## nature portfolio

Corresponding author(s):	Sivaraj Sivaramakrishnan,	Chetan Gadgi
--------------------------	---------------------------	--------------

Last updated by author(s): 2022/01/21

## **Reporting Summary**

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

$\sim$				
ζ.	ta	١Ť١	ıst	100

For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
X	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
×	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
×	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

## Software and code

Policy information about availability of computer code

Data collection Computational calculations were performed in Matlab R2020a, using the equations and parameters outlined in the supplemental materials.

Data analysis Figure preparation, data and statistical analysis was completed using Prism 19

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

## Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The data generated in this study is available in the Source Data File provided with this paper. Entries in the Bionumbers database were referenced in confirming parameter availability (ID: 1030809)

Field-spe	ecific re	porting				
Please select the o	one below that i	s the best fit for your research. If you are not sure, read the appropriate sections before making your selection.				
Life sciences For a reference copy of		Behavioural & social sciences				
Life scier	nces sti	udy design				
		points even when the disclosure is negative.				
Sample size	deltaFRET mea measurements over time. To n	riment, a single n was performed with a membrane preparation derived from one batch of cells or one passage of cells. asurements on a single membrane preparation (n=1)) were repeated 4 times (average of 4 measurements of agonist - 4 s of buffer). For a given membrane preparation (n=1), stopped flow decay curves show the decrease in FRET (525 nm/475nm) measure the FRET, the counts at 525 nm or 475 nm from at least 5 injections were averaged. For cAMP assays, one experiment erage of 4 data points for a given condition.				
Data exclusions	Data was not e	xcluded.				
Replication		tal data was replicated at least 3 times. Computational calculations were performed independently by two of the authors to utational calculations.				
Randomization	Membranes we	ere randomly assigned to the agonist treated and buffer treated groups.				
Blinding		were not blinded due to the technical nature of the experimental protocols and the limited number of personel performing it was not possible for one person to prepare samples and a second to run samples blinded.				
Reportin	ng for sp	pecific materials, systems and methods				
We require informat	tion from authors	about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.				
Materials & ex	perimental s	ystems Methods				
n/a Involved in t	•	n/a Involved in the study				
Antibodie		ChIP-seq				
Eukaryotic cell lines    X   Flow cytometry    X   MRI-based neuroimaging						
Animals and other organisms						
Human research participants						
Clinical data						
<b>x</b> Dual use r	research of conce	n				
<u>Eukaryotic c</u>	cell lines					
Policy information	about <u>cell lines</u>					
Cell line source(s)		Hek293T-Flp-In were obtained from ThermoFischer (catalog number R78007)				
Authentication	Authentication Cell line was not authenticated					
Mycoplasma contamination Cell line was not tested for Mycoplasma contamination		Cell line was not tested for Mycoplasma contamination				

No commonly misidentified cell lines were used.

Commonly misidentified lines (See <u>ICLAC</u> register)