Science Advances

Supplementary Materials for

In vivo mRNA delivery to virus-specific T cells by light-induced ligand exchange of MHC class I antigen-presenting nanoparticles

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Published 23 February 2022, *Sci. Adv.* **8**, eabm7950 (2022) DOI: 10.1126/sciadv.abm7950

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Figs. S1 to S6 Tables S1 and S2



Fig. S1. Synthesis and characterization of antigen-presenting nanoparticles (APNs). (A) Schematic of the generation of APNs using microfluidic mixers and post-insertion technique. (B) Uniform distribution of the synthesized bare nanoparticles (NPs) and APNs functionalized with GP33/D^b APN. (C) Hydrodynamic size, polydispersity index (PDI), zeta potential, and mRNA loading of the produced NPs. Data are presented as mean \pm S.D. n = 3 technical replicates.



Fig. S2. *In vivo* APN-mediated transfection in CD8+ splenocytes isolated from Pmel TCR transgenic mice. ***P < 0.001 between GP33/D^b APNs (non-cognate) and GP100/D^b APNs (cognate); one-way ANOVA and Tukey post-test and correction. All data are means \pm SD; n = 3 biologically independent mice.



Fig. S3. UV tetramers (unexchanged) did not cause non-non-specific binding to CD8+ splenocytes from P14 and Pmel TCR transgenic mice. Representative flow plots of CD8+ splenocytes staining with UV tetramers or UV exchanged cognate tetramers. Frequencies depicted are based on gating on CD8+ cells.



Fig. S4. UV-exchanged tetramers bound to P14 CD8 splenocytes comparably to the folded counterpart. Dot plots of flow cytometry analysis for P14 splenocytes stained with cognate (GP33/D^b) tetramer and non-cognate (GP100/D^b) tetramer prepared by conventional folding protocol (folded) or UV-mediated ligand exchange protocol (UV).



Fig. S5. Biodistribution of DiR-labeled APNs. Quantification of DiR fluorescent signal in each organ. Each symbol indicates one measured organ. All data are means \pm SD; n = 3 biologically independent mice.



Fig. S6. Dose-dependent APN targeting to two immunodominant flu-specific T cells in PR8 model. Infected mice were intravenously injected with a mixture of folded NP366/D^b APNs and PA224/D^b APNs. n.s. = not significant where *P*>0.9999 between PBS and folded APNs (0.03 mg/kg total mRNA dose) as well as between PBS and folded APNs (0.2 mg/kg) in NP366-PA224-T cells; ***P* = 0.0033 between PBS and folded APNs (0.03 mg/kg) in NP366+ flu-specific T cell population (NP366+); *****P* < 0.0001; two-way ANOVA and Sidak post-test and correction. All data are means \pm SD; n = 3 biologically independent mice.

APN formulation	Bare NP	UV PA224/D ^b APN	UV GP33/D ^b APN
pMHC concentration (µg/mL)	13.21	297.97	265.74
pMHC (µg/µg lipids)	0.033	0.745	0.664

Table S1. Quantification of pMHC on APNs using BCA assay kit.

REAGENT							
Antibodies							
Target	Fluorochrome	Clone	Source	Identifier	Working concentration		
Camelid VHH	iFluor488	96A3F5	Genscript	A01862	5 µg/mL		
CD8	APC	53-6.7	Biolegend	100712	$2 \mu g/mL$		
CD8	PE	53-6.7	BD	561095	$2 \mu g/mL$		
CD4	PerCP/Cy5.5	RM4-2	TONBO	65-0042-U100	$2 \mu g/mL$		
NK1.1	PerCP/Cy5.5	PK136	TONBO	65-5941-U100	$2 \mu g/mL$		
B220	PerCP/Cy5.5	RA3-6B2	TONBO	65-0452-U100	$2 \mu g/mL$		
CD31	PerCP/Cy5.5	MEC13.3	Biolegend	102522	$2 \mu g/mL$		
CD45	PE/Cy7	30-F11	Biolegend	103114	$2 \mu g/mL$		
CD11b	BV421	M1/70	Biolegend	101235	$2 \mu g/mL$		
CD11c	BV605	N418	Biolegend	117333	$2 \mu g/mL$		
Ly6c	BV785	HK1.4	Biolegend	128041	$2 \mu g/mL$		
F4/80	BV421	BM8	Biolegend	123131	$2 \mu g/mL$		
Other reagents							
Streptavidin	PE		Invitrogen	S866	10 µg/mL		
Streptavidin	APC		Invitrogen	S868	6.66 µg/mL		
Streptavidin	BV421		Biolegend	405225	2μg/mL		
LIVE/DEAD™ Fixable Aqua Dead Cell Stain Kit			Invitrogen	L34957			

 Table S2. List of staining reagents for flow cytometry analysis.