nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

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Fora	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	$oxed{x}$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🕱 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	🕱 A description of all covariates tested
×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
×	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
X	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
×	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about <u>availability of computer code</u>

Data collection Metamorph 7.7.5

Data analysis Cell Ranger Single Cell Software suite 3.0.2, R(V3.4.3), Seurat (V3.0.2), Signac (V0.2.4), Fiji (V2.0.0), Graphpad (V6.0)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our <u>policy</u>

Single nucleus ATAC-seq data are available in the NCBI Gene Expression Omnibus (GEO) database under accession GSE150065 and 4C-seq data GSE168074

Life sciences study design

All studies must d	lisclose on these points even when the disclosure is negative.			
Sample size	SnATAC-seq experiments have been published already (Dos Santos et al, 2020). For4C-seq experiments, we used more than ten animals per condition. This number of animals is sufficient to obtain statistically significant differences. Significant differences between mean values were evaluated using two-way ANOVA or t-Student tests with Graphpad 6 software.			
Data exclusions	We excluded no data.			
Replication	For all experiments, three biological replicates were used for each condition, giving rise to the same results.			
Randomization	Experiments have been performed randomly. Samples were allocated into experimental groups randomly depending on the condition.			
Blinding	The investigators were not blinded in regard to allocation of samples during experiments and outcome assessment because of technical limitation and impossibility of blinding the investigators. Moreover, the blinding is used mainly in the case of clinical studies where the number and the variations between samples are high. In our study (fundamental research), we use inbred mouse lines reducing the interindividual variability. The different samples were treated a equally as possible.			
Reportir	ng for specific materials, systems and methods			
	tion from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, isted is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.			
Materials & e.	xperimental systems Methods			
n/a Involved in	the study n/a Involved in the study			
X Antibodie	<u></u>			
Palaeont	Palaeontology and archaeology MRI-based neuroimaging			
Animals a	and other organisms			
Human r	esearch participants			
Clinical d	Clinical data			
Dual use	Dual use research of concern			
A sattle a altica				
Antibodies				
Antibodies used	Target Antibody reference Supplier Species dilution			
	Myh7 BA-F8 DHSB mouse IgG2B 1/40.			
	Myh2 SC-71 DHSB mouse IgG1 1/200			
	Myh1 6H1 DHSB mouse IgM 1/40.			
	Myh4 BF-F3 DHSB mouse IgM 1/200 Laminin L9393 Sigma Rabbit 1/500			
	Myh3 BF-45 DHSB mouse IgG1 1/200			
	Myh8 N3.36 DHSB mouse IgG1 1/200			
	Myh13 4A6 DHSB mouse IgM 1/200			
	GFP ab290 Abcam Chicken 1/200			
	Goat anti mouse IgG2b-350 A21140 Invitrogen, 1/500			
	Goat anti mouse IgG1-546 A21123 Invitrogen , 1/1000			
	Goat anti mouse IgM-647 A21238 Invitrogen, 1/1000			

Goat anti rabbit IgG-488 A11008 Invitrogen, 1/1000 Goat anti chicken IgG-488 A11039 Invitrogen, 1/1000

For GFP antibodies ab290 Abcam, PMID: 32123325

PMID: 22530000 for Myh antibodies PMID: 29733324 for Laminin antibodies

Validation

The antibodies have been validated by several studies as specified by:

Animals and other organisms

Policy information about <u>studies involving animals</u>; <u>ARRIVE guidelines</u> recommended for reporting animal research

Laboratory animals 6-8 weeks old C57bl6N females were used in this study. Mice were maintained at temperature 22+/-2 °C, with 30 to 70% humidity

and with a dark/light cycle of 12h/12h.

Wild animals Were used.

Field-collected samples No field collected samples.

Tield collected samples

Ethics oversight

Animal experimentations were carried out in strict accordance with the European STE123 and the French national charter of Ethics of Animal Experimentation. Protocols were approved by the Ethical Committee of Animal experiments of the Institut Cochin, CNRS UMR 8104, INSERM U1016, and by the Ministere de l'Education nationale, de l'enseignement superieur et de la recherche, n° APAFIS

#15699-2018021516569195.

Note that full information on the approval of the study protocol must also be provided in the manuscript.