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#### **Supplemental Information**

A Selectivity Filter Gate Controls

**Voltage-Gated Calcium Channel** 

**Calcium-Dependent Inactivation** 

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#### Supplementary material for

#### A selectivity filter gate controls voltage-gated calcium channel calcium-dependent inactivation

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Figure S1 Exemplar Ca<sup>2+</sup> currents for Ca<sub>V</sub>1.2, Ca<sub>V</sub>1.3, Ca<sub>V</sub>2.1, and mutants co-expressed with Ca<sub>V</sub> $\beta_{2a}$ . Related to Figures 1, 2, and 4. Exemplar traces recorded in A, *Xenopus* oocytes expressing Ca<sub>V</sub>1.2 or the indicated mutants, B, HEK 293 cells expressing Cav1.2 or the indicated mutants, C, HEK 293 cells expressing Ca<sub>V</sub>1.3 or the indicated mutants, and D, HEK 293 cells expressing Ca<sub>V</sub>2.1 or the indicated mutants. Currents were evoked using a multi-step activation protocol (insets).



Figure S2 SFII (+1) mutants do not dramatically reduce Ca<sub>V</sub> selectivity properties. Related to Figures 1, 2, and 4. A, Exemplar recordings at +20 mV in Ca<sup>2+</sup> (black) or Ba<sup>2+</sup> (grey) from *Xenopus* oocytes expressing Ca<sub>V</sub>1.2 or indicated mutants, **B**, from HEK293 cells expressing Ca<sub>V</sub>1.2 or indicated mutants, **C**, from HEK293 cells expressing Ca<sub>V</sub>1.3 or indicated mutants, and **D**, HEK293 cells expressing Ca<sub>V</sub>2.1 or indicated mutants. Right panels represent averaged values of ratios between Ba<sup>2+</sup> and Ca<sup>2+</sup> peak current amplitude at +20 mV or Ca<sub>V</sub>1.2 and Ca<sub>V</sub>1.3 or at +10 mV for Ca<sub>V</sub>2.1. '\*' indicates p<0.01, '\*\*' indicates p<0.001, and 'N.S.' indicates 'not statistically significant' compared to wild-type channels. n values for all bar graphs are in Table 1. All recordings were made with Ca<sub>V</sub>β<sub>2a</sub>.



**Figure S3 Ca<sub>v</sub>1.2 selectivity filter mutations affect CDI independently of Ca<sub>v</sub>\beta. Related to Figure 1. A, Exemplar raw traces recorded at +20 mV in Ca<sup>2+</sup> (black) or Ba<sup>2+</sup> (grey) from** *Xenopus* **oocytes expressing Ca<sub>v</sub>1.2 or the indicated mutants with Ca<sub>v</sub>\beta\_3. <b>B**, Normalized traces from 'A'. **C**, Ratio of normalized I<sub>Ca</sub>/I<sub>Ba</sub> currents (net CDI, (Barrett and Tsien, 2008; Findeisen and Minor, 2009)) showing average plots ± s.e.m. **D**, net CDI 300-ms post-depolarization. '\*' indicates p<0.01 and '\*\*' indicates p<0.001 compared to Cav1.2. n = 5-10.



**Figure S4 DII (+1) aspartate mutations affecting CDI spare VDI. Related to Figure 1. A,** Exemplar two electrode voltage clamp recordings at +20 mV from *Xenopus* oocytes expressing Ca<sub>V</sub>1.2 (black), Ca<sub>V</sub>1.2 D707A (grey), Cav1.2 D707E (light blue), or Cav1.2 D707N (orange) with Ca<sub>V</sub>β<sub>3</sub>. **B**,  $\tau$  inactivation ( $\tau_{inact}$ ) for '**A**'. n = 6-15. **C**, Exemplar two electrode voltage clamp recordings at +20 mV from *Xenopus* oocytes expressing Ca<sub>V</sub>1.2 (black), Ca<sub>V</sub>1.2 D707A (grey), Cav1.2 D707A (grey), Cav1.2 D707A (grey), Cav1.2 D367A (green), Cav1.2 E1119A (blue), or Cav1.2 D1420A (pink) with Ca<sub>V</sub>β<sub>3</sub>. **D**,  $\tau$  inactivation ( $\tau_{inact}$ ) for '**C**'. n = 9-19. '\*' indicates p<0.001, and 'N.S.' indicates 'not statistically significant' compared to Ca<sub>V</sub>1.2. **E**, Voltage-dependent inactivation curves representing channel availability after a steady-state inactivation at the indicated potentials.



**Figure S5 SF (+4) mutations affecting VDI spare CDI. Related to Figure 1. A,** Exemplar traces recorded in *Xenopus* oocytes co-expressing Ca<sub>V</sub>1.2, Ca<sub>V</sub>1.2 E1119A, or Cav1.2 D1420A with Cav $\beta_{2a}$  in response to the indicated protocol. **B**, Exemplar normalized recordings at +20 mV in Ca<sup>2+</sup> (black) or Ba<sup>2+</sup> (grey) from *Xenopus* oocytes expressing Ca<sub>V</sub>1.2, Ca<sub>V</sub>1.2 E1119A, or Cav1.2 D1420A. **C**, Fractional current remaining 300 ms post-depolarization (R<sub>300</sub>) as a function of the membrane potential for channels in '**B**'. **D**, Average fraction CDI (f) at +20 mV. 'N.S.' indicates 'not statistically significant'. n values are found in Table 1. **E**, Voltage-dependent activation curves for Ca<sub>V</sub>1.2 (black circles), Ca<sub>V</sub>1.2 E1119A (blue triangles), and Cav1.2 D1420A (red inverted triangles). **F**, I-V relationships for the indicated channels. Symbols are the same as '**E**'.



Figure S6 Ca<sub>v</sub>1.2 CDI requires a negative charge at +1 position on DII, DIII, or DIV, not DI, regardless of the amount of Ca<sup>2+</sup> influx. Related to Figures 3 and 5. A, Exemplar Ca<sup>2+</sup> currents in response to a +20 mV depolarization from *Xenopus* oocytes, co-expressing Ca<sub>v</sub>1.2 or the indicated mutants with Cav $\beta_{2a}$  in the absence (black) or presence of 5  $\mu$ M Bay K8644 (blue). Raw traces (upper panels) and normalized traces (lower panels) are shown to illustrate Bay K8644 effects on both the peak current and inactivation, respectively. **B**, Exemplar normalized Ba<sup>2+</sup> currents from *Xenopus* oocytes expressing Ca<sub>v</sub>1.2 or the indicated mutants, in response to a +20 mV depolarization in absence (black) or in presence of 5  $\mu$ M Bay K8644 (blue). **C**, Ratio of Ca<sup>2+</sup> current amplitudes in presence, I<sub>Ca</sub>(BayK), and in absence, I<sub>Ca</sub>, of Bay K8644. 'N.S.' indicates not statistically different. n = 4-7. **D** and **E**, Percentage of inactivation 300 ms post-depolarization (t<sub>i300</sub>) in the absence (dark bars) and presence of Bay K8644 (light bars) on recordings performed using **D**, Ca<sup>2+</sup>, or **E**, Ba<sup>2+</sup> as the charge carrier. '\*' indicates p<0.01 and '\*\*' indicates p<0.01. n= 6-11. **F**, Difference in t<sub>i300</sub> induced by Bay K8644 in Ca<sup>2+</sup>(dark bars) and Ba<sup>2+</sup> (light bars).