A Novel Wireless ECG System for Prolonged Monitoring of Multiple Zebrafish for Heart Disease and Drug Screening Studies

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Supplementary document

Supplementary Section 1. Drug Administration

To anesthetize fish, a buffered solution of 200 parts-per-million (ppm) Tricaine (Sigma, USA) was used (Le et al. 2019). Tricaine was dissolved in distilled water to a final concentration of 7,000 parts-per-million (ppm) as a stock, and the pH value was adjusted to 7.2 with sodium hydroxide (Sigma).

Amiodarone (Sigma) was dissolved in water at 65°C for 2 hours and stocked as 900 μ M at 4°C. Before use, the solution was re-dissolved at 65°C for 1 hour (Chen et al. 2012). The fish were immersed in a tank with 100 μ M amiodarone for 1 hour and then returned to fish water for 15 min before conducting experiment.



Supplementary figure 1. Schematic of the ECG acquisition circuit. (a) The analog front-end circuit, including ADS chip with 4 differential channels, an OP-AMP utilized to reduce common mode noise. (b) wireless transmission circuit, including a system-on-chip nRF52832. (c) The power-supply module included a charge management controller (MCP73832, *Microchip Inc*), a buck boost converter TPS63060 (*Texas Instrumentation*) to maintain a 3.5 V output for the digital system regardless of the fluctuation of battery voltage level and two low-dropout regulators (*i.e.*, TPS72325 and TPS73225) provide stable ±2.5 V, respectively.



Supplementary figure 2. The IOT system integrated with the prolonged ECG system. The IoT Core provides the functionalities to manage and configure connected devices conveniently and securely. Once the ECG data are acquired from the measurement device, it is transmitted in real-time to the cloud platform through the Message Queuing Telemetry Transport (MQTT) protocol. The MQTT protocol is designed to maintain a long-lived connection between the device and the client with minimal communications overheads to save bandwidth for data transmission. Cloud Pub/Sub is an asynchronous communication medium between the device and the servers on the IoT cloud. The communication is based on the notion of topics that cache durable messages. Zebrafish ECG published on a certain topic by the device can be pushed to or pulled from the servers that subscribe to the same topic for storage and analytics.

The storage layer is responsible for storing the real-time zebrafish ECG data into the database. As one of the most promising time-series databases, InfluxDB is employed in the proposed system for timely and reliable storage. As an advanced non-relational database, InfluxDB resolves the performance bottleneck of traditionally used databases such as MySQL and provides greater flexibility and read-write speed. In the analytics layer, Cloud Functions, a serverless execution environment, run processing techniques such as denoising, filtering, normalizing, detecting P wave, QRS complex, or T wave, and extracting other useful features. Furthermore, machine learning approaches can be applied to these data with the aid of the Kubernetes Engine to train and deploy the models in containerized applications. Computationally intensive process and analysis tasks are carried out in powerful servers, which greatly eases the burden of local devices. The graphical user interface is responsible for data visualization and management, providing an easy access to the data in the IoT cloud. Grafana is an open platform that is utilized for designing dashboards to represent the ECG data. Users can log onto the cloud to acquire visualized ECG data on either web pages

or mobile applications. Based on the results of data analysis, users can observe and understand the realtime conditions of zebrafishes. In the event of any anomalies or suspicious readings, the IoT cloud will notify users in time.



Supplementary figure 3. Wireless data transmission to mobile application. An Android application developed in Java that connects to the ECG system via BLE communication for data collection, displaying, and logging. An Android smartphone application in Java was developed to connect to the Bluetooth Low Energy (BLE) device for data collection, displaying, and saving. Through BLE protocols, the application connects to the Zebra II and reads multiple channels of data at a rate of 250 Hz. After accumulating 1000 data points, the input data are sent to the connecting cloud server for feature extraction of fish ECG. The results will start appearing on the application interface after at least 10 seconds of initial start of data acquisition in the form of dynamic graphs – 4 ECG channels of zebrafish. The user can disconnect the Zebra II at any time and save the raw data plus the exact time of acquisition through the application as a .txt file with a customizable name in the phone's external storage as well as on the cloud server as a backup.



Supplementary figure 4. Characterization of electrode-skin impedance. (a) Timeline for experiment: Zebrafish were first anesthetized with 200ppm Tricaine in 3 minutes. Zebrafish were then loaded to each apparatus, followed by the tube system continuously providing low Tricaine concentration. The skin-electrode impedance was measured at 5 time points (*i.e.*, every 10 min for 40 min). (b) Averaged impedance magnitude (left side) of 5 time points and phase (right side) of the electrodes placed on fish skin (n=8 fish).



Supplementary figure 5. Comparison of performance between Zebra II system and iWorx system. The ECG measurement was conducted with two systems simultaneously (n = 8 fish). The collected data were then analyzed and compared in terms of correlation on time domain (**a**) and frequency domain (**b**). Correlation of heart rate (HR) (**d**) and QRS interval (**c**) extracted from ECG data collected by Zebra II and iWORX.



Supplementary figure 6. Compare the performance between Zebra II and Iworx with amiodarone treatment. Control group (n = 20 WT fish) and Amiodarone-treated group (n = 16 WT fish) were used to this experiment. The ECG data from two groups were acquired by the two systems and, and features such as HR (a), QTc interval (c), and QRS duration (e) were extracted from ECG data. The Bland Altman analysis was used to compare the agreement level between two systems in response to such features, including HR (b), QTc interval (d), and QRS duration (f). *p <0.05; **p < 0.01 (paired sample T-test). *ns* indicates not significant.



Supplementary figure 7. Characterization of heart rate variation with different Tricaine concentration.

The experiment was conducted with n = 8 fish within 40 minute (min) long ECG measurement, showing the heart rate variation with two Tricaine concentrations in every 5 min. p <0.05; **p < 0.01 (paired sample T-test). *ns* indicates not significant.



Supplementary figure 8. Heart rate comparison between WT fish and *Tg(SCN5A-D1275N)* with different NaCl concentrations. HR value calculated with ECG data acquired from both WT fish and *Tg(SCN5A-D1275N)* mutant fish with the following treatments: without NaCl (a); with 0.1 ‰ NaCl (b); with 0.3 ‰ NaCl (c); with 0.6 ‰ NaCl (d); with 0.9 ‰ NaCl (e); with 1.8 ‰ NaCl (f). Overall, the HR in WT fish did not show significant difference among NaCl concentrations. In contrast, the in mutant fish showed significant changes with the HR decreasing in response to the increase of NaCl dose. p <0.05; **p < 0.01; **p < 0.001 (Student's T-test). *ns* indicates not significant.



Supplementary figure 9. QTc comparison between WT fish and *Tg(SCN5A-D1275N)* with different NaCl concentration. QTc interval calculated with ECG data collected from WT and mutant fish with the following treatments: without NaCl(a); with 0.1 ‰ NaCl (b); with 0.3 ‰ NaCl (c); with 0.6 ‰ NaCl (d); with 0.9 ‰ NaCl (e); with 1.8 ‰ NaCl (f). The significant difference showing in QTc value between two groups was found in 0.1, 0.3 and 1.8 ‰ NaCl. An increase in QTc value responding to the increase of NaCl level was found in both WT fish and mutant fish. p <0.05; **p < 0.01 (Student's T-test). *ns* indicates not significant.

Table S1:

Parameters	Numbers	
Number of channels	4 (can be up to 8)	
Bandwidth	1 Hz – 150 Hz	
Input Range	±2.5V	
Resolution	24 Bit	
Sampling rate	250 Hz	
Common mode rejection ratio (CMRR © 60 Hz)	100 dB	
Input-Referred Noise	1 μV	
Enclosure	Plastic	
Dimensions	6.3"W x 4.8"L x 3.9"H	

Table S2:

Parameters	Mean	STD	RSD (100*STD/mean)
Signal to noise ratio (SNR)	25.36	7.38	29.10
Heart rate	117.28	9.34	7.96
QTc interval	384.21	12.25	3.18
QRS interval	376.47	8.27	2.19