Supplementary Table 1. Multiplex cytokine analysis of the plasma from patient with myopericarditis after COVID-19 vaccine

Soluble factors	Healthy controls	Vaccine controls	Patient day 3	Patient day 4
(ng/ml)	(n=5, median)	(n=5, median)	(n=1)	(n=1)
Increased in patient compared to both healthy and vaccine controls				
IL-18	27.32	109.18	247.54	139.98
IL-27	467.30	1747.30	4556.30	1826.40
CXCL9 (MIG)	1077.30	1780.28	3951.15	2608.25
CXCL10 (IP-10)	75.54	57.95	305.83	99.95
Increased in patient compared to healthy controls at a same level as vaccine controls				
IFN-γ	0.00	15.92	11.82	12.91
IL-1Ra	0.00	24.16	21.42	8.34
IL-6	0.00	7.65	12.15	3.99
IL-7	0.00	1.55	0.99	0.86
IL-10	1.21	10.88	10.64	14.10
IL-12p40	15.80	56.99	40.82	66.33
IL-15	0.00	9.84	5.31	3.03
TNF-α	4.16	36.64	20.31	28.53
CCL4 (MIP-1β)	11.68	24.93	26.42	26.19
Increased in patient compared to healthy controls but lower than vaccine controls				
IFN-α	0.00	43.30	2.70	0.00
IL-1a	0.00	197.11	26.46	17.89
IL-1β	0.00	8.26	0.19	0.00
IL-2	0.00	1.14	0.31	0.26
IL-8	1.27	23.45	2.09	3.12
IL-9	0.00	161.22	74.45	72.69
IL-12p70	0.00	4.82	0.49	0.61
IL-17A	0.00	2.93	0.42	0.39
IL-17E/IL-25	205.36	3759.12	437.43	171.12
IL-17F	0.00	1679.68	62.38	65.94
IL-22	0.00	230.45	28.36	25.52
TNF-β	0.00	17.82	0.91	0.91
$CCL3 (MIP-1\alpha)$	0.00	85.27	16.01	21.37
CCL7 (MCP-3)	0.00	45.48	12.05	17.76
CXCL1 (GROa)	2.38	25.45	3.02	2.43
EGF	0.00	589.59	9.92	7.43
FGF-2	0.00	250.29	55.14	61.12
G-CSF	0.00	18.84	7.95	0.41
M-CSF	0.00	645.58	113.32	13.65
PDGF-AA	100.75	11535.95	440.48	118.8/
PDGF-AB/BB	1821.33	86247.27	5663.03	2826.79
IGF-α	0.00	116.01	5.11	5.38
VEGF-A	0.00	/5.60	6.23	3.48
sCD40L	0.00	545.34	121.03	63.51
Entry in 242.02 59.12 154.05 156.70				
Eotaxin	No difference	Joild botwoon nationt and has	134.23	130.70
II_3		0 00		0.00
IL-5 II -4	0.00	1.44	0.00	0.00
IL- -	2.00	7.01	1.50	1 28
IL-5 II_13	0.00	14.52	0.00	1.30
CCL 2 (MCP 1)	400.14	660 /2	0.00	0.00 101.66
CCL2 (MCF-1)	701.12	2777 62	++7.04 1773 50	404.00 870.00
CCL 22 (MDC)	717.12	120/ 05	502.00	571 76
CX3CL1 (Fractalkine)	24.50	1004.05	392.89 13.70	7/1./0
FI T_3I	24.39	101.29	30.86	21 /2
GM-CSF	0.00	0.00	0.00	0.00



Supplementary Figure 1. Gating strategy for flow cytometry analysis of patient PBMC.



Supplementary Figure 2. Activated circulating T cells in patient with myopericarditis after COVID-19 mRNA vaccine. PMBC was isolated from the patient with myopericarditis on days 3 (D3) and 4 (D4) after COVID-19 mRNA vaccination. For flow cytometry analysis, four healthy controls (Healthy) and three individuals who had been vaccinated in the past 2 to 3 days (Vaccine) were used as control. For RT-PCR, three healthy and three vaccinated individuals were used as control. (A) Histograms of CD107a expression in PBMC NK cells of the patient with COVID-19 mRNA vaccine-related myopericarditis. (B) CD4⁺ and CD8⁺ T cell frequencies among PBMC CD45⁺ cells in the patient with COVID-19 vaccine-related myopericarditis. CD4⁺ T cells were gated on CD45⁺CD3⁺CD4⁺, and CD8⁺ T cells were gated on CD45⁺CD3⁺CD4⁺ (Supplementary Figure 1). (C,D) Flow cytometry plots (C) and frequencies (D) of CD8⁺ T cell subsets in the patient PBMC defined by CCR7⁺CD45RA⁺ (T_N; naïve), CCR7⁺CD45RA⁻ (T_{CM}; central memory), CCR7⁻CD45RA⁻ (T_{EM}; effector memory), and CCR7⁻CD45RA⁺ (T_{EMRA}; effector memory expressing CD45RA).



Supplementary Figure 3. Gating strategy for flow cytometry analysis of cardiac infiltrating immune cells from IL-18-injected mice.



Supplementary Figure 4. Activated cardiac T cells and macrophages in IL-18-injected mice. Mice were intraperitoneally injected with recombinant IL-18 (n=5) or PBS (n=5) every other day for eight days. Mice were sacrificed on day 10. (**A**) Number of CD45⁺ infiltrating cells in hearts of IL-18-injected analyzed by flow cytometry. P-value: 0.0687. (**B**) Number of CD4⁺ T cells (CD45⁺NKp46⁻CD11b⁻CD3⁺CD4⁺), CD8⁺ T cells (CD45⁺NKp46⁻CD11b⁻CD3⁺CD8⁺), B cells (CD45⁺NKp46⁻CD11b⁻CD19⁺), macrophages (CD45⁺NKp46⁻CD11b⁺Ly6G⁻F4/80⁺CD64⁺Ly6C⁻), monocytes (CD45⁺NKp46⁻CD11b⁺Ly6G⁻F4/80⁺CD64⁺Ly6C⁺CCR2⁺), and neutrophils (CD45⁺NKp46⁻CD11b⁺Ly6G⁺) in hearts of IL-18-injected mice. P-value: 0.0196 (neutrophil). (**C**,**D**) Flow cytometry plots (**C**) and frequencies (**D**) of CD8⁺ T cell subsets defined by CD62L⁺CD44⁻ (naïve), CD62L⁻CD44⁺ (effector), and CD62L⁺CD44⁺ (memory) in IL-18injected mouse hearts. P-value: 0.0385 (left). (**E**,**F**) Flow cytometry plots (**E**) and frequencies (**F**) of MHCII expression on macrophages in IL-18-injected mouse hearts. P-value: 0.0011. Student *t*-test was used for statistical analysis. *P < 0.05; **P < 0.005.



Supplementary Figure 5. No involvement of autoantibodies B cells in COVID-19 vaccinerelated myopericarditis. (A) Representative image of indirect immunofluorescence assay for detecting anti-cardiac autoantibodies in the plasma of the patient with COVID-19 vaccine-related myopericarditis. The plasma was collected on day 3 after COVID-19 mRNA vaccine. Scale bars: $200 \ \mu\text{m}$. (B) Plasma levels of antibodies against myosin peptide S2-16 or S2-28 measured by ELISA in the patient with myopericarditis on days 3 (D3) and 4 (D4) after COVID-19 vaccination. As controls, five healthy controls (Healthy) were tested. (C) Frequencies of plasma cells among PBMC CD45⁺ cells from the patient with COVID-19 vaccine-related myopericarditis. Plasma cells were gated on CD45⁺CD19⁺CD38⁺CD27⁺ (Supplementary Figure 1). As controls, four healthy controls (Healthy) and three individuals who had been vaccinated in the past 2 to 3 days (Vaccine) were tested. (D) Frequencies of PBMC B cell subsets from the patient with COVID-19 vaccine-related myopericarditis defined by IgD⁺CD27⁻ (B_N; naïve), IgD⁺CD27⁺ (B_{NSM}; non-class-switched memory), and IgD⁻CD27⁺ (B_{SM}; class-switched memory). B cells were gated on CD45⁺CD19⁺ (Supplementary Figure 1). (E) Plasma levels of IgA and IgG against SARS-CoV-2 spike protein measured by ELISA in the patient with myopericarditis on days 3 (D3) and 4 (D4) after COVID-19 vaccination. As controls, five controls who had been vaccinated in the past 2 to 3 days (Vaccine) were tested.