The American Journal of Human Genetics, Volume 109

Supplemental information

A flexible summary statistics-based colocalization method with application to the mucin cystic fibrosis lung disease modifier locus Fan Wang, Naim Panjwani, Cheng Wang, Lei Sun, and Lisa J. Strug

Supplemental Figures



Figure S1. Empirical power of SS2, SMR, SMR-multi and true positive rates of COLOC and COLOC2 for testing a single gene for colocalization under six alternative scenarios: (A) one shared SNP for GWAS and eQTL study, (B) two independent GWAS SNPs, and one eQTL SNP is shared with the first GWAS SNP, (C) two independent GWAS SNPs, and one eQTL SNP is shared with the second GWAS SNP, (D) two independent eQTL SNPs, one GWAS SNP colocalizes with the eQTL SNP, given the non-overlapped eQTL association is low, (E) two independent eQTL SNPs, one GWAS SNP colocalizes with the eQTL SNP, given the non-overlapped eQTL association is strong, (F) two independent GWAS SNPs, two independent eQTL SNPs and both SNPs are shared. The LD pattern is simulated following the SLC6A14 locus at chromosome X. For SS2, SMR¹ and SMR-multi,² the nominal type 1 error rate is set at alpha = 0.05. For COLOC³ and COLOC2,⁴ the false positive rates are controlled by applying the default 0.8 threshold (as recommended in ⁴) for the colocalization posterior probability. In total, 10⁴ replications are simulated to obtain the empirical power and true positive rates. The x-axis represents parameter values for $\lambda_{T_{c1}}$ or $\lambda_{T_{c2}}$ (3.4, 4.09, 4.45, 5.21, 5.73 and 7.01) such that 0.01, 0.05, 0.1, 0.3, 0.5 and 0.9 power is achieved to detect the eQTL association at significance level 10⁻⁸. The subplot in the bottom right of each plot provides a general visualization of GWAS (orange line) and eQTL (purple line) colocalization patterns in a region of interest.



Figure S2. Heatmaps of colocalization evidence across genes and tissues for a 1Mb region encompassing the peak lung-associated variants at chromosome 3: (A) SS2, (B) SMR, (C) SMR-multi, and (D) COLOC. In each panel, each cell shows the colocalization evidence for the specified tissue and gene calculated from SNPs within 0.1Mb of the peak GWAS variant. The genes on the x-axis are annotated by GENCODE version 26 for hg38 GTEX V8 to 1Mb on either side of the peak GWAS variant and are ordered by their chromosomal positions. Grey indicates insufficient expression levels attained for the gene in the tissue under study. (A): the color intensity corresponds to the SS2 colocalization evidence as measured by $-\log_{10}(SS2 \text{ p-})$

value), with red representing $-\log_{10}(p) = 8.5$ and white representing eQTL evidence for the corresponding gene and tissue does not pass the stage 1 test. (B) and (C): the color intensity corresponds to the SMR and SMR-multi colocalization evidence as measured by -log₁₀(SMR pvalue) and $-\log_{10}(SMR$ -multi p-value), respectively; with red representing $-\log_{10}(p) = 8.5$ and white representing eQTL evidence for the corresponding gene and tissue does not pass the eQTL p-value threshold (5×10^{-8}) . (D): the color intensity corresponds to the COLOC colocalization evidence as measured by colocalization posterior probability (CLPP) ranging from 0 to 1. The eQTL analyses used for all gene/tissue pairs are those conducted by GTEx⁵ version 8 release, except for the HNE eQTLs. eQTL analyses in HNE is conducted using FastQTL⁶ with RNAsequencing of HNE from 94 CF Canadians enrolled in the Canadian CF Gene Modifier study. Esophagus G.J represents Esophagus Gastroesophageal Junction; Esophagus M. represents Esophagus Muscularis; Ileum represents Small Intestine Terminal Ileum; Lymphocytes represents Cells EBV-transformed lymphocytes.



Figure S3. Heatmap of colocalization evidence across genes and tissues for a 1Mb region encompassing the peak lung-associated variants at chromosome 3 by COLOC2. COLOC2 analysis is conducted based on the likelihood from each single gene-by-tissue pair, calculated from SNPs within 0.1Mb of the peak GWAS variants. The color intensity corresponds to the COLOC2 colocalization evidence as measured by colocalization posterior probability (CLPP) ranging from 0 to 1. The genes on the x-axis are annotated by GENCODE version 26 for hg38 GTEX V8 and are ordered by their chromosomal positions. Grey indicates insufficient expression levels attained for the gene in the tissue under study. The eQTL data used for all gene/tissue pairs was sourced from GTEx version 8 release, with exception to the HNE eQTLs. eQTL analysis in HNE is conducted using FastQTL with RNA-sequencing of HNE from 94 CF

Canadians enrolled in the Canadian CF Gene Modifier study. Esophagus G.J represents Esophagus Gastroesophageal Junction; Esophagus M. represents Esophagus Muscularis; Ileum represents Small Intestine Terminal Ileum; Lymphocytes represents Cells EBV-transformed lymphocyte.

Supplemental Tables

Null Scenarios	<i>H</i> ₀₁	<i>H</i> ₀₂	H _{03,(1)}	H _{03,(2)}	H _{04,(1)}	H _{04,(2)}
Parameter	$\lambda_{Z_c} = 0;$	$\lambda_{Z_c} = 0;$	$\lambda_{Z_c} = 6.57;$	$\lambda_{Z_c} = 5.73$	$\lambda_{Z_c} = 6.57;$	$\lambda_{Z_c} = 5.73$
Values	$\lambda_{T_c} = 0$	$\lambda_{T_c} = 5.48$	$\lambda_{T_c} = 0$	$\lambda_{T_c} = 0$	$\lambda_{T_c} = 5.48$	$\lambda_{T_c} = 5.48$

Table S1. Parameter settings under composite null hypothesis when conducting

colocalization tests for a single gene with overlapping samples. Each column corresponds to a specific null scenario when there is no colocalization. The values of λ_{Zc} and λ_{Tc} represent the standardized true effect size of a GWAS associated variant and an eQTL variant, respectively. For example, a GWAS association, λ_{Zc} is set to be 0, 5.73, or 6.57 such that 0, 0.5 or 0.8 power is achieved to detect the signal at the significance level of 10⁻⁸. If there is an eQTL, λ_{Tc} is set to be 5.48 such that 0.4 power is achieved to detect signal at the significance level of 10⁻⁸. H_{01} represents the scenario when there are no SNP-phenotype associations and no eQTL; H_{02} represents the scenario when there are no SNP-phenotype associations but eQTLs are present; $H_{03,(1)}$ and $H_{03,(2)}$ represent two scenarios where SNP-phenotype associations are present but no eQTL; $H_{04,(1)}$ and $H_{04,(2)}$ represent two scenarios where both SNP-phenotype association and eQTL are present, but occur at two independent SNPs. See Section *Simulation Details for Overlapping Samples* for other simulation details.

	Type I error of SS2						
Null Scenarios	Correlation	Correlation	Correlation	Correlation	Correlation		
	=0	=0.3	=0.5	=0.7	=0.9		
H ₀₁	0.0033	0.0031	0.0033	0.0031	0.0030		
H ₀₂	0.0430	0.0426	0.0425	0.0430	0.0425		
H _{03,(1)}	0.0243	0.0271	0.0266	0.0269	0.0259		
H _{03,(2)}	0.0178	0.0202	0.0199	0.0190	0.0202		
H _{04,(1)}	0.0257	0.0281	0.0277	0.0277	0.0269		
H _{04,(2)}	0.0187	0.0208	0.0206	0.0209	0.0217		

Table S2. Empirical type I error rates of SS2 when 50% of the participants in the eQTL study overlap with the GWAS study with varying phenotypic correlations. The LD pattern at the simulated region follows that at the *MUC20/MUC4* locus. Simulations are performed under the null hypothesis of no colocalization and with varying phenotypic correlation (0.3, 0.5, 0.7 and 0.9). For comparison, the type I error rate when no samples are overlapping is also shown (correlation = 0). Table S1 defines the parameters used under each null scenario. The nominal type I error is set at alpha = 0.05. In total, 10⁴ replications are simulated for each null scenario. See Section *Simulation Details for Overlapping Samples* for other simulation details.

Null Comprise	Type I error of SS2						
Null Scenarios	correlation=0.3	correlation=0.5	correlation=0.7	correlation=0.9			
H ₀₁	0.0015	0.0024	0.0038	0.0039			
H ₀₂	0.0373	0.0375	0.0385	0.0382			
H _{03,(1)}	0.0229	0.0242	0.0241	0.0224			
H _{03,(2)}	0.0241	0.0251	0.0254	0.0231			
H _{04,(1)}	0.0183	0.0181	0.0175	0.0174			
H _{04,(2)}	0.0188	0.0186	0.0176	0.0185			

Table S3. Empirical type I error rates of SS2 when 100% of the participants in theeQTL study overlap with the GWAS study, with varying phenotypic correlations.

The LD pattern at the simulated region follows that at the MUC20/MUC4 locus.

Simulations are performed under the null hypothesis of no colocalization and with varying phenotypic correlation (0.3, 0.5, 0.7 and 0.9). Table S1 defines the parameters used under each null scenario. The nominal type I error is set at alpha = 0.05. In total, 10^4 replications are simulated for each null scenario. See Section *Simulation Details for Overlapping Samples* for other simulation details.

NT 11	Type I error of SS2						
Null	eQTL sample	eQTL sample	eQTL sample	eQTL sample	eQTL sample		
Scenarios	size =200	size =200	size =300	size =400	size =500		
H ₀₁	0.0037	0.0040	0.0053	0.0065	0.0086		
H ₀₂	0.0495	0.0478	0.0487	0.0504	0.0493		
H _{03,(1)}	0.0266	0.0265	0.0227	0.0255	0.0243		
H _{03,(2)}	0.0288	0.0274	0.0233	0.0263	0.0255		
H _{04,(1)}	0.0199	0.0165	0.0222	0.0202	0.0261		
H _{04,(2)}	0.0190	0.0166	0.0218	0.0210	0.0255		

Table S4. Empirical type I error rates of SS2 when 100% of the participants in the eQTL study overlap with the GWAS study, with phenotypic correlation 0.5, and varying sample sizes of the eQTL study. The LD pattern at the simulated region follows that at the *MUC20/MUC4* locus. Simulations are performed under the null hypothesis of no colocalization and with varying eQTL sample size (100, 200, 300, 400 or 500). Table S1 defines the parameters used under each null scenario. The nominal type I error is set at alpha = 0.05. In total, 10^4 replications are simulated for each null scenario. See Section *Simulation Details for Overlapping Samples* for other simulation details.

	Type I error of SS2						
Null Scenarios	eQTL sample	eQTL sample	eQTL sample	eQTL sample	eQTL sample		
	size=100	size =200	size =300	size =400	size =500		
H ₀₁	0.0037	0.009	0.0122	0.0177	0.0216		
H ₀₂	0.0495	0.0502	0.0122	0.0572	0.0576		
H _{03,(1)}	0.0266	0.0258	0.0122	0.024	0.0252		
H _{03,(2)}	0.0288	0.0276	0.0122	0.0262	0.0262		
H _{04,(1)}	0.0199	0.0185	0.0122	0.0209	0.0296		
H _{04,(2)}	0.019	0.019	0.0122	0.0211	0.0304		

Table S5. Empirical type I error rates of SS2 when 100% of the participants in the eQTL study overlap with the GWAS study, with strong phenotypic correlation 0.9, and varying sample sizes of the eQTL study. The LD pattern at the simulated region follows that at the *MUC20/MUC4* locus. Simulations are performed under the null hypothesis of no colocalization and with varying eQTL sample size (100, 200, 300, 400 or 500). Table S1 defines the parameters used under each null scenario. The nominal type I error is set at alpha = 0.05. In total, 10⁴ replications are simulated for each null scenario. See Section *Simulation Details for Overlapping Samples* for other simulation details.

Null Scenarios	Parameter Values
H ₀₁	$\lambda_{Z_c} = 0; \ \lambda_{T_c} = 0$
H ₀₂	$\lambda_{Z_c} = 0; \ \lambda_{T_c} = 7.01$
H ₀₃	$\lambda_{Z_c} = 5.73; \ \lambda_{T_c} = 0$
H ₀₄	$\lambda_{Z_c} = 5.73; \ \lambda_{T_c} = 7.01$

Table S6. Parameter settings under composite null hypothesis when conducting colocalization tests for a single gene with independent samples. Each row corresponds to a specific null scenario when there is no colocalization. H_{01} represents the scenario when there is no SNP-phenotype association and no eQTL; H_{02} represents the scenario when there is no SNP-phenotype association but eQTLs are present; H_{03} represents the scenario where SNP-phenotype associations are present but no eQTL; H_{04} represents the scenario where both SNP-phenotype association and eQTL are present, but occur at two independent SNPs. Values of λ_{Z_c} and λ_{T_c} represent the standardized true effect size of a GWAS associated variant and an eQTL variant, respectively. GWAS association, λ_{Z_c} is set to be 5.73 such that 0.5 power is achieved to detect the signal at the significance level of 10⁻⁸. If there is an eQTL, λ_{T_c} is set to be 7.01 such that 0.9 power is achieved to detect the signal at the significance level of 10⁻⁸. See Section Simulation Details for a Single Gene-by-Tissue Pair for other simulation details.

Scenarios	Parameter Values
Scenario 1: one shared SNP for GWAS and eQTL study (Figure 2A and Figure S1A)	$\lambda_{Z_{c1}} = 6.57; \ \lambda_{Z_{c2}} = 0$ $\lambda_{T_{c1}} = 3.40, 4.45, 5.21, 5.73, 6.25 \text{ or } 7.01;$ $\lambda_{T_{c2}} = 0$
Scenario 2: two independent GWAS SNPs, and one eOTL SNP is shared with the first GWAS SNP	$\lambda_{Z_{c1}} = 6.57; \ \lambda_{Z_{c2}} = 5.73$ $\lambda_T = 3.40, 4.45, 5.21, 5.73, 6.25 \text{ or } 7.01;$
(Figure 2B and Figure S1B)	$\lambda_{T_{c2}} = 0$
Scenario 3: two independent GWAS SNPs, and one	$\lambda_{Z_{c1}} = 6.57; \ \lambda_{Z_{c2}} = 5.73$
eQTL SNP is shared with the second GWAS SNP	$\lambda_{T_{c1}} = 0;$
(Figure 2C and Figure S1C)	$\lambda_{T_{c2}} = 3.40, 4.45, 5.21, 5.73, 6.25 \text{ or } 7.01$
Scenario 4: two independent eQTL SNPs, one GWAS SNP colocalizes with the eQTL SNP; the	$\lambda_{Z_{c1}} = 6.57; \ \lambda_{Z_{c2}} = 0$
non-overlapping eQTL association is mild (Figure 2D and Figure S1D)	$\lambda_{T_{c1}} = 3.40, 4.45, 5.21, 5.73, 6.25 \text{ or } 7.01;$ $\lambda_{T_{c2}} = 5.73$
Scenario 5: two independent eQTL SNPs, one	
GWAS SNP co-localizes with the eQTL SNP; the	$\lambda_{Z_{c1}} = 6.57; \ \lambda_{Z_{c2}} = 0$
non-overlapping eQTL association is strong (Figure	$\Lambda_{T_{c1}} = 3.40, 4.45, 5.21, 5./3, 6.25 \text{ or } 7.01;$
2E and Figure S1E)	$n_{T_{c2}} - 7.01$

Scenario 6: two independent GWAS SNPs, two	$\lambda_{Z_{c1}} = 6.57; \ \lambda_{Z_{c2}} = 5.73$
independent eQTL SNPs and both SNPs are shared	$\lambda_{T_{c1}} = 3.40, 4.45, 5.21, 5.73, 6.25 \text{ or } 7.01;$
(Figure 2F and Figure S1F)	$\lambda_{T_{c2}} = 7.01$

Table S7. Parameter settings of the six alternative scenarios simulated to access the power/true positive rate of different methods. $\lambda_{Z_{c1}}$ and $\lambda_{Z_{c2}}$ denote the standardized true effect sizes of two GWAS associated variants, while $\lambda_{T_{c1}}$ and $\lambda_{T_{c2}}$ denote the standardized true effect sizes of two eQTL variants. For the GWAS associated variants, $\lambda_{Z_{c1}}$ is set to be 6.57 such that 0.8 power is achieved to detect the GWAS signal at the significance level 10⁻⁸; $\lambda_{Z_{c2}}$ is set to be 0 or 5.73 such that 0 or 0.5 power is achieved to detect the GWAS signal at the significance level 10⁻⁸. For the eQTL variants, $\lambda_{T_{c1}}$ is set to be 0, 3.4, 4.09, 4.45, 5.21, 5.73 or 7.01 such that 0, 0.01, 0.05, 0.1, 0.3, 0.5 or 0.9 power is achieved to detect the eQTL signal at the significance level 10⁻⁸; same for the value of $\lambda_{T_{c2}}$. See Section *Simulation Details for a Single Gene-by-Tissue Pair* for other simulation details.

	Proportion of		FWER of SS2			
Locus	genes with eQTL association but do not colocalize	100 genes	200 genes	300 genes	400 genes	500 genes
	0%	0.0352	0.0348	0.0364	0.0367	0.0370
	20%	0.0240	0.0272	0.0274	0.0283	0.0283
MUC20/MUC4	40%	0.0247	0.0267	0.0274	0.0291	0.0292
MUC20/MUC4	60%	0.0213	0.0227	0.0253	0.0279	0.0291
	80%	0.0194	0.0223	0.0258	0.0293	0.0309
	100%	0.0213	0.0270	0.0293	0.0311	0.0332
	0%	0.0058	0.0055	0.0055	0.0053	0.0054
	20%	0.0050	0.0031	0.0025	0.0020	0.0018
SLC6414	40%	0.0032	0.0021	0.0016	0.0014	0.0012
SLC0A14	60%	0.0026	0.0017	0.0014	0.0012	0.0010
	80%	0.0021	0.0014	0.0011	0.0010	0.0009
	100%	0.0018	0.0012	0.0010	0.0009	0.0008

 Table S8. Empirical family-wise error rates (FWERs) of SS2 with different number

 of genes. The LD pattern at the simulated region follows that at the *MUC20/MUC4* and

 SLC6A14 loci. Simulations are conducted using a different number of genes (columns)

and a varying proportion of genes with eQTL association (rows). The eQTL peaks are randomly generated from 6 different intervals (50%-60%, 60%-70%, 70%-80%, 80%-90%, 90%-95%, 95%-100% power is achieved to detect the eQTL association at the significance level of 10⁻⁸) with probabilities defined by the proportion of the log10(maximum eQTL p-value) within each interval observed at the corresponding locus. The height of the GWAS peak is set at 5.06 on the -log10p scale such that 10% power is achieved to detect the GWAS association at the significance level of 10⁻⁸. None of the eQTL peaks colocalize with the GWAS peak for FWER evaluation. In total, 10⁵ replications are simulated to evaluate FWER of 0.05. The empirical FWER is calculated by counting the proportion of 10⁵ replications where at least one gene has a false colocalization claim. See Section *Simulation Details for Multiple Genes-Tissue Pairs* for other simulation details.

	Proportion of genes	FWER of SMR				
Locus	with eQTL association but do not colocalize	100 genes	200 genes	300 genes	400 genes	500 genes
	0%	0.0002	0.0004	0.0006	0.0007	0.0008
	20%	0.0021	0.0011	0.0008	0.0006	0.0004
MUC20/MUC4	40%	0.0011	0.0005	0.0003	0.0002	0.0002
MOC20/MOC4	60%	0.0006	0.0003	0.0002	0.0002	0.0001
	80%	0.0005	0.0003	0.0002	0.0001	0.0001
	100%	0.0004	0.0002	0.0002	7.00x10 ⁻⁵	7.00x10 ⁻⁵
	0%	0.0002	0.0003	0.0005	0.0005	0.0007
	20%	0.0052	0.0023	0.0016	0.0012	0.0009
	40%	0.0024	0.0012	0.0008	0.0006	0.0005
SLC6A14	60%	0.0016	0.0008	0.0005	0.0004	0.0003
	80%	0.0012	0.0006	0.0004	0.0002	0.0002
	100%	0.0009	0.0004	0.0003	0.0002	0.0001

Table S9. Empirical family-wise error rates (FWERs) of SMR with different

number of genes. The LD pattern at the simulated region follows that at the

MUC20/MUC4 and *SLC6A14* loci. SMR is conducted under the default setting such that

a SNP is picked if only if the eQTL p-value is less than 5×10^{-8} . Simulations are conducted using a different number of genes (columns) and a varying proportion of genes with eQTL association (rows). The eQTL peaks are randomly generated from 6 different intervals (50%-60%, 60%-70%, 70%-80%, 80%-90%, 90%-95%, 95%-100% power is achieved to detect the eQTL association at the significance level of 10⁻⁸) with probabilities defined by the proportion of the -log10(maximum eQTL p-value) within each interval observed at the corresponding locus. The height of the GWAS peak is set at 5.06 on the -log10p scale such that 10% power is achieved to detect the GWAS association at the significance level of 10⁻⁸. None of the eQTL peaks colocalize with the GWAS peak for FWER evaluation. In total, 10⁵ replications are simulated to evaluate FWER of 0.05. The empirical FWER is calculated by counting the proportion of 10^5 replications where at least one gene has a false colocalization claim. See Section Simulation Details for Multiple Genes-Tissue Pairs for other simulation details.

	Proportion of	FWER of SMR-multi				
Locus	genes with eQTL association but do not colocalize	100 genes	200 genes	300 genes	400 genes	500 genes
	0%	0.0002	0.0004	0.0006	0.0007	0.0008
	20%	0.0015	0.0006	0.0004	0.0003	0.0002
	40%	0.0006	0.0003	0.0002	0.0001	0.0001
MUC20/MUC4	60%	0.0003	0.0002	0.0001	9.00x10 ⁻⁵	7.00x10 ⁻⁵
	80%	0.0002	0.0002	8.00x10 ⁻⁵	6.00x10 ⁻⁵	6.00x10 ⁻⁵
	100%	0.0003	0.0002	7.00x10 ⁻⁵	6.00x10 ⁻⁵	5.00x10 ⁻⁵
	0%	0.0002	0.0004	0.0005	0.0005	0.0007
	20%	0.0035	0.0015	0.0009	0.0007	0.0005
	40%	0.0014	0.0007	0.0004	0.0003	0.0002
SLC6A14	60%	0.0009	0.0004	0.0002	0.0002	0.0002
	80%	0.0007	0.0003	0.0002	0.0002	0.0002
	100%	0.0004	0.0002	0.0002	0.0001	0.0001

Table S10. Empirical family-wise error rates (FWERs) of Multi-SNP-based SMR

test (SMR-multi) with different number of genes. The LD pattern at the simulated region follows that at the *MUC20/MUC4* and *SLC6A14* loci. Multi-SNP-based SMR test

(SMR-multi) is conducted under the default setting such that a SNP is picked if only if the eQTL p-value is less than 5×10^{-8} . Simulations are conducted using a different number of genes (columns) and a varying proportion of genes with eQTL association (rows). The eQTL peaks are randomly generated from 6 different intervals (50%-60%, 60%-70%, 70%-80%, 80%-90%, 90%-95%, 95%-100% power is achieved to detect the eQTL association at the significance level of 10^{-8}) with probabilities defined by the proportion of the -log10(maximum eQTL p-value) within each interval observed at the corresponding locus. The height of the GWAS peak is set at 5.06 on the -log10p scale such that 10% power is achieved to detect the GWAS association at the significance level of 10⁻⁸. None of the eQTL peaks colocalize with the GWAS peak for FWER evaluation. In total, 10⁵ replications are simulated to evaluate FWER of 0.05. The empirical FWER is calculated by counting the proportion of 10^5 replications where at least one gene has a false colocalization claim. See Section Simulation Details for Multiple Genes-Tissue Pairs for other simulation details.

	Proportion of	False positive rate of COLOC				
Locus	genes with eQTL association but do not colocalize	100 genes	200 genes	300 genes	400 genes	500 genes
	0%	0.0265	0.0489	0.0704	0.0912	0.1118
	20%	0.0223	0.0406	0.0587	0.0758	0.0927
MUC20/MUC4	40%	0.0180	0.0320	0.0460	0.0597	0.0729
MUC20/MUC4	60%	0.0134	0.0232	0.0330	0.0424	0.0515
	80%	0.0088	0.0142	0.0197	0.0252	0.0300
	100%	0.0009	0.0017	0.0022	0.0030	0.0037
	0%	0.0350	0.0634	0.0902	0.1157	0.1400
	20%	0.0269	0.0509	0.0735	0.0952	0.1162
	40%	0.0218	0.0408	0.0592	0.0766	0.0938
SLC6A14	60%	0.0163	0.0304	0.0441	0.0574	0.0703
	80%	0.0111	0.0201	0.0289	0.0373	0.0458
	100%	0.0057	0.0093	0.0128	0.0164	0.0198

Table S11. Empirical false positive rates of COLOC with different number of genes.

The LD pattern at the simulated region follows that at the *MUC20/MUC4* and *SLC6A14* loci. Simulations are conducted using a different number of genes (columns) and a

varying proportion of genes with eQTL association (rows). The eQTL peaks are randomly generated from 6 different intervals (50%-60%, 60%-70%, 70%-80%, 80%-90%, 90%-95%, 95%-100% power is achieved to detect the eQTL association at the significance level of 10^{-8}) with probabilities defined by the proportion of the log10(maximum eQTL p-value) within each interval observed at the corresponding locus. The height of the GWAS peak is set at 5.06 on the -log10p scale such that 10% power is achieved to detect the GWAS association at the significance level of 10⁻⁸. None of the eQTL peaks colocalize with the GWAS peak for FWER evaluation. In total, 10⁵ replications are simulated to evaluate the false positive rates by applying the 0.8 threshold (as recommended by ⁴) for the colocalization posterior probability. The empirical false positive rate for COLOC is calculated by counting the proportion of 10^5 replications where at least one gene has a false colocalization claim. See Section Simulation Details for Multiple Genes-Tissue Pairs for other simulation details.

	Proportion of	False positive rate of COLOC2					
Logus	genes with eQTL						
Locus	association but do	100 genes	200 genes	300 genes	400 genes	500 genes	
	not colocalize						
	0%	0.0020	0.0034	0.0048	0.0061	0.0073	
	20%	0.0023	0.0023	0.0023	0.0023	0.0023	
MUCDO/MUCA	40%	0.0025	0.0026	0.0027	0.0027	0.0027	
MUC20/MUC4	60%	0.0033	0.0033	0.0033	0.0034	0.0034	
	80%	0.0038	0.0040	0.0040	0.0040	0.0040	
	100%	0.0044	0.0044	0.0044	0.0044	0.0044	
	0%	2.00x10 ⁻⁵	5.00x10 ⁻⁵	9.00x10 ⁻⁵	0.0001	0.0002	
	20%	0.0046	0.0047	0.0047	0.0048	0.0049	
SLC6A14	40%	0.0054	0.0055	0.0055	0.0056	0.0056	
	60%	0.0068	0.0069	0.0070	0.0070	0.0070	
	80%	0.0079	0.0079	0.0080	0.0080	0.0080	
	100%	0.0088	0.0088	0.0089	0.0089	0.0089	

Table S12. Empirical false positive rates of COLOC2 with different number of

genes. The LD pattern at the simulated region follows that at the *MUC20/MUC4* and *SLC6A14* loci. Simulations are conducted using a different number of genes (columns)

and a varying proportion of genes with eQTL association (rows). The eQTL peaks are randomly generated from 6 different intervals (50%-60%, 60%-70%, 70%-80%, 80%-90%, 90%-95%, 95%-100% power is achieved to detect the eQTL association at the significance level of 10⁻⁸) with probabilities defined by the proportion of the -log10(maximum eQTL p-value) within each interval observed at the corresponding locus. The height of the GWAS peak is set at 5.06 on the -log10p scale such that 10% power is achieved to detect the GWAS association at the significance level of 10⁻⁸. None of the eQTL peaks colocalize with the GWAS peak for FWER evaluation. In total, 10⁵ replications are simulated to evaluate the false positive rates by applying the 0.8 threshold (as recommended by ⁴) for the colocalization posterior probability. The empirical false positive rates for COLOC2 are calculated by counting the proportions of 10⁵ replications where at least one gene has a false colocalization claim. See Section Simulation Details for Multiple Genes-Tissue Pairs for other simulation details.

	eQTL height	Power of SS2					
	for the 5% of						
Locus	genes that	100 genes	200 genes	300 genes	400 genes	500 genes	
	colocalize						
	5.48-5.73	0.8880	0.8668	0.8508	0.8379	0.8268	
	5.73-5.98	0.8891	0.8671	0.8512	0.8381	0.8269	
MUC20/MUC4	5.98-6.25	0.8893	0.8672	0.8508	0.8376	0.8261	
	6.25-6.57	0.8891	0.8667	0.8499	0.8368	0.8255	
	6.57-7.01	0.8885	0.8658	0.8488	0.8352	0.8235	
SLC6A14	5.48-5.73	0.7126	0.7104	0.7018	0.6938	0.6855	
	5.73-5.98	0.7450	0.7337	0.7200	0.7090	0.6992	
	5.98-6.25	0.7642	0.7468	0.7306	0.7183	0.7076	
	6.25-6.57	0.7752	0.7547	0.7376	0.7247	0.7138	
	6.57-7.01	0.7819	0.7601	0.7424	0.7295	0.7184	

Table S13. Power of SS2 with different number of genes. The LD pattern at the

simulated region follows that at the *MUC20/MUC4* and *SLC6A14* loci. The height of the GWAS peak is set at 5.06 on the -log10p scale such that 10% power is achieved to detect the GWAS association at the significance level of 10⁻⁸. Simulations are conducted using a different number of genes (columns) and a varying range of the eQTL height for the 5%

of genes that colocalize (rows). The eQTL peaks are set with 5 different intervals such that 40%-50%, 50%-60%, 60%-70%, 70%-80%, 80%-90% power is achieved to detect the eQTL association at the significance level of 10^{-8} . The 95% of genes that do not colocalize are simulated under a mixture of H_{03} and H_{04} , and details are demonstrated in the Supplemental Methods. The definitions of H_{03} and H_{04} are provided in Table S6. In total, 10^5 replications are simulated to evaluate power at 0.05 significance level. The power is calculated by counting the proportion of 10^5 replications where at least one gene is correctly identified with colocalization. See Section *Simulation Details for Multiple Genes-Tissue Pairs* for other simulation details.

	eQTL height	Power of SMR					
_	for the 5% of						
Locus	genes that	100 genes	200 genes	300 genes	400 genes	500 genes	
	colocalize						
	5.48-5.73	0.8299	0.7907	0.7540	0.7257	0.7015	
MUC20/MUC4	5.73-5.98	0.8467	0.7993	0.7633	0.7359	0.7126	
	5.98-6.25	0.8561	0.8072	0.7717	0.7448	0.7225	
	6.25-6.57	0.8633	0.8149	0.7808	0.7542	0.7322	
	6.57-7.01	0.8704	0.8234	0.7906	0.7646	0.7435	
	5.48-5.73	0.8005	0.7486	0.7067	0.6749	0.6519	
SLC6A14	5.73-5.98	0.8184	0.7599	0.7186	0.6879	0.6652	
	5.98-6.25	0.8292	0.7699	0.7293	0.6995	0.6767	
	6.25-6.57	0.8379	0.7796	0.7398	0.7102	0.6886	
	6.57-7.01	0.8449	0.7886	0.7504	0.7213	0.7005	

Table S14. Power of SMR with different number of genes. The LD pattern at the simulated region follows that at the *MUC20/MUC4* and *SLC6A14* loci. SMR is conducted under the default setting such that a SNP is picked if only if the eQTL p-value is less than $5x10^{-8}$. The height of the GWAS peak is set at 5.06 on the -log10p scale such

that 10% power is achieved to detect the GWAS association at the significance level of

10⁻⁸. Simulations are conducted using a different number of genes (columns) and a varying range of the eQTL height for the 5% of genes that colocalize (rows). The eQTL peaks are set with 5 different intervals such that 40%-50%, 50%-60%, 60%-70%, 70%-80%, 80%-90% power is achieved to detect the eQTL association at the significance level of 10⁻⁸. The 95% of genes that do not colocalize are simulated under a mixture of H_{03} and H_{04} , and details are demonstrated in the Supplemental Methods. The definitions of H_{03} and H_{04} are provided in Table S6. In total, 10⁵ replications are simulated to evaluate power at 0.05 significance level. The power is calculated by counting the proportion of 10⁵ replications where at least one gene has correctly identified with colocalization. See Section *Simulation Details for Multiple Genes-Tissue Pairs* for other simulation details.

	eQTL height	Power of SMR-multi					
T	for the 5% of						
Locus	genes that	100 genes	200 genes	300 genes	400 genes	500 genes	
	colocalize						
MUC20/MUC4	5.48-5.73	0.7955	0.7432	0.6964	0.6615	0.6316	
	5.73-5.98	0.8077	0.7477	0.7009	0.6655	0.6363	
	5.98-6.25	0.8133	0.7510	0.7050	0.6702	0.6410	
	6.25-6.57	0.8162	0.7536	0.7074	0.6734	0.6453	
	6.57-7.01	0.8161	0.7542	0.7094	0.6758	0.6481	
	5.48-5.73	0.7659	0.7003	0.6483	0.6077	0.5785	
SLC6A14	5.73-5.98	0.7789	0.7068	0.6548	0.6155	0.5859	
	5.98-6.25	0.7850	0.7118	0.6604	0.6215	0.5923	
	6.25-6.57	0.7883	0.7154	0.6652	0.6266	0.5983	
	6.57-7.01	0.7884	0.7166	0.6686	0.6311	0.6036	

Table S15. Power of Multi-SNP-based SMR test (SMR-multi) with different number of genes. The LD pattern at the simulated region follows that at the *MUC20/MUC4* and *SLC6A14* loci. SMR-multi is conducted under the default setting such that a SNP is picked if only if the eQTL p-value is less than 5×10^{-8} . The height of the GWAS peak is set at 5.06 on the -log10p scale such that 10% power is achieved to detect the GWAS

association at the significance level of 10^{-8} . Simulations are conducted using a different number of genes (columns) and a varying range of the eQTL height for the 5% of genes that colocalize (rows). The eQTL peaks are set with 5 different intervals such that 40%-50%, 50%-60%, 60%-70%, 70%-80%, 80%-90% power is achieved to detect the eQTL association at the significance level of 10^{-8} . The 95% of genes that do not colocalize are simulated under a mixture of H_{03} and H_{04} , and details are demonstrated in the Supplemental Methods. The definitions of H_{03} and H_{04} are provided in Table S6. In total, 10^5 replications are simulated to evaluate power at 0.05 significance level. The power is calculated by counting the proportion of 10^5 replications where at least one gene is correctly identified with colocalization. See Section *Simulation Details for Multiple Genes-Tissue Pairs* for other simulation details.

	eQTL height	True positive rate of COLOC2					
	for the 5% of						
Locus	genes that	100 genes	200 genes	300 genes	400 genes	500 genes	
	colocalize						
	5.48-5.73	0.4976	0.5516	0.5783	0.5968	0.6105	
	5.73-5.98	0.5193	0.5690	0.5942	0.6118	0.6249	
MUC20/MUC4	5.98-6.25	0.5353	0.5817	0.6051	0.6219	0.6343	
	6.25-6.57	0.5477	0.5908	0.6129	0.6287	0.6405	
	6.57-7.01	0.5581	0.5981	0.6184	0.6333	0.6439	
	5.48-5.73	0.5801	0.6258	0.6407	0.6494	0.6555	
SLC6A14	5.73-5.98	0.5979	0.6364	0.6497	0.6581	0.6638	
	5.98-6.25	0.6118	0.6443	0.6565	0.6643	0.6697	
	6.25-6.57	0.6232	0.6516	0.6625	0.6693	0.6745	
	6.57-7.01	0.6330	0.6572	0.6669	0.6734	0.6781	

Table S16. True positive rates of COLOC2 with different number of genes. The LD pattern at the simulated region follows that at the *MUC20/MUC4* and *SLC6A14* loci. The height of the GWAS peak is set at 5.06 on the -log10p scale such that 10% power is achieved to detect the GWAS association at the significance level of 10⁻⁸. Simulations are conducted using a different number of genes (columns) and a varying range of the eQTL

height for the 5% of genes that colocalize (rows). The eQTL peaks are set with 5 different intervals such that 40%-50%, 50%-60%, 60%-70%, 70%-80%, 80%-90% power is achieved to detect the eQTL association at the significance level of 10^{-8} . The 95% of genes that do not colocalize are simulated under a mixture of H_{03} and H_{04} , and details are demonstrated in the Supplemental Methods. The definitions of H_{03} and H_{04} are provided in Table S6. In total, 10^5 replications are simulated to evaluate the true positive rates by applying the 0.8 threshold (as recommended by ⁴) for the colocalization posterior probability. The true positive rates for COLOC2 are calculated by counting the proportion of 10^5 replications where at least one gene is correctly identified with colocalization. See Section *Simulation Details for Multiple Genes-Tissue Pairs* for other simulation details.

Null Scenarios	False positive rate of COLOC2						
	eQTL sample size =	eQTL sample size =	eQTL sample size =	eQTL sample size =			
	200	500	1000	2000			
H ₀₁	0.0022	0.0025	0.0024	0.0021			
H ₀₂	0.0468	0.0468	0.0467	0.0467			
H ₀₃	0.0880	0.0692	0.0550	0.0417			
H ₀₄	0.0012	0.0012	0.0012	0.0012			

Table S17. False positive rates of COLOC2 for a single hypothesis test with different sample sizes for the eQTL study. The LD pattern at the simulated region follows that at the *SLC6A14* locus. Sample size is fixed at 2000 participants in the GWAS study, and is varied for the eQTL study (columns). Each row corresponds to a specific null scenario when there is no colocalization. See Table S6 for the explanations of each null scenario with the corresponding parameter settings. The false positive rates are calculated by applying the 0.8 threshold (as recommended by ⁴) for the colocalization posterior probability. In total, 10⁴ replications are simulated for each null scenario. See Section *Simulation Details for a Single Gene-by-Tissue Pair* for other simulation details.

Null Scenarios	Type I err	or of SMR	Type I error of SMR-multi		
	(eQTL p<0.01)	(eQTL p<0.05)	(eQTL p<0.01)	(eQTL p<0.05)	
H ₀₁	0.0067	0.0071	0.003	6.00x10 ⁻⁴	
H ₀₂	0.0392	0.0392	0.0162	0.006	
H ₀₃	0.0766	0.0934	0.0843	0.0807	
H ₀₄	0.0404	0.0404	0.0483	0.0606	

Table S18. Empirical type I error rates of SMR and Multi-SNP-based SMR test

(SMR-multi) for a single hypothesis test under eQTL p-value thresholds 0.01 and

0.05. SMR and Multi-SNP-based SMR test (SMR-multi) are conducted under the setting such that a SNP is picked if only if the eQTL p-value is less than 0.01(eQTL p<0.01) or 0.05 (eQTL p<0.05). The LD pattern at the simulated region follows that at the *MUC20/MUC4* locus. Each row corresponds to a specific null scenario when there is no colocalization. Table S6 defines the parameters used under each null scenario. The nominal type I error is set at alpha = 0.05. In total, 10^4 replications are simulated for each null scenario.

	Type I error of SS2						
INUII Georgenies	Correlation	Correlation	Correlation	Correlation	Correlation		
Scenarios	=0	=0.3	=0.5	=0.7	=0.9		
<i>H</i> ₀₁	0.0033	0.0091	0.0249	0.0422	0.0489		
H ₀₂	0.0430	0.052	0.0643	0.0884	0.1404		
H _{03,(1)}	0.0243	0.0222	0.0254	0.0282	0.0320		
H _{03,(2)}	0.0178	0.0300	0.0338	0.0405	0.0501		
H _{04,(1)}	0.0257	0.0226	0.0258	0.0288	0.0317		
H _{04,(2)}	0.0187	0.0312	0.0336	0.0397	0.0478		

Table S19: Empirical type I error rates of SS2 when all of 2000 participants in the

eQTL study overlap with the GWAS study, with varying phenotypic correlations.

The LD pattern at the simulated region follows that at the *MUC20/MUC4* locus. Simulations are performed under the null hypothesis of no colocalization and with varying phenotypic correlation (0.3, 0.5, 0.7 and 0.9). For comparison, the type I error rate when no samples are overlapping is also shown (correlation = 0). Table S1 defines the parameters used under each null scenario. The nominal type I error is set at alpha = 0.05. In total, 10^4 replications are simulated for each null scenario. See Section *Simulation Details for Overlapping Samples* for other simulation details.

Supplemental Methods

Covariance of Summary Statistics from Meta-analyses with Related

Individuals within Sub-studies

Suppose the GWAS meta-analysis consists of C sub-studies with sample sizes n_c , c =

1,2,...*C*. Let
$$\hat{\beta}_c = (\hat{\beta}_{c,1}, ..., \hat{\beta}_{c,m})^{\mathsf{T}}$$
 and $Z_c = (Z_{c,1}, Z_{c,2}, ..., Z_{c,m})^{\mathsf{T}}$ denote the

marginal effect sizes and Z-scores, respectively, from study c. Let Z_{meta} =

 $(Z_{\text{meta},1}, Z_{\text{meta},2}, \dots, Z_{\text{meta},m})^{\mathsf{T}}$, where $Z_{\text{meta},j}$ represents the Z-score obtained from the meta-analysis for SNP *j*. Then

$$Z_{meta,j} = \frac{\sum_{c=1}^{C} w_{c,j} \hat{\beta}_{c,j}}{\sqrt{\sum_{c=1}^{C} w_{c,j}}}$$
(2)
$$= \frac{\sum_{c=1}^{C} \sqrt{w_{c,j}} Z_{c,j}}{\sqrt{\sum_{c=1}^{C} w_{c,j}}},$$
(3)

where $w_{c,j}$ represents the weight for study c and can take different forms (i.e. the inverse variance of $\hat{\beta}_{c,j}$). In particular, for the sample size weighted meta-analysis, $w_{c,j} = n_c$ and $Z_{meta,j} = \frac{\sum_{c=1}^{C} \sqrt{n_c z_{c,j}}}{\sqrt{n\lambda_c}}$, where λ_c is the genomic control.⁷ For SNP *j* and SNP *l*,

$$cov (Z_{meta,j}, Z_{meta,l}) = cov \left(\frac{\sum_{c=1}^{C} \sqrt{w_{c,j}} Z_{c,j}}{\sqrt{\sum_{c=1}^{C} w_{c,j}}}, \frac{\sum_{c=1}^{C} \sqrt{w_{c,l}} Z_{c,l}}{\sqrt{\sum_{c=1}^{C} w_{c,j}}} \right) \\
= \frac{\sum_{c=1}^{C} \sqrt{w_{c,j}} \sqrt{w_{c,l}}}{\sqrt{\sum_{c=1}^{C} w_{c,j}}} cov(Z_{c,j}, Z_{c,l}).$$
(4)

Therefore, the covariance of meta-Z scores between two SNPs is a weighted sum of covariance measures for the sub-studies.

If all participants in the sub-study c are independent, we consider fitting the following single-marker model with additional covariates to obtain the marginal summary statistics:

$$Y = G_i \beta + X\alpha + \epsilon \text{ with } \epsilon \sim N(0, \sigma^2 I),$$
(5)

for SNP $j.Y = (y_1, ..., y_{n_c})^{\mathsf{T}}$ denote a $n_c \times 1$ vector of phenotypic values, where each component y_i is the phenotype for the *i*th individual. Assume there are m SNPs at the locus, $G_j = (G_{j1}, ..., G_{jn_c})^{\mathsf{T}}$ is a $n_c \times 1$ vector of genotype for SNP *j*, where each component G_{ij} is the genotype for the *i*th individual. *X* is a $n_c \times q$ matrix of covariates (i.e. sex and age) and α is a $q \times 1$ vector of fixed effects for the covariates, including the intercept.

We define $U_j = (G_j, X)$, which is a $n_c \times (q + 1)$ matrix that contains the genotype for SNP *j* and additional covariates. Let B = (1, 0, 0, ..., 0), a $1 \times (1 + q)$ vector with the first component to be 1 and other components to be 0. We obtain $\hat{\beta}_{c,j}$ and $Z_{c,j}$ by the least square method:

$$\hat{\beta}_{c,j} = B \left(U_j^{\mathsf{T}} U_j \right)^{-1} U_j^{\mathsf{T}} Y \quad ; \tag{6}$$

$$Z_{c,j} = \frac{B(U_j^{\top} U_j)^{-1} U_j^{\top} Y}{\sqrt{\hat{\sigma}_j^2 B(U_j^{\top} U_j)^{-1} B^{\top}}},$$
(7)

where $\hat{\sigma}_j^2$ is the maximum likelihood estimator (MLE) of σ_j in model (5). For SNP *j* and SNP *l*,

$$\begin{aligned} \operatorname{cov}\left(Z_{c,j}, Z_{c,l}\right) &= \operatorname{cov}\left(\frac{B\left(U_{j}^{\mathsf{T}}U_{j}\right)^{-1}U_{j}^{\mathsf{T}}Y}{\sqrt{\hat{\sigma}_{j}^{2}B\left(U_{l}^{\mathsf{T}}U_{j}\right)^{-1}B^{\mathsf{T}}}}, \frac{B\left(U_{l}^{\mathsf{T}}U_{l}\right)^{-1}U_{l}^{\mathsf{T}}Y}{\sqrt{\hat{\sigma}_{l}^{2}B\left(U_{l}^{\mathsf{T}}U_{l}\right)^{-1}B^{\mathsf{T}}}}\right) \\ &= \frac{B\left(U_{j}^{\mathsf{T}}U_{j}\right)^{-1}U_{j}^{\mathsf{T}}}{\sqrt{\hat{\sigma}_{j}^{2}B\left(U_{l}^{\mathsf{T}}U_{l}\right)^{-1}B^{\mathsf{T}}}} \sigma^{2}I \frac{U_{l}\left(U_{l}^{\mathsf{T}}U_{l}\right)^{-1}B^{\mathsf{T}}}{\sqrt{\hat{\sigma}_{l}^{2}B\left(U_{l}^{\mathsf{T}}U_{l}\right)^{-1}B^{\mathsf{T}}}}, \end{aligned}$$

where $\hat{\sigma}_i^2$ and $\hat{\sigma}_l^2$ approaches σ^2 in probability. By Slutsky's theorem, the asymptotically covariance is

$$\operatorname{cov}(Z_{c,j}, Z_{c,l}) = \frac{B(U_j^{\mathsf{T}} U_j)^{-1} U_j^{\mathsf{T}} U_l (U_l^{\mathsf{T}} U_l)^{-1} B^{\mathsf{T}}}{\sqrt{B(U_j^{\mathsf{T}} U_j)^{-1} B^{\mathsf{T}} B (U_l^{\mathsf{T}} U_l)^{-1} B^{\mathsf{T}}}}.$$
(8)

Zheng et al⁸ showed equation (8) can be simplified such that

$$\operatorname{cov}\left(Z_{c,j}, Z_{c,l}\right) = \frac{G_{j}^{\top}(I - P_{X})G_{l}}{\sqrt{G_{j}^{t}(I - P_{X})G_{j}}\sqrt{G_{l}^{t}(I - P_{X})G_{l}}}$$

$$= \operatorname{cor}\left(\hat{\epsilon}_{G_{j}|X}, \hat{\epsilon}_{G_{l}|X}\right),$$
(9)

where $P_X = X(X^T X)^{-1} X^T$. $\hat{\epsilon}_{G_j|X}$ denotes the residuals obtained by regressing G_j on X, for j = 1, ...m. In particular, if both phenotypes and genotypes are standardized, and there are no additional covariates expect the genotypes (B = 1 and $U_j = G_j$), equation (9) can be simplified as

$$\operatorname{cov}(Z_{c,j}, Z_{c,l}) = \frac{G_j^{\mathsf{T}} G_l}{\sqrt{G_j^{\mathsf{T}} G_j G_l^{\mathsf{T}} G_l}}.$$

The asymptotic covariance of Z scores between two SNPs is their pairwise Pearson correlation coefficient.^{8,9}

If the sub-study c contains related samples, the first approach to obtain the summary statistics (for SNP j) is considering a linear fixed-effect model:

$$Y = G_i \beta + X\alpha + \epsilon, \text{ with } \epsilon \sim N(0, \Sigma_{\epsilon}), \tag{10}$$

where Σ_{ϵ} denote the matrix that contains information about the sample relatedness (i.e. the kinship coefficient matrix). If Σ_{ϵ} is known, let *H* denote the Cholesky decomposition of Σ_{ϵ}^{-1} such that $H^{t}H = \Sigma_{\epsilon}^{-1}$. We define $Y^{*} = HY, X^{*} = HX, G_{j}^{*} =$ $HG_{j}, U_{j}^{*} = HU_{j}$ and $\epsilon^{*} = H\epsilon$. By the generalized least square (GLS) method, the effect size and Z-score for SNP *j* is

$$\hat{\beta}_{c,j} = B(U_{j}^{*\top}U_{j}^{*})^{-1}U_{j}^{*\top}Y^{*}
= B(U_{j}^{\top}\Sigma_{\epsilon}^{-1}U_{j})^{-1}U_{j}^{\top}\Sigma_{\epsilon}^{-1}Y;$$

$$Z_{c,j} = \frac{B(U_{j}^{*\top}U_{j}^{*})^{-1}U_{j}^{*}Y^{*}}{\sqrt{B(U_{j}^{*\top}U_{j}^{*})^{-1}B^{\top}}}
= \frac{B(U_{j}^{\top}\Sigma_{\epsilon}^{-1}U_{j})^{-1}U_{j}^{\top}\Sigma_{\epsilon}^{-1}Y}{\sqrt{B(U_{j}^{\top}\Sigma_{\epsilon}^{-1}U_{j})^{-1}B^{\top}}}.$$
(12)

Similar to equation (9),

$$\operatorname{cov}\left(Z_{c,j}, Z_{c,l}\right) = \frac{G_{j}^{*^{\top}}(I - P_{X^{*}})G_{l}^{*}}{\sqrt{G_{j}^{*^{\top}}(I - P_{X^{*}})G_{j}^{*}}\sqrt{G_{l}^{*^{\top}}(I - P_{X^{*}})G_{l}^{*}}}, \text{ with } P_{X^{*}} = X^{*}(X^{*^{\top}}X^{*})^{-1}X^{*^{\top}};$$
$$= \frac{G_{j}^{\top}P^{*}G_{l}}{\sqrt{G_{j}^{\top}P^{*}G_{j}}\sqrt{G_{l}^{\top}P^{*}G_{l}}}, \text{ with } P^{*} = \Sigma_{\epsilon}^{-1} - \Sigma_{\epsilon}^{-1}X(X^{\top}\Sigma_{\epsilon}^{-1}X)^{-1}X^{\top}\Sigma_{\epsilon}^{-1}. (13)$$

If Σ_{ϵ} is unknown, we can get a consistent estimator of Σ_{ϵ} first,¹⁰ say $\hat{\Sigma}_{\epsilon}$, and then obtain the effect size and Z-score by using $\hat{\Sigma}_{\epsilon}$:

$$\hat{\beta}_{c,j}(\hat{\Sigma}_{\epsilon}) = B \left(U_j^{\mathsf{T}} \hat{\Sigma}_{\epsilon}^{-1} U_j \right)^{-1} U_j^{\mathsf{T}} \hat{\Sigma}_{\epsilon}^{-1} Y;$$

$$= D \left(U_j^{\mathsf{T}} \hat{\Sigma}_{\epsilon}^{-1} U_j \right)^{-1} U_j^{\mathsf{T}} \hat{\Sigma}_{\epsilon}^{-1} Y;$$

$$(14)$$

$$Z_{c,j}(\hat{\Sigma}_{\epsilon}) = \frac{B(U_j^{\top} \Sigma_{\epsilon}^{-1} U_j) - U_j^{\top} \Sigma_{\epsilon}^{-1} Y}{\sqrt{B(U_j^{\top} \hat{\Sigma}_{\epsilon}^{-1} U_j)^{-1} B^{\top}}}.$$
(15)

By Slutsky theorem, equation (13) is the asymptotic covariance of Z-scores. In practice, we estimate the covariance by $\hat{\Sigma}_{\epsilon}$:

$$\operatorname{cov}(Z_{c,j}, Z_{c,l}) = \frac{G_j^{\top} \hat{P}^* G_l}{\sqrt{G_j^{\top} \hat{P}^* G_j} \sqrt{G_l^{\top} \hat{P}^* G_l}}, \text{ with } \hat{P}^* = \hat{\Sigma}_{\epsilon}^{-1} - \hat{\Sigma}_{\epsilon}^{-1} X \left(X^{\top} \hat{\Sigma}_{\epsilon}^{-1} X \right)^{-1} X^{\top} \hat{\Sigma}_{\epsilon}^{-1}.$$
(16)

Secondly, the linear mixed-effect model (LMM) with a random intercept is another commonly used approach for related samples:^{11,12}

$$Y = G_j \beta + X\alpha + Q + \epsilon, \text{ with } Q \sim N\left(0, \sum_{k=1}^K \tau_k \Phi_k\right) \text{ and } \epsilon \sim N(0, \sigma^2 I).$$
(17)

Q is the random intercept; K is the number of correlation matrices (Φ_k) included for a given individual and τ_k is the corresponding variance component parameter, as defined in ¹¹. For example, Φ_1 can be the standard genetic correlation matrix (GRM). Then Φ_2 could be the environmental relationship matrix, which captures the variance associated with shared environmental effects between family members.¹³ Q and ϵ are assumed to be independent. α no longer includes the intercept in this case since the equation is supposed to have a random intercept. Excluding the intercept, X becomes a $n_c \times (q - C_{cont})$

1) matrix of covariates and $U_j = (G_j, X)$ which becomes a $n_c \times q$ matrix that contains the genotype for SNP j and covariates. Given the observed covariates X and G_j (j = 1, ..., m), the marginal distribution of Y is $N(G_j\beta + X\alpha, \Sigma_{\varepsilon})$, where $\Sigma_{\varepsilon} = \sum_{k=1}^{K} \tau_k \Phi_k + \sigma^2 I$.

When Σ_{ϵ} is known, the MLE of β based on the marginal distribution of Y can be written in the same form as the estimate in (11), which is also the minimum variance unbiased estimate.¹⁴ Therefore, we can estimate the covariance of Z-scores according to equation (13) under this case, which is the same as the covariance derived under linear fixed-effect model in (10) except excluding the column corresponding to the intercept in X.

When Σ_{ϵ} is unknown, the variance parameters contained in Σ_{ϵ} can be estimated by EM algorithm for maximum likelihood (ML) estimates or restricted maximum likelihood (REML) estimates.^{14,15} Then the effect size is calculated by equation (14) (without the intercept in X),¹⁴ and this procedure has been implemented in the R package nlme.¹⁶ Therefore, the Z-scores and its covariance can be calculated by equation (15) and equation (16) (without the intercept in X), respectively, using variance component estimates from MLE or REML. Other types of summary statistics such as the score-based statistics have been implemented in R packages GMMAT based on the LMM assumption, with the covariance estimated by AI-REML algorithm.¹⁷

Independence of the stage 1 and the stage 2 test

We assume the vector of summary statistics $(Z_1, Z_2, ..., Z_m, T_1, T_2, ..., T_m)$ has multivariate normal distribution. When there is no overlapping or related samples between two studies, the GWAS summary statistics and eQTL summary statistics are pairwise independent (i.e. $cov(Z_j, T_l) = 0$ for j = 1, 2, ..., m and l = 1, 2, ..., m), and thus

$$\operatorname{cov} \begin{bmatrix} \begin{pmatrix} Z_1 \\ Z_2 \\ \vdots \\ Z_m \end{pmatrix}, \begin{pmatrix} T_1 \\ T_2 \\ \vdots \\ T_m \end{pmatrix} \end{bmatrix} = \begin{pmatrix} \Sigma & 0_{m \times m} \\ 0_{m \times m} & \Sigma \end{pmatrix}.$$

Because of the multivariate normality of $(Z_1, Z_2, ..., Z_m, T_1, T_2, ..., T_m)$, the vector of GWAS summary statistics $(Z_1, Z_2, ..., Z_m)$ is independent of the vector of eQTL summary statistics $(T_1, T_2, ..., T_m)$ when there is no overlapping or related samples between two studies.

Recall that the stage 2 test statistic is defined as $\sum_{j=1}^{m} a_j Z_j^2$, where $a_j = \frac{t_j - \bar{t}}{\sum_j t_j^2 - m \bar{t}^2}$ as a function of eQTL evidence. The distribution of $\sum_{j=1}^{m} a_j Z_j^2$ is accessed by treating $a'_j s$ as the constants, where the variation of the eQTL summary statistics is only considered in the stage 1 test. Therefore, the stage 2 test statistic $\sum_{j=1}^{m} a_j Z_j^2$ and the stage 1 test statistic $\sum_{j=1}^{m} T_j^2$ are measurable functions of $(Z_1, Z_2, ..., Z_m)$ and $(T_1, T_2, ..., T_m)$, respectively, and they are independent due to the independence of $(Z_1, Z_2, ..., Z_m)$ and $(T_1, T_2, ..., T_m)$.

Type I Error Rate Control for a Single Hypothesis Testing

Let H_0 denote the null hypothesis that there is no colocalization given a specific geneby-tissue pair, including multiple different scenarios: H_{01}, H_{02}, H_{03} , and H_{04} . The definitions of H_{01}, H_{02}, H_{03} , and H_{04} are provided in Table S6. For a specific gene-bytissue pair, let R_1 and R_2 denote the event that the stage 1 eQTL test and the stage 2 SS test, respectively, provide significant p-values. Let R_{SS2} denote the event where the SS2 test rejects the null hypothesis of no colocalization. Thus, $R_{SS2} = R_1 \cap R_2$. The type I error rate for testing a single gene-by-tissue pair is

$$Pr(R_{SS2} | H_0) = Pr(R_2 | R_1, H_0) \times Pr(R_1 | H_0)$$

= $\sum_{i=1}^{4} Pr(R_{SS2} | H_{0i}) \times Pr(H_{0i} | H_0)$
= $\sum_{i=1}^{4} Pr(R_2 | R_1, H_{0i}) \times Pr(R_1 | H_{0i}) \times Pr(H_{0i} | H_0)$ (18)

$$= \sum_{i=1}^{4} Pr(R_2 \mid H_{0i}) \times Pr(R_1 \mid H_{0i}) \times Pr(H_{0i} \mid H_0)$$
(19)

$$\leq \sum_{i=1}^{4} \alpha \times Pr(H_{0i} \mid H_0)$$

$$= \alpha$$
(20)

From equation (18) to equation (19), $Pr(R_2 | H_{0i}) = Pr(R_2 | R_1, H_{0i})$ because the stage 2 test is independent of the stage 1 test. We access the distribution of the stage 2 test statistics by conditioning on the observed eQTL summary statistics, where the variation of the eQTL summary statistics is only considered in the stage 1 test. In equation (20), for i = 1 or 3 when there is no eQTL, the stage 1 test controls the type I error rate within α

and thus $Pr(R_1 | H_{0i}) \le \alpha$; for i = 2 or 4, the stage 2 test controls the type I error rate within α and thus $Pr(R_2 | H_{0i}) \le \alpha$. Therefore, $Pr(R_{SS2} | H_0) \le \alpha$.

Upper Bound of Family-wise Error Rate for Multiple Hypothesis Testing

To understand the challenge of Family-wise error rate (FWER) control in this mixture context, we first demonstrate a simple example of testing two genes at a locus, and then formally derive a crude upper bound that works for any number of tests at a locus, which would not change as the number of tests increases.

Consider one simple scenario where the locus of interest was identified through GWAS as in the CF example, and there are two genes, A under H_{03} (no eQTL) and B under H_{04} (eQTL but not overlapping with GWAS). Let α denote the nominal significance level for the SS2 test and thus the stage 1 eQTL test is conducted at the $\frac{\alpha}{2}$ significance level for each gene. While testing the colocalization for gene A under H_{03} , let α^* and α_2^* denote the empirical false positive rate of the SS (stage 2) test at significance level α and $\frac{\alpha}{2}$, respectively. For gene B, let 1- β denote the power of the eQTL test in stage 1.

FWER = Pr(SS2 rejects at least one gene)
= Pr(stage 1 rejects gene A but not gene B)

 \times Pr(stage 2 rejects gene *A* |stage 1 rejects gene *A* but not gene *B*)

+ Pr(stage 1 rejects gen*e B* but not gene *A*)

× Pr(stage 2 rejects gene *B* |stage 1 rejects gene *B* but not gene *A*)

+ Pr(stage 1 rejects both genes)

× Pr(stage 2 rejects at least one gene |stage 1 rejects both genes)

By definition of α^* and the independence of the stage 1 and the stage 2 tests, Pr(stage 2 rejects gene *A* | stage 1 rejects gene *A* but not gene *B*) = Pr(stage 2 rejects gene *A*) = α^* , which could be greater or equal to α . Similarly, Pr(stage 2 rejects gene *A* | stage 1 rejects both genes)= α_2^* , which could be greater or equal to $\frac{\alpha}{2}$. Thus, we can obtain

$$\begin{aligned} \text{FWER} &\leq \frac{\alpha}{2} \times \beta \times \alpha^* + (1 - \beta) \times \left(1 - \frac{\alpha}{2}\right) \times \alpha + \frac{\alpha}{2} \times (1 - \beta) \times \left(1 - \left(1 - \frac{\alpha}{2}\right)(1 - \alpha_2^*)\right). \end{aligned}$$

$$\begin{aligned} \text{If } \alpha^* &= \alpha \text{ and } \alpha_2^* = \frac{\alpha}{2}, \left(1 - \left(1 - \frac{\alpha}{2}\right)(1 - \alpha_2^*)\right) \leq \alpha, \text{ then} \end{aligned}$$

$$\begin{aligned} \text{FWER} &\leq \frac{\alpha}{2} \times \beta \times \alpha + (1 - \beta) \times \left(1 - \frac{\alpha}{2}\right) \times \alpha + \frac{\alpha}{2} \times (1 - \beta) \times \alpha \end{aligned}$$

$$\begin{aligned} &= \alpha \times \left(\frac{\alpha}{2} \times \beta + (1 - \beta) \times \left(1 - \frac{\alpha}{2}\right) + \frac{\alpha}{2} \times (1 - \beta)\right) \end{aligned}$$

$$\leq \alpha. \end{aligned}$$

When the magnitude of α^* and α_2^* are unknown, a crude upper bound for this FWER can be specified by assuming the maximum value of α^* and α_2^* to be 1, leading to FWER $\leq \frac{\alpha}{2} + \alpha - \frac{\alpha^2}{2}$; this is not necessarily less than or equal to α but provides a benchmark for the worst case scenario. Now we consider deriving the upper bound of the FWERs when we are interested in testing colocalization for K gene-by-tissue pairs at a locus. Let α denote the nominal significance level for the SS2 test. Let $H_{0,K}$ denote the null hypothesis that there are no colocalizations for the K gene-by-tissue pairs at the locus. If there is no SNP-phenotype association (GWAS) at the locus (denoted by $H_{0,K,nsp}$), the K tests are under H_{01} or H_{02} ; if there is SNP-phenotype association (GWAS) at the locus (denoted by $H_{0,K,sp}$), the K tests are under H_{03} or H_{04} . Let R_{SS2}^* denote the event that the SS2 test rejects at least one tests among the K tests. Therefore, for a locus with K tests, the FWER of the SS2 test is

$$Pr(R_{SS2}^{*} | H_{0,K}) = Pr(R_{SS2}^{*} | H_{0,K,nsp}) \times Pr(H_{0,K,nsp} | H_{0,K})$$
(21)
+
$$Pr(R_{SS2}^{*} | H_{0,K,sp}) \times Pr(H_{0,K,sp} | H_{0,K}).$$
(22)

 $Pr(H_{0,K,sp} | H_{0,K})$ represents the probability that the locus has phenotype-SNP association given there are no colocalizations for the *K* gene-by-tissue pairs, which is unknown in practice. Therefore, we need to find the maximum values for both $Pr(R_{SS2}^* | H_{0,K,nsp})$ in (21) and $Pr(R_{SS2}^* | H_{0,K,sp})$ in (22) to obtain the upper bound for $Pr(R_{SS2}^* | H_{0,K})$. Let $R_{1,P}$ represent the event that the stage 1 test has *P* significant geneby-tissue pairs. Let $R_{2,P}^*$ represents the event that the stage 2 test rejects at least one gene-by-tissue pairs among the *P* significant gene-by-tissue pairs from the stage 1 test. For a locus with no phenotype-SNP association (under $H_{0,K,nsp}$), the K tests consist of mixture of H_{01} or H_{02} . If there are P significant gene-by-tissue pairs from the stage 1 test, the stage 2 test is implemented at significant level $\frac{\alpha}{P}$ per test by Bonferroni

correction, and thus $Pr(R_{2,P}^* | R_{1,P}, H_{0,K,nsp}) \leq \frac{\alpha}{P} \times P = \alpha$. Therefore,

$$Pr(R_{SS2}^{*} | H_{0,K,nsp}) = \sum_{P=1}^{K} Pr(R_{2,P}^{*} | R_{1,P}, H_{0,K,nsp}) \times Pr(R_{1,P} | H_{0,K,nsp})$$

$$\leq \sum_{P=1}^{K} \left(\frac{\alpha}{P} \times P\right) \times Pr(R_{1,P} | H_{0,K,nsp})$$

$$= \alpha \times \sum_{P=1}^{K} Pr(R_{1,P} | H_{0,K,nsp})$$

$$\leq \alpha$$

$$(23)$$

For a locus with phenotype-SNP association (under $H_{0,K,sp}$), we need to consider three cases separately: the *K* tests are all under H_{04} ; the *K* tests are all under H_{03} ; the *K* tests consist of a mixture of H_{03} and H_{04} . Let $H_{0,K,h4}$, $H_{0,K,h3}$ and $H_{0,K,h34}$ denote the three cases. respectively. Therefore,

$$Pr(R_{SS2}^{*} | H_{0,K,sp}) = Pr(R_{SS2}^{*} | H_{0,K,h4}) \times Pr(H_{0,K,h4} | H_{0,K,sp}) + Pr(R_{SS2}^{*} | H_{0,K,h3}) \times Pr(H_{0,K,h3} | H_{0,K,sp}) + Pr(R_{SS2}^{*} | H_{0,K,h34}) \times Pr(H_{0,K,h34} | H_{0,K,sp}).$$
(24)

Under $H_{0,K,h4}$, when all the K tests are under H_{04} , $Pr(R_{2,P}^* | R_{1,P}, H_{0,K,nsp}) \leq$

 $\frac{\alpha}{P} \times P = \alpha$. Thus,

$$Pr(R_{SS2}^{*} | H_{0,K,h4}) = \sum_{P=1}^{K} Pr(R_{2,P}^{*} | R_{1,P}, H_{0,K,h4}) \times Pr(R_{1,P} | H_{0,K,h4})$$

$$\leq \sum_{P=1}^{K} (\frac{\alpha}{P} \times P) \times Pr(R_{1,P} | H_{0,K,h4})$$

$$= \alpha \times \sum_{P=1}^{K} Pr(R_{1,P} | H_{0,K,h4})$$

$$\leq \alpha.$$
(25)

Under $H_{0.K,h3}$ when all the *K* tests are under H_{03} , the stage 1 test for each gene-bytissue pair is implemented under the null that there is no eQTL, and adjust the α for the total number of tests by Bonferroni correction, $\frac{\alpha}{K}$. Thus,

$$Pr(R_{SS2}^* | H_{0,K,h3}) \leq Pr(\text{ the stage 1 test rejects at least one gene-by-tissue pairs } | H_{0,K,h3})$$
$$\leq \frac{\alpha}{K} \times K$$
$$= \alpha.$$
(26)

So far we have proved that when all the K tests are under H_{03} or H_{04} , the upper bound on the FWER of the two-stage SS2 test is α .

Under $H_{0,K,h34}$ when K tests consist of H_{03} and H_{04} , let H_{0,K_1,K_2} denote the event that there are K_1 tests under H_{04} and K_2 tests under H_{03} . We define R_{1,P_1,P_2} to be the event that there are P_1 significant stage 1 eQTL tests among those K_1 tests and P_2 significant stage 1 eQTL tests among those K_2 tests. In this case,

$$Pr(R_{SS2}^{*} | H_{0,K,h_{34}}) = \sum_{K_{1} \ge 1,K_{2} \ge 1} Pr(R_{SS2}^{*} | H_{0,K_{1},K_{2}}) \times Pr(H_{0,K_{1},K_{2}} | H_{0,K,h_{34}}),$$

$$Pr(R_{SS2}^{*} | H_{0,K_{1},K_{2}}) = \sum_{P_{1} \ge 1,P_{2} \ge 0} Pr(R_{2,p}^{*} | R_{1,P_{1},P_{2}}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,P_{1},P_{2}} | H_{0,K_{1},K_{2}})$$

$$+ \sum_{P_{1} \ge 0,P_{2} \ge 1} Pr(R_{2,p}^{*} | R_{1,P_{1},P_{2}}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,P_{1},P_{2}} | H_{0,K_{1},K_{2}})$$

$$+ \sum_{P_{1} \ge 1,P_{2} \ge 1} Pr(R_{2,p}^{*} | R_{1,P_{1},P_{2}}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,P_{1},P_{2}} | H_{0,K_{1},K_{2}})$$

$$= \sum_{P_{1} \ge 1,P_{2} \ge 1} Pr(R_{2,p_{1}}^{*} | R_{1,P_{1},0}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,P_{1},0} | H_{0,K_{1},K_{2}})$$

$$+ \sum_{P_{2} \ge 1} Pr(R_{2,p_{2}}^{*} | R_{1,0,P_{2}}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,0,P_{2}} | H_{0,K_{1},K_{2}})$$

$$(27)$$

$$+ \sum_{P_{2} \ge 1} Pr(R_{2,p_{2}}^{*} | R_{1,0,P_{2}}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,0,P_{2}} | H_{0,K_{1},K_{2}})$$

$$(27)$$

The term (27) demonstrates the scenario when all the significant stage 1 eQTL tests

are within the K_1 gene-by-tissue pairs under H_{04} . In this scenario, $P = P_1$ and the stage 2 test is implemented at significance level $\frac{\alpha}{P_1}$ per test. Therefore, term (27)

$$\sum_{P_{1}=1}^{K_{1}} Pr(R_{2,p_{1}}^{*} | R_{1,P_{1},0}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,P_{1},0} | H_{0,K_{1},K_{2}})$$

$$\leq \left(\frac{\alpha}{P_{1}} \times P_{1}\right) \times \sum_{P_{1}=1}^{K_{1}} Pr(R_{1,P_{1},0} | H_{0,K_{1},K_{2}})$$

$$= \alpha \times \sum_{P_{1}=1}^{K_{1}} Pr(R_{1,P_{1},0} | H_{0,K_{1},K_{2}})$$

$$= \alpha \times S_{(1,0)}, \qquad (30)$$

Where $S_{(1,0)} = \sum_{P_1=1}^{K_1} Pr(R_{1,P_1,0} \mid H_{0,K_1,K_2})$ which represents the probability that there is at least one significant stage 1 eQTL test among the K_1 gene-by-tissue pairs but no significant stage 1 eQTL test among the K_2 gene-by-tissue pairs. Let α_p^* denote the empirical false positive rate of the stage 2 test at the significance level $\frac{\alpha}{p}$ under H_{03} at the locus of interest. We define $\tilde{\alpha}$ to be $\max_{p=1,\dots,K} (\alpha_p^* \times P)$, which can be interpreted as an upper bound of the empirical probability that the stage 2 test rejects at least one gene-by-tissue pairs among those *P* significant stage 1 tests at the locus. In particular when $\alpha_p^* = \frac{\alpha}{p}$ (for $P = 1, \dots, K$), $\tilde{\alpha} = \alpha$, and $\tilde{\alpha}$ could be bigger than α if α_p^* is inflated. The term (28) demonstrates the scenario when all the significant stage 1 eQTL tests are within the K_2 gene-by-tissue pairs under H_{03} . In this scenario, $P = P_2$; the stage 2 test is implemented at significance level $\frac{\alpha}{P_2}$ per test.

$$\sum_{P_{2}=1}^{K_{2}} Pr(R_{2,p_{2}}^{*} | R_{1,0,P_{2}}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,0,P_{2}} | H_{0,K_{1},K_{2}})$$

$$\leq \sum_{P_{2}=1}^{K_{2}} (\alpha_{P_{2}}^{*} \times P_{2}) \times \sum_{P_{2}=1}^{K_{2}} Pr(R_{1,0,P_{2}} | H_{0,K_{1},K_{2}})$$

$$\leq \tilde{\alpha} \times S_{(0,1)}$$
(31)

, where $S_{(0,1)} = \sum_{P_2=1}^{K_2} Pr(R_{1,0,P_2} \mid H_{0,K_1,K_2})$ which represents the probability that there are at least one significant stage 1 eQTL tests among the K_2 tests but no significant stage 1 eQTL tests among the K_1 tests.

The term (29) demonstrates the scenario when those significant stage 1 eQTL tests present both in the K_1 and K_2 gene-by-tissue pairs. In this scenario,

$$\sum_{P_{1}\geq 1,P_{2}\geq 1} Pr(R_{2,P}^{*} \mid R_{1,P_{1},P_{2}}, H_{0,K_{1},K_{2}}) \times Pr(R_{1,P_{1},P_{2}} \mid H_{0,K_{1},K_{2}}) \leq \sum_{P_{1}\geq 1,P_{2}\geq 1} (\alpha_{P}^{*} \times P) \times \leq \tilde{\alpha} (1 - S_{(0,0)} - S_{(0,1)} - S_{(1,0)}),$$
(32)

where $S_{(0,0)}$ represent the probability that under H_{0,K_1,K_2} , there are no significant stage 1 eQTL tests among the *K* gene-by-tissue pairs. Note that

 $S_{(0,1)} = Pr($ there are at least one significant stage 1 tests among the K tests $| H_{0,K_1,K_2})$ -Pr(there are at least one significant stage 1 tests among the K_1 tests $| H_{0,K_1,K_2})$ $\ge 1 - S_{(0,0)} - \alpha,$

and therefore

$$\alpha \ge 1 - S_{(0,0)} - S_{(0,1)} . \tag{33}$$

We define δ to be $\tilde{\alpha} - \alpha$ and combine the results in (30), (31), (32) and (33) to obtain

$$Pr(R_{SS2}^{*} | H_{0,K_{1},K_{2}}) \leq \alpha S_{(0,1)} + \tilde{\alpha} S_{(1,0)} + \tilde{\alpha} (1 - S_{(0,0)} - S_{(1,0)} - S_{(0,1)})$$

$$= \alpha S_{(0,1)} + \tilde{\alpha} (1 - S_{(0,0)} - S_{(0,1)})$$

$$= \alpha S_{(0,1)} + (\alpha + \delta) (1 - S_{(0,0)} - S_{(0,1)})$$

$$= \alpha (1 - S_{(0,0)}) + \delta (1 - S_{(0,0)} - S_{(0,1)})$$

$$\leq \alpha (1 - S_{(0,0)}) + \delta \times \alpha$$

$$\leq \alpha + \delta \times \alpha$$

$$= \alpha \times (1 + \delta).$$
(34)

Therefore under the $H_{0,K,h_{34}}$,

$$Pr(R_{SS2}^{*} | H_{0,K,h_{34}}) = \sum_{K_{1} \ge 1, K_{2} \ge 1} Pr(R_{SS2}^{*} | H_{0,K_{1},K_{2}}) \times Pr(H_{0,K_{1},K_{2}} | H_{0,K,h_{34}})$$

$$\leq \sum_{K_{1} \ge 1, K_{2} \ge 1} \alpha (1 + \delta) \times Pr(H_{0,K_{1},K_{2}} | H_{0,K,h_{34}})$$

$$\leq \alpha \times (1 + \delta).$$
(35)

So far, we have shown that when the K tests consist of a mixture of H_{03} and H_{04} , the upper bound is $\alpha \times (1 + \delta)$; by assuming the maximum value of $\tilde{\alpha}$ to be 1,

 $\alpha \times (1 + \delta) = 2\alpha - \alpha^2$, which is bigger than α . This upper bound works for any value of *K*, which would not change as the number of tests increases.

Taking into consideration all of the scenarios, the FWER of the SS2 in (21) and (22)

can be calculated by

$$\begin{aligned} Pr(R_{SS2}^* | H_{0,K}) &= Pr(R_{SS2}^* | H_{0,K,nsp}) \times Pr(H_{0,K,nsp} | H_{0,K}) \\ &+ \{Pr(R_{SS2}^* | H_{0,K,h_4}) \times Pr(H_{0,K,h_4} | H_{0,K,sp}) \\ &+ Pr(R_{SS2}^* | H_{0,K,h_3}) \times Pr(H_{0,K,h_3} | H_{0,K,sp}) \\ &+ Pr(R_{SS2}^* | H_{0,K,h_3}) \times Pr(H_{0,K,h_34} | H_{0,K,sp}) \} \times Pr(H_{0,K,sp} | H_{0,K}) \\ &\leq \alpha \times \{Pr(H_{0,K,nsp} | H_{0,K}) + Pr(H_{0,K,h_4} | H_{0,K}) + Pr(H_{0,K,h_3} | H_{0,K}) \} \\ &+ \alpha (1 + \delta) \times Pr(H_{0,K,h_{34}} | H_{0,K}) \\ &\leq \alpha \times (1 + \delta). \end{aligned}$$

If we want to strictly control the upper bound of the FWER at a certain level such as 0.05, $\alpha \times (1 + \delta) = 0.05$, where $\delta = \tilde{\alpha} - \alpha$. This crude upper bound can be used to find the nominal level for performing the test in practice. By assuming a maximum value of $\tilde{\alpha}$ to be 1, we can solve $\alpha = 0.0253$. However, this procedure is found to be unnecessary from our empirical studies, because $\tilde{\alpha}$ never approaches 1 and we did not observe a single iteration with the FWER greater than the specified significance level without this procedure.

Simulation Details for a Single Gene-by-Tissue Pair

We generate the GWAS summary statistics Z and the eQTL summary statistics T from $N(\Sigma\Lambda_Z, \Sigma)$ and $N(\Sigma\Lambda_T, \Sigma)$, respectively, where Σ is the LD matrix from the locus of interest. $\Lambda_Z = (\lambda_{Z_1}, \lambda_{Z_2}, ..., \lambda_{Z_c}, ..., \lambda_{Z_m})^T$ and $\Lambda_T = (\lambda_{T_1}, \lambda_{T_2}, ..., \lambda_{T_c}, ..., \lambda_{T_m})^T$ are vectors with each component being the true standardized effect size of the corresponding SNP,

respectively for GWAS and eQTL analyses. In particular, λ_{Z_c} and λ_{T_c} represent the value for the associated SNP from the GWAS and eQTL studies, respectively. When there is SNP-phenotype association, we use the lead GWAS SNP from our CF study^{18,19} as the associated SNP with value λ_{Z_c} , and we define a SNP (r < 0.002 and > -0.002 with the associated) SNP as an independent eQTL SNP with value λ_{T_c} .

For type I error evaluation, we consider all four scenarios of the composite null hypothesis. For example, under H_{01} when there is no SNP-phenotype association and no eQTL, $\lambda_{Z_c} = 0$ and $\lambda_{T_c} = 0$. In contrast, under H_{04} when both SNP-phenotype association and eQTL are present but occurring at independent SNPs, $\lambda_{Z_c} \neq 0$ and $\lambda_{T_c} \neq$ 0, and we set $\lambda_{Z_c} = 5.73$ and $\lambda_{T_c} = 7.01$ to be consistent with the previous simulation study in ¹⁸. Table S6 provides details for the parameter settings under the four null scenarios. Under each scenario, 10^4 replications are simulated.

To study power, we simulate six scenarios. For scenarios with two association peaks, we used the lead GWAS SNP from our CF study at the locus to be the first associated SNP, while the second associated SNP is defined as the next adjacent SNP with r < 0.002 and > -0.002 with the first associated SNP. We vary the magnitude of eQTL evidence to measure the relationship between power and colocalization strength. Table S7 provides details for the parameter settings under the six alternative scenarios. Under each scenario, 10^4 replications are simulated.

Simulation Details for Multiple Genes-Tissue Pairs

The GWAS summary statistics are generated from N($\Sigma \Lambda_Z, \Sigma$) with $\lambda_{Z_c} = 4.5$, and Σ is simulated based on the LD within the MUC20/MUC4 and SLC6A14 loci. The two loci are defined by including SNPs within 0.1Mb of either side of the top associated GWAS SNP from the CF study. For FWER evaluation with one GWAS association at the simulated locus, we generate 600 sets of independent eQTL summary statistics corresponding to eQTL analyses of 600 genes of interest under H_{03} or H_{04} , where different proportions of genes with eQTL evidence under H_{04} are considered (0%, 20%, 40%, 60%, 80% and 100%). For a given gene with eQTL signals under H_{04} , the eQTL summary statistics are generated from N($\Sigma \Lambda_T, \Sigma$) where the value of λ_{T_c} is randomly generated from 6 different intervals (50%-60%, 60%-70%, 70%-80%, 80%-90%, 90%-95%, 95%-100% power is achieved to detect the eQTL association at the significance level of 10^{-8}) with probabilities according to the proportion of the -log10 (maximum eQTL p-value) within each interval observed at the locus. For other genes with eQTL under H_{03} , the eQTL summary statistics are generated from $N(0, \Sigma)$.

For power evaluation, with the same GWAS peak, we set 5% of genes with eQTL signals as colocalizing with the GWAS signal (Scenario 1 in Figure 1). For the *SLC6A14* locus, we set 47.5% of genes as having no eQTL signal (under H_{03}), and 47.5% of genes

as having eQTL signals that are independent from the GWAS signal (under H_{04}); while modeling the *MUC20/MUC4* locus, we set 19% of genes under H_{03} , and 76% of genes under H_{04} . The GWAS associated SNP and the eQTL associated SNPs are selected in the same way in section *Simulation Details for a Single Gene-by-Tissue Pair*. For genes under the alternative, the eQTL summary statistics are generated from N($\Sigma\Lambda_T$, Σ), where the value of λ_{T_c} is randomly selected from an interval (i.e. [5.48, 5.73]). For genes with the eQTL under H_{04} , λ_{T_c} is selected from 4.45 to 9.48 such that 10% to 100% power is achieved at genome-wide significance level 10⁻⁸. We evaluate power as the average increase in strength of eQTL among the 5% of genes under the alternative, and consider 5 different intervals: [5.48, 5.73], [5.73, 5.98], [5.98, 6.25], [6.25, 6.57] and [6.57, 7.01]. Under each scenario, 10⁵ replications are simulated.

Simulation Details for Overlapping Samples

Given the sample size n_{GWAS} and n_{eQTL} for a GWAS and eQTL study, respectively, we randomly select n_{GWAS} individuals from the genotype data of the CF lung GWAS study.¹⁹ A region is defined which included the SNPs 0.1Mb around the top GWAS SNP at the *MUC4/MUC20* locus. All or half of the participants in the eQTL study are randomly selected and simulated such that they are overlapping with the participants in the GWAS. For individual *i*, let g_{1i} and g_{2i} denote the genotype of the associated SNP for the GWAS and eQTL study respectively. We simulate a pair of phenotypes for each individual $\begin{pmatrix} y_{1i} \\ y_{2i} \end{pmatrix} = \begin{pmatrix} g_{1i}\beta_1 + \epsilon_{1i} \\ g_{2i}\beta_2 + \epsilon_{2i} \end{pmatrix}$, where β_1 and β_2 denote the effect sizes for the GWAS and eQTL studies, respectively. Hormozdiari et al²⁰ has shown that

$$\lambda_{Z_c} = \frac{\beta_1}{\sqrt{\operatorname{var}(\epsilon_{1i})}} \times \sqrt{n_{GWAS}};$$

$$\lambda_{T_c} = \frac{\beta_2}{\sqrt{\operatorname{var}(\epsilon_{2i})}} \times \sqrt{n_{eQTL}}.$$
 (1)

We set different values for λ_{Z_c} and λ_{T_c} under the composite null hypothesis, and then calculate the corresponding values for β_1 and β_2 according to equation (1). For example, when the GWAS signal and eQTL signal are distinct (H_{04}), we set $\Lambda_{T_c} = 5.48$ to mimic the observed eQTL peak for *MUC4* in HNE. Given 100 participants in the eQTL study ($n_{eQTL} = 100$) and $var(\epsilon_{2i}) = 1$, we obtain $\beta_1 = 0.123$. We consider two different values of λ_{Z_c} (5.73 or 6.57) such that 0.5 or 0.8 power is achieved to detect the GWAS association at the genome-wide significance level of 10⁻⁸. Given 2000 participants in the GWAS study and $var(\epsilon_{2i}) = 1$, we obtain $\beta_1 = 0.128$ and 0.147, respectively. Parameter settings for λ_{Z_c} and λ_{T_c} under the composite null hypothesis is provided in Table S1. $\binom{\epsilon_{1i}}{\epsilon_{2i}} \sim MVN (\binom{0}{0}, \Sigma_{\epsilon})$, where Σ_{ϵ} is the covariance matrix with the diagonal elements set to be 1. If an individual is included in both studies, the off-diagonal terms are set to C which represents the phenotypic correlation; otherwise it is set to be 0. Simulation studies are conducted for different values of phenotypic correlation, C= 0.3, 0.5, 0.7 or 0.9. We perform univariate linear regression to obtain the marginal summary statistics and then apply SS2. Under each scenario, 10^4 replications are simulated.

Supplemental References

- Zhu, Z., Zhang, F., Hu, H., Bakshi, A., Robinson, M.R., Powell, J.E., Montgomery, G.W., Goddard, M.E., Wray, N.R., Visscher, P.M., and Yang, J. (2016). Integration of summary data from GWAS and eQTL studies predicts complex trait gene targets. Nature Genetics 48, 481-487. 10.1038/ng.3538.
- Wu, Y., Zeng, J., Zhang, F., Zhu, Z., Qi, T., Zheng, Z., Lloyd-Jones, L.R., Marioni, R.E., Martin, N.G., Montgomery, G.W., et al. (2018). Integrative analysis of omics summary data reveals putative mechanisms underlying complex traits. Nature Communications 9, 918. 10.1038/s41467-018-03371-0.
- Giambartolomei, C., Vukcevic, D., Schadt, E.E., Franke, L., Hingorani, A.D., Wallace, C., and Plagnol, V. (2014). Bayesian Test for Colocalisation between Pairs of Genetic Association Studies Using Summary Statistics. PLOS Genetics 10, e1004383. 10.1371/journal.pgen.1004383.
- Dobbyn, A., Huckins, L.M., Boocock, J., Sloofman, L.G., Glicksberg, B.S., Giambartolomei, C., Hoffman, G.E., Perumal, T.M., Girdhar, K., Jiang, Y., et al. (2018). Landscape of Conditional eQTL in Dorsolateral Prefrontal Cortex and Colocalization with Schizophrenia GWAS. Am J Hum Genet *102*, 1169-1184. 10.1016/j.ajhg.2018.04.011.
- Lonsdale, J., Thomas, J., Salvatore, M., Phillips, R., Lo, E., Shad, S., Hasz, R., Walters, G., Garcia, F., Young, N., et al. (2013). The Genotype-Tissue Expression (GTEx) project. Nature Genetics 45, 580-585. 10.1038/ng.2653.
- Ongen, H., Buil, A., Brown, A.A., Dermitzakis, E.T., and Delaneau, O. (2016).
 Fast and efficient QTL mapper for thousands of molecular phenotypes.
 Bioinformatics 32, 1479-1485. 10.1093/bioinformatics/btv722.
- Bulik-Sullivan, B.K., Loh, P.-R., Finucane, H.K., Ripke, S., Yang, J., Patterson, N., Daly, M.J., Price, A.L., and Neale, B.M. (2015). LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. Nature genetics 47, 291-295.
- Xu, Z., Duan, Q., Yan, S., Chen, W., Li, M., Lange, E., and Li, Y. (2015). DISSCO: direct imputation of summary statistics allowing covariates. Bioinformatics 31, 2434-2442.
- Hormozdiari, F., van de Bunt, M., Segrè, A.V., Li, X., Joo, J.W.J., Bilow, M., Sul, J.H., Sankararaman, S., Pasaniuc, B., and Eskin, E. (2016). Colocalization of GWAS and eQTL Signals Detects Target Genes. Am J Hum Genet 99, 1245-1260. 10.1016/j.ajhg.2016.10.003.
- Baltagi, B.H. (2008). Econometrics, 4th ed. 2008. Edition (Springer Berlin Heidelberg). 10.1007/978-3-540-76516-5.

- 11. Park, J.Y., Wu, C., Basu, S., McGue, M., and Pan, W. (2018). Adaptive SNP-set association testing in generalized linear mixed models with application to family studies. Behavior genetics *48*, 55-66.
- 12. Park, J.Y., Wu, C., and Pan, W. (2018). An adaptive gene-level association test for pedigree data. BMC genetics *19*, 39-43.
- Hill, W.D., Arslan, R.C., Xia, C., Luciano, M., Amador, C., Navarro, P., Hayward, C., Nagy, R., Porteous, D.J., and McIntosh, A.M. (2018). Genomic analysis of family data reveals additional genetic effects on intelligence and personality. Molecular psychiatry 23, 2347-2362.
- Laird, N.M., and Ware, J.H. (1982). Random-effects models for longitudinal data. Biometrics, 963-974.
- 15. Lindstrom, M.J., and Bates, D.M. (1988). Newton—Raphson and EM algorithms for linear mixed-effects models for repeated-measures data. Journal of the American Statistical Association *83*, 1014-1022.
- 16. Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., Heisterkamp, S., Van Willigen, B., and Maintainer, R. (2017). Package 'nlme'. Linear and nonlinear mixed effects models, version *3*.
- 17. Chen, H., Conomos, M.P., and Chen, M.H. (2019). Package 'GMMAT'.
- Gong, J., Wang, F., Xiao, B., Panjwani, N., Lin, F., Keenan, K., Avolio, J., Esmaeili, M., Zhang, L., He, G., et al. (2019). Genetic association and transcriptome integration identify contributing genes and tissues at cystic fibrosis modifier loci. PLOS Genetics 15, e1008007. 10.1371/journal.pgen.1008007.
- Corvol, H., Blackman, S.M., Boëlle, P.-Y., Gallins, P.J., Pace, R.G., Stonebraker, J.R., Accurso, F.J., Clement, A., Collaco, J.M., Dang, H., et al. (2015). Genomewide association meta-analysis identifies five modifier loci of lung disease severity in cystic fibrosis. Nature Communications *6*, 8382. 10.1038/ncomms9382.
- Hormozdiari, F., Kostem, E., Kang, E.Y., Pasaniuc, B., and Eskin, E. (2014). Identifying causal variants at loci with multiple signals of association. Genetics 198, 497-508.