BT1917				81.37	63.85	51.74	
BT1917	Biotin-dependent carboxylase, carboxytransferase subunit	NP 810830	Q8A6G7	511			BT1915-BT1917		Succinate	Propionate	94.73	73.61	62.12	This work
BT1969	Malate dehydrogenase (NADP-dependent)	NP 810882	Q8A6B5	763				Yes	Malate	Pyruvate	14.54	19.15	19.05	This work

BT1972	Phosphoenolpyruvate synthase	NP 810885	Q8A6B3	990			Yes	Pyruvate	PEP	5.27	4.02	3.25	
BT2070	Citrate synthase	NP 810983	Q8A616	447		BT2070-BT2072		Acetyl-CoA	Citrate	8.70	15.50	70.24	
BT2071	Isocitrate dehydrogenase	NP 810984	Q8A615	396		BT2070-BT2072		Isocitrate	2-oxo glutarate	5.89	12.45	63.38	
BT2072	Aconitase	NP 810985	Q8A614	747		BT2070-BT2072		Citrate	Isocitrate	5.54	11.85	54.16	
BT2090	Methylmalonyl-CoA mutase (large subunit)	NP 811003	Q8A5Z6	715		BT2090-BT2091		Succinate	Propionate	17.89	24.16	27.71	This work
BT2091	Methylmalonyl-CoA mutase (small subunit)	NP 811004	Q8A5Z5	633		BT2090-BT2091	Yes	Succinate	Propionate	17.75	22.69	21.46	This work
BT2256	Fumarase, class I, homodimeric	NP 811169	Q8A5I3	544			Yes	Malate	Fumarate	94.12	140.72	79.26	(3)
BT2510	Malate dehydrogenase (NAD-dependent)	NP 811423	Q8A4T7	333			Yes	Oxaloacetate	Malate	24.30	56.09	67.76	
BT2790	Phosphoenolpyruvate carboxykinase	NP 811702	Q8A414	535			Yes	PEP	Oxaloacetate	523.59	766.60	316.40	
BT2836	2-oxoglutarate/2-oxoacid ferredoxin oxidoreductase subunit alpha	NP 811748	Q8A3W8	616		BT2836-BT2837	Yes	2-oxo glutarate	Succinyl-CoA	24.61	24.81	9.56	
BT2837	2-oxoglutarate/2-oxoacid ferredoxin oxidoreductase subunit beta	NP 811749	Q8A3W7	336		BT2836-BT2837		2-oxo glutarate	Succinyl-CoA	23.39	22.28	9.49	
BT2841	Pyruvate kinase	NP 811753	Q8A3W3	485				PEP	Pyruvate	70.72	47.98	19.38	
BT2955	Formate acetyltransferase 2 [Pyruvate formate-lyase]	NP 811867	Q8A3J9	714		BT2955-BT2956		Pyruvate	Formate	0.04	0.07	0.09	This work, (2)
BT2956	Pyruvate formate lyase activating enzyme	NP 811868	Q8A3J8	299		BT2955-BT2956		Pyruvate	Formate	0.05	0.00	0.09	
BT3053	Fumarate reductase (cytochrome b subunit)	NP 811965	Q8A3A1	234	5	BT3053-BT3055	Yes	Fumarate	Succinate	165.87	220.34	63.42	(4)
BT3054	Fumarate reductase (flavoprotein subunit)	NP 811966	Q8A3A0	659		BT3053-BT3055		Fumarate	Succinate	142.68	198.90	58.45	(4)
BT3055	Fumarate reductase (iron sulfur protein subunit)	NP 811967	Q8A399	251		BT3053-BT3055		Fumarate	Succinate	116.09	157.01	49.08	(4)
BT3186	Dihydrolipoamide dehydrogenase (E3 subunit, 2-oxoacid dehydrogenase complex)	NP 812098	Q8A2W9	447			BT3185			5.04	4.76	2.05	
BT3193	Coenzyme A transferase	NP 812105	Q8A2W2	499				Propionyl-CoA	Succinyl-CoA	28.38	21.45	5.09	This work
BT3692	Phosphate acetyltransferase	NP 812603	Q8A1G9	339		BT3692-BT3693	Yes	Acetyl-CoA	Acetyl phosphate	33.06	38.00	7.96	This work
BT3693	Acetate kinase	NP 812604	Q8A1G8	399		BT3692-BT3693		Acetyl phosphate	Acetate	28.12	32.05	8.46	This work
BT3755	Acetyl-coenzyme A synthetase	NP 812666	Q8A1B1	551			BT3757	Acetate	Acetyl-CoA	17.51	11.31	13.59	
BT3911	Malate dehydrogenase (NAD-dependent)	NP 812822	Q8A0W0	313			Yes	Oxaloacetate	Malate	200.37	543.45	473.80	

BT4455	L-lactate dehydrogenase, Fe-S oxidoreductase subunit	NP 813366	Q89ZC2	246		BT4455-BT4457	Yes	Lactate	Pyruvate	22.41	31.12	63.69	This work
BT4456	L-lactate dehydrogenase, Iron-sulfur cluster-binding subunit	NP 813367	Q89ZC1	461		BT4455-BT4457		Lactate	Pyruvate	20.53	25.60	61.81	This work
BT4457	L-lactate dehydrogenase, hypothetical protein subunit	NP 813368	Q89ZC0	193		BT4455-BT4457		Lactate	Pyruvate	15.77	21.79	60.98	This work
BT4737	Pyruvate formate lyase activating enzyme	NP 813648	Q89YJ4	242		BT4737-BT4738		Pyruvate	Formate	39.20	33.02	3.22	
BT4738	Formate acetyltransferase [Pyruvate formate-lyase]	NP 813649	Q89YJ3	742		BT4737-BT4738	Yes	Pyruvate	Formate	186.40	117.01	16.28	This work, (2)

^a TH: Predicted transmembrane helices (IMG)

^b TSS: 5' transcriptional start site (from Theta-Base, (5))

^c RPKM EEP: Normalized gene expression of *Bt* in early exponential phase (Theta-Base, (5))

^d RPKM MEP: Normalized gene expression of *Bt* in mid exponential phase (Theta-Base, (5))

^e RPKM Stat: Normalized gene expression of *Bt* in stationary phase (Theta-Base, (5))

Table S3. Growth summary data for strains grown in rich medium (TYG) and minimal medium (MMG)

Strain	TYG Max OD600 ^a	TYG Specific Rate ^b	MMG Max OD600	MMG Specific Rate
Growth curve summary data from Figures 3-7; n = 3 except WT, n = 6				
WT	1.36 ± 0.06	0.1647 ± 0.0110	1.09 ± 0.11	0.0813 ± 0.0253
ΔBT2955	1.32 ± 0.09	0.1712 ± 0.0065	1.00 ± 0.03	0.0803 ± 0.0290
ΔBT4738	1.26 ± 0.09	0.1619 ± 0.0075	0.98 ± 0.07	0.0668 ± 0.0243
ΔBT2955/4738	1.28 ± 0.09	0.1608 ± 0.0120	0.99 ± 0.08	0.0669 ± 0.0248
ΔBT1575	1.42 ± 0.04	0.1644 ± 0.0213	1.15 ± 0.11	0.0812 ± 0.0348
ΔBT4455	1.42 ± 0.03	0.1593 ± 0.0181	1.15 ± 0.11	0.0802 ± 0.0319
ΔBT1575/4455-7	1.43 ± 0.02	0.1632 ± 0.0155	1.17 ± 0.11	0.0861 ± 0.0357
ΔBT1450	1.34 ± 0.08	0.1672 ± 0.0091	0.99 ± 0.05	0.0758 ± 0.0232
ΔBT1686-9	1.30 ± 0.04	0.1286 ± 0.0093	0.96 ± 0.05	0.0797 ± 0.0184
ΔBT1917	1.35 ± 0.05	0.1816 ± 0.0166	1.02 ± 0.03	0.0814 ± 0.0226
ΔBT2090-1	1.30 ± 0.06	0.1566 ± 0.0217	1.04 ± 0.13	0.0861 ± 0.0410
ΔBT3193	1.26 ± 0.07	0.1410 ± 0.0116	0.33 ± 0.19	0.0059 ± 0.0040
ΔBT1969	1.43 ± 0.02	0.1534 ± 0.0075	1.12 ± 0.14	0.0750 ± 0.0309
ΔBT1820	1.42 ± 0.02	0.1538 ± 0.0063	1.15 ± 0.11	0.0814 ± 0.0285
ΔBT1822	1.39 ± 0.05	0.1502 ± 0.0200	1.20 ± 0.09	0.0846 ± 0.0275
ΔBT1820/1822	1.41 ± 0.02	0.1459 ± 0.0168	1.15 ± 0.07	0.0793 ± 0.0273
Growth curve summary data from Figure 8; n = 3				
WT	1.45 ± 0.02	0.1463 ± 0.0044	0.90 ± 0.05	0.0406 ± 0.0049
P_BfP2E3	1.29 ± 0.02	0.0874 ± 0.0085	0.90 ± 0.08	0.0296 ± 0.0066
P_BfP5E4	1.40 ± 0.02	0.1354 ± 0.0069	0.92 ± 0.03	0.0402 ± 0.0029

^a OD600 = optical density at 600 nm

^b Specific rate: (Maximum - Minimum OD600) / time

Table S4. Organic acid concentrations (mM \pm standard deviation) at 48 hours for the wild-type (WT), various mutant strains and their complements (n = 3)

Strain	Acetate	Formate	Lactate	Malate	Propionate	Succinate
WT	16.48 \pm 0.35	7.55 \pm 0.28	2.09 \pm 0.25	ND ^a	7.74 \pm 0.64	9.47 \pm 0.64
Δ BT1686-9	17.60 \pm 0.28	8.75 \pm 1.26	2.26 \pm 0.32	1.52 \pm 0.08	ND	17.33 \pm 0.42
Comp ^b 1685-9	16.67 \pm 0.19	8.08 \pm 0.43	2.51 \pm 0.63	ND	8.70 \pm 0.26	8.95 \pm 0.77
Δ BT2090-1	16.82 \pm 0.27	10.13 \pm 0.85	2.62 \pm 0.63	1.56 \pm 0.06	ND	17.49 \pm 0.32
Comp 2090-1	17.09 \pm 0.07	7.25 \pm 1.48	1.47 \pm 0.21	ND	8.52 \pm 1.58	8.61 \pm 1.90
Δ BT4738	15.04 \pm 0.07	1.93 \pm 0.66	1.16 \pm 0.06	ND	7.77 \pm 0.92	12.50 \pm 0.16
Comp 4738	16.00 \pm 0.51	6.45 \pm 1.25	1.73 \pm 0.37	0.55 \pm 0.95	9.27 \pm 1.26	9.93 \pm 1.66

^a Not detected. α -ketoglutarate or pyruvate were also not detected in any strain or sample.

^b Comp: Mutant strain from the row above complemented with the listed genes

Table S5. Organic acid concentrations (mM \pm standard deviation) at 48 hours for the wild-type (WT) and Δ BT1969 strains (n = 6)

Strain	Acetate	Formate	Lactate	Malate	Propionate	Succinate
WT	14.62 \pm 0.42	7.72 \pm 0.75	1.83 \pm 0.13	ND ^a	8.76 \pm 0.60	6.11 \pm 0.48
Δ BT1969	13.03 \pm 0.25	9.22 \pm 1.14	1.40 \pm 0.11	1.39 \pm 0.68	8.30 \pm 0.57	8.43 \pm 0.56

^a Not detected. α -ketoglutarate or pyruvate were also not detected in any strain or sample.

Table S6. Organic acid concentrations (mM \pm standard deviation) at 48 hours for the wild-type (WT) and the promoter replacement strains (n = 3)

Strain	Acetate	Formate	Lactate	Malate	Propionate	Succinate
WT	17.43 \pm 0.17	11.60 \pm 0.52	2.26 \pm 0.05	1.60 \pm 0.01	10.92 \pm 0.37	6.84 \pm 0.22
P_BfP2E3	17.13 \pm 0.20	16.76 \pm 14.10	3.13 \pm 1.22	0.64 \pm 1.11	9.46 \pm 0.26	6.79 \pm 1.01
P_BfP5E4	17.12 \pm 0.25	11.26 \pm 0.29	2.44 \pm 0.17	1.54 \pm 0.02	9.89 \pm 0.10	7.23 \pm 0.16

Not detected for any strain or sample: α -ketoglutarate or pyruvate

Table S7. Strains and plasmids used in this study.

Name	Genotype	Ref.	Notes
Strains			
WT <i>Bt</i>	<i>tdk</i> - (ΔBT2275)	(6)	Parent strain of other <i>Bt</i> strains; derived from ATCC 29148/VPI-5482
ΔBT2955	<i>tdk</i> - ΔBT2955		
ΔBT4738	<i>tdk</i> - ΔBT4738		
ΔBT2955/4738	<i>tdk</i> - ΔBT2955/4738		
ΔBT1575	<i>tdk</i> - ΔBT1575		
ΔBT4455-7	<i>tdk</i> - ΔBT4455-7		
ΔBT1575/4455-7	<i>tdk</i> - ΔBT1575/4455-7		
ΔBT1450	<i>tdk</i> - ΔBT1450		
ΔBT1686-9	<i>tdk</i> - ΔBT1686-9	(1)	
ΔBT1917	<i>tdk</i> - ΔBT1917		
ΔBT2090-1	<i>tdk</i> - ΔBT2090-1		
ΔBT3193	<i>tdk</i> - ΔBT3193		
ΔBT1969	<i>tdk</i> - ΔBT1969		
ΔBT1820	<i>tdk</i> - ΔBT1820		
ΔBT1822	<i>tdk</i> - ΔBT1822		
ΔBT1820/1822	<i>tdk</i> - ΔBT1820/1822		
Comp 1685-9	<i>tdk</i> - ΔBT1686-9 ::pNBU2-bla-tetQb::BT1685-9		pNBU2-bla-tetQb chromosomally integrates at one of two tRNAs ^{er} sites in <i>Bt</i>
Comp 2090-1	<i>tdk</i> - ΔBT2090-1 ::pNBU2-bla-tetQb::BT2090-1		pNBU2-bla-tetQb chromosomally integrates at one of two tRNAs ^{er} sites in <i>Bt</i>
Comp 4738	<i>tdk</i> - ΔBT4738 ::pNBU2-bla-tetQb::BT4738		pNBU2-bla-tetQb chromosomally integrates at one of two tRNAs ^{er} sites in <i>Bt</i>
PBT3692::P Bfp2E3	<i>tdk</i> - PBT3692::P Bfp2E3		Promoter replaces 100 bp sequence immediately upstream of BT3692
PBT3692::P Bfp5E4	<i>tdk</i> - PBT3692::P Bfp5E4		Promoter replaces 100 bp sequence immediately upstream of BT3692
<i>Escherichia coli</i> S17-1 <i>λpir</i>			Shuttles plasmids to <i>Bacteroides</i>
<i>Escherichia coli</i> BW27343	<i>pir</i> ⁺ derivative of DH5α		Transformation and maintenance of plasmids; CGSC strain #7924
Plasmids			
pExchange- <i>tdk</i>		(6)	Counters selectable suicide plasmid for gene deletions
pExchange::ΔBT2955			Deletion of BT2955
pExchange::ΔBT4738			Deletion of BT4738
pExchange::ΔBT1575			Deletion of BT1575
pExchange::ΔBT4455-7			Deletion of BT4455-7
pExchange::ΔBT1450			Deletion of BT1450
pExchange::ΔBT1686-9		(1)	Deletion of BT1686-9
pExchange::ΔBT1917			Deletion of BT1917

pExchange::ΔBT2090-1			Deletion of BT2090-1
pExchange::ΔBT3193			Deletion of BT3193
pExchange::ΔBT1969			Deletion of BT1969
pExchange::ΔBT1820			Deletion of BT1820
pExchange::ΔBT1822			Deletion of BT1822
pNBU2-bla-tetQb		(7)	pNBU2-bla-tetQb chromosomally integrates at one of two tRNA ^{ser} sites in <i>Bt</i>
pNBU2-bla-tetQb::BT1685-9			BT1685-9 gene complementation
pNBU2-bla-tetQb::BT2090-1			BT2090-1 gene complementation
pNBU2-bla-tetQb::BT4738			BT4738 gene complementation
pNTP201			Vector for integrating Golden Gate-compatible parts
pNTP202			pExchange2, part A
pNTP203			pExchange2, part B
pNTP221			pExchange2 with PBT3692::PBT1431 for PBT3692 replacement
pNTP224			pExchange2 with PBT3692::BfP3E1/RBS8 for PBT3692 replacement
pNTP225			pExchange2 with PBT3692::BfP2E2/RBS8 for PBT3692 replacement
pNTP226			pExchange2 with PBT3692::P_BfP2E3/RBS8 for PBT3692 replacement
pNTP227			pExchange2 with PBT3692::P_BfP5E4/RBS8 for PBT3692 replacement
pNTP228			pExchange2 with PBT3692::PBT1925 for PBT3692 replacement
pNTP230			pExchange2 with PBT3692::PBT3110 for PBT3692 replacement
pNTP231			pExchange2 with PBT3692::PBT3321 for PBT3692 replacement
pNTP232			pExchange2 with PBT3692::PBT4618 for PBT3692 replacement

Ref.: Reference

Table S8. Primers, promoters and ribosome binding site used in this study.

Name	Notes	Sequence (restriction sites underlined)
Deletion constructs		
BT1450 left F		GCGTCTAGAG <u>CCC</u> CAGATGAACCACATTG
BT1450 left R		TAATAAAATGGAAAATGGAAAGTTG
BT1450 right F		TTTCCATTTTCCATTTATTACATAGGCTGTTT GTATTAATGTTATG
BT1450 right R		GCGG <u>T</u> CGACCATATTTTCAGACGGGCGTAAC
BT1917 left F		GCGTCTAGAGCTTTATCACCAAGAACGG
BT1917 left R		CATGATGCAATCCGTTTTTAGTA
BT1917 right F		TAAAAACGGATTGCATCATGTAAGTTATAG GCGATTACCTATAA
BT1917 right R		GCGG <u>T</u> CGACCGGTCCAATAAGTAGGGTG
BT2090 left F		GCGTCTAGAGACAATCAATGCCAGCGTAG
BT2090 left R		TAAGTACGGAATTTAATCATTTTTTTC
BT2091 right F		ATGATTAAATTCCGTACTIONTAGTCTGCCATAG TTTATTTAAAATTTAAAAC
BT2091 right R		GCGG <u>T</u> CGACGGAGTGTTTCGTCACAGC
BT3193 left F		GCGTCTAGACCTGTAACGAATCAATGACTT G
BT3193 left R		TAAGGCATGAAACGTCTGTC
BT3193 right F		GACAGACGTTTCATGCCTTACATAAATAGTA TAATTTTCGCGTTAAATG
BT3193 right R		GCGG <u>T</u> CGACCTTGAGTTCAGGCATTATTCT G
BT4738 left F		GCGTCTAGACTTTGAATACCTCCGCAGC
BT4738 left R		TAATCTATTACTCTATATGATGATAAACG
BT4738 right F		TCATATAGAGTAATAGATTACATAAAATGA TAAAATTAGAGGTTGTAG
BT4738 right R		GCGG <u>T</u> CGACGTACTIONCACGCTTCTTTCCAG
BT1575 left F		GCGTCTAGACTTGCCCCTAACTGCAAAC
BT1575 left R		CATAATATTTTCTGTTTTTATTCTTATTAATA AACG
BT1575 right F		TAAAAACAGAAAATATTATGTAGAACCCTT ACCTGACTTCCC
BT1575 right R		GCGG <u>T</u> CGACCAATAAGATATGTGAGTCCTT CAATG
BT1820 left F		GCGTCTAGAGTCTGCTGATGCTTGAATAG
BT1820 left R		CATATTTCTAATAGGTTTAGGATTGATTCTT AAT
BT1820 right F		CTAAACCTATTAGAAATATGTAAAAGAGTT CTTTGAAATATTTGAAATTTAGTT
BT1820 right R		GCGG <u>T</u> CGACCTCTGACAACAAAGTAGGCTT C
BT1822 left F		GCGTCTAGAGCATAACATTATATGATGTGGA AACAC
BT1822 left R		CATTAGTGTCCCATTTAAAATTGGGA

BT1822 right outer F	Outer PCR used as template for BT1822 right flank	GCAATCCGGAAGAAGTACTGA
BT1822 right outer R	Outer PCR used as template for BT1822 right flank	GGAATATGTTGACGTCCCTAAGG
BT1822 right F		ATTTTAATGGGACACTAATGTGACAGTTCCT TCTTCAACGA
BT1822 right R		GCGGTCGACGTTTTGTCGGCTGAGGGT
BT2955 left F		GCGTCTAGACCCAGATAATCAATTCCTTTTC CG
BT2955 left R		TAAATCATATTCACTCTAAAAATCTTATATC ATGA
BT2955 right F		TTTAGAGTGAATATGATTTACTCGTAATTCT TCATTTTTTGTATTGTT
BT2955 right R		GCGGTCGACCCATTGTTTCTCTTGATAACG GT
BT1969 left F		GCGTCTAGACCATGCTATGTACTTCCCGATA
BT1969 left R		GGTTATTTTAGCCATGGTATTGGTA
BT1969 right F		ATACCATGGCTAAAATAACCTGAGAAGGCC GTTATCAAAATC
BT1969 right R		GCGGTCGACCGTTTGACTTACCACGTGG
BT4455-57 left F		GCGTCTAGACCGCAATGCTATAAAGTAGAA GTG
BT4455-57 left R		CATAATTATATTACGTAAAATTTAGATGCAT CCT
BT4455-57 right F		TTTTACGTAATATAATTATGGTCATATTGAC GTAACTCCGC
BT4455-57 right R		GCGGTCGACGACCTTTATCGGACAGGAGAC
Complementations		
BT1685-9 F		GCGTCTAGAGCTTTATTCCCAATAATTTAAA TGCA
BT1685-9 R		GCGGTCGACGAGAAGATAACATAATGATAA GTGCG
BT2090-1 F		GCGTCTAGAGCTTTAAGTTGCATGGAGATTC T
BT2090-1 R		GCGGTCGACCATCCCGCCTTCAGCC
BT4738 F		GCGTCTAGACTAATCTTTTCCTAAGGATAGC AATT
BT4738 R		GCGGTCGACATAGAGTAATAGATTACATAC GTTTCGTG
Golden Gate flank primers		
PBT3692 GG left-F		GCGGGTCTCCGTTCTGTAGCTTGTCGGACAA GTG
PBT3692 GG left-R		GCGGGTCTCGTAGATATTCTCCAAGTCTAAA ATATTATAATATTTAACACT
PBT3692 GG right-F		GCGGGTCTCCATGCTTAATTTAATCAACCAA ATCGTG
PBT3692 GG right-R		GCGGGTCTCGCACTTTCTTACCATACTCGGG AGC
qPCR primers		
BT0899 qPCR-F	<i>gyrA</i> as reference gene	CCATGCGTTATACGGAGGCT
BT0899 qPCR-R	<i>gyrA</i> as reference gene	TCACCTTCGGCTCTGTCAAC
BT3692 qPCR-F		CCGCAACACAACAGGCAA
BT3692 qPCR-R		ACACAAGTGATGCCCGGA
BT3693 qPCR-F		ATGCCGGTGCAGTAACATTC

BT3693 qPCR-R		TGATACGCCGGAAATACCCA
Promoters and Ribosome Binding Sites (RBS)		
PBT3692	Native promoter sequence; replaced by other promoters in this study	AAATTAGCCGGAATTGTAATGTGGATATTG CAAATTGAGTACTTTTGCAGCGCAAATTAC AAATCGTAACAAGGTTTATAATATTATTCAA ATATATCT
P BfP3E1	Promoter from pWW3852, (8)	GATAAAACGAAAGGCTCAGTCGAAAGACTG GGCCTTTCGTTTTACAATTGGGCTACCTTTTT TTTGTAATAAAAAAACCCCGCCCCTGACAGG GCGGGGTTTTTTTTTTCACTTGAACCTTCAA ATAATGTTCTTATAAAACCAGTGTCGAAAG AAACAAAGTAG
P BfP2E2	Promoter from pWW3853, (8)	GATAAAACGAAAGGCTCAGTCGAAAGACTG GGCCTTTCGTTTTACAATTGGGCTACCTTTTT TTTGTAATAAAAAAACCCCGCCCCTGACAGG GCGGGGTTTTTTTTTTCACTTGAACCTTCAA ATAATGTTCTTATATATGCAGTGTCGAAAGA AACAAAGTAG
P BfP2E3	Promoter from pWW3818, (8)	GATAAAACGAAAGGCTCAGTCGAAAGACTG GGCCTTTCGTTTTACAATTGGGCTACCTTTTT TTGTTTTGTTTGCAATGGTTAATCTATTGTT AAAATTTAAAGTTTCACTTGAACCTTCAAAT AATGTTCTTATATGTGCAGTGTCGAAAGAA ACAAAGTAG
P BfP5E4	Promoter from pWW3821, (8)	GATAAAACGAAAGGCTCAGTCGAAAGACTG GGCCTTTCGTTTTACAATTGGGCTACCTTTTT TTGTTTTGTTTGCAATGGTTAATCTATTGTT AAAATTTAAAGTTTCACTTGAACCTTCAAAT AATGTTCTTCTATTTGCAGTGTCGAAAGAAA CAAAGTAG
RBS8	RBS used with Whitaker <i>et al.</i> 2017 synthetic promoters, (8)	GATCGTCCATCAATTTAAAATTTAAAAT
PBT1431	Promoter from <i>Bt</i>	GCCAATTCATGTATTTTGCAAAAAATATCAT GTACATTTGTGTTTCAAAAAACAGCAAAAC GC
PBT1925	Promoter from <i>Bt</i>	GGGGTATTAGTATTGGGGAGCTTGCTCTCAA ATAAAAGGTTACAGAATTGACGGTAGTGCG CCTCTGCCGGAATTTGAGGGTAAAATGGTTT AC
PBT3110	Promoter from <i>Bt</i>	GACCAAATTAATAAATTGGAGGGTGACTGT TTTACAACCACCATCCTTTGCTGGTTATTGT TCTAAGTTAGTAATTTTATTCACCTTTACTTT TGCGTTTCAATTGTAAGCATCTTTTTTTTC ATAACTTTGTCCTCTGTATCCTTATTGTCTTT TTCTGTCCGGAATATAACTTAATGATCCGGGA TACTTCATAACAT
PBT3321	Promoter from <i>Bt</i>	AGTCGCAAAGATAAGATTTTTATTACATTTG CAGTGCACATTAACCTTTCTGTTAAC
PBT4618	Promoter from <i>Bt</i>	TAGAGGTGTTTCAGTAAAGAATGTGTTAGTC CAAACAGTCGTTTATCCCCATTTATTATGT TGCCCAATAAATTTTCGTATTAATGAAACTT AAATCTGTCAAGTGGTAAATGAGCGGTTGAT TGGTGGCGGTCAAGCACCTATTTAGCACCTA TAAGCATCCCTATACTTTTGCGGCATAAATT TTAATACGAAGAAAA

References

1. Kovatcheva-Datchary P, Nilsson A, Akrami R, Lee YS, De Vadder F, Arora T, Hallen A, Martens E, Björck I, Bäckhed F. 2015. Dietary fiber-induced improvement in glucose metabolism is associated with increased abundance of *Prevotella*. *Cell Metab* 22:971–982.
2. Khademian M, Imlay JA. 2020. Do reactive oxygen species or does oxygen itself confer obligate anaerobiosis? The case of *Bacteroides thetaiotaomicron*. *Mol Microbiol* 1–15.
3. Lu Z, Sethu R, Imlay JA. 2018. Endogenous superoxide is a key effector of the oxygen sensitivity of a model obligate anaerobe. *Proc Natl Acad Sci U S A* 115:E3266–E3275.
4. Lu Z, Imlay JA. 2017. The fumarate reductase of *Bacteroides thetaiotaomicron*, unlike that of *Escherichia coli*, is configured so that it does not generate reactive oxygen species. *MBio* 8:1–11.
5. Ryan D, Jenniches L, Reichardt S, Barquist L, Westermann AJ. 2020. A high-resolution transcriptome map identifies small RNA regulation of metabolism in the gut microbe *Bacteroides thetaiotaomicron*. *Nat Commun* 11.
6. Bogachev A V., Bertsova Y V., Bloch DA, Verkhovsky MI. 2012. Urocanate reductase: Identification of a novel anaerobic respiratory pathway in *Shewanella oneidensis* MR-1. *Mol Microbiol* 86:1452–1463.
7. Martens EC, Chiang HC, Gordon JI. 2008. Mucosal glycan foraging enhances fitness and transmission of a saccharolytic human gut bacterial symbiont. *Cell Host Microbe* 4:447–57.
8. Whitaker WR, Shepherd ES, Sonnenburg JL. 2017. Tunable expression tools enable single-cell strain distinction in the gut microbiome. *Cell* 169:538-546.e12.