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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a Confirmed
The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
A description of all covariates tested
A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.
For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\square Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated
Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.
Software and code
Policy information about <u>availability of computer code</u>

Data collection

LightCycler 480 software (version 1.5.1.62) for qPCR data.

PacBio Sequel CtrlVer 6.0.1.52258 and SigProcVer 6.0.0.47712 for PacBio SMRT subreads.bam files.

Data analysis

Software used for the PLASTER pipeline is available in a docker container hosted on docker hub at bahlolab/plaster:v0.0, from which individual software versions can be obtained.

For the qPCR data analysis, the following software was used: R version 3.6.1 with packages dplyr version 1.0.2, ggplot2 version 3.3.2, magrittr version 1.5, readr version 1.3.1, tidyr version and mixtools version 1.2.

Custom software used is available at https://github.com/bahlolab/PLASTER.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about <u>availability of data</u>

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

		ng stage) is available through the NCBI Sequence Read Archive (SRA) (BioProject Accession tHub repository (https://github.com/bahlolab/PLASTER).	
Field-spe	ecific reporting		
Please select the o	ne below that is the best fit for your	research. If you are not sure, read the appropriate sections before making your selection.	
Life sciences	Behavioural & social s	ciences Ecological, evolutionary & environmental sciences	
For a reference copy of t	the document with all sections, see <u>nature.co</u>	n/documents/nr-reporting-summary-flat.pdf	
Life scier	nces study desig	า	
All studies must dis	sclose on these points even when th	e disclosure is negative.	
Sample size	The Solomon Islands Cohort was gathered as part of clinical trial that is not discussed or reported on in this manuscript. The sample size was determined based on the requirements of the clinical trial.		
Data exclusions	Samples were excluded when insufficient sequencing depth was attained for allele-typing.		
Replication	10 Control samples with previously chassessed.	aracterized CYP2D6 alleles from the Coriell Biobank were included, and concordance of results was	
Randomization	N/A		
Blinding	N/A		
-	-	aterials, systems and methods aterials, experimental systems and methods used in many studies. Here, indicate whether each material,	
		ot sure if a list item applies to your research, read the appropriate section before selecting a response.	
Materials & ex	perimental systems	Methods	
n/a Involved in th	·	n/a Involved in the study	
Antibodies		ChIP-seq	
Eukaryotic		Flow cytometry	
Palaeontology and archaeology MRI-based neuroimaging Animals and other organisms		MKI-based neuroimaging	
	search participants		
Clinical dat	·		
	esearch of concern		

Human research participants

Policy information about studies involving human research participants

Population characteristics

Population from the Tetere region of the Solomon Islands.

Recruitment

Individuals were recruited through local health care centers, inclusion criteria was positive diagnosis of vivax malaria. Exclusion criteria were infancy (< 1 year), pregnancy, a positive G6PD deficiency test or refusal or inability to provide informed written consent. The inclusion/exclusion criteria are unlikely to bias the results of the CYP2D6 allelic characterisation of this population.

Ethics oversight

This study was approved by the Solomon Islands Health Research Ethics Review Board (HRE #041/16) and the Walter and Eliza Hall Institute Human Research Ethics Committee (HREC #16/02).

Note that full information on the approval of the study protocol must also be provided in the manuscript.