

Supplementary Figures

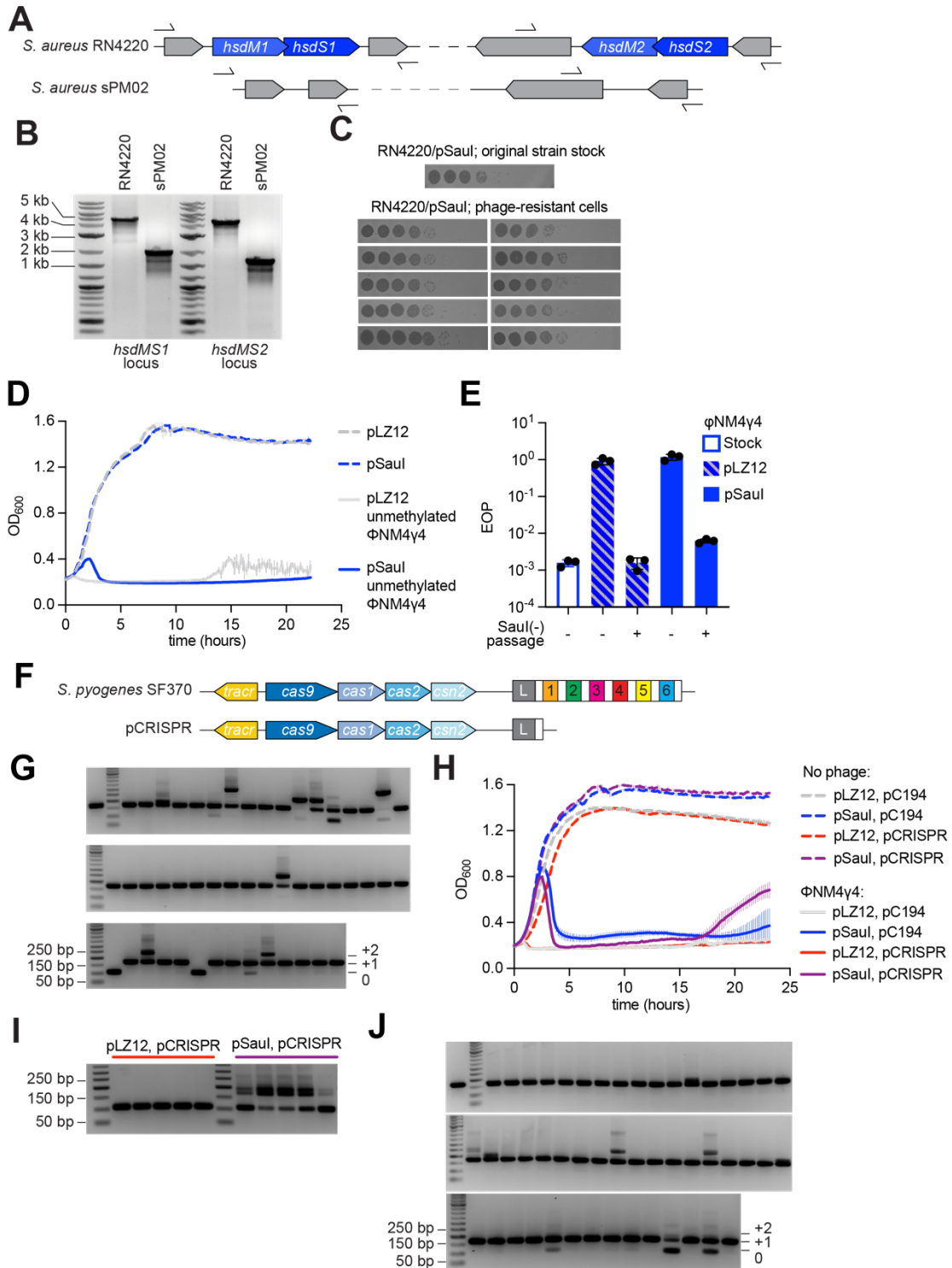


Figure S1. Construction of *S. aureus* sPM02 lacking the Saul restriction-modification system (related to Figure 1). (A) The two *saul-hsdMS* operons present in *S. aureus* RN4220 were removed (in-frame deletion) to generate strain sPM02. Arrows; primers used to check for the presence of the deletion. (B) Agarose gel

electrophoresis of PCR products obtained after amplification of the *hsdMS1* and *hsdMS2* loci using template DNA from RN4220 and sPM02 strains, and the primers shown in (A). (C) Plaque assays using bacterial lawns seeded with cells from 10 single colonies recovered from phage-resistant cells at the end of the growth curve shown in Fig. 1B, in which staphylococci harboring pSaul were infected with unmethylated Φ NM4 γ 4. 10-fold dilutions of unmethylated phage were spotted on the lawns. A lawn of the original stock strain carrying pSaul was used as control. (D) Growth of staphylococci expressing Saul or carrying a vector control in the presence or absence of Φ NM4 γ 4 infection, measured as the OD₆₀₀ of the cultures over time. MOI ~250. Mean of three biological replicates \pm SD are reported. (E) EOP of a phage stock, or of phages obtained at the end of the growth curve shown in (D), amplified or not through the non-methylating strain sPM02, after plating on lawns of staphylococci expressing Saul, relative to PFUs obtained with cells carrying a vector control. Mean of three biological replicates \pm SD are reported. (F) Schematic of the *S. pyogenes* type II-A CRISPR-*cas* locus. Grey rectangle, leader sequence ('L'); white rectangle, repeat; colored, numbered rectangles, spacers. It was cloned into the staphylococcal vector pC194 without spacers, just a single repeat. (G) Agarose gel electrophoresis of PCR products obtained after the amplification of the CRISPR array from 50 individual colonies recovered from one of the pSaul/pCRISPR cultures infected with Φ NM4 γ 4 at the end of experiment in Fig. 1D. (H) Growth of staphylococci harboring different plasmid combinations, in the presence or absence of Φ NM4 γ 4 infection, measured as the OD₆₀₀ of the cultures over time. MOI ~10. Mean of three biological replicates \pm SD are reported. (I) Agarose gel electrophoresis of PCR products obtained after amplification of the CRISPR array using DNA obtained from the cultures shown in (H). (J) Same as (G) after the amplification of the CRISPR array from 50 individual colonies recovered from one of the pSaul/pCRISPR cultures infected with Φ NM4 γ 4 at the end of the experiment shown in (H).

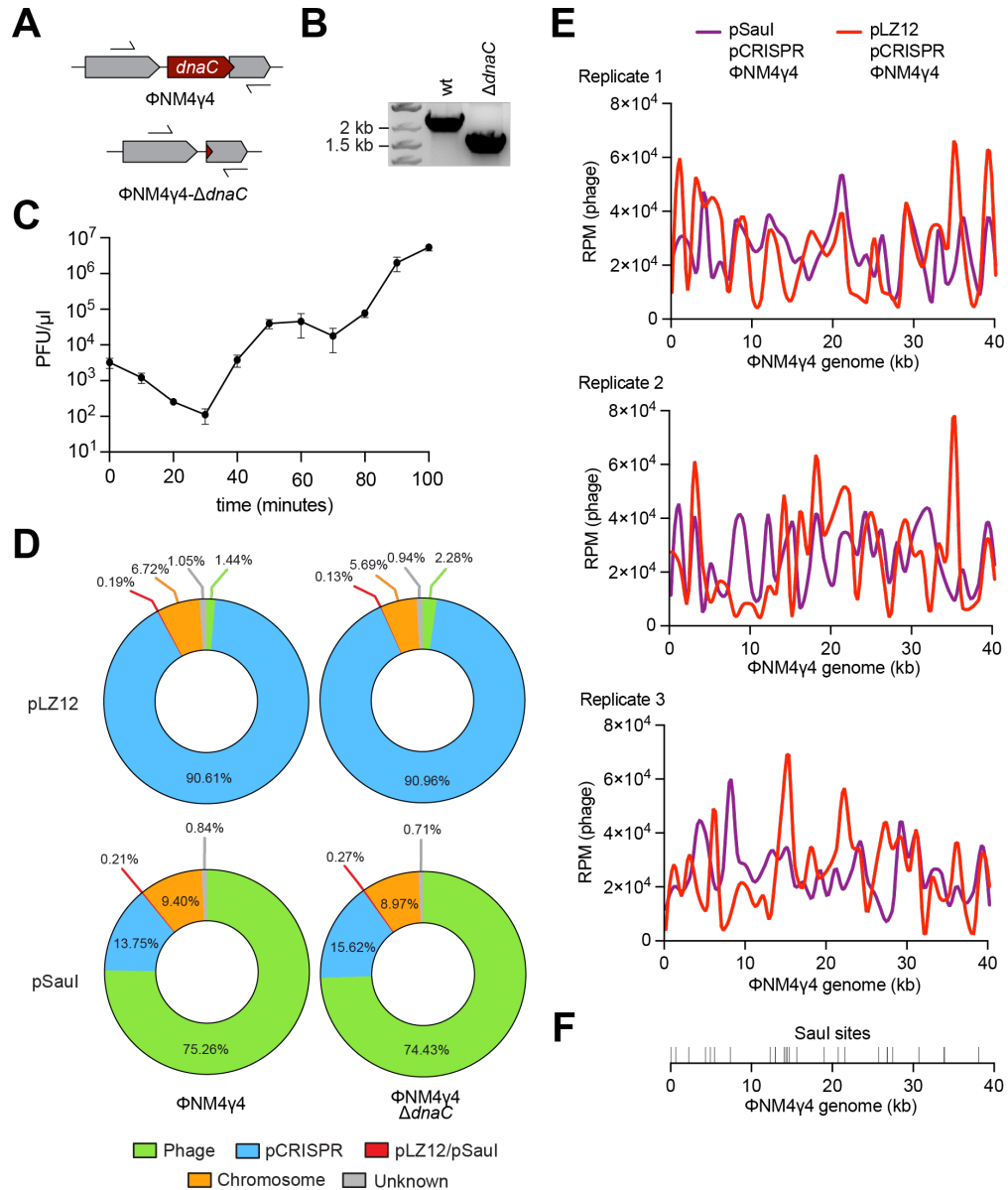


Figure S2. Spacer acquisition from the $\Phi\text{NM4}\gamma_4$'s genome in the presence of Saul restriction (related to Figure 2). (A) The *dnaC* gene of $\Phi\text{NM4}\gamma_4$ was removed (in-frame deletion) to generate $\Phi\text{NM4}\gamma_4-\Delta\text{dnaC}$. Arrows; primers used to check for the presence of the deletion. (B) Agarose gel electrophoresis of PCR products obtained after amplification of the *dnaC* locus using template DNA from $\Phi\text{NM4}\gamma_4$ and $\Phi\text{NM4}\gamma_4-\Delta\text{dnaC}$, and the primers shown in (A). (C) Quantification of the number of phage in the supernatant after 10-minute intervals following infection of *S. aureus* RN4220 with $\Phi\text{NM4}\gamma_4$ at MOI \sim 0.1. Mean of three biological replicates \pm SD are reported. (D) Quantification, using NGS reads, of spacers derived from the DNA substrates indicated with different colors, relative to total new spacers, acquired 30 minutes after infection of *S. aureus* harboring pCRISPR and expressing Saul or carrying a vector control, pLZ12, with $\Phi\text{NM4}\gamma_4$ or $\Phi\text{NM4}\gamma_4-\Delta\text{dnaC}$ phages. MOI \sim 250. Mean of three biological replicates

are reported. **(E)** Distribution of spacer abundance (measured as RPM of phage-matching reads) obtained in Fig. 2B across the Φ NM4 γ 4 genome. Maps for three independent replicates are shown. **(F)** Schematic representation of the Φ NM4 γ 4 genome showing its 26 Saul sites (grey lines).

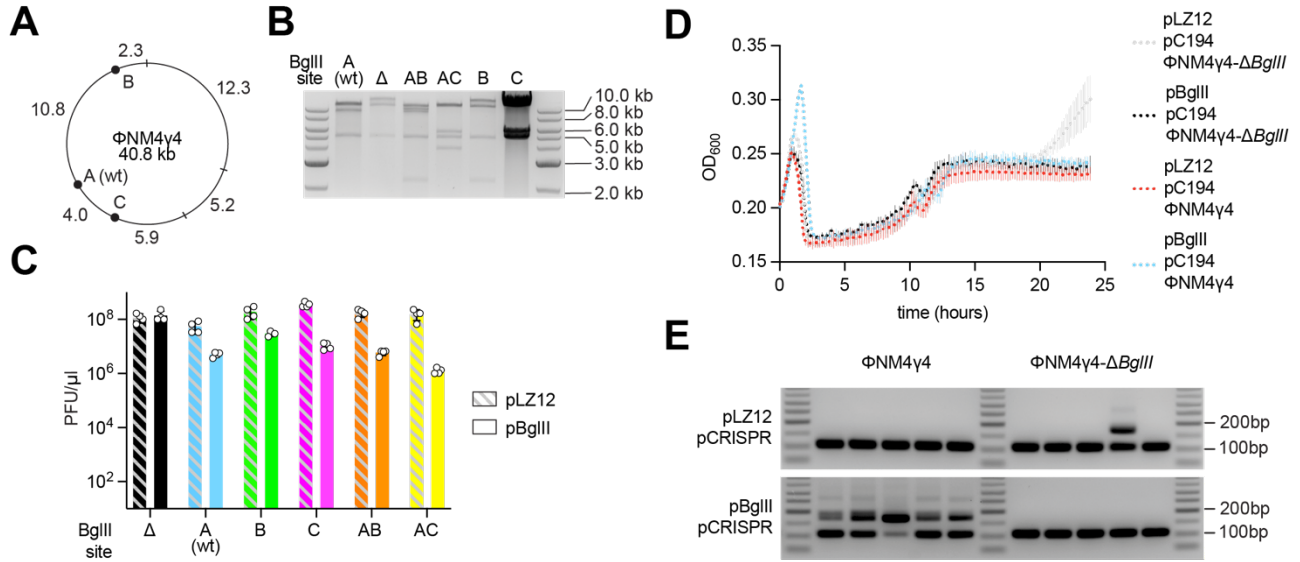


Figure S3. BgIII restriction of $\Phi\text{NM4}\gamma\text{4}$ promotes spacer acquisition (related to Figure 3). (A) Map of the $\Phi\text{NM4}\gamma\text{4}$ genome showing the different BgIII sites analyzed in this study (black circles, “A”, “B”, “C”). The three BssHII sites used for restriction mapping are shown as well (black lines). (B) Agarose gel electrophoresis of restriction fragments of different $\Phi\text{NM4}\gamma\text{4}$ phages, after digestion with BssHII and BgIII. The sizes of molecular weight markers are shown. (C) Enumeration of PFU generated by different $\Phi\text{NM4}\gamma\text{4}$ phage stocks on lawns of staphylococci expressing BgIII or carrying an empty vector control. Mean of four biological replicates \pm SD are reported. (D) Growth of staphylococci expressing BgIII or carrying a vector control after infection with $\Phi\text{NM4}\gamma\text{4}$ or $\Phi\text{NM4}\gamma\text{4}-\Delta\text{BgIII}$ phages, measured as the OD₆₀₀ of the cultures over time. MOI \sim 10. Mean of five biological replicates \pm SD are reported. (E) Agarose gel electrophoresis of PCR products obtained after amplification of the CRISPR array using DNA obtained from the cultures used in Fig. 3B.

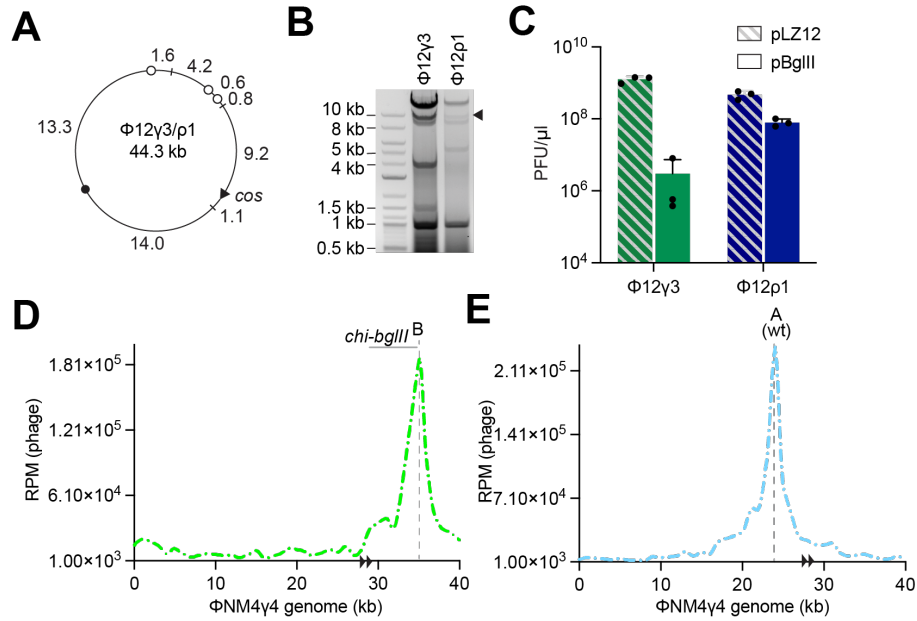


Figure S4. Analysis of BglIII restriction of Φ NM4 γ 4 and $\Phi 12\rho 1$ (related to Figure 4).

(A) Map of the $\Phi 12\gamma 3$ and $\Phi 12\rho 1$ genomes showing the BglIII present in both phages (black circles), as well as those removed from $\Phi 12\gamma 3$ to generate $\Phi 12\rho 1$ (white circles). The three BsrBI sites used for restriction mapping (black lines), as well as the *cos* site (black arrowhead), are also shown. (B) Agarose gel electrophoresis of restriction fragments of $\Phi 12\gamma 3$ and $\Phi 12\rho 1$ phage DNA, after digestion with BsrBI and BglIII. The sizes of molecular weight markers are shown. The black arrowhead marks a restriction fragment produced after the annealing of the *cos* site. (C) Enumeration of PFU generated by $\Phi 12\gamma 3$ or $\Phi 12\rho 1$ on lawns of staphylococci expressing BglIII or carrying an empty vector control. Mean of three biological replicates \pm SD are reported. (D) Distribution of spacer abundance (measured as RPM of phage-matching reads) across the Φ NM4 γ 4-B genome, using NGS data collected after infection of *addA*ⁿ mutant staphylococci cells carrying pCRISPR and pBglIII. (E) Same as (D) using data from Φ NM4 γ 4-A(wt) infection experiments.

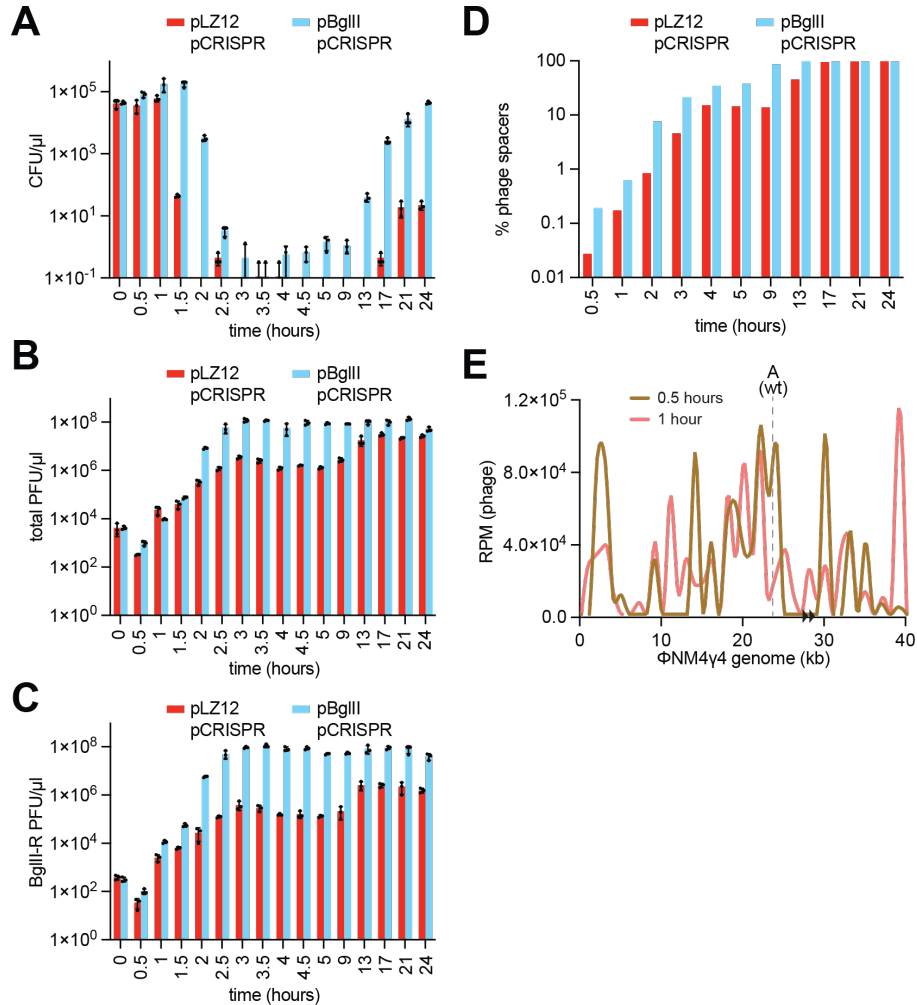


Figure S5. Dynamics of Φ NM4 γ 4 restriction, methylation, and spacer acquisition (related to Figure 5). (A) Enumeration of CFUs present at different times following infection of *S. aureus* cells harboring pCRISPR and expressing BgIII or carrying a vector control with Φ NM4 γ 4 (MOI ~1). Mean of three technical replicates \pm SD are reported. (B) Enumeration of total PFUs present in the cultures of the experiment described in (A), after plaquing on the CRISPR(-), RM(-) strain RN4220. Mean of three technical replicates \pm SD are reported. (C) Same as (B) but plaquing on staphylococci expressing BgIII to enumerate PFUs resistant to restriction. Mean of three technical replicates \pm SD are reported. (D) Quantification of phage-derived spacers, relative to total new spacers, acquired at the different time points of the experiment shown in (A). (E) Distribution of spacer abundance (measured as RPM of phage-matching reads) obtained after NGS of the CRISPR locus present in the cultures used in (A) across the Φ NM4 γ 4 genome, using data from the 0.5- and 1-hour time points.

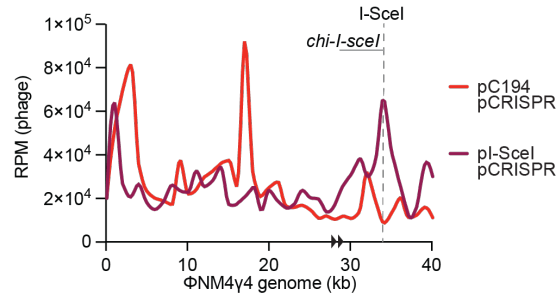


Figure S6. Scel restriction of Φ NM4 γ 4 promotes spacer acquisition at the cleavage site (related to Figure 6). Distribution of spacer abundance (measured as RPM of phage-matching reads) across the Φ NM4 γ 4-I-Scel genome, using NGS data collected after infection of staphylococcal cells carrying pCRISPR and pl-Scel or the control vector pC194.