

1 **Supplemental Figure Legends**

2

3 **Figure S1. The neutralization activity of DH1047, ADG-2, and REGN 10933 against**
4 **sarbecoviruses.** Neutralization is shown for SARS-CoV-2 B.1.351, SARS-CoV urbani, and
5 RsSHC014. Data are representative of two technical replicates.

6

7 **Figure S2. The binding activity of cross-reactive antibodies against MERS-CoV and human**
8 **common-cold coronaviruses.** The binding activity of four broadly neutralizing antibodies
9 against SARS-CoV-2 NTD, MERS-CoV spike protein, HCoV-OC43 spike protein, HCoV-NL63
10 spike protein, and HCoV-229E spike protein is shown for DH1235, DH1073, DH1046, and
11 DH1047. Data are representative of two technical replicates. OD, optical density.

12

13 **Figure S3. Coronavirus cross-reactive antibodies exhibit distinct binding modes to SARS-**
14 **CoV-2 receptor binding domain (RBD).** (A) Three-dimensional reconstruction from negative
15 stain electron microscopic images of DH1073 (purple) in complex with Hexapro stabilized
16 SARS-CoV-2 Spike ectodomain (gray). On the left and center, the map is shown in two
17 orthogonal views. On the right, the map is shown as a transparent surface, rigidly fit with a 1-
18 RBD-up spike model, PDB ID: 6ACK. (B) Three-dimensional reconstruction of DH1235 (blue)
19 in complex with Hexapro stabilized Spike ectodomain (gray). The rigidly fit 3-RBD-up spike
20 model on the right was generated from PDB ID: 6ACK. Fit models in (A) and (B) were used to
21 compare the binding of DH1073 and DH1235 to known RBD-directed nAbs. (C) Four
22 orthogonal views of RBD binding by DH1073 (purple) and DH1235 (blue), compared to known
23 RBD nAbs, shown as molecular surface: C105 (pale cyan, Class 1, PDG ID 6XCN and 6XCA),

24 DH1041 (light blue, Class 2, PDB ID: 7LAA), S309 (wheat, Class 3, PDB ID: 7BEP), CR3022
25 (pink, Class 4, PDB ID: 6W41), and DH1047 (green, PDB ID: 7LD1). The last two views
26 correspond to the same views shown in Fig. 3D. DH1073 overlaps with DH1235, C105 (Class
27 1) and CR3022 (Class 4). DH1235 overlaps DH1073, DH1047, C105 and CR3022. **(D)**
28 Antibody competition for binding to SARS-CoV-2 spike protein stabilized with two prolines (S-
29 2P) is shown by results from surface plasmon resonance (SPR) assays. The antibody used to
30 capture S-2P is shown as the row title. The antibody flowed over the captured spike is shown as
31 column titles. + indicates blocking and competition; - indicates no blocking of binding and no
32 competition.

33

34 **Figure S4. Negative Stain Electron Microscopy (NSEM) of DH1047 bound to bat**

35 **RsSHC014 and SARS-CoV spike ectodomains.** (A) Representative 2D class averages of bat
36 RsSHC014 S-2P ectodomain bound to DH1047 Fab are shown. (B) An overlay of 3D
37 reconstructions of DH1047 bound to bat RsSHC014 2P (gray) and SARS-CoV-2 HexaPro
38 (purple) spike ectodomains is shown. (C) Representative 2D class averages of SARS-CoV S-2P
39 ectodomain bound to DH1047 Fab are shown. (D) An overlay of 3D reconstruction of DH1047
40 bound to bat SARS-CoV 2P (gray) and SARS-CoV-2 HexaPro (purple) spike ectodomains is
41 shown. The red boxes in (A) and (C) indicate the classes that show DH1047 Fab bound to spike.

42

43 **Figure S5. Cryo-EM data processing for the SARS-CoV spike ectodomain bound to**

44 **DH1047.** (A) A representative cryo-EM micrograph is shown. (B) Cryo-EM contrast transfer
45 function (CTF) fit is shown. (C) Representative 2D class averages from Cryo-EM dataset are
46 shown. (D and E) Ab initio reconstruction (D) and a refined map (E) are shown. (F) Fourier

47 shell correlation curves are shown. **(G)** A refined cryo-EM map colored by local resolution is
48 shown. **(H)** Zoom-in images show the SD1, NTD, HR1/CH and RBD/Fab contact regions in the
49 structure. The cryo-EM map is shown as a blue mesh and the fitted model is in cartoon
50 representation, with residues shown as sticks.

51

52 **Figure S6. The affinity data of DH1047 against SARS-CoV and RsSHC014 spikes. SPR**
53 binding experiments of DH1047 against SARS-CoV-2 Toronto and RsSHC014. Binding affinity
54 measurements are shown in the tables and response units (RU) as a function of time in seconds
55 (s) is shown for both SARS-CoV and RsSHC014. SPR experiments were repeated twice.

56

57 **Figure S7. Hematoxylin and eosin staining of lungs from SARS-CoV infected mice collected**
58 **at 4 days post infection.**

59 **(A to D)** mice were treated with control CH65 antibody prophylactically **(A)** or therapeutically
60 **(B)** or treated with DH1047 antibody prophylactically **(C)** or therapeutically **(D)**. Prophylactic
61 treatment was administered 12 hours before infection, and therapeutic treatment was
62 administered 12 hours after infection. Pathologic features of acute lung injury were scored using
63 two separate tools: the American Thoracic Society Lung Injury Scoring (ATS ALI) system.
64 Using this ATS ALI system, we created an aggregate score for the following features:
65 neutrophils in the alveolar and interstitial space, hyaline membranes, proteinaceous debris filling
66 the air spaces, and alveolar septal thickening. Three randomly chosen high power ($\times 60$) fields of
67 diseased lung were assessed per mouse. Representative images are shown. All images were taken
68 at the same magnification. The black bar indicates 100 μ m scale.

69

70 **Figure S8. Comparison of DH1047 binding footprint to that of other RBD-directed Abs**
71 **that bind overlapping epitopes.**

72 (A) RBD is shown in black and ACE2 binding footprint is colored yellow. Binding footprints of
73 antibodies DH1047 (PDB: 7LD1), CR3022 (PDB: 6YLA), EY6A (PDB: 6ZCZ), H014 (PDB:
74 7CAH), COVA1-16 (PDB: 7JMW) and S304 (PDB: 7JW0) are shown in green. (B) The overlay
75 shows DH1047 bound RBD and RBD bound to nanobodies VHH V (PDB: 7KN6), VHH W
76 (PDB: 7KN7) and VHH-72 (PDB: 6WAQ).

77 **Figure S9. Binding of DH1047 mutants to the SARS-CoV-2 S-2P ectodomain.**

78 (A) Binding assays were performed by Biolayer Interferometry using alanine scanning mutants
79 of DH1047. (Top) Interferometry curves are shown. (Bottom) Binding of DH1047 IgG and mutants
80 to SARS-CoV-2 S-2P ectodomain was normalized to the binding observed for DH1047 WT IgG
81 (dotted line). (B) Binding of purified DH1047 WT IgG and selected mutants to SARS-CoV-2 S-
82 2P ectodomain was measured by SPR. The spike was captured by its C-terminal TwinStreptactin
83 tag on a SA chip. Binding to the antibodies was measured by flowing over 200 nM solutions of
84 the IgG in HBS-EP+ buffer. (C) The binding interface of DH1047 to RBD (gray) is shown. The
85 DH1047 residues that were mutated are shown as sticks and labeled.

86

87 **Figure S10. DH1047 and ADG-2 bind the RBD of SARS-CoV and SARS-CoV-2 spike**
88 **ectodomains using a similar footprint.**

89 (A) A cartoon representation is shown for DH1047 (colored in pale green) bound to the RBD
90 (gray surface, ACE2 binding site in yellow) of SARS-CoV spike ectodomain and for ADG-2
91 (cyan) bound to SARS-CoV-2 spike ectodomain. The homologous Fab ADI-19425 (PDB 6APC)

92 was docked in the ADG-2 cryo-EM map (EMD-23160) to generate the model. **(B)** DH1047 and
93 ADG-2 bind partially overlapping binding sites on the RBD. **(C)** The therapeutic activity of
94 DH1047, ADG-2, and control mAb in aged mice infected with SARS-CoV-2 B.1.351 was
95 measured. Animals were treated at 6 hours post infection with the indicated antibodies and
96 monitored for percent of starting weight (left) as well as lung viral replication titers (middle), and
97 lung pathology (right) at 4 days post infection. Data are presented as mean±SEM (weight) or as
98 mean (lung viral titer and lung pathology). Change in weight was compared using a two-way
99 ANOVA followed by Tukey's multiple comparisons test, and differences in lung viral load and
100 pathology were evaluated using a one-way ANOVA followed by Dunnett's multiple
101 comparisons test. Differences in survival were measured by a Mantel-Cox test. *p<0.05;
102 **p<0.01; ***p<0.001.

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111 **Table S1. Monoclonal antibody neutralization screen against SARS-CoV-2 2AA MA,**
112 **SARS-CoV, WIV-1, and RsSHC014**

113

114 **Table S2. Cryo-EM data collection and refinements statistics.**

115

116 **Table S3. Immunogenetic characteristics of broadly cross-reactive mAbs.**

117

Figure S1

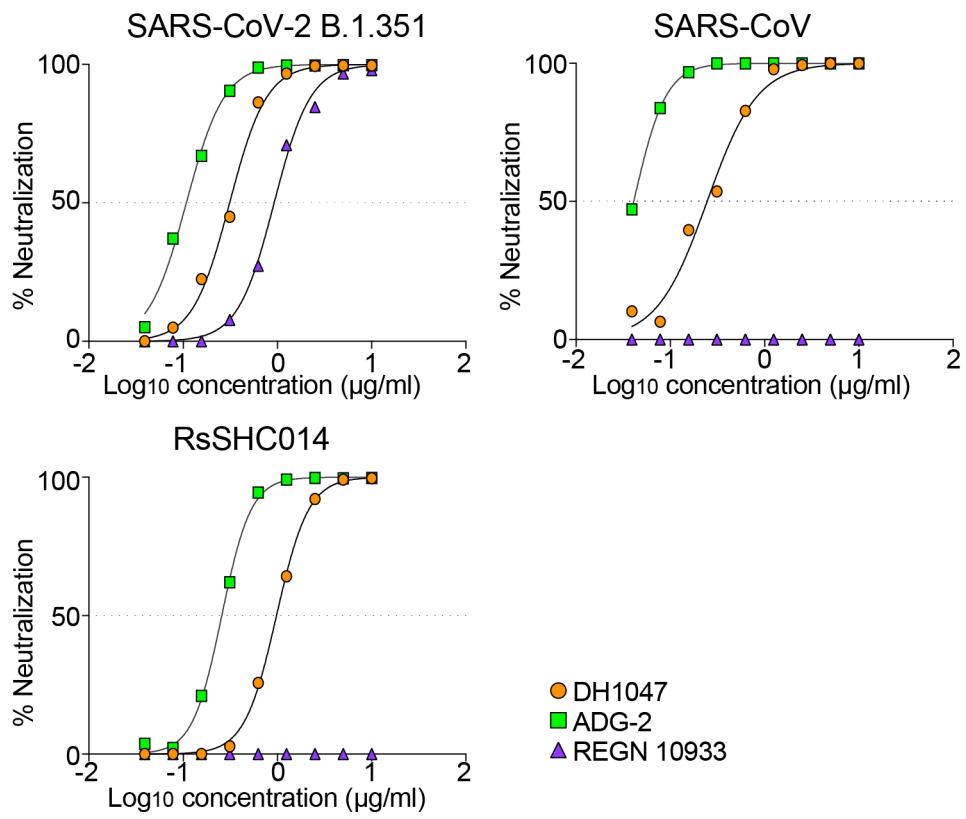


Figure S2

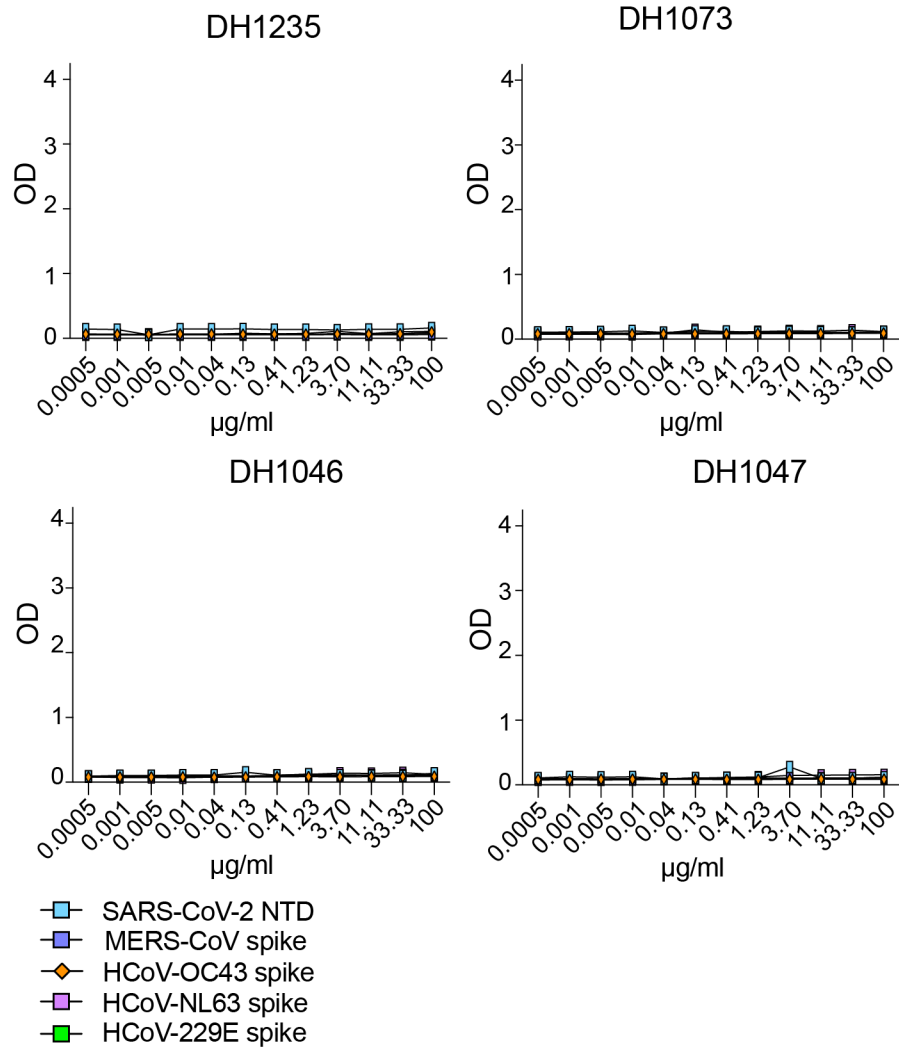
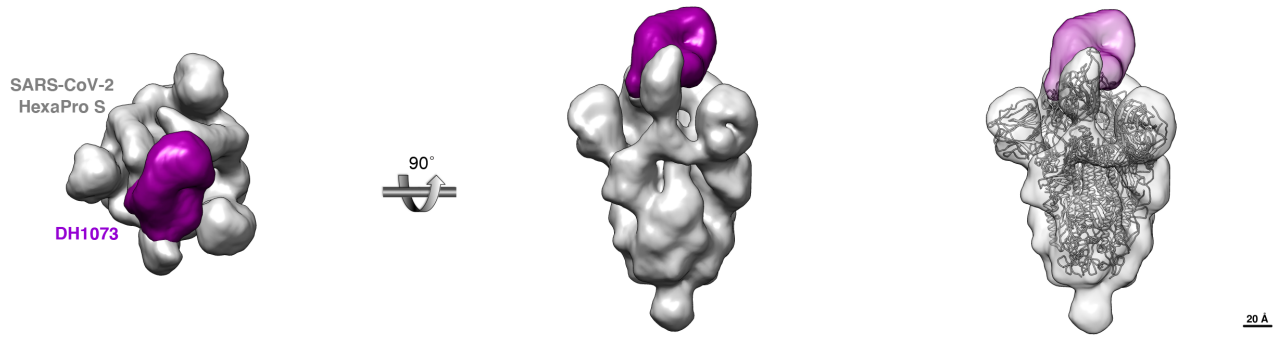
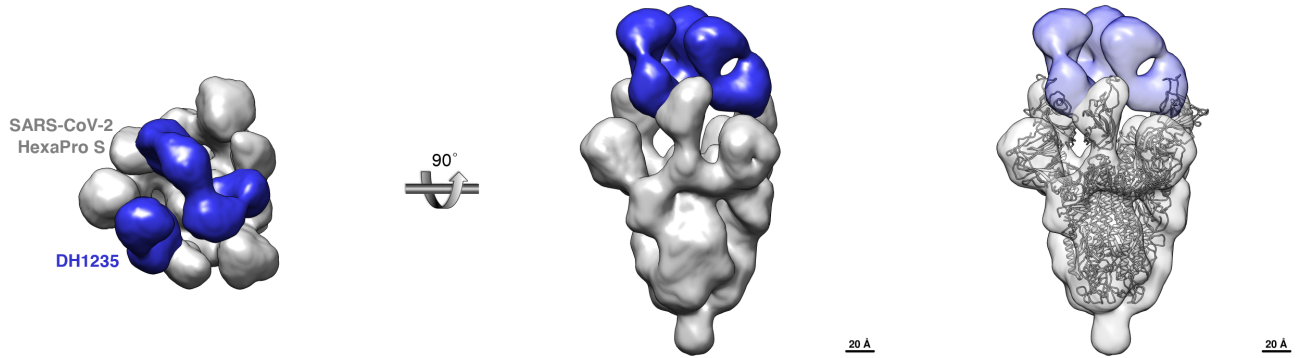


Figure S3

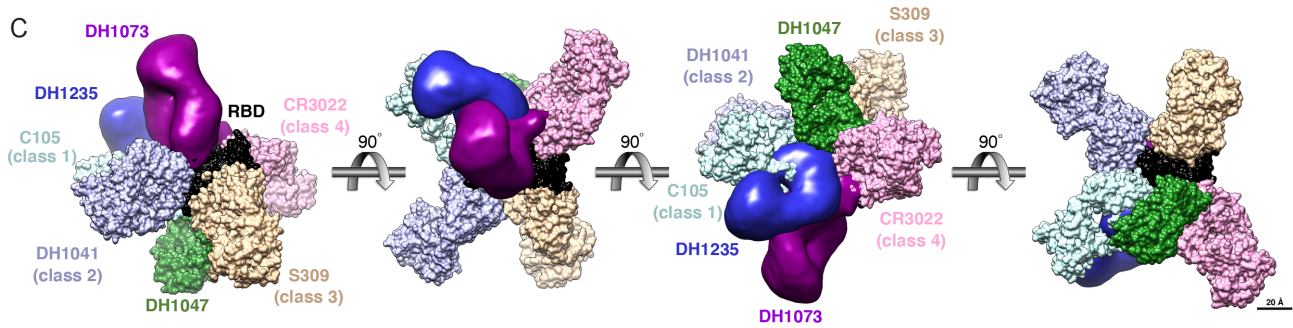
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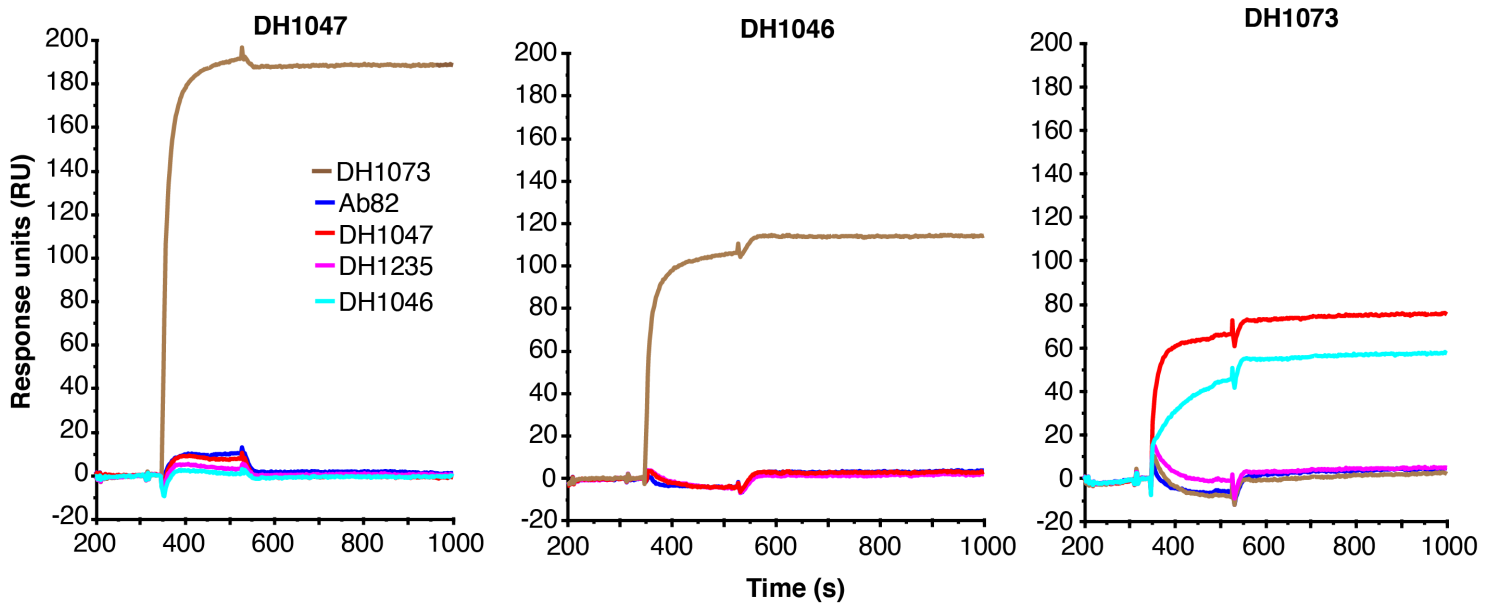
B



C



D

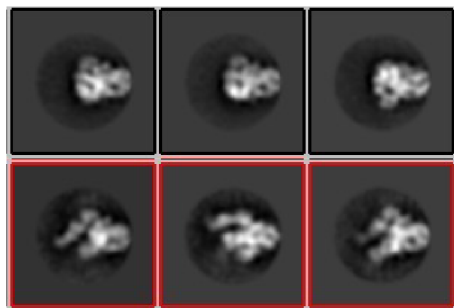


mAb	DH1046	DH1047	DH1073	DH1235
DH1046	+	+	-	+
DH1047	+	+	-	+
DH1073	-	-	+	+

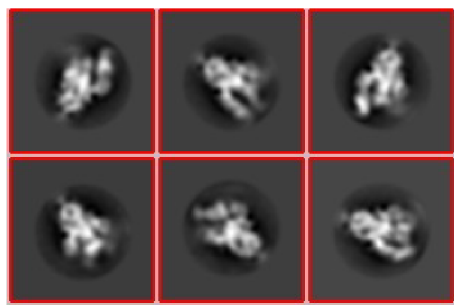
- no competition
+ no competition

Figure S4

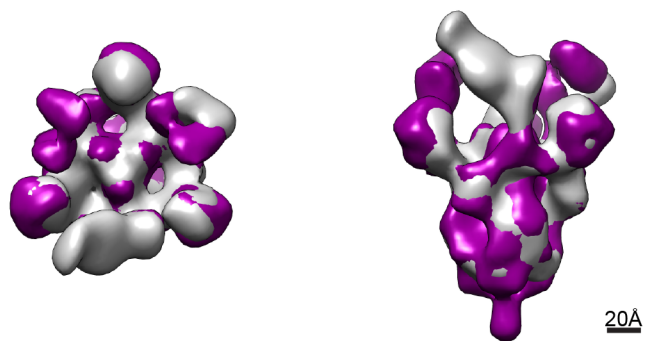
A



C



B



D

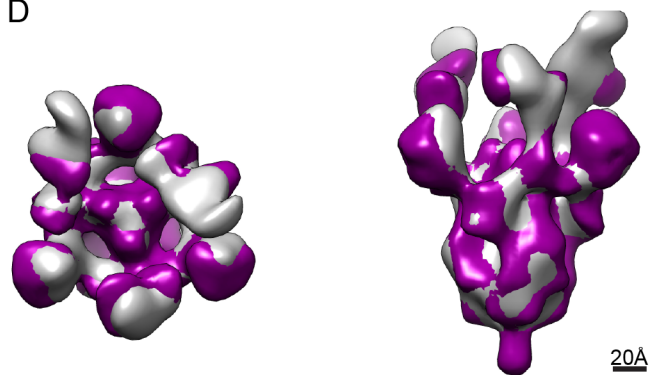


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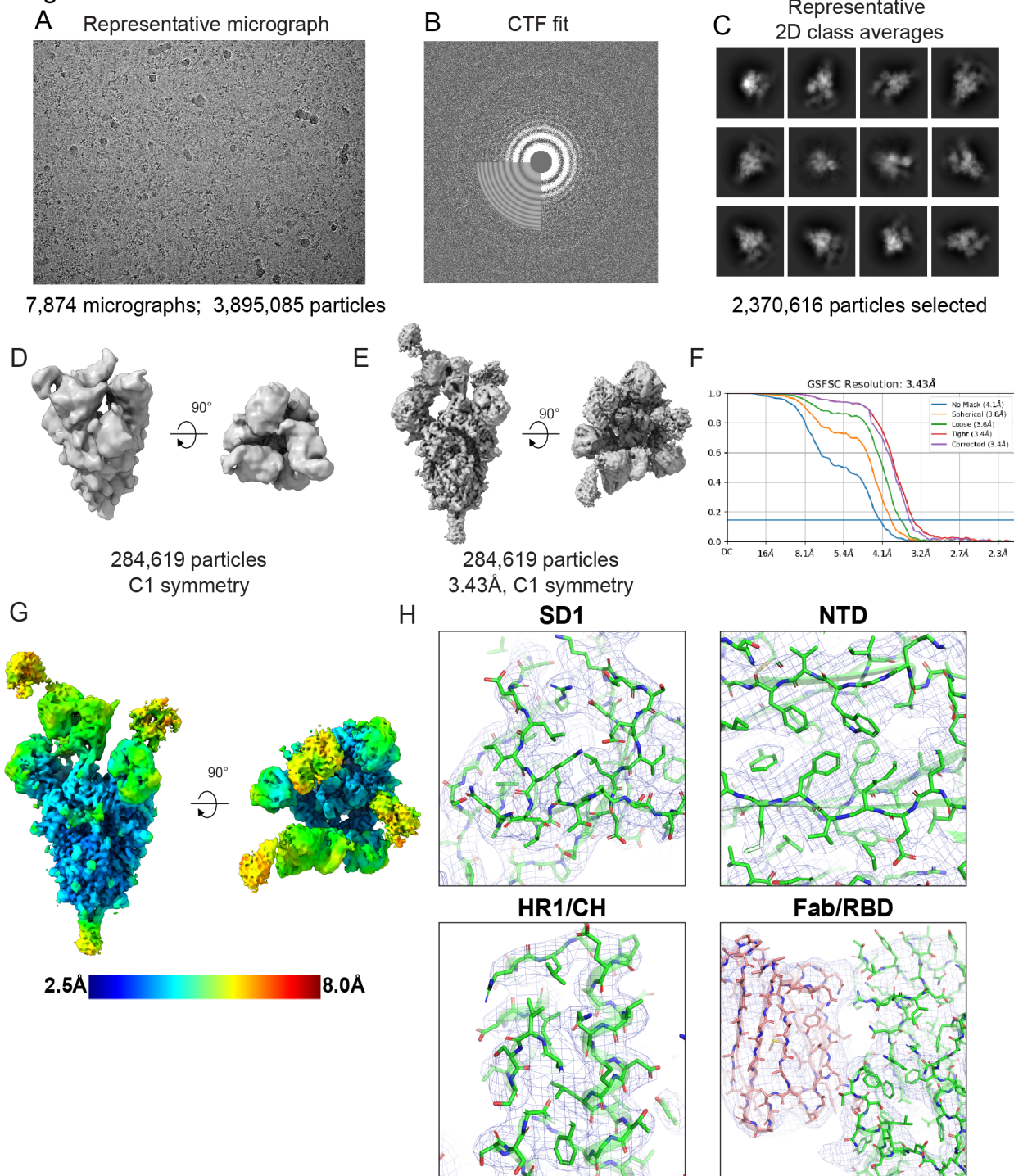
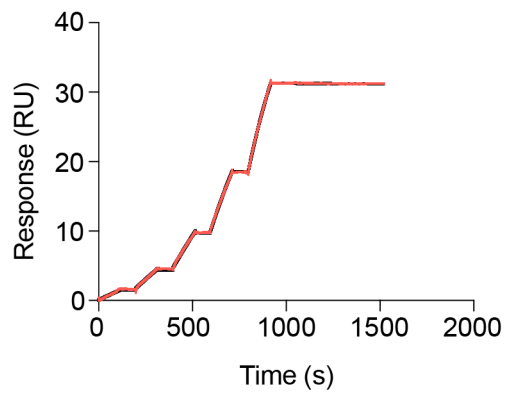


Figure S6

SARS-CoV Toronto spike			
Fab	ka (1/Ms)	kd (1/s)	KD(nM)
DH1047	9.622E+4	<1.0E-5	<0.1

RsSCH014 spike			
Fab	ka (1/Ms)	kd (1/s)	KD(nM)
DH1047	8.602E+4	<1.0E-5	<0.1

DH1047 Fab vs. SARS-CoV Toronto



DH1047 Fab vs. RsSCH014

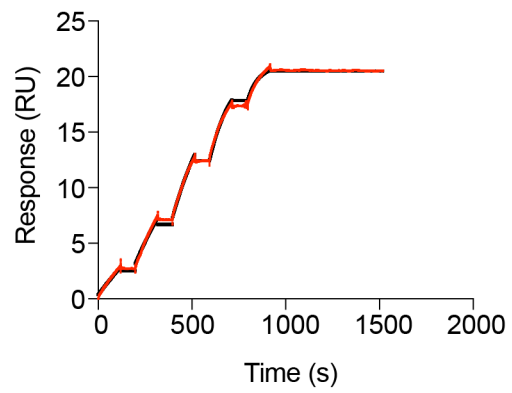
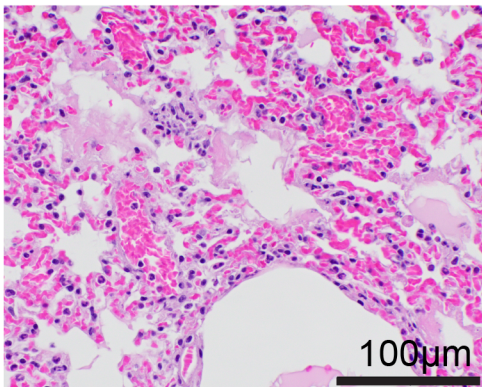
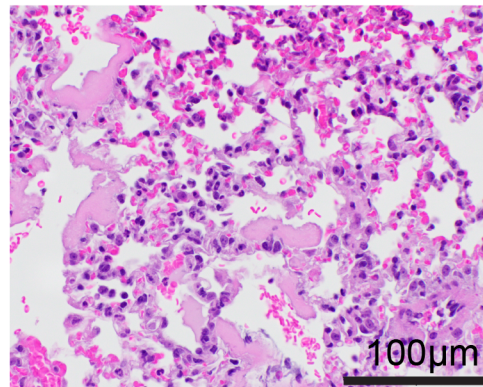


Figure S7

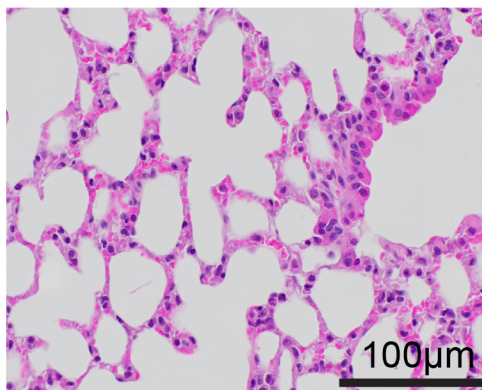
A CH65 control -12hr prophylaxis



B CH65 +12hr therapy



C DH1047 -12hr prophylaxis



D DH1047 +12hr therapy

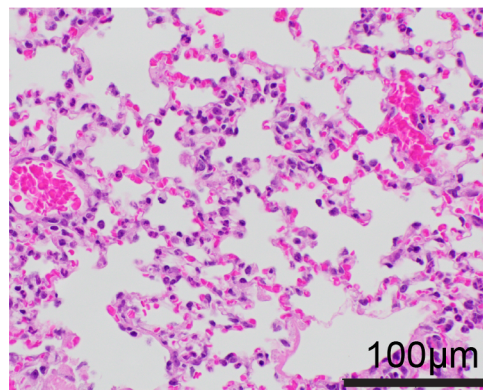
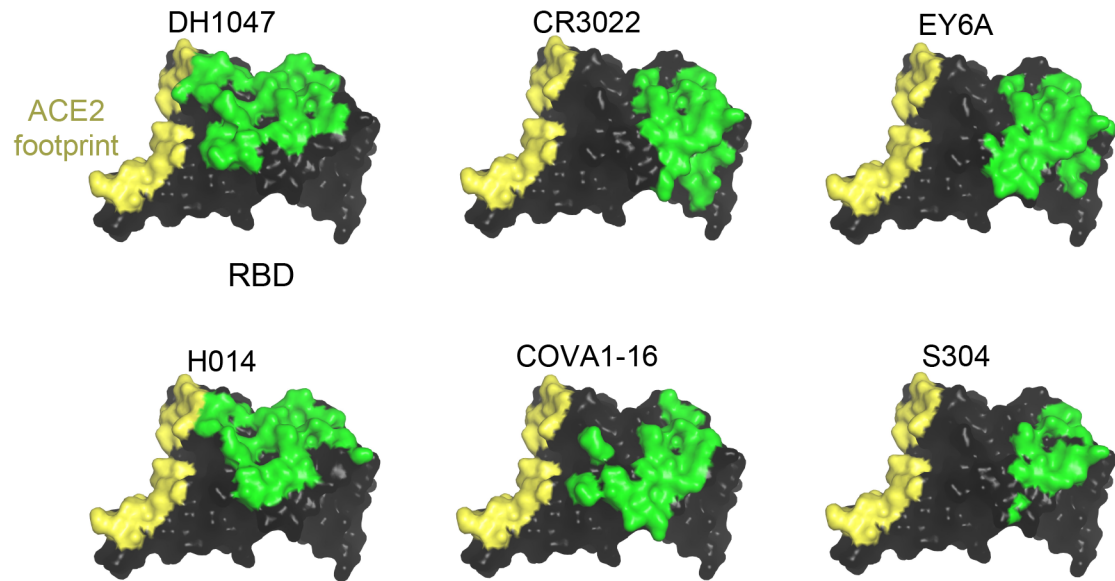


Figure S8

A



B

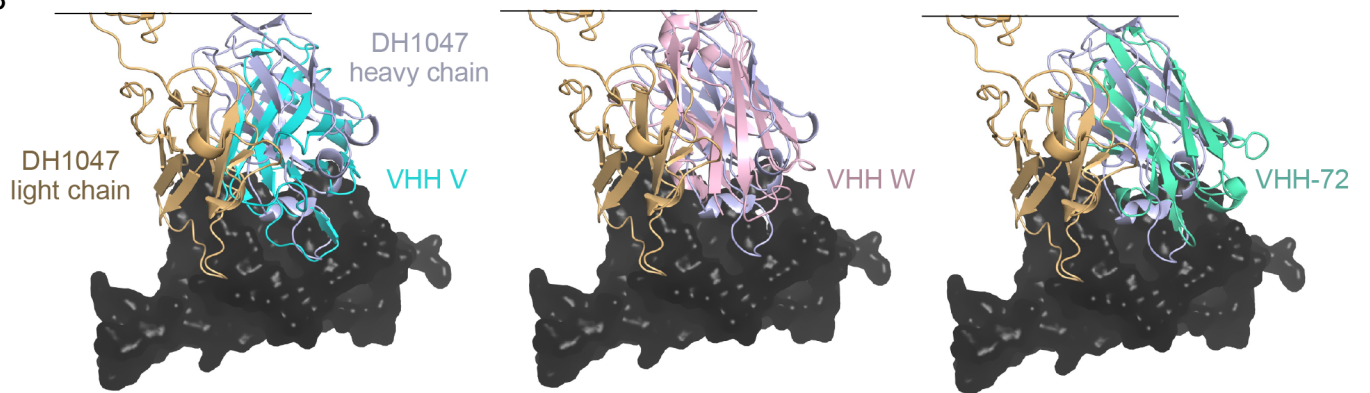
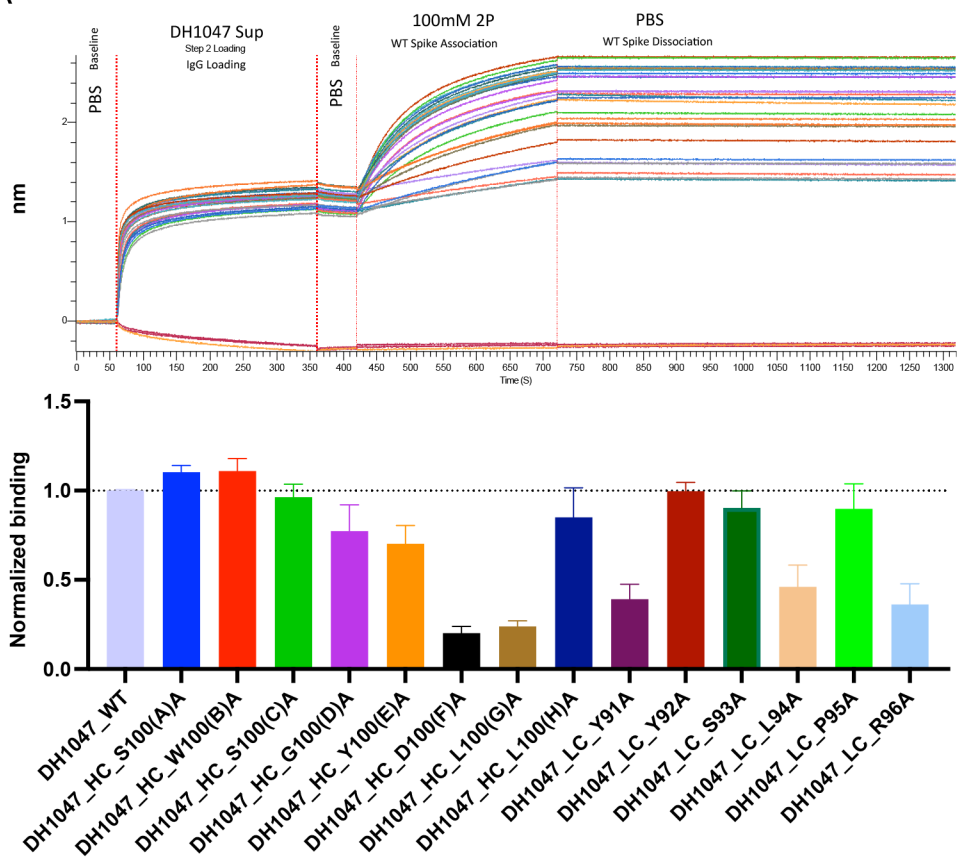
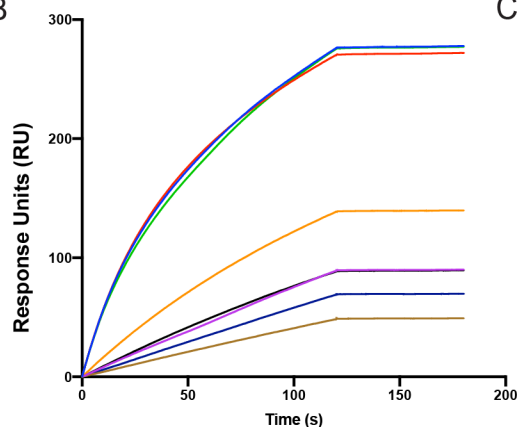


Figure S9

A



B



- DH1047 WT
- DH1047 HC S100(A)A
- DH1047 HC W100(B)A
- DH1047 HC D100(F)A
- DH1047 HC L100(G)A
- DH1047 LC Y91A
- DH1047 LC L94A
- DH1047 LC R96A

C

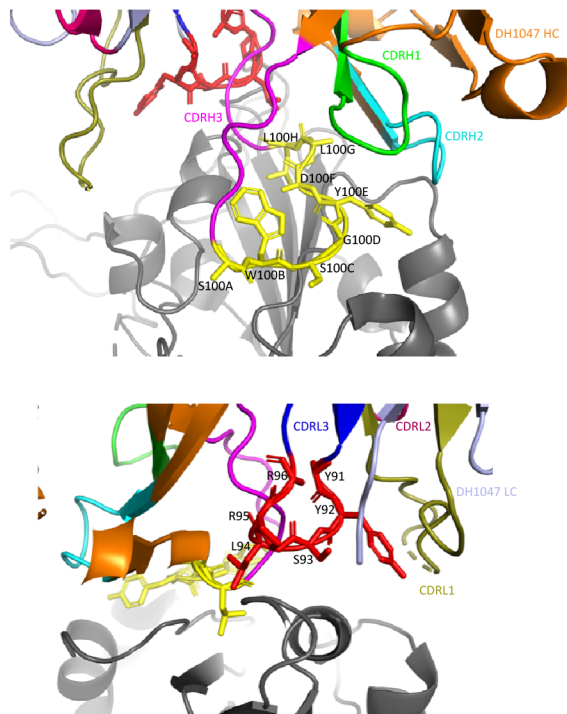
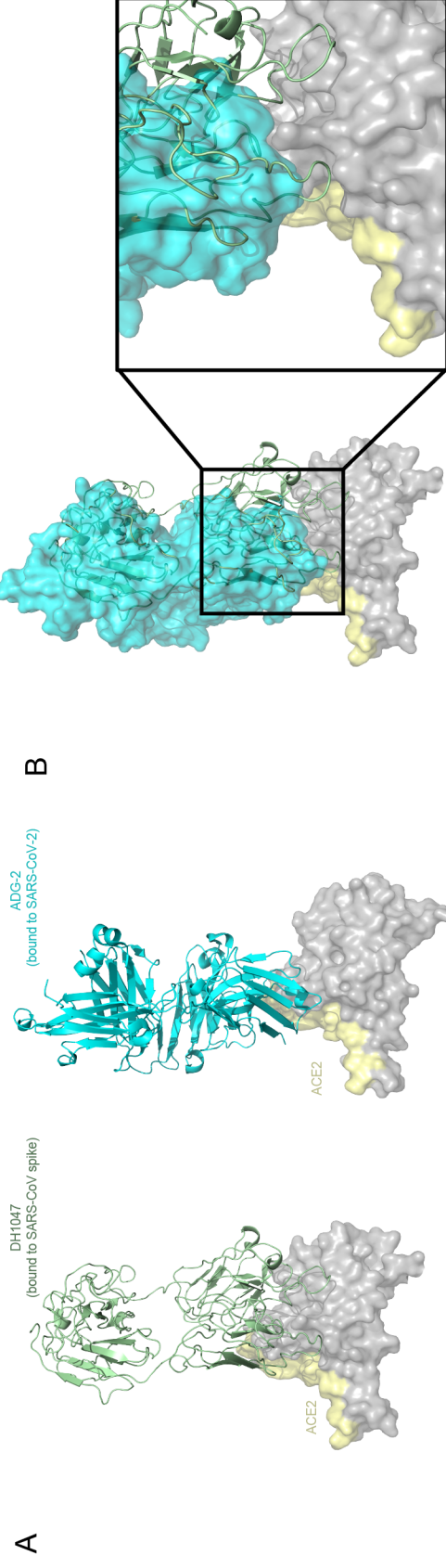
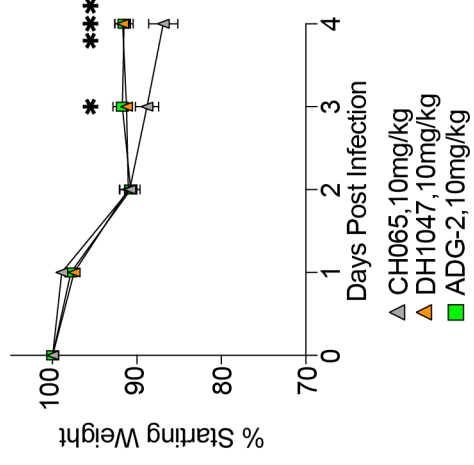


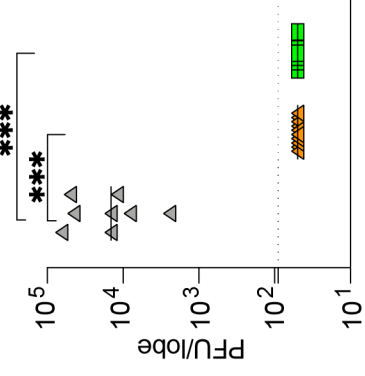
Figure S10



C SARS-CoV-2 B.1.351



SARS-CoV-2 B.1.351 Titer 4dpi



Lung Discoloration 4dpi

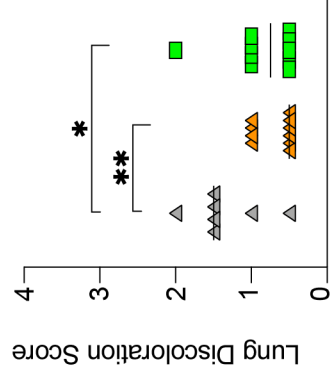


Table S1. Monoclonal antibody screen against SARS-CoV-2 2AA MA, SARS-CoV, WIV-1, and RsSHC014

mAb #	DH #	Specificity to SARS-CoV-2	ELISA cross-reactivity	Live virus neutralization IC ₅₀ (µg/ml)			
				SARS-CoV-2 2AA MA	SARS-CoV	WIV-1	RsSHC014
1	DH1058	S2	SARS-CoV-1, MERS-CoV, 229E, NL63, HKU1, OC43	>10	>10	>10	>10
2	DH1057	S2	SARS-CoV-1, OC43	>10	>10	>10	>10
3	DH1047	RBD	SARS-CoV, SARS-CoV-2, and bat CoVs	0.3979	0.0287	0.191	0.2005
4	DH1203	RBD	SARS-CoV and SARS-CoV-2	3.768	0.04781	>10	>10
5	DH1127	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
6	DH1059	no binding	SARS-CoV-2	>10	>10	>10	>10
7	DH1081	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
8	DH1085	no binding	SARS-CoV-2	>10	>10	>10	>10
9	DH1080	RBD	SARS-CoV and SARS-CoV-2	>10	0.0059	0.0330	>10
10	DH1061.1	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
11	DH1065	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
12	DH1066	NTD	only SARS-CoV	>10	>10	>10	>10
13	DH1064	RBD	SARS-CoV and SARS-CoV-2	>10	0.0216	>10	>10
14	DH1067	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
15	DH1069	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
16	DH1046	RBD	SARS-CoV, SARS-CoV-2, and bat CoVs	2.857	0.1033	0.4248	1.274
17	DH1068	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
18	DH1086	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
19	DH1071	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
20	DH1088	no binding	SARS-CoV-2	>10	>10	>10	>10
21	DH1073	RBD	SARS-CoV, SARS-CoV-2, and bat CoVs	0.8088	0.0161	0.267	>10
22	DH1235	RBD	SARS-CoV, SARS-CoV-2, and bat CoVs	0.1226	0.0403	0.0602	>10
23	DH1306	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
24	DH1193	RBD	SARS-CoV and SARS-CoV-2	4.345	>10	>10	>10
25	DH1152	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
26	DH1171	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
27	DH1109	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
28	DH1208	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
29	DH1166	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
30	DH1191	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
31	DH1120	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
32	DH1110	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
33	DH1106	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
34	DH1112	RBD	SARS-CoV and SARS-CoV-2	>10	0.0023	0.1617	>10
35	DH1117	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
36	DH1115	RBD	SARS-CoV and SARS-CoV-2	>10	0.0083	0.1614	>10
37	DH1093	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
38	DH1095	RBD	SARS-CoV	>10	>10	>10	>10
39	DH1113	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
40	DH1114	NTD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
41	DH1098	RBD	SARS-CoV and SARS-CoV-2	>10	0.0052	0.0318	>10
42	DH1101	RBD	SARS-CoV and SARS-CoV-2	>10	0.0012	>10	>10
43	DH1298	RBD	SARS-CoV and SARS-CoV-2	>10	0.0399	0.4312	>10
44	DH1299	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
45	DH1300	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
46	DH1301	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
47	DH1302	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
48	DH1303	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
49	DH1304	RBD	SARS-CoV and SARS-CoV-2	>10	>10	>10	>10
50	DH1305	RBD	SARS-CoV and SARS-CoV-2	9.877	>10	>10	>10

PDB ID 7SG4

EMDB ID EMD-25105

Data collection and processing

Microscope	FEI Titan Krios
Detector	Gatan K3
Magnification	81,000
Voltage (kV)	300
Electron exposure (e-/Å ²)	54.1
Defocus range (µm)	~0.75-2.50
Pixel size (Å)	1.08
Reconstruction software	cryoSparc
Symmetry imposed	C1
Initial particle images (no.)	2,370,616
Final particle images (no.)	284,619
Map resolution (Å)	3.43
FSC threshold	0.143

Refinement

Initial model used	7LD1
Model resolution (Å)	3.43
FSC threshold	0.143

Model composition

Nonhydrogen atoms	28,712
Protein residues	3,737

R.m.s. deviations

Bond lengths (Å)	0.013
Bond angles (°)	1.906

Validation

MolProbity score	1.C9
Clashscore	0.2
Poor rotamers (%)	2.01
EM ringer score	2.83

Ramachandran plot

Favored (%)	88.89
Allowed (%)	9.53
Disallowed (%)	1.58

Table S3. Immunogenetic characteristics of broadly cross-reactive mAbs

DH#	Antibody ID	Binding Specificity	Cross Reactivity	Antibody Gene Analysis									
				Donor ID	Time Point	HCDR3 Length	Heavy chain mutation	VH_Gene	JH_Gene	LCDR3 Length	Light chain mutation	VL_Gene	JL_Gene
DH1235	Ab026319_LS	RBD	SARS-CoV-1	SARS-CoV-2 convalescent	Day 36	21	1.68	IGHV3-48	IGHJ4	9	1.75	IGLV4-60	IGLJ2
DH1073	Ab026258_LS	RBD	SARS-CoV-1	SARS-CoV convalescent	Year 17	15	9.06	IGHV1-46	IGHJ6	11	2.92	IGKV3-11	IGKJ1
DH1046	Ab026204_LS	RBD	SARS-CoV, PCoV GXP4L, Bat CoV RsSHC014, Bat CoV RaTG13	SARS-CoV convalescent	Year 17	24	4.70	IGHV3-23	IGHJ6	9	3.65	IGKV1-5	IGKJ2
DH1047	Ab712384_LS	RBD	SARS-CoV, PCoV GXP4L, Bat CoV RsSHC014, Bat CoV RaTG13	SARS-CoV convalescent	Year 17	24	8.05	IGHV1-46	IGHJ4	9	2.05	IGKV4-1	IGKJ1