

Supplementary Online Content

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This supplementary material has been provided by the authors to give readers additional information about their work.

eTable 1. Antibody Cocktail to Identify Adaptive Immune Cell Populations

| Antibody Cocktail 1 | Supplier | Antibody Cocktail 2 | Supplier |
|---------------------|-----------------------------------|---------------------|-----------------------------------|
| CD3-BUV395 | BD Bioscience, San Diego, CA, USA | CD3-Percp/Cy5.5 | BD Bioscience, San Diego, CA, USA |
| CD4-BV421 | BD Bioscience, San Diego, CA, USA | CD4-BV510 | BioLegend, San Diego, USA |
| CD8-BUV805 | BD Bioscience, San Diego, CA, USA | CXCR5-APCR700 | BD Bioscience, San Diego, CA, USA |
| CD45RA-Percp/Cy5.5 | BD Bioscience, San Diego, CA, USA | PD-1-PEcy7 | BD Bioscience, San Diego, CA, USA |
| CCR7-BV785 | BioLegend, San Diego, USA | CD19-BV785 | BioLegend, San Diego, USA |
| CD69-BV650 | BD Bioscience, San Diego, CA, USA | CD20-BV421 | BioLegend, San Diego, USA |
| HLA-APC-H7 | BD Bioscience, San Diego, CA, USA | CD27-BUV737 | BD Bioscience, San Diego, CA, USA |
| Zombie NIR | BioLegend, San Diego, USA | CD19-BV785 | BioLegend, San Diego, USA |
| | | CD38-BUV496 | BD Bioscience, San Diego, CA, USA |
| | | CD24-BV711 | BioLegend, San Diego, USA |
| | | IgG-BV605 | BD Bioscience, San Diego, CA, USA |
| | | IgM-FITC | BioLegend, San Diego, USA |
| | | Zombie NIR | BioLegend, San Diego, USA |

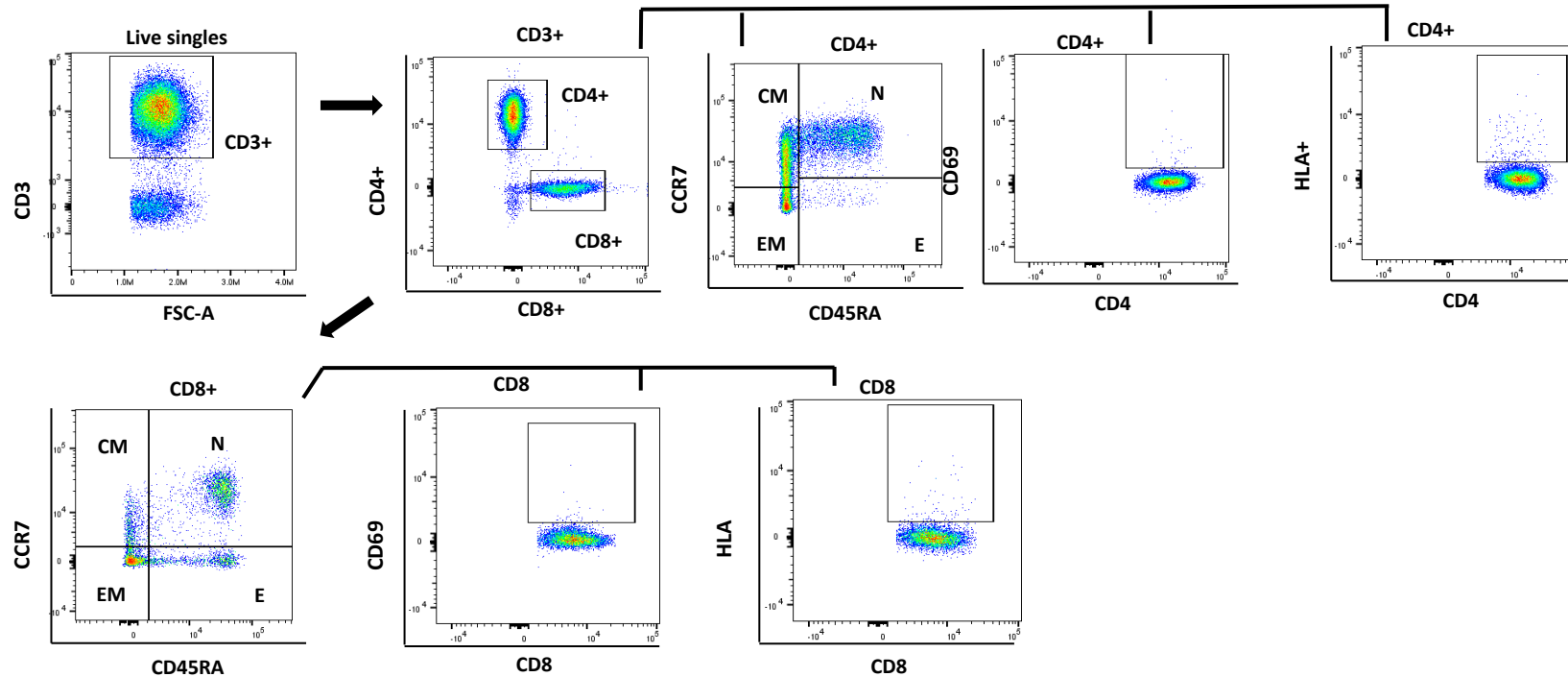
eTable 2. Antibody Cocktail to Identify Innate Immune Cell Populations

| Surface Marker | Fluorophore | Clone | Final Dilution |
|-----------------------|--------------------|--------------|-----------------------|
| CD14 | BV786 | M5E2 | 1:50 |
| CD45 | BV711 | HI30 | 1:100 |
| CD56 | BUV737 | NCAM16.2 | 1:100 |
| CD11c | PE-Cy7 | B-ly6 | 1:100 |
| CD3 | BB515 | UCHTI | 1:100 |
| CD15 | PE-CF594 | W6D3 | 1:200 |
| HLA-DR | V500 | G46-6 | 1:200 |
| CD19 | BV605 | SJ25C1 | 1:200 |
| CD16 | BUV395 | 3G8 | 1:400 |
| Live/dead | N-IR | | |

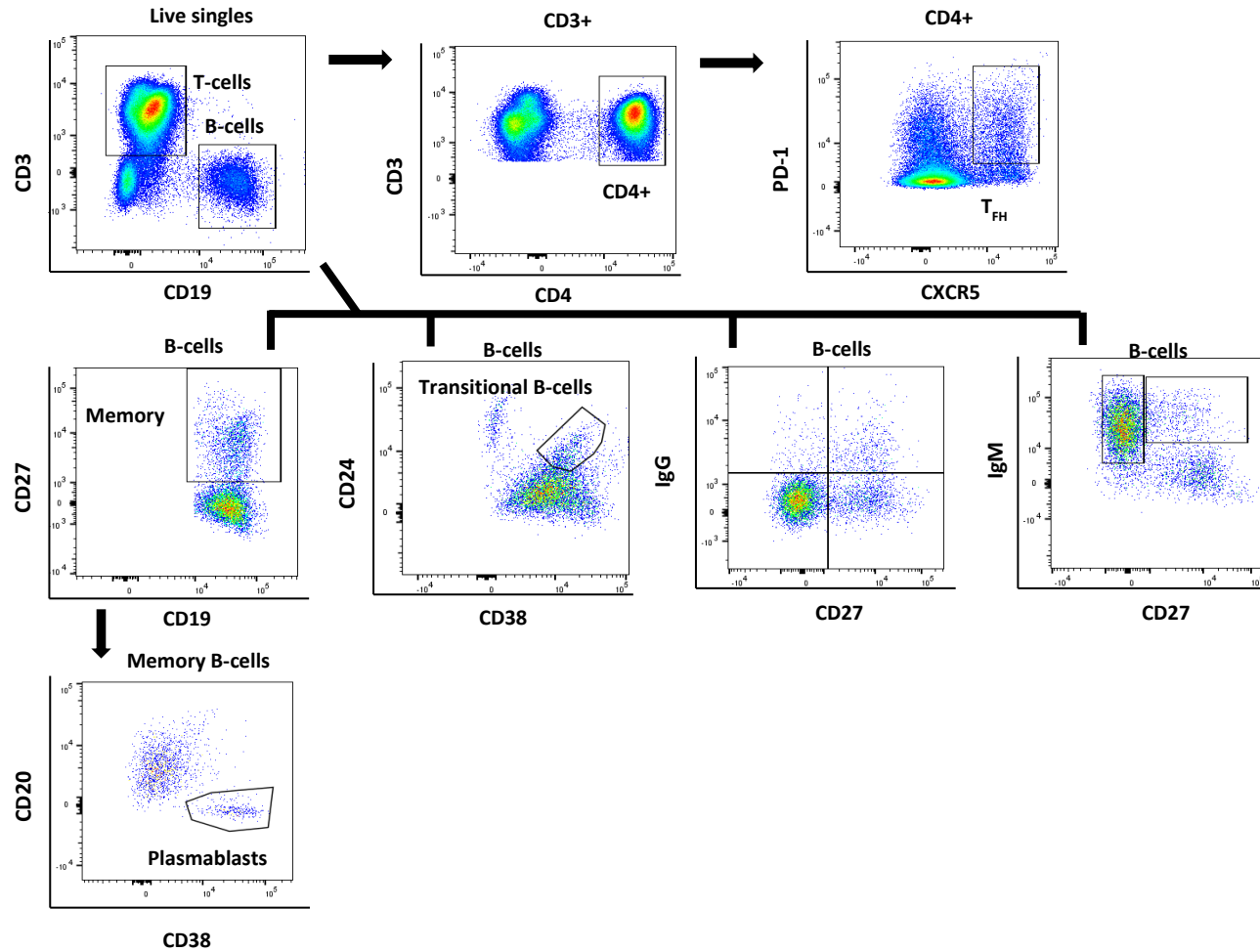
eTable 3. Concordance of 3 Serological Assays for All Samples at Convalescent Period From the Whole Household Cohort Study

| | | Diasorin | | | | Diasorin | | | | Wantai | |
|-----------------------|----------|----------|----------|---------------|----------|----------|----------|-----------------------|----------|----------|----------|
| | | Positive | Negative | | | Positive | Negative | | | Positive | Negative |
| In-house ELISA | Positive | 56 | 10 | Wantai | Positive | 54 | 10 | In-house ELISA | Positive | 61 | 3 |
| | Negative | 3 | 164 | | Negative | 3 | 162 | | Negative | 3 | 162 |
| | Total | | 233 | | Total | | 229 | | Total | | 229 |
| Agreement | | 0.94 | | | | 0.94 | | | | 0.97 | |

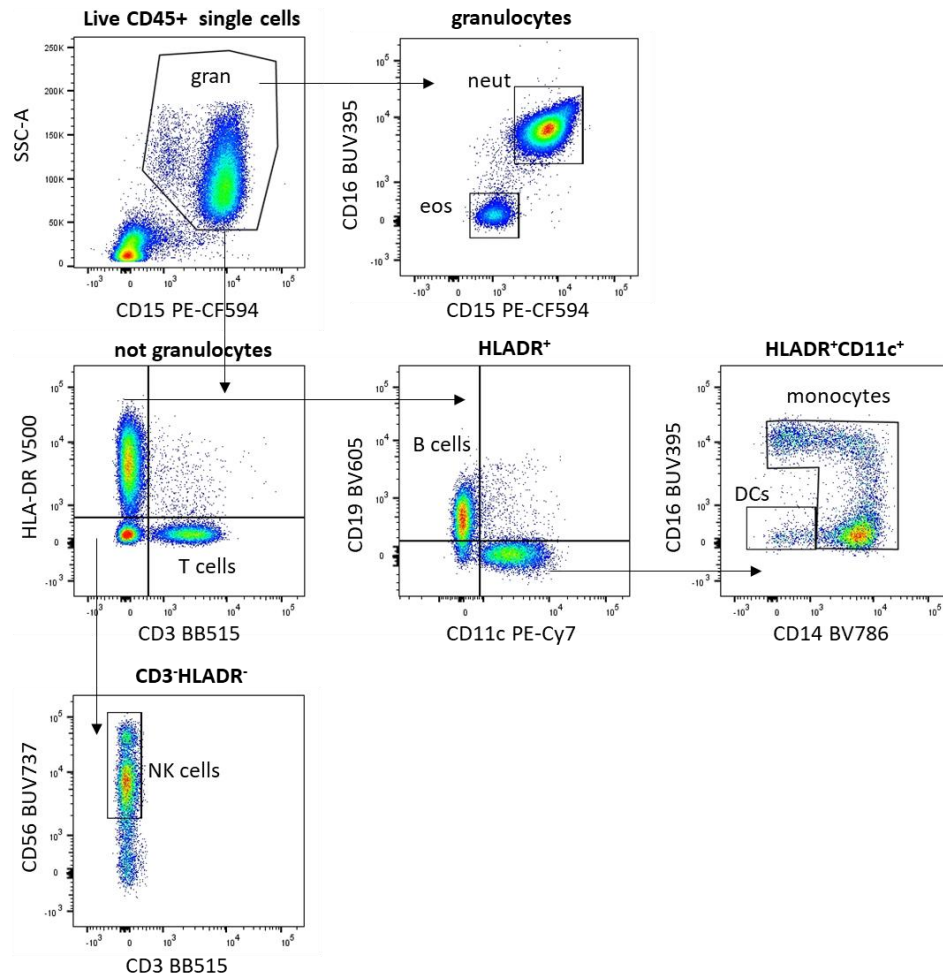
eFigure 1. Gating Strategy to Identify T-cell Subsets From live single cells, T-cells were identified by positive CD3 expression. CD4+ and CD8+ T-cells were identified from the CD3+ population. CD4+ T-cells and CD8+ T-cells were characterised into naïve (N), effector (E), central memory (CM) and effector memory (EM) based off CCR7 and CD45RA expression. These were CCR7+CD45RA+, CCR7-CD45RA+, CCR7+CD45RA- and CCR7-CD45RA- respectively. CD69+ and HLA+ expression was also characterised on CD4+ and CD8+ T-cells.



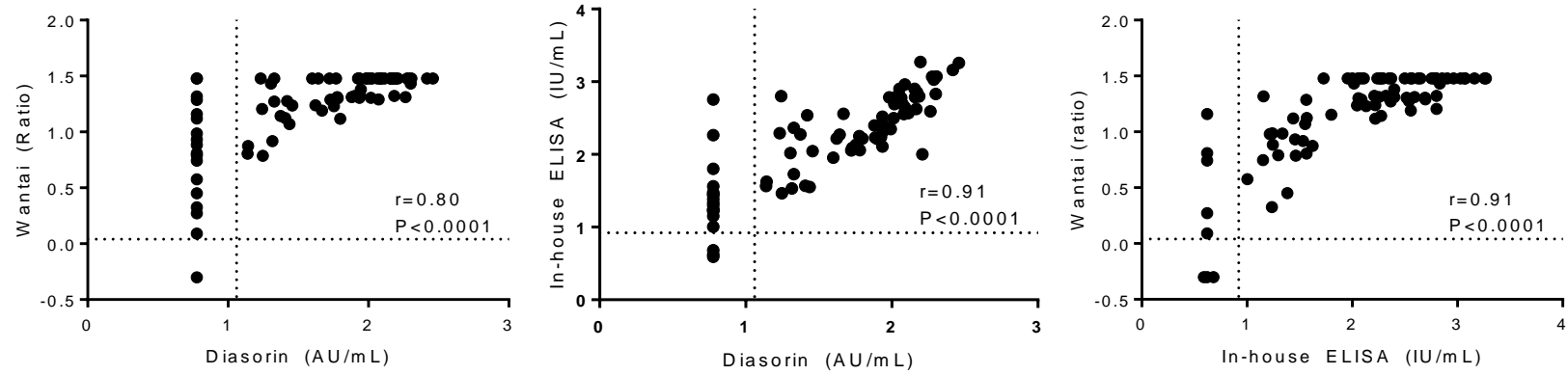
eFigure 2. Gating Strategy to Identify B-cell/T_{FH} Subsets From live single cells, T-cells were identified as CD3+CD19- and B-cells were identified as the CD3-CD19+ population. From the CD3+ population, T_{FH} was characterised as CD4+CXCR5+PD-1+ expressing T-cells. From the B-cell population, memory was identified as CD27+, transitional B-cells as CD24+CD38+ and plasmablasts as CD27+CD20-CD38+. IgM and IgG expression were also characterised on B-cell populations.



eFigure 3. Gating Strategy for Innate Cell Populations Granulocytes were selected within CD45⁺ leukocytes based on their SSC profile and CD15 expression. Neutrophils were CD15⁺CD16⁺ and eosinophils were CD15⁺CD16⁻. Within the non-granulocyte fraction, CD3 T cells were identified and B cells were identified based on CD19 and HLA-DR expression. CD11c⁺CD14⁺ monocytes were gated within the non-T cell and non-B cell fraction. Total dendritic cells were HLADR⁺CD11c⁺CD14⁻ and NK cells were HLADR⁻CD3⁻CD56⁺ cells.

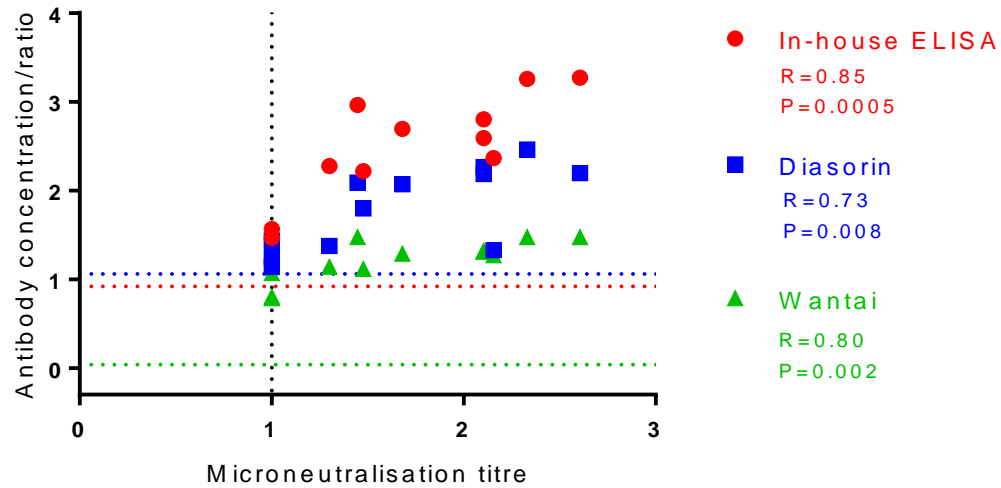


eFigure 4. Correlation Analysis of 3 Serological Assays



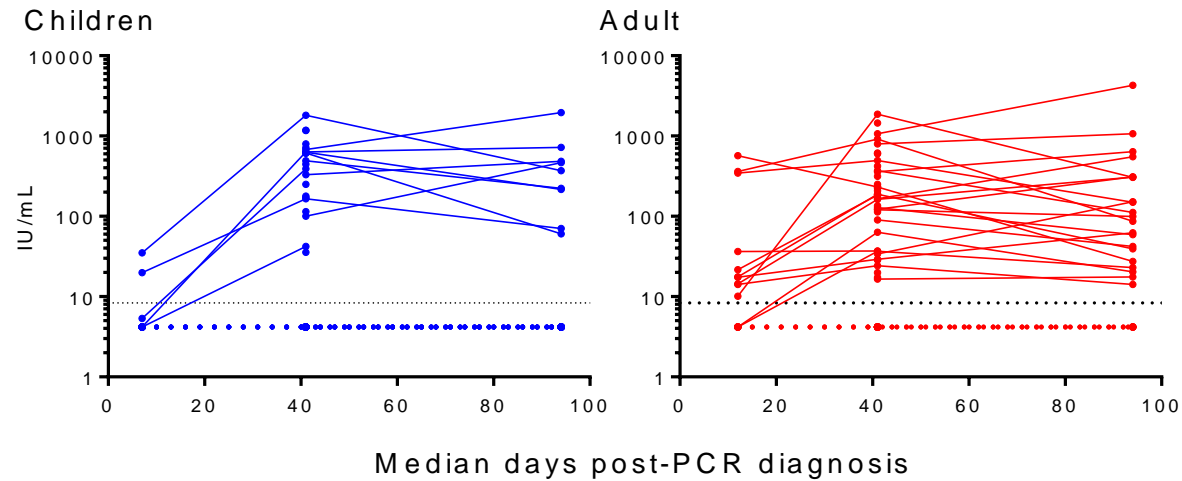
Antibody levels were log-transformed and analysed using Pearson correlation analyses. N=138-145.

eFigure 5. Correlation Analysis of 3 Serological Assays Against SARS-CoV-2 Microneutralization Assay



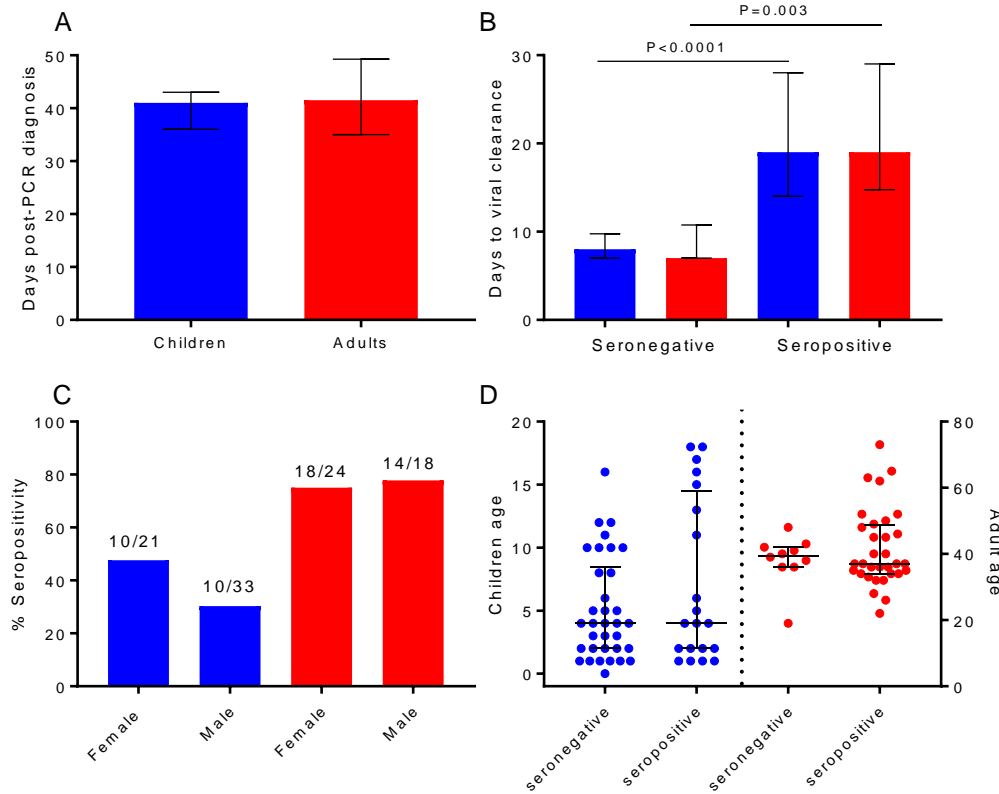
Antibody levels were log-transformed and analysed using Pearson correlation analyses. Coloured dotted line represents assay specific cut-off for seropositivity. N=12.

eFigure 6. SARS-CoV-2 IgG Levels Over Time in Children and Adults Using an In-house ELISA



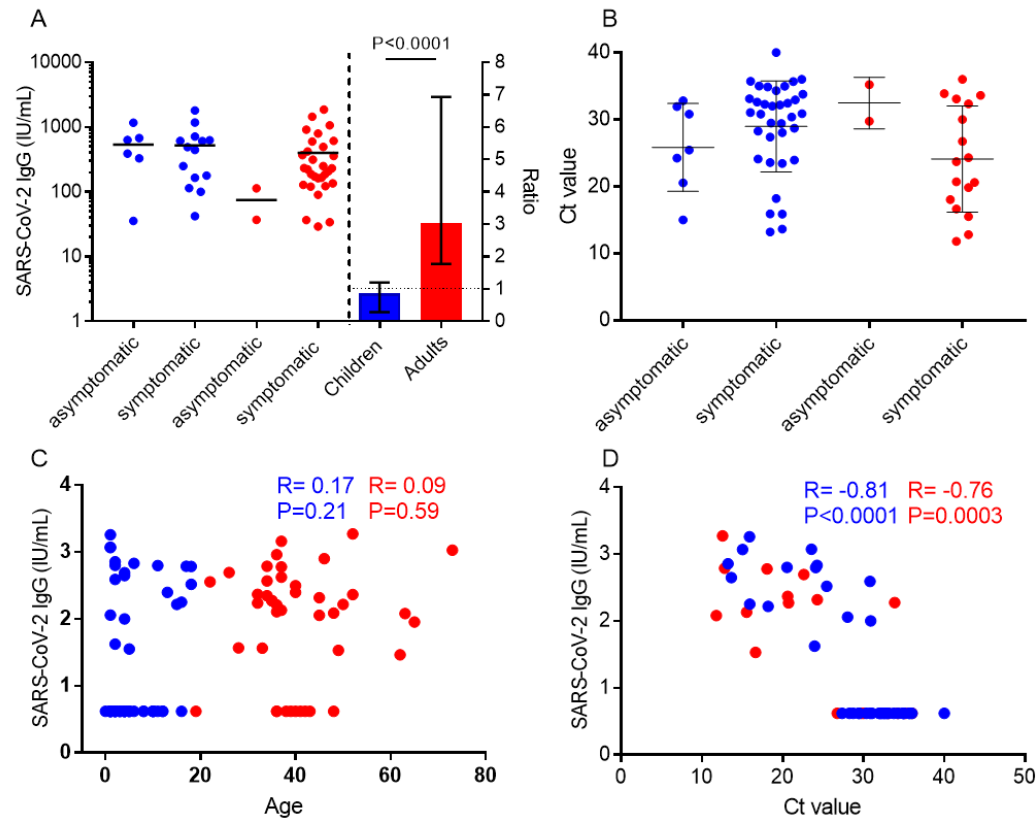
Number of samples per timepoint: Children, Day 7 (N=13), Day 41 (N=59), Day 94 (N=26); Adult, Day 12 (N=20), Day 41 (N=57), Day 94 (N=29).

eFigure 7. Duration of Post-PCR Diagnosis and Viral Clearance, Sex, and Age Associated With SARS-CoV-2 Antibody Responses



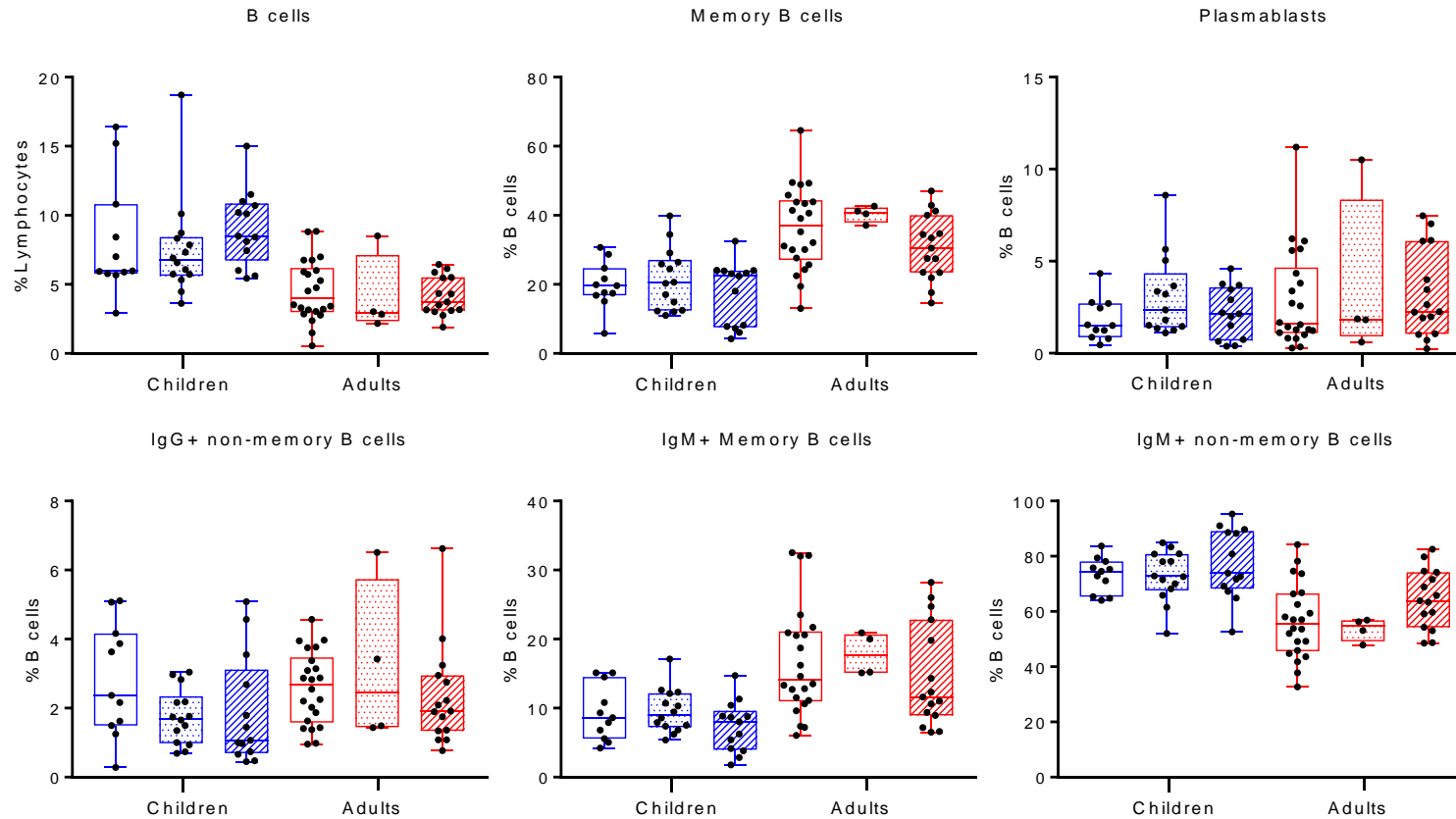
(A) Median days (IQR) between positive PCR diagnosis and convalescent blood sampling between children (N=54) and adult (N=42). (B) Duration of viral clearance (median days, IQR) stratified by serostatus (seronegative children, N=20, seropositive children, N=7; seronegative adult, N=4, seropositive adult, N=10). (C) Seropositivity rate stratified by sex. (D) Age of children and adults stratified by serostatus (Children, N=54; Adult, N=42) (Median, IQR).

eFigure 8. Symptoms and Correlation Analysis Associated With SARS-CoV-2 Antibody Responses



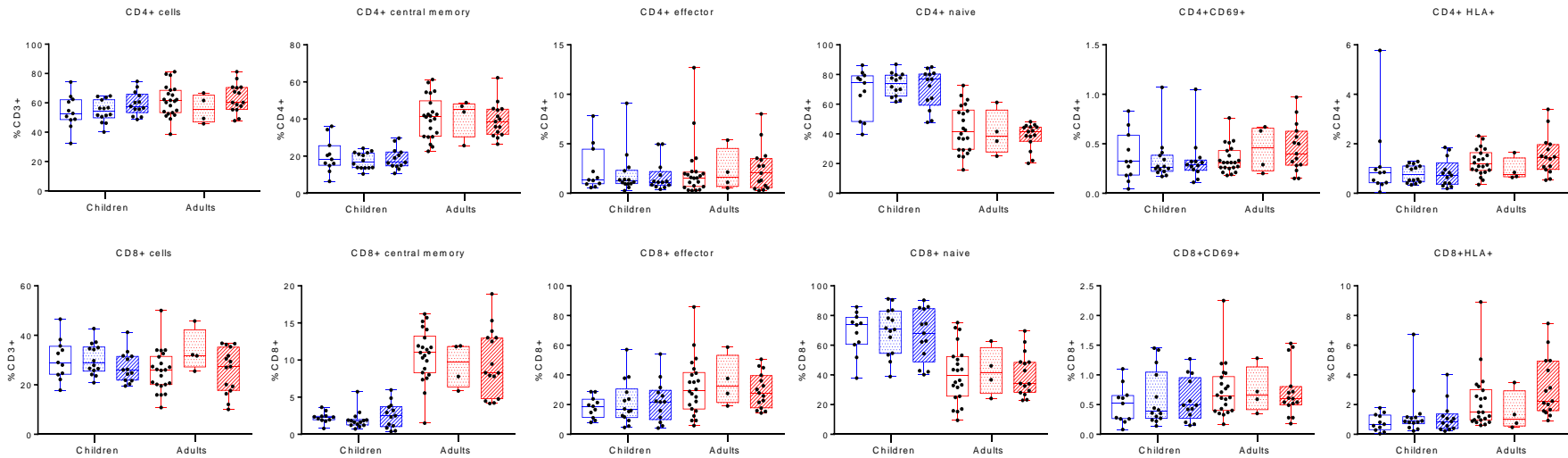
(A) Median antibody levels (IQR) based on symptoms (left y-axis) and median fold-change in antibody levels between asymptomatic and symptomatic in children (N=6 vs N=14) and adults (N=2 vs N=30) (right y-axis). (B) Mean viral load (SD) stratified based on symptoms in children (asymptomatic, N=7 vs symptomatic, N=35) and adults (asymptomatic, N=2 vs symptomatic, N=17). (C) Correlation between antibody levels and viral load. (D) Correlation between antibody levels and age. Blue dots/bars represent children and red dots/bars represent adults. Seropositivity was defined as seropositive by all three assays. Pearson's correlation analysis was used to examine association. Ct value: cycle threshold.

eFigure 9. Humoral Immune Cells Profile During Convalescence Period in Children and Adults Following SARS-CoV-2 Infection



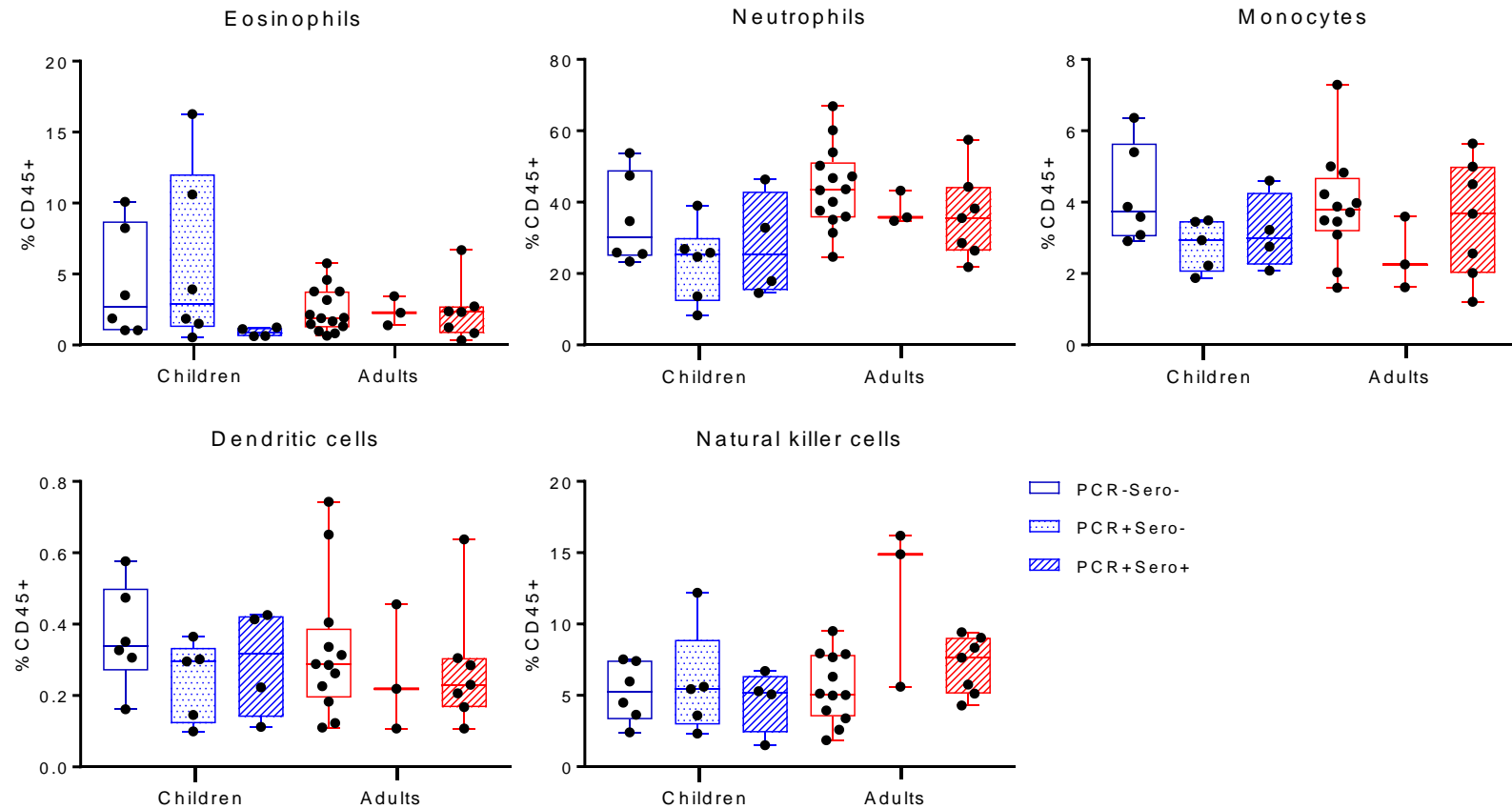
Convalescence period (median day 41) in children (PCR+sero-, N=14; PCR+sero+, N=13) and adults (PCR+sero-, N=4, PCR+sero+, N=15) following SARS-CoV-2 infection. An uninfected control group was included for comparison (PCR-sero-: children, N=11; adults, N=22). Bars represent median and range.

eFigure 10. Cellular Immune Profile (T Cells) During Convalescence Period in Children and Adults Following SARS-CoV-2 Infection



Convalescence period (median day 41) in children (PCR+sero-, N=14; PCR+sero+, N=13) and adults (PCR+sero-, N=4, PCR+sero+, N=15) following SARS-CoV-2 infection. An uninfected control group was included for comparison (PCR-sero-: children, N=11; adults, N=22). Bars represent median and range.

eFigure 11. Innate Cell Profiles During Acute Phase for Children and Adults Following SARS-CoV-2 Infection



Acute phase (day 7-12) for children (PCR+sero-, N=7; PCR+sero+, N=4) and adults (PCR+sero-, N=3, PCR+sero+, N=8) following SARS-CoV-2 infection. An uninfected control group was included for comparison (PCR-sero-: children, N=6; adults, N=16). Bars represent median and range.