# nature research

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### **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section

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n/a	Confirmed	
	$oxed{x}$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement	
	🕱 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly	
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.	
×	A description of all covariates tested	
×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons	
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)	
x	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>	
×	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings	
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes	
×	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated	

#### Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

### Software and code

Policy information about <u>availability of computer code</u>

Data collection

Äkta system: UNICORN 7 (Cytiva); ITC: MicroCal PEAQ-ITC (@Malvern Panalytical); MST system: NanoTemper Control Version 2.0.2.29 (NanoTemper); X-Ray Data Collection: MxCube3, MxCube2 (https://github.com/mxcube); Cryo-EM: relion's 3.1 implementation of MotionCor2, CTFFIND 4.1.13, Gautomatch v0.56 (http://www.mrc-lmb.cam.ac.uk/kzhang/), RELION 3.1

Data analysis

Microsoft Excel for Mac 2011 Version 14.6.8; GraphPad Prism 5 for Mac OS X; ImageQuant (GE Healthcare); ITC: MicroCal PEAQ-ITC Analysis Software (@Malvern Panalytical); MST system: NanoTemper Analysis Version 1.5.37 and 1.2.009 (NanoTemper); Origin8G (OriginLab); Pymol Version 1.8.2.0 (Schroedinger); Coot Version 0.8.9.2 and 0.9.4.1 (doi:10.1107/S0907444910007493); Phenix Version 1.17.1-3660 and 1.19 (https://doi.org/10.1107/S2059798319011471); XDS Version January 31, 2020; Cryosparc v3.2.0; Chimera Version 1.13.1, ChimeraX version 1.1; crYOLO version 1.7.6

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

#### Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Data availability. Coordinates and structure factors of the crystal structures and coordinates of the cryo-EM structure have been deposited at the Protein Data Bank with the accession codes: 709F (https://doi.org/10.2210/pdb709F/pdb), 709G (https://doi.org/10.2210/pdb709H/pdb), 709H (https://doi.org/10.2210/pdb709H/pdb), 709H (https://doi.org/10.2210/pdb709H/pdb), 709H (https://doi.org/10.2210/pdb709H/pdb), 709H (https://doi.org/10.2210/pdb709H/pdb), 709H (https://doi.org/10.2210/pdb709H/pdb)

pdb), 709I (https://doi.org/10.2210/pdb709I/pdb), 705B (https://www.rcsb.org/structure/705B). Cryo EM maps have been deposited at the EMDB with the accession codes: 12734 (https://www.emdataresource.org/EMD-12734), 12735 (https://www.emdataresource.org/EMD-12735), 13839 (https://			
www.emdataresource.org/EMD-13839), 13840 (https://www.emdataresource.org/EMD-13840). All other data generated in this study are provided in the Supplementary Information/Source Data file or are available upon request. Requests for materials and correspondence should be addressed to LC and GB. Already published datasets used in this study are: 10KK (https://www.rcsb.org/structure/10KK), 3NG1 (https://www.rcsb.org/structure/3NG1), 2YHS (https://www.rcsb.org/structure/3J9W), 4UE4 (https://www.rcsb.org/structure/4UE4), 5GAF			
(https://www.rcsb.org/structure/5GAF).			
Field-specific reporting			
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Life sciences			
For a reference copy of the document with all sections, see <a href="mailto:nature.com/documents/nr-reporting-summary-flat.pdf">nature.com/documents/nr-reporting-summary-flat.pdf</a>			
Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences			

### Life sciences study design

field. NOT APPLICABLE

Sample size	Experiments were performed in (at least) two (biological) replicates (see also; replication).
Data exclusions	We did not exclude any data from consideration.
Replication	Fig. 1 and 2: Membrane insertion and floatation analyses have been performed in at least n=3 independent experiments. In case of LepB (Fig. 2) n=4 independent experiments were performed.  Fig. 3 c and d: GTPase data represent mean values (± SD) of n=3 replicates.
	Fig. 3 e: ITC measurments have been performed at least twice and MST data are presented as mean values +/- SD of n=2 biologically independent samples and each three technically independent measurements.
Randomization	No randomization was necessary for this study because investigators were comparing well studied specific proteins under well controlled conditions (e.g. addition of different nucleotides). No human or animal subjects were used in the study. Randomization is not generally used in this field.
Blinding	Investigators were not blinded. Blinding during collection was not needed because conditions were well controlled. Blinding is also not

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

necessary because the results are quantitative and did not require subjective judgment or interpretation. Blinding is not typically used in the

Materials & experimental systems	Methods	
n/a Involved in the study	n/a Involved in the study	
Antibodies	ChIP-seq	
<b>x</b> Eukaryotic cell lines	Flow cytometry	
Palaeontology and archaeology	MRI-based neuroimaging	
Animals and other organisms	'	
Human research participants		
Clinical data		
Dual use research of concern		
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