Supplementary Table 1. List of initially screened computationally identified lead compounds as PTHR ligands. Indicated are predicted computed binding scores, and area under the curve (A.U.C.) of cAMP time courses induced by 1 nM PTH in the presence of 10 μ M of small molecules (mean \pm S.D. of *N* independent experiments), normalized to the PTH response without compounds and expressed as % values. In brackets is alternative nomenclature used in Source Data Files.

	Integrate		ted	Ν		
Compound			cAMP			
	MolPort ID	score	$(\text{mean} \pm \text{s.d.})$			
1 (Pitt8)	002-583-206	-8.41	32.3	±	9.5	3
2 (Pitt12)	039-313-655	-8.47	39.4	±	10.6	3
3 (Pitt1)	000-837-761	-8.16	82.7	±	10.0	2
4 (Pitt2)	000-749-199	-9.28	72.4	±	8.1	2
5 (Pitt3)	002-764-247	-8.24	50.6	±	10.2	2
6 (Pitt4)	019-692-354	-8.23	118.8	±	8.9	2
7 (Pitt5)	006-815-377	-8.62	79.6	±	8.9	2
8 (Pitt6)	002-582-026	-8.17	64.9	±	7.5	2
9 (Pitt7)	002-527-649	-8.31	70.3	±	6.3	2
10 (Pitt9)	000-829-806	-8.21	85.3	±	14.3	2
11 (Pitt10)	028-805-319	-8.33	92.5	±	12.0	2
12 (Pitt11)	016-916-811	-8.38	106.2	±	12.9	2
13 (Pitt13)	030-037-643	-8.42	96.1	±	10.9	2
14 (Pitt14)	003-269-448	-8.39	137.2	±	5.9	2
15 (Pitt15)	023-186-594	-8.33	41.9	±	18.9	2
16 (Pitt16)	010-715-401	-8.28	76.7	±	12.8	2
17 (Pitt17)	010-715-395	-8.33	99.9	±	9.2	2
18 (Pitt18)	010-807-325	-8.5	71.1	±	10.9	2
19 (Pitt19)	010-807-350	-8.5	75.0	±	7.7	2
20 (Pitt20)	002-369-638	-9.33	63.5	±	14.3	2
21 (Pitt21)	005-944-940	-8.54	62.0	±	5.5	2
22 (Pitt22)	021-757-743	-8.92	73.6	±	9.3	2
23 (Pitt23)	020-207-515	-8.61	111.2	±	10.7	2
24 (Pitt24)	020-198-951	-8.80	117.2	±	7.5	2
25 (Pitt25)	005-011-602	-8.40	119.1	±	11.6	2
26 (Pitt26)	002-284-377	-7.19	81.6	±	2.9	2
27 (Pitt27)	002-169-278	-7.71	100.9	±	11.2	2
28 (Pitt28)	007-831-482	-10.39	152.0	±	7.9	2
29 (Pitt29)	006-598-622	-9.74	135.2	±	4.8	2
30 (Pitt30)	010-693-446	-8.85	140.4	\pm	7.0	2

Compound	Structure	SMILES	Molecular weight	Γυαδ
1		Cc1ccc(cc1)C1=NN(C(C1)c1cc2 cc3OCCOc3cc2nc1Cl)C(=O)CC C(O)=O	479.92	4.51
2		COc1ccc(cc1)[C@@H]1CN(C[C @H]1C(O)=O)C(=O)OCC1c2ccc cc2-c2cccc12	443.499	4.744
3		[O-][N+](=O)c1cccc(c1)C1N2[C@H](Cc3c1[nH]c1ccccc31)C(=O)N(Cc1ccncc1)CC2=O	467.485	3.356
4		[O-][N+](=O)c1cccc(c1)N1C(=O)C2 C(C3N(N=Cc4cccc34)C2C(=O) c2ccc(F)cc2)C1=O	484.443	3.495
5	H,C,C,C,C,C,N,N,N,N,N,N,N,N,N,N,N,N,N,N,	Cc1ccc(cc1C)-c1cc(- c2ccc(C)c(C)c2)n(Cn2ccc(n2)[N +]([O-])=O)n1	401.47	5.061
6		COc1ccc(cc1)C1CC(=NN1C(=O)CN=[N+]=[N-])c1cccc2ccccc12	385.427	4.836
7		CC(=O)Nc1ccc2c(c[nH]c2c1)C(N1CCN(CC1)c1cc(C)c2ccccc2n1)C(O)=O	457.534	4.58

Supplementary Table 2. Detailed chemical properties of hit compounds.

8	F	[O-	483.455	4.132
	0][N+](=O)c1cccc(c1)C(=O)[C@ @H]1[C@H]2[C@@H](C3N1C		
		= $Cc1ccccc31)C(=O)N(C2=O)c1c$		
		ccc(F)c1		
9	HO	Cc1ccc2c(NC(=O)C22NC(CCC(481.93	2.742
		O = O [C(a)(a)H] [C(a)(a)H] [C(a)H] [C(a)H] [C(a)(a)H] [C(a)(a)(a)H] [C(a)(a)H] [C(a)(a)H] [C(a)(a)H] [C(a)(a)H] [C(a)(
	HE			
10		COc1ccccc1CN1CC(=O)N2[C@	496.523	3.97
		H](Cc3c([nH]c4ccccc34)C2c2ccc c(c2)[N+]([O-])=O)C1=O		
11	HJC	Cc1cccc(c1)C1CC(=NN1C(=O)C CC(O)=O)c1c(C)nc2ccccc2c1-	477.564	6.061
	H,C N-N	c1ccccc1		
	HO			
12	r start	CN(CC(=0)N1N=C(CC1c1cccc(478.456	4.323
	CH3	C1)[N+]([O-])=O)c1ccc(F)cc1)C(=O)c1cccc(
		F)c1		
	N N N N			
	, second se			
13	F HO FO	CCOc1cccc2cc(C3CC(=NN3C(=	465.93	5.138
		O)CCC(O)=O)c3ccc(C)cc3)c(Cl) nc12		
	CH ₃			
14	\bigcirc	CC1CCCN(C1)S(=0)(=0)c1ccc(492.64	3.634
		2)c(c1)[N+]([O-])=O		
	CH ₃			
15		COc1ccc2nc(Cl)c(cc2c1)C1CC(= NN1C(=O)CCC(O)=O)c1ccc(F)c	455.87	4.578
		c1		
	но			

16		OC(=O)c1cc(NS(=O)(=O)c2ccc(F)c(F)c2)ccc1N1CCN(CC1)c1cc c(F)cc1	491.49	3.93
17		CCc1ccc(cc1)S(=O)(=O)Nc1ccc(N2CCN(CC2)c2ccc(F)cc2)c(c1) C(O)=O	483.56	4.214
18		OC(=O)c1ccc(N2CCCN(CC2)c2 ccc(Cl)nn2)c(NC(=O)c2ccc(Cl)c c2)c1	486.35	4.451
19		OC(=O)c1ccc(N2CCCN(CC2)c2 ccc(Cl)nn2)c(NC(=O)c2ccc(F)cc 2)c1	469.9	3.936
20		Cc1ccc(cc1)C1=NN(C(C1)c1cn(nc1-c1ccc(C)cc1)- c1ccccc1)C(=O)CCC(O)=O	492.579	5.699
21		CC1(C)CC[C@@]2(CC[C@]3(C)C(=CCC4[C@@]5(C)C[C@@H](O)[C@H](O)C(C)(C)C5CC[C @@]34C)[C@@H]2[C@@H]1 O)C(O)=O	488.709	5.175
22	HN CH3 NH	Cc1c(- c2cc(NCc3ccncc3)nc3[nH]ccc23) c(=O)[nH]c2cccc12	381.439	4.387
23	NH NH HN CH ₃	COCCNc1cc(- c2ccc3c(c2)[nH]c2c3cn[nH]c2= O)c2cc[nH]c2n1	374.404	3.006
24		Nc1nc(cc(- c2ccc(O)c(O)c2)c1C#N)- c1cc(F)cc(c1)C(F)(F)F	389.31	4.439

25		Fc1cccc(c1)- c1n[nH]c2CCN(Cc12)C(=O)CCn 1ccc(=O)[nH]c1=O	383.383	1.041
26		Cc1cc(\C=C(/C#N)c2nc3ccccc3[nH]2)c(C)n1- c1sc(C)c(C)c1C(O)=O	416.5	5.411
27		[O-][N+](=O)c1cccc(\C=C2/CCCC3 C(NN=C23)c2cccc(c2)[N+]([O-])=O)c1	378.388	4.387
28	$H_3C \longrightarrow N \longrightarrow N \longrightarrow N \longrightarrow N \longrightarrow CH_3$ $H_3C \longrightarrow N \longrightarrow N \longrightarrow CH_3$ $H_3C \longrightarrow CH_3$	Cc1n(nc2c(nnc(C)c12)N1CCN(C C1)C(=O)Nc1cc(C)cc(C)c1)- c1ccc(C)cc1	469.593	4.712
29		Cc1cccc(c1)C1N(CCc2ccc(OCc3 nc(co3)C(=O)NC3CC3)cc12)C(= O)c1ccco1	497.551	4.835
30		CCC(=O)Nc1cccc(c1)- c1noc(n1)C1CCCN(C1)C(=O)c1 cc2cccc2[nH]1	443.507	4.586



Supplementary Figure 1. Effect of computationally identified small molecules on PTH-induced cAMP production. Averaged cAMP time-courses following brief stimulation with 1 nM PTH₁₋₃₄ without (*black*) or with (*red*) 10 μ M corresponding compounds measured by FRET changes from HEK293 cells stably expressing the recombinant human PTHR. The bars graph represents the area under the curve (AUC) of cAMP time-courses. Data were normalized to the maximal forskolin induced cAMP response, which is set to 100%. Error bars represent the mean values \pm s.d. of N = 3 (Pitt8 and Pitt12) or N = 2 (all the rest compounds) independent experiments.



Supplementary Figure 2. Basal cAMP controls. (a, b) Effects of Pitt8 and Pitt12 on Fsk-mediated cAMP in HEK-293 cells recorded via either the Glo-sensor (a) or FRET (b) assays. Mean \pm s.d. of N = 4 (Pitt12), and 3 (Pitt8) experiments carried out in duplicate for (a), and N = 3 experiments for (b).



Supplementary Figure 3. Selectivity of compounds. (a–c) Averaged cAMP time-courses following brief stimulation with either isoproterenol (Iso), vasopressin (AVP) or PTHrP₁₋₃₆ without (*black*) or with (*colored*) 10 μ M compounds measured by FRET changes from HEK293 cells expressing the β_2 -adrenergic receptor (β_2 AR, panel a), a HA-tagged vasopressin type 2 receptor (V2R, panel b), or the PTHR (panel c) and a FRET-based cAMP sensor Epac^{CFP/YFP}. Bars represent the corresponding quantitation of cAMP responses by measuring the area under the curve (A.U.C.) from 0 to 20 min. Data are the mean \pm s.e.m. of N = 3 independent experiments with n = 16 (DMSO), 21 (Pitt8), 16 (Pitt12) cells examined in panel a, n = 41 (DMSO), 29(Pitt8), 36 (Pitt12) cells examined in panel b, and n = 54 (DMSO), 49 (Pitt8), 48 (Pitt12) cells examined in panel c. P values were assessed by two-tailed Student's *t*-test and are *P=0.037, ***P=0.012.



Supplementary Figure 4. Effect of Selected Pitt molecules on native PTHR. (a, b) Averaged cAMP time-courses following brief stimulation with PTH_{1-34} without (*black*) or with (*colored*) 10 µM Pitt molecules measured by FRET changes from RPTEC (a) or osteoblasts (b) expressing the FRET-based cAMP sensor Epac^{CFP/YFP}. Bars represent the corresponding quantitation of cAMP responses by measuring the area under the curve (A.U.C.) from 0 to 20 min. Data are the mean ± s.e.m. of N = 3 independent experiments with n = 29 (DMSO), 44 (Pitt8) and 33 (Pitt12) cells examined in panel a, and n = 23 (DMSO), 26 (Pitt8) and 26 (Pitt12) cells examined in panel b. *P* values were assessed by Paired two-tailed Student's *t*-test and are **P*=0.041 (Pitt8) and **P*=0.023 (Pitt12).



Supplementary Figure 5. Effect of small molecules on G-protein coupling to PTHR. (a, b) Timecourse recorded by single cell FRET assay in HEK-293 cells expressing PTHR-CFP and mini-G proteins mGs-YFP (a) or mGq-YFP (b) in response to PTH \pm the indicated Pitt molecule. Mean \pm s.e.m. of N = 3 experiments.